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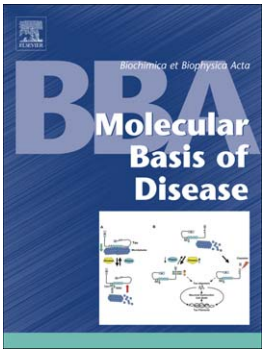


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# MIS-SPLICING OF TAU EXON 10 IN MYOTONIC DYSTROPHY TYPE I IS REPRODUCED BY OVEREXPRESSION OF CELF2 BUT NOT BY MBNL1 SILENCING

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**ABSTRACT (250 words)**

Tau is the proteinaceous component of intraneuronal aggregates common to neurodegenerative diseases called Tauopathies, including myotonic dystrophy type I (DM1). In DM1, the presence of microtubule-associated protein Tau aggregates is associated with a mis-splicing of Tau. A toxic gain-of-function at the RNA level is a major etiological factor responsible for the mis-splicing of several transcripts in DM1. These are probably the consequence of a loss of MBNL1 function or gain of CELF1 splicing function. Whether these two dysfunctions occur together or separately, and whether all mis-splicing events in DM1 brain result from one or both of these dysfunctions remains unknown. Here, we analyzed the splicing of Tau exons 2 and 10 in the brain of DM1 patients. Two DM1 patients showed a mis-splicing of exon 10 whereas exon 2-inclusion was reduced in all DM1 patients. In order to determine the potential factors responsible for exon 10 mis-splicing, we studied the effect of the splicing factors MBNL1, CELF1, CELF2 and CELF4 or a dominant-negative CELF factor on Tau exon 10 splicing by ectopic expression or siRNA. Interestingly, the inclusion of Tau exon 10 is reduced by CELF2 whereas it is insensitive to the loss-of-function of MBNL1, CELF1 gain-of-function or a dominant-negative of CELF factor. Moreover, we observed an increased expression of CELF2 only in the brain of DM1 patients with a mis-splicing of exon 10. Taken together, our results indicate the occurrence of a mis-splicing event in DM1 that is neither induced by a loss of MBNL1 function nor a gain of CELF1 function, but is rather associated to CELF2 gain-of-function.

Highlights
> Herein, we showed a mis-splicing of Tau exon10 in myotonic dystrophy brain > This implies a deregulation of the splicing factor CELF2 > Implication of MBNL1 or CELF1 is ruled out > Our work demonstrated a deregulation of CELF2 in myotonic dystrohpy brain

**Keywords:** Myotonic dystrophy, neurofibrillary degeneration, triplet expansion disease, microtubule-associated protein Tau, splicing, CELF splicing factor family, MBNL1

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## INTRODUCTION

Myotonic dystrophy type I (Steinert's disease) is an inherited neuromuscular disease affecting multiple organs, including skeletal and smooth muscle (myotonia), the heart (arrhythmias, conduction defects), the endocrine system (hyperinsulinemia) and the eyes (cataracts) [1]. The involvement of the central nervous system in DM1 was reported long ago [2, 3], but systematic studies to analyze the brain structures and cognitive functions involved have only recently been initiated. The available clinical data suggest that patients experience focused executive and visuoconstructive difficulties [4-7] and frontal cognitive impairment including attentional ability worsening with aging [8, 9]. In addition, impairments in facial emotion recognition have been associated with the adult form of DM1 [5]. These clinical findings suggest the involvement of the cerebral cortex as well as subcortical areas, such as the amygdala. In line with these observations, biomarkers of Alzheimer's disease including amyloid-beta peptide and Tau protein concentrations in the cerebrospinal fluid were shown to be significantly reduced and increased, respectively, in DM1 when compared to aged-matched control patients [10]. Structural neuroimaging in DM1 patients reveals the presence of brain atrophy and white matter lesions [11-13].

At the neuropathological level, neurofibrillary tangles (NFTs) are found in the limbic system of DM1 patients, including the hippocampus and isocortical areas such as the temporal and frontal cortices [14-16]. NFTs are intraneuronal aggregates made of hyper- and abnormally phosphorylated isoforms of microtubule-associated Tau (For review see [17]). Tau proteins belong to the family of microtubule-associated proteins and are encoded by a single *MAPT* gene located on chromosome 17. The alternative splicing of Tau exons 2, 3 and 10 gives rise to six adult human brain isoforms that are principally expressed in neurons and localized preferentially in the axons. The alternative splicing of Tau is developmentally regulated, tissue specific and deregulated in several Tauopathies [18]. In frontotemporal dementia, *MAPT* gene mutations modify the splicing of Tau, leading to the development of NFTs in adulthood. In DM1, we reported a modified splicing pattern for Tau, essentially characterized by the reduced inclusion of exons 2 and 3 [15, 19]. A reduction in the inclusion of Tau exon 10 has also been reported in several brains from DM1 patients [20] as well as in a transgenic animal model of DM1 [21]. However, the molecular mechanism underlying the mis-splicing of exon 10 in DM1 and its functional consequences remain unknown. This is of importance since the Tau exon 10 encodes for a fourth microtubule-binding domain. Thus, Tau proteins with four

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microtubule-binding domains (4R) bind more avidly and further stabilize microtubules than those with three binding domains (3R). The increased expression of 3R Tau isoforms in DM1 may modify the axonal transport and plasticity, since Tau is suggested to regulate the motor proteins transport along microtubules [22]. Moreover, a growing body of evidences suggests that Tau proteins may also localize to spines of dendrites and regulate glutamatergic transmission [23, 24]. The splicing of N-Methyl D-Aspartate receptor subunit one is altered in DM1 brains [19, 20], which together with the mis splicing of Tau, may also deregulate the glutamatergic synaptic transmission. Thus, the reduced Tau protein isoforms expressed in DM1 brains and the development of NFT may contribute the cognitive impairments in DM1.

A toxic gain-of-function at the RNA level is the main pathophysiological hypothesis to explain the development of the disease. DM1 is a genetically inherited disease characterized by a dynamic and unstable trinucleotide repeat expansion in the 3' UTR of the *DMPK* gene [25]. DMPK transcripts bearing long CUG repeats are retained in the nucleus and accumulate to form ribonuclear protein inclusions called foci. This nuclear retention of mutated transcripts of DMPK is responsible for DMPK haploinsufficiency. In addition, the sequestration inside foci of proteins such as members of the MBNL family of splicing factors results in a partial loss-of-function of these splicing factors [26, 27]. Moreover, a gain-of-function of CUG-binding protein and ETR3-like factor 1 (CUGBP1/CELF1) in skeletal or heart muscle, through its stabilization and phosphorylation by PKC, is also instrumental to DM1 pathophysiology [28-30]. CELF1 and embryonic lethal abnormal vision-like RNA-binding protein-3 (ETR3/CELF2) like factors (CELF) belong to a family of RNA-binding proteins that includes 6 members. While both CELF1 and CELF2 are ubiquitously expressed, the latter is highly expressed in the brain and heart of rodents [31, 32]. The four other members, CELF3, 4, 5 and 6, are enriched in the rodent brain. However, less is known about their expression in the human brain and their potential implication in DM1.

Dysfunctions of MBNL1 and CELF1 together are responsible for the modified splicing of several transcripts in DM1 muscle. However, whether the two dysfunctions occur concomitantly or separately, and whether CELF family members other than CELF1 are involved in DM1 brain, remain to be determined. We previously suggested that a loss of MBNL1 or a gain of CELF2 function could repress Tau exon 2-inclusion as observed in human disease [19, 33]. In the present study, we extended our analysis to additional DM1

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patients, and observed that, in addition to the presence of NFTs in all DM1 patients, certain patients displayed a mis-splicing of exon 10 in addition to that of exon 2. We therefore investigated the factors potentially responsible for this difference between patients as well as the mechanism involved in the deregulation of Tau exon 10 splicing.

## RESULTS

*Clinical and genetic presentation*

In the present study, we first analyzed the presence of *DMPK* CTG-expansions, the brain distribution of NFTs, and Tau protein expression in the brain tissue of two new DM1 patients (DM1a and DM1e), in order to determine whether their phenotype was comparable to that of the DM1 patients previously described [15]. Using small pool PCR with 0.5 ng of DNA, we have observed CTG-expansion in five brain regions in DM1a (Fig. 1A, left panel) and six regions in DM1e (Fig. 1A, right panel). The smaller expanded allele of patient DM1a, visible in the hippocampus and frontal and entorhinal cortices, was of 150 CTG (Fig. 1A, left panel), likely corresponds to the progenitor mutant allele. The largest allele of patient DM1e, seen in the hippocampus, contained 2900 CTG repeats, but several alleles with more than 1500 CTG repeats were observed in the other brain regions (Fig. 1A, right panel). However, the number of expanded alleles detected is well below that expected relative to the quantity of input DNA. The number of expanded alleles and CTG repeat lengths were comparable to those observed in the DM1 patients studied previously [15]. However, these data suggest that a high proportion of brain cells may contain alleles too large to be efficiently amplified by PCR, resulting in a probable underestimation of the true level of diversity within each sample.

