

A distinct CDV genotype causing a major epidemic in Alpine wildlife

Isabella Monne, Alice Fusaro, Viviana Valastro, Carlo Citterio, Manuela Dalla Pozza, Federica Obber, Karin Trevisiol, Mariapia Cova, Paola de Benedictis, Marco Bregoli, et al.

▶ To cite this version:

Isabella Monne, Alice Fusaro, Viviana Valastro, Carlo Citterio, Manuela Dalla Pozza, et al.. A distinct CDV genotype causing a major epidemic in Alpine wildlife. Veterinary Microbiology, 2011, 150 (1-2), pp.63. 10.1016/j.vetmic.2011.01.009 . hal-00687334

HAL Id: hal-00687334 https://hal.science/hal-00687334

Submitted on 13 Apr 2012 $\,$

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Accepted Manuscript

Title: A distinct CDV genotype causing a major epidemic in Alpine wildlife

Authors: Isabella Monne, Alice Fusaro, Viviana Valastro, Carlo Citterio, Manuela Dalla Pozza, Federica Obber, Karin Trevisiol, Mariapia Cova, Paola De Benedictis, Marco Bregoli, Ilaria Capua, & Giovanni Cattoli



PII:	S0378-1135(11)00013-7
DOI:	doi:10.1016/j.vetmic.2011.01.009
Reference:	VETMIC 5147
To appear in:	VETMIC
Received date:	29-6-2010
Revised date:	28-12-2010
Accepted date:	10-1-2011

Please cite this article as: Monne, I., Fusaro, A., Valastro, V., Citterio, C., Pozza, M.D., Obber, F., Trevisiol, K., Cova, M., De Benedictis, P., Bregoli, M., Capua, I., Cattoli, G., A distinct CDV genotype causing a major epidemic in Alpine wildlife, *Veterinary Microbiology* (2010), doi:10.1016/j.vetmic.2011.01.009

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1	A distinct CDV genotype causing a major epidemic in Alpine wildlife					
2						
3	Isabella Monne ¹ , Alice Fusaro ¹ , Viviana Valastro ¹ , Carlo Citterio ¹ , Manuela Dalla					
4	Pozza ¹ , Federica Obber ¹ , Karin Trevisiol ¹ , Mariapia Cova ² , Paola De Benedictis ¹ ,					
5	Marco Bregoli ¹ , Ilaria Capua ¹ & Giovanni Cattoli ¹					
6						
7	¹ Istituto Zooprofilattico Sperimentale delle Venezie, Viale dell'Università, 10, 35020,					
8	Legnaro (PD), Italy.					
9	² Istituto Zooprofilattico Sperimentale della Lombardia e dell' Emilia Romagna, Via P.					
10	Rovelli, 53, 24100 Bergamo (BG), Italy.					
11						
12	Corresponding author: Isabella Monne. Mailing address: Istituto Zooprofilattico					
13	Sperimentale delle Venezie, Viale dell'Università, 10, 35020, Legnaro (PD). E-mail:					
14	imonne@izsvenezie.it.					
15	Phone number: 0039-0498084381, Fax number: 00390498084360.					
16						

1 ABSTRACT

2 Canine distemper virus (CDV) infection represents an important conservation threat to 3 many carnivore species and has contributed to the population decline of several wild 4 terrestrial and aquatic mammalian species. Since 2006, the Alpine region of North-5 Eastern (NE) Italy has been experiencing a severe and widespread outbreak of CDV affecting the wild carnivore population. In this study we performed an extensive 6 7 phylogenetic and molecular evolutionary analysis of CDV identified during the recent 8 wildlife epidemic in the Alpine region. Our analysis yielded data on the evolutionary 9 dynamics of the Alpine wildlife CDV epidemic and revealed the emergence and spread 10 of a single genetic cluster of CDV. The wide distribution of the novel cluster combined 11 with the identification of a specific amino acid mutation, which is believed to increase the ability of the virus to replicate in a wider host range, raises concerns over the 12 13 possible implications of the spread of this virus on the conservation of endangered 14 wildlife species.