*Brain distribution of NFTs and Tau isoform expression*

In order to analyze Tau pathology and its topographic distribution in the brain of DM1a and DM1e, the presence or absence of NFTs was established in several brain regions, including the entorhinal, the hippocampus, the amygdala, the basal and dorsolateral frontal, the parietal and the occipital cortices, using immunohistochemistry (Fig. 1B). NFTs composed of hyperphosphorylated Tau could be seen using phospho-specific AT8, AD2, AT100 and S199P antibodies and a pan TauCter antibody. Similar staining was observed in the amygdala and hippocampus of patient DM1e (not shown). The immunohistochemical observations were correlated with western blotting studies of Tau

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pathology. Using the phospho-specific Tau antibody AD2, we observed the typical pathological Tau quartet of 60, 64, 69 and 74 kDa in the sample from an Alzheimer's disease (AD) patient (Fig. 1C, lane 1 and 2). Tau proteins were not detected using the AD2 antibody in control brain tissue (Fig. 1C and S1A, lane 3 and 2, respectively). Pathological Tau proteins were detected and consisted of two bands of 60 and 64 kDa in the hippocampus (Fig. 1C, lane 5) and temporal amygdala (Fig. 1C, lane 6) and a single band of 60 kDa in the entorhinal cortex (Fig. 1C, lane 4) of DM1a, the insula and temporal pole of DM1e (Fig. S1A, lanes 3-6). We next used an antibody to the N-terminus of Tau (Tau Nter), to analyze the overall expression of Tau protein isoforms. Full-length Tau isoforms in the adult brain were resolved as four bands in the control case (Fig 1C, lane 3 and Fig S1A. lane 2, dots). The bands under the full-length isoforms correspond to catabolic products. In all brain regions studied in both DM1a and DM1e, two main bands were detected at 55 and 60 kDa, whereas the 64 kDa Tau band was faintly labeled, and the upper bands were not detected (Fig. 1C and Fig S1A). In order to determine the Tau isoforms expressed in DM1 brains, we used Tau exon-specific antibodies (Fig. S1B). In the control case, three bands (55, 60, 64 kDa) corresponding to full-length Tau isoforms were detected using an antibody to Tau isoforms lacking exon 10 (Fig. S1B, panel Exon 10-). In contrast, the exon 10-specific antibody stained strongly the 60 kDa band and to a lesser extent, the bands at 55 and 64 kDa (Fig. S1B, panel Exon10+, lane Ctrl). The exon 2-specific antibody stained the 55 and 60 kDa bands and to a lesser extent the 64 kDa band. In both DM1a and DM1e, there was a strong reduction of Tau isoforms stained with antibodies specific to exon 10 and 2 (Fig. S1B, panels Exon 10+ and Exon 2+), suggesting an alteration in Tau protein isoform expression. We next determined the composition of the Tau isoforms found in NFTs by immunohistochemistry, using Tau antibodies specific to exons 2 and 10 (Fig. S1C). As a positive control, NFTs in an AD brain were stained with the AD2 phospho-Tau antibody as well as with Tau exon-specific antibodies (Fig. S1C). In sharp contrast, in serial sections of the amygdala of patient DM1a, no NFTs or neuropil threads were detected with exon-specific Tau antibodies (Fig. S1C). This result suggests that no other isoforms than those lacking the exons 2 and 10 are found in NFTs of DM1 brains.

*Tau splicing modification in DM1 patients*

Immunohistochemical and western blot analyses indicate a modified expression of Tau protein isoforms in the brain of patients DM1a and DM1e, including the loss of Tau isoforms containing the sequence encoded by the exon 10. We therefore analyzed Tau

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isoform expression at the mRNA level in five brain regions in comparison with previously studied DM1 patients (Fig. 2 and Fig. S2). With regard to Tau exon 2/3 splicing, in the control brain, exon 2 was included in 53 ± 1 % of Tau transcripts (Fig. 2A, histogram). In the brain of DM1a and DM1e, this proportion significantly dropped to 35 ± 3% ($p = 0.0079$) and 31 ± 2% ($p < 0.0001$) respectively, demonstrating a preferential skipping of exon 2 (Fig. 2, histogram). To estimate Tau exon 10-inclusion, primers in exons 9 and 11 were used. The splicing of exon 10 was measured in the hippocampus, temporal, parietal entorhinal and frontal cortices of a control, DM1a and DM1e patients (Fig. S2). On average the % of Tau exon 10 inclusion in the control brain regions was of 47 ± 2%, consistent with the fact that half of the six Tau isoforms contain the exon 10. On average, in both DM1a and DM1e, a reduced inclusion of exon 10 was observed when compared to the control (DM1a: 23 ± 4%; DM1e 31 ± 2%). Apart from the parietal cortex of DM1a, the reduced inclusion of exon 10 was similar between brain regions analyzed. We further compared the splicing pattern of exon 10 in the temporal cortex between four controls and five DM1 brains at our disposal. In the temporal cortex of the four controls, the percentage of Tau exon 10-inclusion was of 46.5 ± 1.5% and homogeneous. In DM1a, there was a significant reduction of Tau exon 10-inclusion to 37 ± 3% ($p = 0.0013$) (Fig. 2B). Tau exon 10-inclusion was also reduced in the brain of DM1e (38 ± 2%, $p = 0.002$). In contrast, this reduction was not observed in DM1b, c and d. Thus, the modified splicing of Tau exon 2 is a robust and common phenotype in all DM1 brains analyzed so far, and it is also homogenous among brain areas. In contrast, the reduced inclusion of exon 10 is less pronounced and not constant in DM1 patients. One of the possible factors known to influence Tau exon 10 splicing is *MAPT* gene haplotype (H1 or H2), which has been suggested to modify the level of exon 10-inclusion [34]. Thus, we genotyped all patients included in the present study. DM1 patients were all H1/H1 (data not shown), excluding the possibility that variation of Tau exon 10 splicing could have been related to polymorphisms of *MAPT* gene haplotype.

*DMPK CTG repeats reduce Tau exon 10 inclusion*

The effect of *DMPK* long CTG repeats on Tau exon 10 splicing was next investigated (Fig. 3A). Tau exon 10 splicing is developmentally regulated - the switch between exclusion and inclusion of exon 10 is postnatal. Here, we used HeLa cells or T98 glioblastoma cells to analyze the trans-dominant effect of long CUG repeats because at the opposite to neuroblastoma cell-line SY5Y, the exon 10 is alternatively spliced in HeLa and

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T98 but completely exclude in SY5Y [33]. Under control conditions, exon 10 was included in 33 ± 0.5 % and 36 ± 2% of HeLa and T98 Tau transcripts, respectively (Fig. 3A and S3A). We transfected HeLa and T98 with $CTG_5$, $CTG_{200}$ and $DT_{960}$ constructs, all under the same CMV promoter. The splicing of exon 10 was not significantly modified by $CTG_5$ or $CTG_{200}$ whereas it was moderately but significantly reduced to 29.5 ± 0.5% ($p<0.05$) by $DT_{960}$ in both HeLa and T98 cells (Fig. 3A and S3A). The splicing of exon 10 was similarly modified by $CTG_{960}$ lacking the preceding *DMPK* exons 11 to 14, which are present in DT960 construct (Fig. 3A and S3B) or remained unchanged with the *DMPK* 3'UTR lacking the mutated allele (data not shown). Modification of Tau exon 10 splicing was not related to the presence or absence of DMPK exons. Moreover, despite the differences observed among patients, Tau exon 10 splicing shares the same sensitivity towards long CUG repeats as other cerebral transcripts.

*Differential effect of MBNL1 and CELF proteins on Tau splicing*

The loss-of-function of MBNL1 is instrumental in the pathophysiology of DM1 due to its sequestration into ribonucleoprotein aggregates. As shown in figure 3B, the inactivation of MBNL1 expression with two siRNA duplexes – siMBNL1a and siMBNL1b -significantly reduced the inclusion of Tau exon 2, attesting of the efficacy of siRNA duplexes. Incremental doses from 50 to 200 nM of siMBNL1a duplex were also evaluated and the repression of Tau exon 2-inclusion was already observed at 50 nM (Fig. S3C). However, MBNL1 siRNA did not significantly modified the inclusion of Tau exon 10, regardless of the duplex used (Fig. 3C, histogram and PCR) or the concentration used (Fig. 3D and S3D).

The CELF family also plays a role in DM1 pathophysiology. Moreover, we previously demonstrated the repressive role of CELF2 and CELF4 proteins in Tau exon 2-inclusion [33]. Moreover, CELF3 and CELF4 splicing factors promote the inclusion of Tau exon 10 [18, 35]. However, we do not know whether Tau exon 10 splicing is sensitive to CELF1 or CELF2 splicing factors. We therefore analyzed the effect of CELF1, CELF2 or CELF4 overexpression on the splicing of Tau exons 2 and 10. For siRNA experiments two independent CELF1 siRNA were tested and incremental concentrations were evaluated (Fig. S4A-B). CELF1 mRNA and protein expression were more strongly reduced by siCELFa than with siCELFb (Fig. S4A). The former was therefore selected for the subsequent experiments. Neither CELF1 overexpression (Fig. 4A) nor silencing (Fig. S4C-D) had an effect upon the splicing of Tau exons 2 and 10. In contrast, CELF2 and CELF4

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significantly reduced Tau exon 2-inclusion by 17% (p = 0.0001), and by 12% (p = 0.0012), respectively (Fig. 4B and C). Interestingly, we observed opposing effects of CELF2 and CELF4 on Tau exon 10 splicing. The overexpression of CELF2 significantly reduced exon 10-inclusion from 37.5 to 31.7% (p = 0.0031) (Fig. 4B), whereas CELF4 promoted the inclusion of exon 10 by 8% (p = 0.044, Fig. 4C). CELF4 was described previously as an enhancer of Tau exon 10-inclusion [36]. To exclude a potential artifactual effect due to the efficacy of the vectors or cell-type used, we co-transfected CELF1, CELF2 or CELF4 with the hcTNT exon 5-minigene or a dominant-negative CELF vector (DN-CELF) and reproduced the experiments using both HeLa and T98 cells (Fig. S5B-C). All CELF proteins were expressed as assessed by Western-blotting using a V5-epitope antibody for CELF1, 2 and 4 factors or Xpress-epitope antibody for the DN-CELF factor (Fig. S5A). All CELF factors except the DN-CELF promoted the inclusion of exon 5, as previously demonstrated [37] in both HeLa and T98 cells (Fig. S5B and C). Finally, Tau exon-10 inclusion was promoted by CELF4 and repressed by CELF2 but neither by CELF1 or DN-CELF in both T98 and HeLa cells (Fig. S6).

As several CELF family members appear to be modulators of Tau exon 10 splicing, we finally analyzed the expression of CELF1, CELF2 and CELF4 in brain tissues of DM1 patients as compared to controls (Fig. 5). CELF1, CELF2 and CELF4 were more strongly detected in the temporal, parietal and frontal cortices of DM1a and DM1e than in samples from the control or patients DM1b, c or d (Fig. 5A, B and C). In sharp contrast, in the three latter cases, while CELF4 levels were similar to those observed in the control, CELF2 and CELF1 levels were diminished.

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## DISCUSSION

Our data together suggest that the reduced inclusion of Tau exon 10 is not a ubiquitous splicing event in DM1 brains and is probably not related to a loss of MBNL1 or gain of CELF1 functions but rather to a gain of CELF2 function, shown herein to repress Tau exon 10 inclusion. Moreover, the loss of MBNL1 function and gain of CELF function, as far as the brain is concerned, can occur simultaneously or individually, and may explain the discrepancies observed in splicing events such as Tau exon 10 splicing.

*Presentation of cases with classic DM1 neuropathology but moderate symptoms*

In this study, we described the topographical distribution of NFTs in two patients. These patients presented with a classic adult form of DM1. DM1a was diagnosed at 46 years, and displayed muscular abnormalities and early-onset bilateral cataract. He had no cardiac or endocrine disorders, nor any cognitive impairment from a clinical point of view. The only neurological symptom reported was diurnal somnolence. The anamnesis was marked by frequent gastrointestinal disorders leading to a grade 3 adenocarcinoma of the colon, which was the cause of death. DM1e was diagnosed at the age of 45 years, and had a round, myopathic face, myotonia of both hands, type II diabetes, hypercholesterolemia and suffered from obesity. Although, we do not know the cognitive status of these patients, the topographical distribution of NFTs was very similar to that of DM1 patients already characterized [15]. In our DM1 cohort, we show that Tau pathology is observed early in adulthood and that the hippocampus and temporal cortex, as well as the amygdala and insula, are the most vulnerable regions affected in DM1. Importantly, the amygdala is the brain region controlling fear and facial emotion recognition. In DM1, the latter cognitive function is altered [5]. Moreover, brain imaging also points to the temporo-insular region as a vulnerable brain area [11, 38].

*Analysis of splicing modifications in DM1 brains*

Although the presence of NFTs is restricted to the hippocampus and temporal amygdala in our study, Tau splicing is altered in all regions analyzed, suggesting that NFTs only affect a subset of neuronal populations, as observed in other Tauopathies [17, 39, 40]. Our results show that all DM1 patients, in all brain regions and regardless of the size of the expansion, present a reduction in the inclusion of Tau exon 2. In addition, we were able to confirm a similar ubiquitous increase in the expression of the APP fetal splice variant lacking exons 7 and 8, an increase in the inclusion of NMDAR1 exon 5 [20] and an

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increase in the expression of the longest MBNL1 isoform, $MBNL1_{43}$ [19] (not shown). In contrast to these splicing mechanisms, we found that Tau exon 10 skipping is not a consistent feature of DM1. Indeed, only two among six patients presented an abnormal inclusion of Tau exon 10. The ten DM1 patients previously investigated by Jiang and collaborators also displayed Tau exon 10 mis-splicing [20]. *MAPT* polymorphism has been reported to influence Tau expression and splicing. In fact, H1 carriers have an increased susceptibility to neurodegeneration, and the H1 chromosome results in higher levels of exon 10+ MAPT mRNA than H2 [34]. We systematically genotype all patients in our cohort, and all our cases - DM1 and controls - were H1/H1. Moreover, the *MAPT* gene between exon 9 to exon 10 was sequence in DM1a and DM1b to exclude any unknown polymorphism. Thus, even if NFTs were observed, the *MAPT* haplotype could not explain the differential inclusion of exon 10 in DM1 patients. However, exon 10 splicing is sensitive to the identity of its flanking exons. Indeed, exon 9 promotes skipping whereas exon 11 is repressive [36, 41].

The mechanism underlying the mis-splicing of Tau exon 10 in DM1 remains completely ill-defined. An alternative explanation, that exon 10 mis-splicing is not directly related to CTG expression, can be ruled out. *In vitro* experiments using long CUG tracts lead to a modification in Tau exon 10 splicing, as observed in the human brain. Moreover, a reduced inclusion of Tau exon 10 has also been reported in a mouse model of DM1 [21]. The CUG expansion sequesters MBNL1, resulting in the mis-splicing of transcripts regulated by this splicing factor. The loss of MBNL1 function can be mimicked using specific siRNAs that target MBNL1 transcripts. Surprisingly, Tau exon 10 splicing is not significantly modulated by a loss of MBNL1 expression, in contrast to Tau exon 2 [19]. The splicing event that we demonstrate here thus appears to be a misregulation induced by long CUG repeats but probably independent of an MBNL1 loss-of-function, as previously suggested for splicing events regulated by CELF1 but not by MBNL1 [30].

*Analysis of CELF proteins: expression and splicing regulation*

The present study reports CELF protein expression in DM1 brains. We show that CELF1, CELF2 and CELF4 proteins are expressed in the human brain. Interestingly, an increase in protein levels of CELF1 and CELF2 was observed in the brains of patients DM1a and DM1e but not in the 3 other DM1 patients. GAPDH and CELF4 levels were similar in DM1 and control cases suggesting that the differences observed were not due to the post mortem delay or the quality of the tissue. Post-translational modifications of

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splicing factors can modulate their activity. With regard to Tau exon 10 splicing, phosphorylation of the splicing factor hTra21 or SR proteins by the kinases CLK2 or GSK3 promotes the skipping of exon 10 [42-44]. The stability and activity of CELF1 protein is regulated by phosphorylation [30]. Notably, we also observed an increase in CELF1 protein steady-state levels in DM1a and DM1e. CELF1 and CELF2 share more than 70% sequence homology [45] and preliminary data using 2D electrophoresis and western blotting suggest that CELF2 is also hyperphosphorylated in DM1 brains (data not shown). Therefore, in addition to protein expression, phosphorylation should also be considered to be a modulator of splicing activity in DM1 heart, muscle and brain.

The splicing of several transcripts (NMDAR1 [20]; cTNT [46]; IR [47]; MTMR1 [48] and ClC-1 [49]) is modulated by CELF1. However, we found that the overexpression or inactivation of CELF1 does not modulate the splicing of Tau exons 2 and 10. CELF4 promotes Tau exon 2 splicing whereas CELF2 represses it [33]. CELF4 has also been shown to promote Tau exon 10 inclusion [36] but the potential regulation of Tau exon 10 splicing by CELF2 remains unexplored. In this paper, we used HeLa and T98 cells, in which the endogenous expression of CELF4 and CELF2 is lacking [33, 50], to show that the ectopic expression of CELF4 promotes Tau exon 10 inclusion whereas CELF2 represses it. Thus, whereas these three CELF proteins promote hcTNT exon 5-inclusion, the role of each CELF in Tau splicing was distinct from, and sometimes the opposite of, that of the others, even with respect to a single exon, as reported for α-actinin [51] in non-muscle cells. Therefore, we demonstrate here for the first time that CELF1 has no effect on Tau exons 2 or 10, CELF2 is a repressor of both exons whereas CELF4 is a repressor of exon 2 and an activator of exon 10 inclusion. Together with other data, we demonstrate that two CELF family splicing factors are strong modulators of Tau splicing [18, 33, 35]. In addition to these results obtained *in vitro*, we observed a potential relationship between CELF2 overexpression in DM1 and the mis splicing of Tau exon 10. Since CELF2 levels in DM1b, DM1c and DM1d were lower than in DM1a and DM1e, and since the former patients showed no change in exon 10, we suggest that Tau exon 10-inclusion is likely related to CELF2 expression or activity in the DM1 brain. It has previously been shown that high levels of CELF2 protein in the forebrain are associated with a skipping of the NMDAR1 NI exon and inclusion of the CI exon, and that these splicing patterns are reversed in the hindbrain where CELF2 is deficient [52]. A competitive effect of CELF2 and CELF4 is therefore possible. As CELF2 protein is

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expressed at higher levels than CELF4 in the brains of DM1a and DM1e, the repressive effect is stronger than the activator effect. In DM1b, c and d, the disparity between protein levels is not as great, and may explain why the promoting effect of CELF4 is in balanced with the repressive activity of CELF2.