15

16 Keywords: Canine distemper virus; wildlife; wild carnivores; evolutionary dynamics;

17 Haemagglutinin gene (H)

18

19

1

2 Introduction

3 Canine Distemper virus (CDV) belongs to genus Morbillivirus, family Paramyxoviridae and is the causative agent of a severe infectious disease affecting a broad range of wild 4 5 and domestic carnivores. This virus possesses a non-segmented single stranded negative 6 RNA genome encoding six proteins. Among them, the haemagglutinin (H) protein is a 7 key component of the virus, being responsible for the first virus-host cell interaction and 8 initial virus entry (Appel, 1987; Martella et al., 2008). The H protein binds to the 9 signalling lymphocyte activation molecule (SLAM) on the membrane of the host target 10 cells resulting in cellular infection. Mapping of the distribution of mutations at sites 11 under positive selection pressure has shown that key amino acid substitutions associated 12 with host specificity fall into the SLAM-binding region of the H protein (McCarthy et 13 al., 2007).

The H gene and its encoded protein are subject to a higher genetic and antigenic variation respectively than other viral genes. For this reason it has been the primary focus of previous molecular epidemiological studies of CDV. Seven clusters have been identified based on the genetic relationship between complete H gene sequences of distinct CDV strains, namely America-1, America-2, Arctic- Like, Asia-1, Asia-2, Europe-1 and Wildlife (Martella et al, 2006; McCarthy et al., 2007).

20 CDV infection represents an important conservation threat to many carnivore species 21 and has contributed to the population decline of several wild terrestrial and aquatic 22 mammalian species (Murray et al., 1999; Kennedy et al, 2000). The mountainous region 23 located in South Central Europe (the Alps) is home to several large endangered 24 carnivore species such as the Eurasian lynx (*Lynx lynx*), the wolf (*Canis lupus*) and the 25 brown bear (*Ursus arctos*), all susceptible to CDV infection (Daoust et al., 2009; 26 Marsilio et al., 1997; Martella et al., 2008). Since 2006, the Alpine region of North-

1 Eastern Italy has been experiencing CDV outbreaks affecting the wild carnivore 2 population. Several foxes, badgers and a stone marten, which were found dead or 3 showed neurological and respiratory signs, tested positive for CDV infection. The early 4 CDV cases were identified only in the North-Eastern part of the Italian Alps, but since 5 2007 infections have spread southwest reaching the Pre-Alps and urbanized areas of the 6 region, thus posing a risk to other wild carnivores and susceptible pets (Figure 1). To 7 date it is unclear how CDV reached these regions, propagated and evolved and whether 8 multiple viral strains of the virus have been involved in this epidemic.

9 This study describes the dynamics of the recent CDV wildlife epidemic through an
10 extensive molecular and phylogenetic analysis of the Italian CDVs identified between
11 2006 and 2009.

12

13 Materials and methods

14

15 Samples

16 The 96 CDV positive samples analyzed in this study were collected between October 17 2006 and March 2009 from three regions namely Veneto, Friuli Venezia Giulia and 18 Trentino Alto Adige located in the north-eastern part of Italy (figure 1). All the samples 19 were collected from wild animals, in particular foxes (n=66) (Vulpes vulpes), badgers 20 (n=29) (Meles meles) and one stone marten (Martes foina). 75 out of 96 animals testing 21 positive for CDV were found dead. The remaining animals displayed one or more of the 22 following clinical signs: myoclonus, ataxia, ocular-nasal discharge and dyspnea. In 23 addition, 23 CDVs detected in domestic dogs from the same regions between 2000 and 24 2007 were included in this study. Information regarding the wildlife CDVs viruses 25 analyzed here (collection date and location) are recorded in table S1 in the supplemental 26 material.

1

2 Nucleotide sequencing

3 Viral RNA was extracted from organ samples (lungs and brains) using the Nucleospin 4 RNA II Kit (Machery-Nagel, Duren, Germany). One step RT-PCR amplification 5 (Qiagen, Hilden, Germany) was performed using specific primers (primer sequences 6 available on request). Sequences were generated using the Big Dye Terminator v3.1 7 cycle sequencing kit (Applied Biosystem, Foster City, CA - USA). The products of the sequencing reactions were cleaned-up using PERFORMA DTR Ultra 96-Well kit (Edge 8 9 BioSystems, Gaithersburg, MD - USA) and sequenced in a 16-capillary ABI PRISM 10 3130xl Genetic Analyzer (Applied Biosystem, Foster City, CA - USA). Sequences of 11 the H gene (from nucleotide 120 to 1824) were aligned and compared with 118 12 sequences representative of viruses belonging to the phylogenetic clades identified so 13 far and available on GenBank (Europa-1, Wildlife, Arctic-like, Asia-1, Asia-2, Asia-3, 14 America-1 and America-2).

Representative CDV nucleotide sequences obtained in this study are available fromGenBank under accession numbers from HM443704 to HM443726.