In conclusion, we show that the altered splicing of Tau exon 10 is likely related to a gain of CELF2 function and neither to a loss of MBNL1 nor a gain of CELF1 functions. Our results strengthen the idea that the CELF proteins are involved in the neuropathology of myotonic dystrophy type I. Moreover, since neurofibrillary degeneration is a feature common to DM1 patients, particular attention should be paid to cognitive dysfunction related to the temporo-insular brain region.

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## Materials and methods

*DM1 patients and human brain tissue samples*

DM1a was a patient followed at the Department of Neurology, Donostia Hospital, San Sebastian, Spain. This patient was a male from the Basque Country in the north of Spain, and diagnosed with Myotonic Dystrophy at the age of 46, with a smear expansion of 333 to 833 CTG triplets as established from blood cells. The initial symptom reported was a cataract operated at the age of 32. In association with typical DM1 symptoms, the patient presented diurnal somnolence, trigeminal neuralgia and hepatic steatosis. In the last five years, he presented severe proximal weakness that brought him to a wheelchair-bound state. The patient died at 60 years from a disseminated colon neoplasia. DM1e was a woman diagnosed at the age of 46 years old. She presented a myopathic face, suffered from myotonia of both hands and had type II diabetes, hypercholesterolemia, obesity and severe restrictive ventilation. Cognitive status was not assessed in either case. Brain tissue samples were obtained at autopsy in accordance with the protocol of the local ethics committee. The other three DM1 patients, DM1b, DM1c and DM1d, have been previously described ([15], and correspond to case 3, case 1 and case 2 respectively; [19]). These three patients were 64, 61 and 53 years old respectively at the time of death. Three control individuals were included in the study. Post-mortem delays for tissue sampling were below 48h. One cerebral hemisphere was frozen and stored at -80°C and the other was formalin fixed for histological and immunohistochemical studies. Six cortical brain regions, the temporal, occipital, frontal, entorhinal and parietal cortex and the hippocampus, were dissected for analysis. Due to the limited amount of tissue for certain brain regions, such as the hippocampus and entorhinal cortex, biochemical analyses were performed using the temporal, frontal and parietal cortices. For genetic analysis, all brain regions were included.

*Immunohistochemical analysis*

Immunostaining was performed with paraffin-embedded sections. Sections (5µm-thick) were deparaffinized with xylene and rehydrated through decreasing concentrations of ethanol. Endogenous peroxidase activity was quenched with $H_2O_2$ and non-specific staining blocked with horse serum using the Vectastain ABC Universal kit according to the manufacturer's protocol (Vector laboratories). Brain tissue slices were incubated overnight

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with the following primary antibodies: AT8, AD2, AT100, S199P, Exon 2, Exon 10 and Tau-Cter. AT8, AT100, Exon 2 (Clone DC 39 1N, Sigma) and Exon 10 (DC 4R, Sigma) are mouse monoclonal antibodies. Tau-Cter and S199P antisera were developed in rabbits against synthetic peptides [15]. AT8 and AT100 label phosphorylated Tau epitopes: AT8 specifically detects phospho-serine 202 and phospho-threonine 205, while AT100 recognizes the pathological phospho-Tau epitopes at serine 212 and threonine 214. Antibodies were diluted 1/100 to 1/1000 in PBS.

*Determination of CTG repeats size and Tau haplotype*

Genomic DNA was prepared from brain tissue by phenol-chloroform extraction. The genetic diagnosis of DM1 was confirmed by PCR amplification of the CTG region according to Cheng's protocol [53], using an Expand Long Template PCR System™ (Roche Diagnostics Gmbh, Mannheim, Germany). Each 50 µl reaction consisted of 1X Expand Long PCR buffer, 0.7 mM each of dATP, dCTP and dTTP, 0.06 mM dGTP, 0.24 mM 7-deaza dGTP, 1 µM of primers DMK1 (5'-CACAGGCTGAAGTGGCAGTTCCA-3') and DMK2 (5'-TGTCGGGGTCTCAGTGCATCCA-3'), 3.5 U of *Taq* DNA polymerase, 1.75 mM $MgCl_2$, and 0.01 to 5 ng of total genomic DNA. Amplifications were performed in a PTC200 thermocycler (MJ Research, Prolabo, France). Following denaturation for 5 min at 95°C, 35 cycles of amplification were performed: denaturation at 95°C for 1 min, annealing at 62°C for 30s and extension at 68°C for 10 min. PCR products were resolved on 1% agarose gels. Southern blot transfer and probe-mediated detection was carried out as follows: agarose gels were denatured in 0.4 M NaOH, 0.6 M NaCl and rinsed twice in 2x SSC before being transferred to Hybond $N^+$ nylon membranes. Blots were pre-hybridized in DIG Easy Hyb solution (Roche) for 30 min at 59°C. Hybridization was performed in a fresh solution containing 100 pmol 3'-DIG-$(CTG)_{10}$ (DIG Oligonucleotide terminal 3' labeling Kit, Roche) for 2 h at 59°C followed by two washes in 2x SSC, 0.1% SDS at room temperature for 5 min and two washes in 0.5x SSC, 0.1% SDS for 15 min at 59°C. Probe binding was detected using a DIG Luminescence Detection Kit (Roche) according to the manufacturer's instructions (Amersham-Pharmacia Biotech, Orsay, France). Film exposure times ranged from 2 to 5 min. DNA molecular weight standards were load onto each agarose gel (DNA marker IV, Boehringer Mannheim Gmbh, Germany). Tau H1/H2 genotyping was performed with primers flanking the 236bp insertion/deletion polymorphism in intron 9. The forward (5'-

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GGAAGACGTTCTCACTGATCTG-3') and reverse (5'-AGGAGTCTGGCTTCAGTCTCTC-3') primers used have been previously described [54].

*Semi-quantitative analysis*

Total RNA was isolated from brain tissue using RNAgents (Promega, Madison USA) according to the manufacturer's instructions. Total RNA was isolated from cells using the Nucleospin® RNA II kit (Macherey Nagel, Düren, Germany). RNA concentration was determined by absorption at 260 nm with Nanodrop ND1000 technology (Labtech France). RT-PCR was performed in triplicate with 1 µg of total RNA using random hexamers (5µM/l) and M-MLV reverse transcriptase (Invitrogen, California, USA) according to standard protocols. No DNA amplification was observed in RT controls. PCR was carried out in a final volume of 25 µl, with 10 or 15 pM of each primer (primer sequences are detailed in supplementary Table 1), 1.5 mM $MgCl_2$ and 1 U of Taq polymerase (Invitrogen), under the following conditions: 5 min at 94°C, 22 to 30 cycles of a 1 min denaturation step at 94°C, annealing for 2 min at the temperature indicated in Table 1, 2 min of extension and 7 min of final extension at 72°C. 18S was used as an internal control. Reaction products were resolved by electrophoresis using a 4%, 5% or 8% polyacrylamide gel, and bands were stained with SYBR gold (Invitrogen). The intensity of SYBR gold luminescence was measured using a FluoroImager scanner (Claravision, France). The mRNA signals were normalized to that of 18S mRNA.

*Protein extraction and western blot analysis*

DM1 and control brains were homogenized with a dounce homogenizer in RIPA buffer (Invitrogen, California, USA) with the addition of 1mM $Na_3VO_4$, 125 nM okadaic acid and protease inhibitors. The resulting homogenates were sonicated and spun at 13,000g for 20 min at 4°C. The supernatants were transferred and the concentration of proteins measured. The pellets were sonicated again, warmed and centrifuged. The supernatants were diluted with 2x Laemmli lysis Buffer (LDS buffer, Invitrogen). The protein extracts (5 µg each) were used for SDS-PAGE (4-12%; Nupage® Bis-Tris gels, Invitrogen, California, USA) and transferred onto Hybond nitrocellulose membranes (G&E Healthcare) using the XCell™ II Blot Module (Invitrogen). Proteins were reversibly stained with Ponceau red in

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order to control for the amount of protein loaded. After blocking with 5% skimmed milk in TBS-T (20 mM Tris-HCl pH 8.0, 150 mM NaCl and 0.1% Tween-20), membranes were incubated overnight at 4°C with the following antibodies in the same buffer: anti-CELF1 (Clone 3B1, mouse monoclonal IgG, 1:500; Upstate Biotechnology, Lake Placid, New York, USA), anti-CELF4 (Anti-Brunol4, mouse polyclonal, 1:500; Abcam, Cambridge, UK), anti-CELF2 (anti-CUGBP2, mouse monoclonal, 1:1000; Sigma-Aldrich, Saint Louis, Missouri, USA), and anti-GAPDH as an internal control (anti-FL-335, 1:10000; Tebu-Bio, Le Perray en Yvelines, France). To label Tau isoforms, we used antibodies specific to exon 2+ (Clone DC 39 1N, 1:1000) Sigma), exon 10+ (DC 4R, Sigma) and Tau exon 10- (Anti-Tau 3R-repeat isoform RD3, clone 8E6/C11, 1:1000, Millipore). Membranes were subsequently incubated with a peroxidase-conjugated goat anti-mouse IgG (Sigma-Aldrich; 1:50000 in the same buffer), or with peroxidase-conjugated goat anti-rabbit IgG (Sigma-Aldrich; 1:4000 in the same buffer), and labeling revealed using ECL™ and Hyperfilms (G&E Amersham) according to the manufacturer's instructions. Western blots were carried out at least twice.

*Cell culture and transfection*

HeLa cells or T98 glioblastoma cells were grown in monolayer cultures in 6-well plates in Dulbecco's Modified Essential Medium (DMEM) (GIBCO, Invitrogen, California, USA) supplemented with 10% fetal calf serum, 50 IU/ml Penicillin, 50 µg/ml Streptomycin and 4 mmol/L Glutamine, at 37°C in a humidified 5% $CO_2$ incubator. Cells grown to ~ 50% confluence were transiently transfected with 1 or 3 µg of plasmid DNA, using FuGENE HD transfection reagent (Roche Diagnostics, Indianapolis, USA). Total RNA was analyzed 48h post-transfection, with 18S RNA as an internal loading standard.

CELF1, CELF4 and CELF2 cDNA in TOPO vectors were sub-cloned using the Gateway system (Invitrogen, France) and inserted into pcDNA3.1/nV5-DEST™ (Invitrogen, France). A V5-epitope was fused to the N-terminal region of CELF factors and their expression was analyzed using both CELF antibodies and a V5-specific antibody (Invitrogen, 1:10000 dilution). The dominant-negative CELF factor in pcDNA 3.1-His vector (Invitrogen, France) was kindly provided by Thomas Cooper (Baylor College of Medicine, Houston, TX) and corresponds to a CELF4 deletion that has dominant negative activity on itself and other CELF proteins [37]. The protein expression was verified with an Xpress™-specific antibody (Invitrogen) which epitope is localized at the amino-terminus

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of the protein. The pcDNA 3.1 empty vector was used as a control. Plasmids containing the 3'UTR of DMPK with 5 or 200 CTG repeats were derived from previously described constructs under a rodent ROSA promoter controlling GFP fused to the 3'UTR of *DMPK* [55]. The constructs were sub-cloned into pcDNA 3.1 vector (Invitrogen, France) under the control of the CMV promoter. The plasmid containing the 3'UTR of *DMPK* with 960 interrupted CTGs was a minigene vector also under the control of the CMV promoter named DT960. It contained the last 5 exons of the *DMPK* gene followed by the full length *DMPK* 3'UTR, with 960 CTG repeats interrupted at every 20$^{th}$ repeat by a CTCGA motif [56], named herein CTG960. The conservation of the number of CUG repeats was verified by a long PCR assay. The RTB300-minigene containing exon 5 of human cTNT (hcTNT) has been previously described [57].

For silencing experiments, siRNAs were synthesized by Eurogentec (Belgium). The sense strand sequences of the siRNAs designed to target MBNL1 mRNA transcripts were: siMBNL1a 5'-CGCAGUUGGAGAUAAAUGG-3', and siMBNL1b 5'-CAGACAGACUUGAGGUAUG-3' [58]. To target CELF1 mRNA transcripts, the sense strand sequences of the siRNAs were: siCELF1a 5'-GUUACGACAAUCCUGUUUCdTdT-3' (described by Paul et al., 2006) and siCELF1b 5'-CACCCGUAAAGCUGCAUUdTdT-3'. Scrambled siRNAs and siRNA against β-actin were also designed as controls. The siRNAs were transfected using Oligofectamine Reagent (Invitrogen) according to the manufacturer's instructions. Increasing concentrations were used (10, 50, 100 and 200 nM) to determine the dose response curve otherwise siRNA concentration used was 100 nM.

*Statistical analysis*

Statistical analyses were performed using the non-parametric Mann-Whitney test or One-way ANOVA, using Prism Software (GraphPad Software Inc., San Diego, USA).

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**Figure legends**

**Figure 1: CTG expansion mosaicism, neurofibrillary degeneration and Tau protein expression in DM1 brain tissue**

**A.** CTG repeat lengths were determined from the genomic DNA of cortical (frontal (Fr), temporal (Tp), amygdala (Tp am), insula (Ins), hippocampus (Hip), entorhinal (Ent) and occipital (Occ)) brain tissues of DM1a and DM1e using long PCR followed by Southern blotting for expanded alleles. Results are shown for 0.5 ng of input DNA. Lane 1 (DM1a panel) corresponds to expanded control DNA (1300 CTG repeats) from a congenital form of DM1. DNA molecular weight markers are indicated on the right. Dots indicate the longest expanded alleles for each brain region analyzed. **B.** Top: Location of epitopes recognized by the antibodies used with respect to the longest brain Tau isoform (441 aa isoform). Bottom: Neurofibrillary tangles (NFTs) in the amygdala of DM1a. NFTs composed of abnormal and hyperphosphorylated Tau are stained by AT8, AD2, TauCter, AT100 and S199P antibodies. **C.** Spatial distribution of hyperphosphorylated Tau (PHF-Tau) in DM1a brain. Using the phospho-specific Tau antibody AD2, hyperphosphorylated Tau isoforms (PHF-Tau) were detected only in brain regions with NFTs. Seven cortical brain regions of DM1a (lanes 4 to 10: entorhinal (Ent), hippocampus (Hip), temporal amygdala (Tp am), basal frontal (Fr bas), dorsolateral frontal (Fr D-lat), parietal (Par) and occipital (Occ)) were analyzed by Western blotting using a Tau N-terminal antibody to detect all isoforms, and compared to the frontal cortex of an Alzheimer's disease patient (lane 1 and 2), and to the same region of a control individual (lane 3). In the control brain, Tau isoforms were detected as four bands between 50 and 67 kDa (black dots). Note that the bands under the 60kDa Tau band detected with antibody TauNter correspond to amino-terminal catabolic products of Tau protein isoforms. The vertical bar indicates the position of the pathological Tau protein quartet in lanes 1 and 2.

**Figure 2: Tau splicing in controls and DM1 patients**

**A.** Exclusion of Tau exon 2 in the brain of DM1a and DM1e. Left panel (DM1e not shown): Tau exon 2/3 forward and reverse primers allowed the simultaneous amplification of Tau isoforms with exons 2 and 3 (2+3+), with exon 2 but not 3(2+3-), or lacking both exons (2-3-). PCR was performed using samples from the hippocampus (Hip), temporal (Tp), parietal (Par), entorhinal (Ent) and frontal (Fr) cortical brain regions of a control

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individual and patient DM1a. 18S RNA was used as the internal loading standard. Right panel: histogram showing the percentage of transcripts containing exon 2 in the brain of DM1a and DM1e. The mean and standard deviation were calculated from three individual RT-PCR experiments resulting from the study of four brain regions of two controls (in black) and patients DM1a and DM1e (in white). ***: p<0.0001. **B.** Tau exon 10 splicing in DM1 brain. RT-PCR analyses of Tau exon 10 inclusion in the temporal cortex of 4 controls and five DM1 patients. Histograms indicate the percentage of transcripts including exon 10, from three independent experiments. Tau exon10 inclusion is significantly reduced in DM1a and DM1e but not in DM1b, c or d. **: p<0.01.

**Figure 3: Effect of long CTG repeats and MBNL1 loss-of-function on Tau splicing**

**A.** Overexpression of long CUG repeats in HeLa cells mimics the Tau exon 10-exclusion observed in the brain of DM1a and DM1e. The 3'UTR of the human *DMPK* (1µg) containing 5 (CTG5), 200 (CTG200) or 960 CTG (DT960), subcloned in a pcDNA3.1 plasmid, were transiently transfected in HeLa cells (left panel). HeLa were also transfected with empty pcDNA 3.1 plasmid DT960, and CTG960 vector, which lacking *DMPK* exons 11 to 14 and only containing the CTG repeats (right panel). Total RNA was isolated 48h post-transfection. Primers Tau exon 10 forward and reverse allowed the simultaneous amplification of Tau isoforms with or without exon 10. 18S RNA was used as the internal loading standard. After polyacrylamide gel electrophoresis, bands were quantified and results are expressed as the percentage of Tau exon 10 inclusion. Histogram represents the mean ± SEM of four individual experiments. *: p<0.05; ns: non–significant; Mann-Whitney U test. **B.** Loss of MBNL1 expression reduces Tau exon 2-inclusion. The histogram is representative of three individual experiments and values are expressed the mean ± SEM. **: p<0.01 **C.** Tau exon 10 splicing is insensitive to MBNL1 loss of expression using siRNA. The PCR gel represents the results obtained with two RNA duplexes targeting human MBNL1, siMBNL1a (lane 6-8) and siMBNL1b (lane 9-11), and one scrambled duplex (siMBNL1 Scr, lane 3-5) were used for transfection into HeLa cells. Lane 1-2: control (Ctrl), transfection agent alone. ns: non-significant; Mann-Whitney U test. Means and standard deviations were calculated for three individual experiments. **D.** Evaluation of incremental concentrations (siMBNL1 50, 100 and 200 nM) of siMBNL1a upon Tau exon 10 splicing. The histogram represent the average ± SEM of % of Tau exon 10-inclusion obtained from three independent experiments (ns: non-significant).

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**Figure 4: CELF2 and CELF4, but not CELF1, regulate Tau exon 2 and Tau exon 10 splicing**

**A.** CELF1 expression has no effect on Tau exon 2 and Tau exon 10 splicing (means ± SEM of four independent experiments performed in triplicate). Histograms indicate the percentage of Tau exon 2 and exon 10-inclusion in Controls (Ctrl) and in HeLa cells ectopically expressing CELF1. ns: non-significant; Mann-Whitney U test. **B.** Overexpression of CELF2 reduces the inclusion of Tau exons 2 and 10 (means ± SEM of five independent experiments performed in triplicate). **: $p<0.001$, ***: $p<0.0001$; Mann–Whitney U test. **C.** Overexpression of CELF4 decreases Tau exon 2-inclusion and increases Tau exon 10-inclusion (means ± SEM of four independent experiments performed in triplicate). **: $p<0.001$; Mann–Whitney U test. The control experiments correspond to the transfection of the pcDNA 3.1 vector.