17

18 *Phylogenetic analysis*

19 A phylogenetic tree was inferred for the H gene using the maximum likelihood (ML) 20 method available in PAUP* (Swofford, 2003). The best-fit model of nucleotide 21 substitution was identified by MODELTEST (Posada et al., 1998) as the general time 22 reversible GTR + Γ_4 + I model, with the frequency of each substitution type, the gamma 23 distribution of among site rate variation with four rate categories (Γ_4) and the proportion 24 on invariant site (I) estimated from the empirical data (parameter values available from 25 the authors on request). Because of the very large size of the dataset, the nearest-26 neighbor-interchanges branch swapping method was employed. To assess the robustness

of individual nodes on the phylogenetic tree, we performed a bootstrap resampling
 analysis (1000 replications) using the neighbor-joining method, incorporating the ML
 substitution model.

4

5 Substitution rates and selection pressure

6 Rates of nucleotide substitution per site, per year and the time of the most recent 7 common ancestor (tMRCA) of the samples analyzed in this study were estimated using 8 Bayesian Markov Chain Monte Carlo (MCMC) approach implemented in the BEAST 9 package version 1.4.8 (Drummond & Rambaut, 2007). Uncertainty in the data is 10 reflected in the 95% highest probability density (HPD) values. Chain lengths were run 11 for sufficient time to achieve coverage as assessed using the Tracer v1.4 program 12 (Rambaut & Drummond, 2007). Two molecular clock models, namely strict (constant) 13 and uncorrelated lognormal relaxed clock, were compared analyzing values of the 14 coefficient of variation (CoV) in Tracer (Rambaut & Drummond, 2007), such that CoV 15 values >0 are evidence of non-clock-like evolutionary behaviour. Because of the 16 typically complex population dynamics we used the Bayesian Skyline plot as a 17 coalescent prior. The GTR + Γ_4 model of nucleotide substitution was used.

To determine the nature of selection pressures acting on H gene of CDV, we computed the mean ratio of nonsynonymous (d_N) to synonymous (d_S) substitutions per site (d_N/d_S) using the Single Likelihood Ancestor Counting and the Fixed Effects Likelihood methods available through DATAMONKEY web interface (Pond et al., 2005).

Fixed amino acid changes along major branches of the phylogeny were identified using
the parsimony algorithm available in the MacClade program (Maddison & Maddison,
2000).

25

26

1 Results

2 Phylogenetic analysis

3 The H gene of 96 CDVs detected in the North-Eastern (NE) part of Italy from wild 4 species, mainly foxes and badgers, between 2006 and 2009 was analyzed. The 5 phylogenetic tree revealed that all the isolates clustered together and with 4 viruses 6 detected in Bavarian wildlife (South Germany) in 2008 (similarity ranged from 99.7% 7 to 100%) within the clade designated as Europe 1 (figure 2). This clade contains viruses 8 detected in the domestic canine population from various European countries, such as 9 Denmark, Germany, Turkey, United Kingdom, Hungary and also Italy. Italian and 10 German wildlife CDVs (Sekulin et al., 2010) were the only viruses belonging to this 11 clade collected from wild animals. Within this clade, all the H gene sequences of CDVs 12 obtained from Italian and Bavarian wildlife formed a separate monophyletic group 13 (figure 2) supported by high boostrap value (100). The highest genetic relationship 14 (from 99.1% to 99.5%) for the strains collected in wild species was found with the H 15 gene sequence of a CDV strain identified in an Hungarian puppy in 2004 (Demeter et al, 16 2007). To facilitate epidemic description, the group formed by the Italian and Bavarian 17 wild CDVs and the Hungarian strain will be termed in the present study as "Wildlife 18 Europe 2006-2009" (WE/06-09).

19 Five CDV viruses, identified between 2000-2007 from Italian domestic dogs (two CDV 20 sequences were obtained in the present study), grouped within the Europe 1 clade but 21 clustered in a separate branch of the phylogenetic tree showing a percentage of 22 similarity lower than 98% with the H gene of CDV strains identified in wild carnivores. 23 The remaining sequences of 21 strains identified from the domestic dog population of 24 the North-Eastern Italian area fell in the Arctic lineage, previously reported in 2 25 distemper episodes in Southern Italy (Martella et al., 2006). This lineage was originally 26 detected in the susceptible population of the Arctic ecosystem and it was recently

1 identified in Eastern Europe (Demeter et al., 2007).