**Figure 5: CELF protein expression in brain tissues**

Western blots revealing an overexpression of CELF1 (**A**) and CELF2 (**B**) proteins in DM1a and DM1e, using brain tissue lysates from the temporal (T), parietal (P) and frontal cortices (F) of a control individual and patients DM1a, b, c, d and e. Membranes were striped and probed with a GAPDH antibody for standardization.

**Table 1: Clinical, genetic and neuropathological comparison of four DM1 patients**

**Table 2: List of primers used.** The first column indicates the sequence that was amplified. The second and third columns are the sequences of the forward and reverse primers used and their exon location. The concentration of $MgCl_2$, annealing temperature and size of the amplified fragments are also indicated.

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**Supplementary Figure 1: Splicing of brain transcripts in controls and in patients DM1a and DM1e**

**A.** Distribution of hyperphosphorylated Tau (labeled by the AD2 antibody for PSer 396-404) and total Tau (labeled by an antibody to Tau N-terminus) in brains regions of DM1e. As a positive control for pathological Tau labeling, a sample from an AD brain was loaded onto the first lane. Pathological Tau is not detected in lane 2 in which a brain tissue homogenate from a control individual (Ctrl) was loaded. **B.** Western-blot analysis of Tau protein isoforms. A control (Ctrl) brain homogenate was loaded onto the first lane, followed by brain homogenates from frontal and temporal cortices of patients DM1a and DM1e, respectively. Blots were performed in triplicate and incubated with Tau exon 10-, exon 10+ and exon 2+ specific antibodies. Tau full-length proteins are indicated by dots. **C.** Aggregated Tau proteins lack isoforms with exon 2 and 10 in the brain of DM1a. Three upper panels: NFTs detected by AD2 are also recognized by antibodies to exons 2 and 10 in the frontal cortex of an AD patient. Two lower panels: in the amygdale of DM1a, exon-specific Tau antibodies do not label NFTs.

**Supplementary Figure 2: Reduced inclusion of Tau exon 10 in DM1a and DM1e**

Primers in the exon 9 and 11 were used to analyze the splicing of Tau exon 10 in four brain regions (Hip: hippocampus; Tp: temporal; Par: parietal and Ent: entorhinal) of a control and DM1a and DM1e patients. The two PCR products corresponding to the inclusion (10+) or exclusion of exon 10 (10-) are indicated on the left of the gel. The percentage of Tau exon 10-inclusion in each brain region is represented on the histogram.

**Supplementary Figure 3: Effect of CTG repeats and MBNL1 loss-of-function on Tau exon 2 and 10 splicing.**

**A.** Long CUG repeats repress exon 10-inclusion in T98 glioblastoma cell lines. The pcDNA 3.1 empty vector, CTG5, CTG200 and DT960 vectors were transfected in T98 glioblastoma and Tau exon 10 splicing was analyzed 48h post-transfection. The left histogram represents the percentage of Tau exon 10-inclusion (mean ± SEM; ns: non-significant; **: $p<0.001$, Mann-Whitney U-test). **B.** Tau exon 10-inclusion is repressed

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similarly by DT960 and CTG960 vectors, the latter of which is lacking *DMPK* exons 11 to 14. The histogram is representative of three individual experiments. The percentage of Tau exon 10-inclusion is express as the mean ± SEM (**: p<0.001, Mann-Whitney U-test). Increasing concentrations of the RNA duplex siMBNL1a (50, 100 and 200 nM) were used to transfect HeLa cells. RT-PCR was performed for three independent experiments with primers surrounding Tau exons 2 and 3 (**C**) and Tau exon 10 (**D**). In controls (Ctrl), transfection agent was used alone. The loss of MBNL1 expression appears to increase the mRNA species that do not contain Tau exon 2, but does not influence the inclusion of Tau exon 10.

**Supplementary Figure 4: Loss of CELF1 expression has no effect on Tau splicing.**

RNA duplexes used for transfection of HeLa cells were: two duplexes against human CELF1 (siCELF1a lane 3, and siCELF1b lane 4) and one scrambled duplex (Scrambled, lane 2). In the control experiment (Ctrl, lane 1), the transfection agent was used alone. **A. siCELF1a and siCELF1b reduce CELF1 mRNA expression** relative to 18S RNA, while the scrambled siRNA does not (means ± SEM for three independent experiments). *: p<0.05, **: p<0.001; Mann–Whitney U-test. Upper panel: The efficacy of siCELF1a in reducing CELF1 expression in cells transfected with RNA duplexes (lane 1) vs. cells without siCELF1a (Mock, lane 2) is determined by Western-blot using CELF1 antibody (˜ CELF1). **B, C and D. Incremental loss of CELF1 mRNA expression has no effect on Tau exon 2 and Tau exon 10 splicing. B.** CELF1 mRNA expression is determined by RT-PCR and expressed as the ratio of CELF1 / 18S. *: p<0.05; Mann-Whitney U-test. Tau exon 2 (**C**) and 10 (**D**) inclusions were analyzed by RT-PCR in HeLa cells transfected with the transfecting agent (Ctrl), a Scrambled siRNA or with incremental concentrations (from 10 to 200 nM) of siCELF1a siRNA. Histograms are representative of the percentage of Tau exon 2 and Tau exon 10-inclusion (mean ± SEM for three independent experiments). ns: non significant.

**Supplementary Figure 5: The expression vectors for CELF factors regulate hcTNT exon 5**

To confirm the efficacy of the CELF1, CELF2 and CELF4 expression vectors, hcTNT exon 5-inclusion was analyzed following transfection of HeLa or T98 cells with CELF1,

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CELF2 and CELF4 or a dominant-negative CELF factor (DN-CELF, not shown for HeLa cells). **A.** The protein expression was verified by Western-blotting using a V5-specific antibody since CELF1, CELF2 and CELF4 are fused to a N-terminal V5 epitope and antibodies specific to each of the CELF factor. The expression of the DN-CELF protein was performed using an anti-Xpress™ antibody. Note that CELF factors are not detected in the control condition (pcDNA 3.1 empty vector). We co-transfected HeLa (**B**) or T98 (**C**) cells with 1µg of each splicing factor and 1µg of the RTB300 minigene containing the exon 5 of the human *cTNT* gene (hcTNT). A pcDNA 3.1 empty vector was used as a control. RT-PCR was performed 48h after transfection. Following amplification of hcTNT exon 5, samples were run on an 8% polyacrylamide gel. The upper band contains exon 5 (Exon 5+) while the lower one does not (Exon 5-). The histograms represent the percentage of exon 5 inclusion. Note that the inclusion of hcTNT inclusion is promoted by CEFL4, CELF2 and CELF1 in both HeLa and T98 cells while the DN-CELF does not promote the inclusion.

**Supplementary Figure 6: CELF2 repressed specifically the inclusion of Tau exon 10**

HeLa cells (upper histogram) or T98 cells (lower histogram) were transfect with a pcDNA 3.1 empty vector, CELF1, CELF2, CELF4 or the dominant-negative CELF vectors. Histograms represent the percentage of Tau exon 10-inclusion (mean ± SEM of a minimum of two experiments in triplicate for each condition). The splicing of Tau exon 10 is analyzed by RT-PCR using a forward primer in the exon 9 and a reverse primer in the exon 10. On the PCR gel (middle panel) Tau isoforms having or not the exon 10 are indicated by exon 10+ or exon 10-, respectively. Note that CELF4 expression promotes Tau exon 10-inclusion in both HeLa and T98 cells, while the dominant-negative CELF factor (DN-CELF) does not modify Tau exon 10 splicing.