2

3 Evolutionary dynamics of Europe 1.1 CDV

4 The rate of evolutionary change, recorded as nucleotide substitution per site per year 5 (subs/site/year), was estimated using a bayesian coalescent method for the Italian viruses belonging to WE/06-09 cluster. The lower 95% HPD of CoV values of the 6 7 relaxed molecular clock were approximately 0; therefore, a strict molecular clock model was used to estimate evolutionary dynamics. The mean substitution rate of 10.53×10^{-4} 8 9 subs/site/year (95% HPD $5.2-16 \times 10^{-4}$ subs/site/year) was within the range typically 10 observed for CDV (Pomeroy et al., 2008). To further explore the evolutionary origin of 11 this group we estimated the time of the most recent common ancestor (tMRCA). The 12 mean tMRCA was May 2006 (95% HPD June 2005 – September 2006), approximately five months before the identification of the first virus (6th October 2006) belonging to 13 14 this cluster, suggesting a recent introduction of this strain into Northern Italy.

15

16 Selection pressure and molecular characterization

Using the codon-based maximum likelihood methods SLAC (Single Likelihood Ancestor Counting) and FEL (Fixed Effects Likelihood) we computed the ratio of nonsynonymous (d_N) to synonymous (d_S) substitutions (d_N/d_S) at every codon in the alignment of the H gene of 214 sequences of CDV representative of the seven clusters (sequences available in Genbank) and in the alignment of WE/06-09 (101 sequences). In both cases there was no evidence of positive selection. However, the d_N/d_S was higher in WE/06-09 cluster (0.44) than in all the 214 CDV sequences (0.26).

Analysis of the complete alignment using the FEL method allowed to identify nine residues under influence of positive selection at a significance value of p < 0.1 and five at the more stringent significance value of p < 0.05. Three positive selected sites at a

1	significance value of $p < 0.1$ and one at a significance value of $p < 0.05$ were confirmed
2	under the SLAC method (table 1). Interestingly, viruses of the WE/06-09 cluster
3	possessed four amino acid signatures (substitutions L266F, I315V, S/T590I, R597H,
4	Y549H) and three of them were at sites identified under the influence of positive
5	selection (positions 266, 590 and 549). In particular, residue 549 is located in propeller
6	beta-sheet 5 of the SLAM-binding domain and has been demonstrated to be involved in
7	modulation of the fusion activity in CHOdogSLAM cells (von Messling et al., 2005).
8	Analysis of 567 amino acids (40-607) of the Italian isolates sequenced in this study
9	showed 7 potential N-linked glycosylation sites (N-X-S/T) at positions 149-151, 309-
10	311, 391-393, 422-424, 456-459, 587-589 and 603-605. Only one sequence possesses 6
11	potential glycosylation sites (the site at position 603 is missing).
12	Several amino acid substitutions (range: 48-51 substitutions) were observed between
13	Italian viruses isolated from non-dog species and vaccine strains (Onderstepoort,
14	Lederle, Convac, Snyder Hill) in the H protein portion analysed in this study.

15

16 Discussion

17

18 Before 2006, all CDV isolates from wild animals in Europe fell within the Wildlife and 19 Arctic like clades (Martella et al., 2006; Mamaev et al., 1995). The only reported Italian 20 virus which was isolated from a fox (Martella et al., 2006) belonged to the Wildlife 21 clade. Our study describes, for the first time, the emergence and spread of a novel 22 genetic cluster of CDV in wildlife populations of the Alps. This novel genetic group, 23 herein denoted as WE/06-09 cluster, consists almost exclusively of viruses identified 24 from wild carnivores. A single virus from a domestic species, detected in an Hungarian 25 dog in 2004 (Demeter et al, 2007), falls within this monophyletic group although it is 26 distributed in a well distinct branch of the tree. Phylogenetic analysis of available

1 sequences in Genbank revealed that the circulation of the WE/06-09 cluster is not 2 confined to Italian wildlife species. Four CDV sequences obtained from Bavarian wild 3 carnivores (three foxes and one badger) in 2008 (Sekulin et al., 2010) clustered with the Italian CDV, suggesting a wide geographic spread of this new CDV variant in European 4 5 wildlife. These findings raise concerns over the potential impact of infection on 6 conservation programs for endangered carnivores in Europe. Furthermore, the 7 identification of substitution Y549H in the H protein of all CDV strains analyzed 8 underlines the urgent need to provide more insight into the pathogenic properties of this 9 novel strain. A sequence-based study published by McCarthy et al. (2007) showed that 10 the Y549H substitution occurs predominantly in CDV isolates obtained from non-dog 11 host species. It was also reported in a highly virulent strain for ferrets, obtained by 12 repeated passages in these animals (von Messling et al., 2003). Interestingly, this 13 mutation was not observed in the Hungarian canine virus and in the sequences of CDV 14 identified in Italian dogs belonging to Europe 1 clade, suggesting that it appeared only 15 after the introduction of the strain into the wild population.