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**Table 1: Clinical, genetic and neuropathology characteristics of four DM1 patients**

| Case | Age at death | Phenotype class | Intellectual and cognitive status | Neuropathology | Cortical area and size range of the longest (CTG)n expansion |
|---|---|---|---|---|---|
| AD | - | Alzheimer's disease | Demented | NFT | - |
| DM1a | 60 | Adult form | Not evaluated | NFT | Entorhinal 150-1180 |
| DM1b | 64 | Adult form | Cognitive impairment | NFT | Temporal 150-3300 |
| DM1c | 61 | Adult form | Normal | NFT | Temporal 150-2800 |
| DM1d | 53 | Congenital form | Mental retardation, progressive cognitive impairment | NFT | Temporal 1000-2400 |
| DM1e | 61 | Adult form | Not evaluated | NFT | Temporal amygdala 140-1820 |

NFT: neurofibrillary tangles

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Figure 1

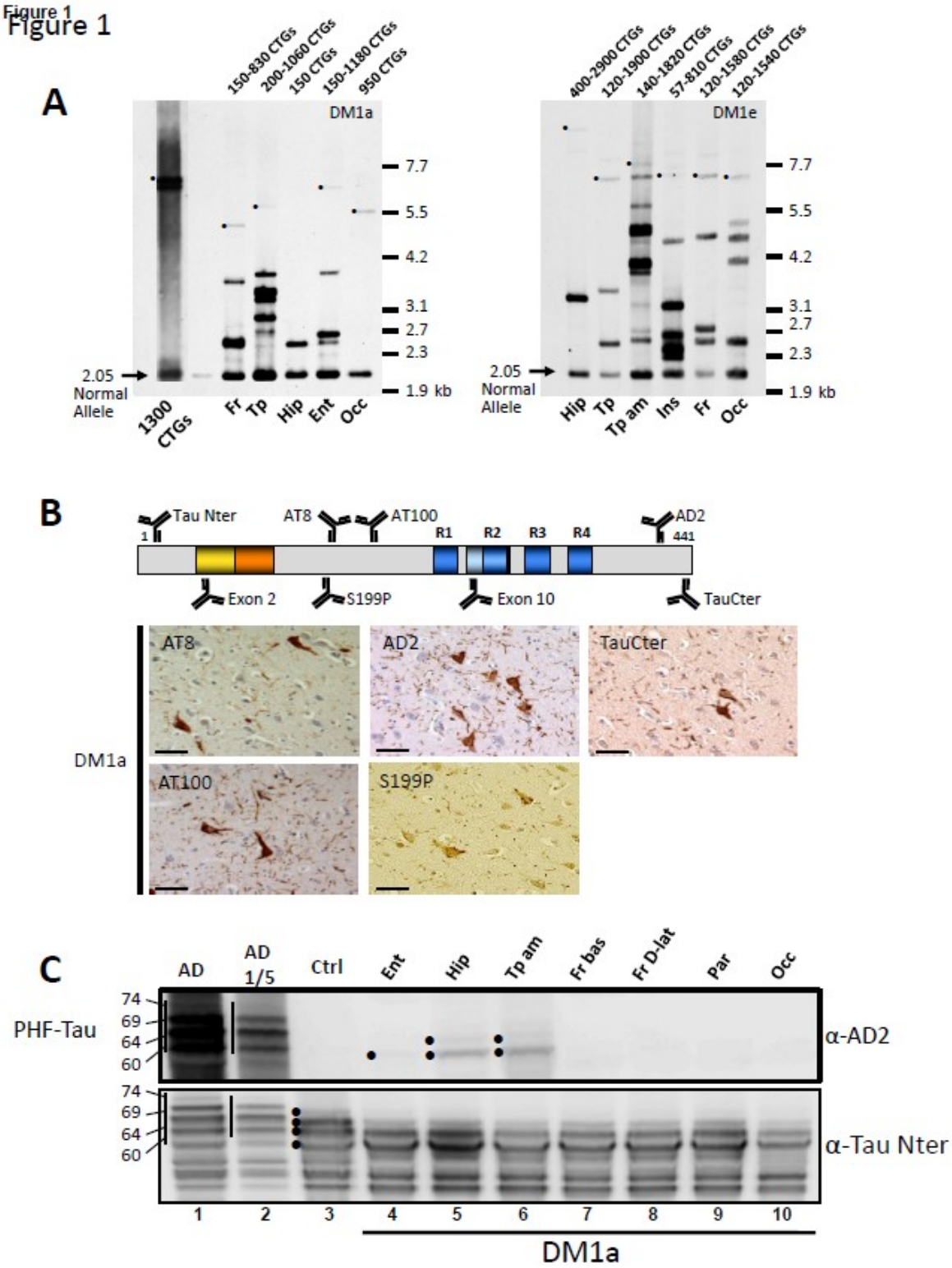

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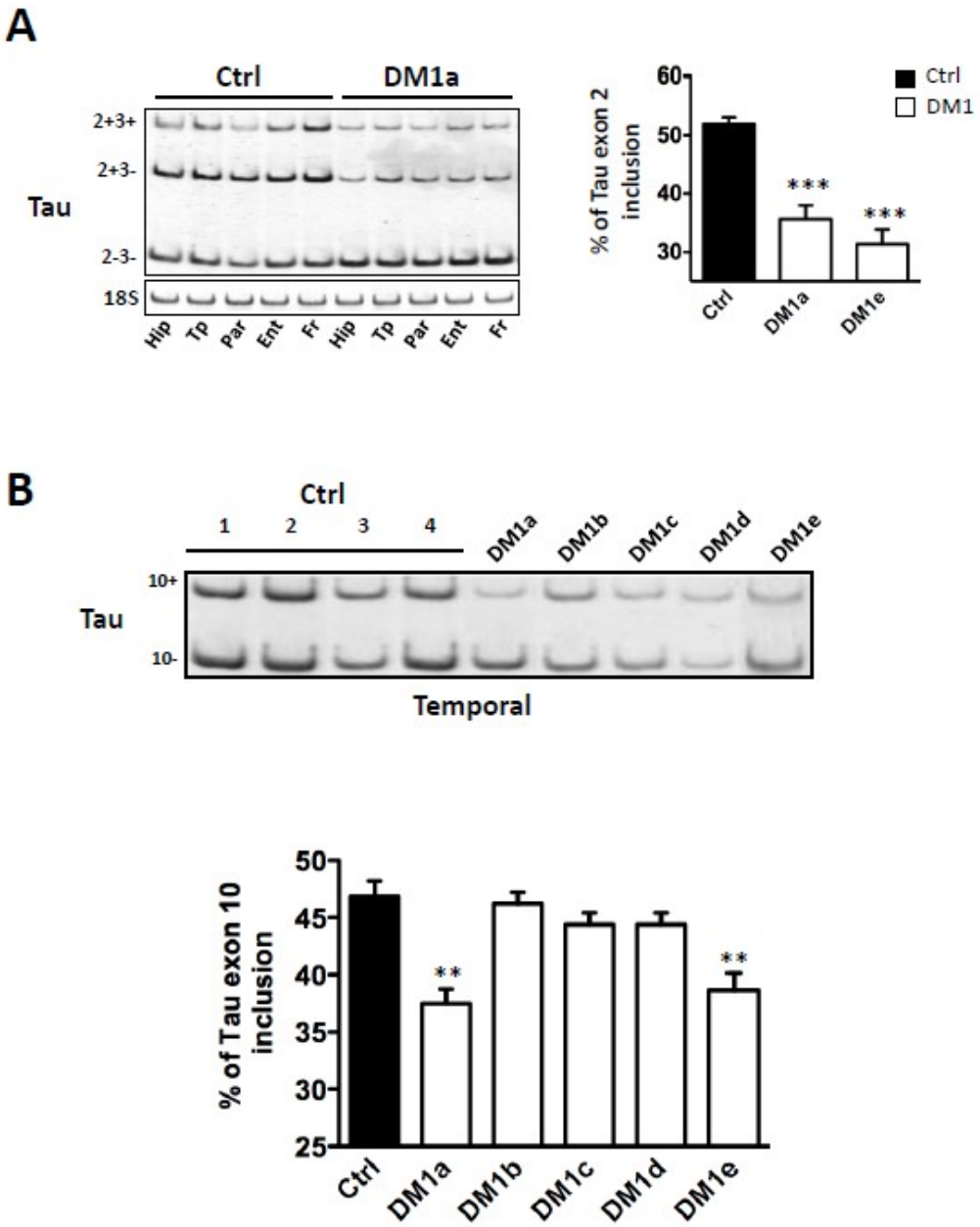
Figure 2



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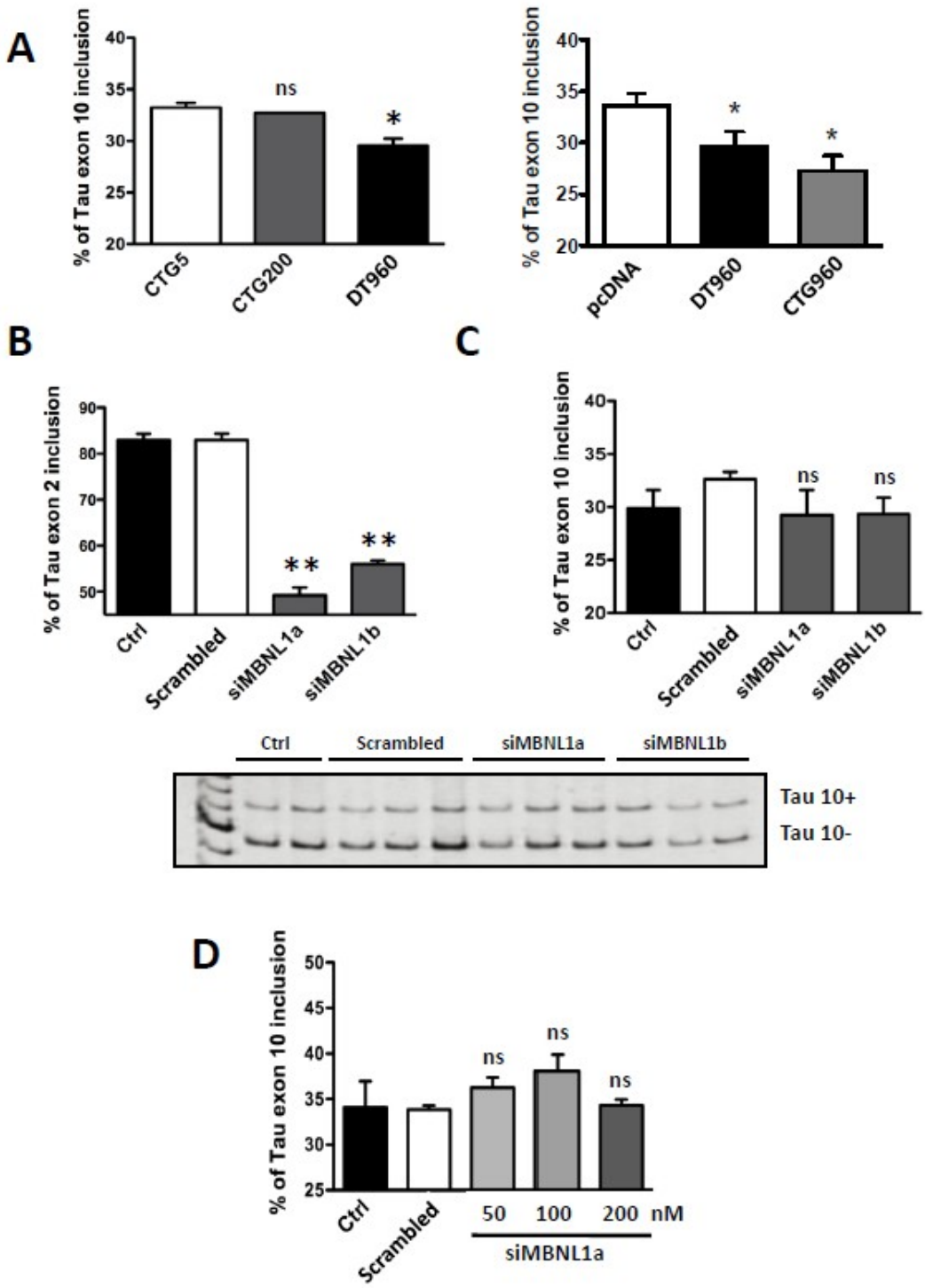

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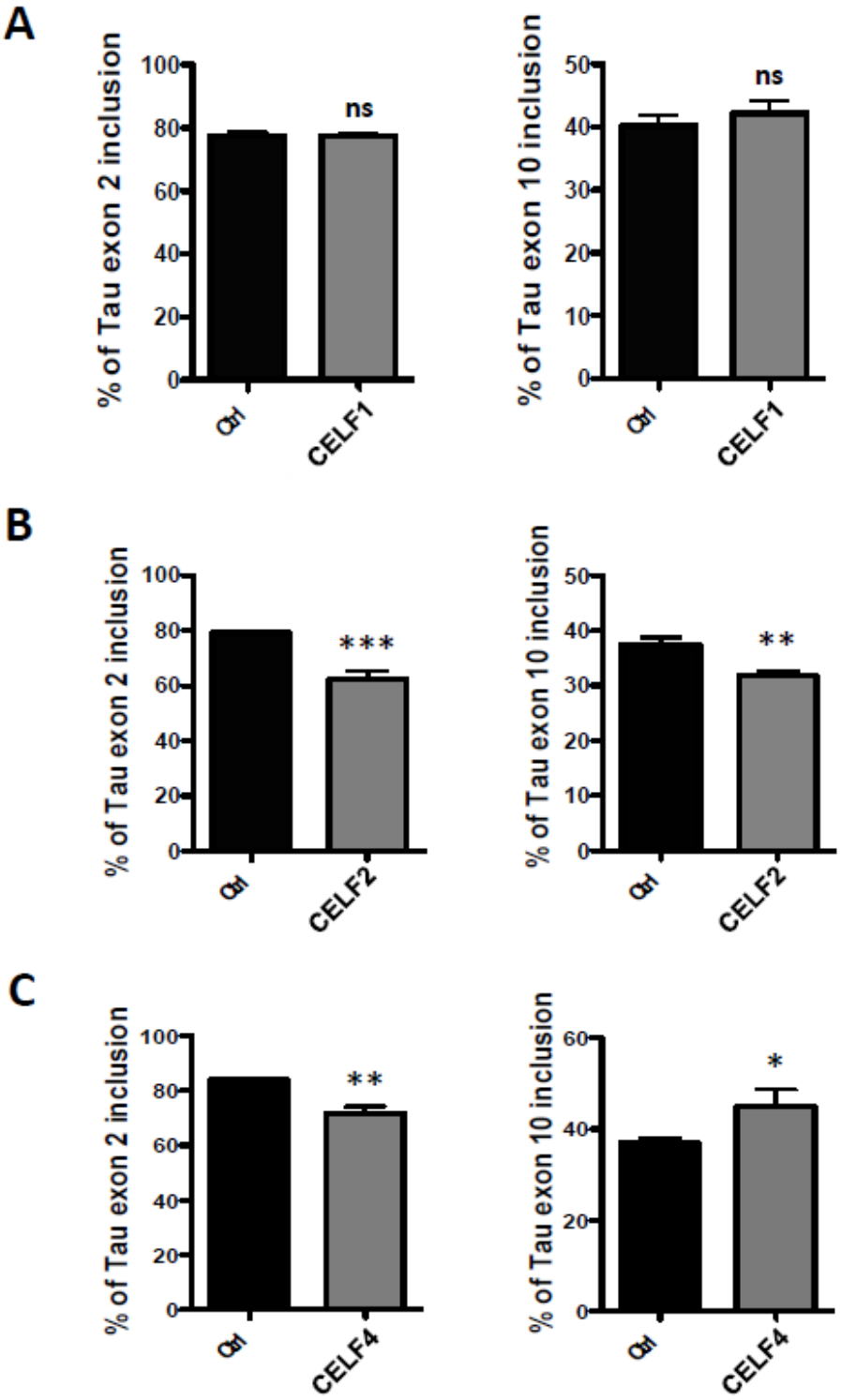

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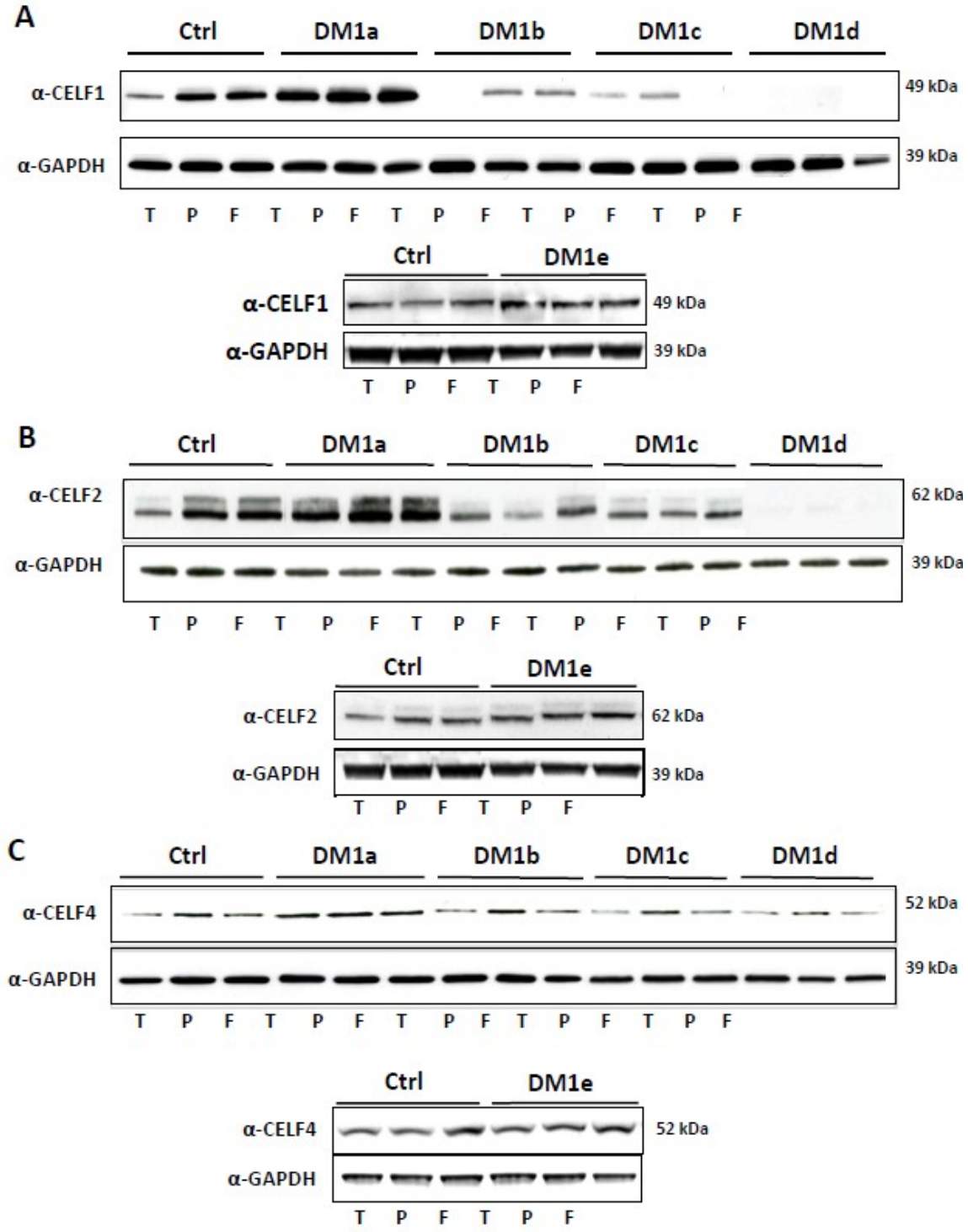
Figure 5