16 Large scale sequence analysis of CDV obtained during the Italian outbreak revealed that 17 the wildlife species of the North Eastern region of the Italian Alps have been affected by 18 an epidemic wave caused by viruses belonging to a single genetic clade. According to 19 our evolutionary analysis, the common ancestor of the CDV strains, involved in the 20 Italian outbreak, dates back to May 2006 (95% HPD June 2005 – September 2006) 21 approximately five months before the first confirmed case of CDV infection (6th 22 October 2006) in Alpine carnivores was reported. Molecular dating suggests that the 23 ancestor of the WE/06-09 cluster was surprisingly recent and that within a few months 24 this virus had managed to spread over a wide geographical area in Northern Italy. Ten 25 months following its initial detection, the spread of the WE/06-09 cluster had already 26 involved 4 provinces and 21 municipalities of three distinct Italian regions (figure 1).

1 Evaluation of the phylogenetic relationships demonstrated that the CDV described in 2 this study possesses a high genetic similarity (99.1% - 99.5%) with a virus isolated from 3 a dog in Hungary in 2004, indicating that the novel cluster circulated in Eastern or 4 Central Europe at that time. Also of note is that viruses isolated from dogs in Italy 5 between 2000 and 2008 and belonging to the Arctic clade were in the same cluster as 6 the viruses from Hungary. This supports the hypothesis of a significant movement of 7 viruses between Italy and East-Central Europe and intense legal and illegal trade of 8 dogs involving several Eastern European countries may explain the diffusion of the 9 Arctic-like strain in Italy (Martella et al., 2006).

10 Our data does not shed light on how the novel virus was introduced into the wild 11 population of carnivores (e.g. by human-related activities, contacts with domestic 12 carnivores or movement of wild animals). However, data obtained in this study suggest 13 that their subsequent spread was primarily linked to the transboundary movements of 14 wildlife species.

15 A variety of factors may have influenced the emergence and the spread of this large 16 epidemic event, such as demographic dynamics or population immunity. While no data 17 were available on the immune status of the susceptible population in the area before the 18 epidemic, some factors with relation to population dynamics can be considered. The last 19 few decades have seen a dramatic increase in Alpine wild ungulate populations, 20 especially in those areas where human activities linked to the forest have been 21 abandoned (Rizzoli, et al 2009). The increase in ungulate populations has represented an 22 important factor for the re-colonization by large carnivores in Europe (Linnell, et al, 23 2007), and it is likely to have favored the abundance of medium sized carnivores such 24 as foxes. This medium sized opportunistic carnivore can adapt its home range size and 25 contours to resource availability, thus dynamically influencing the total abundance of a 26 population insisting over an area (Lucherini et al., 1996). As frequently occurs in highly

contagious and virulent viruses such as CDV (Morner et al., 2002), the increase in host
 concentration may have enhanced the efficacy of virus transmission.

In Italy, the WE/06-09 cluster has been detected circulating among foxes, badgers and a stone marten, without any apparent spill over to the domestic canine population. In fact, CDVs circulating in dogs obtained from the same geographic area and period grouped in well separate branches of the phylogenetic tree. This may be due to the lack of contact between the two populations or to the strong adaptation of this strain to the wild hosts.

9 Available data concerning CDV outbreaks in the European wild carnivore populations 10 are limited to a few reports from Germany and Italy (Frolich et al., 2000; Martella et al., 11 2002; Sekulin et al., 2010). Furthermore, the available information reports phylogenetic 12 analyses of a small number of samples and so there are little or no data concerning the 13 evolution of this virus in Europe. This appears to be the first comprehensive molecular study on a large collection of CDV from wild carnivores in the European continent, 14 15 providing important basic information on the evolutionary and epidemiological 16 dynamics of an infection which threatens endangered species and pets. This work also 17 prompts a call for improved CDV surveillance in European wildlife populations to 18 obtain a reliable picture of the spread of the novel CDV genotype in this continent.

19

20

21 Acknowledgments

This work was financially supported by the Italian Ministry of Health through the RC
IZS VE 07/08 and RC IZS VE 06/09 projects.

We are grateful to Luigi Bertolotti (University of Studies of Torino, Faculty of Veterinary Medicine, Grugliasco, TO, Italy) and Francesca Cagnacci (Research and Innovation Centre, Environment and Natural Resources Area, Edmund Mach

SCRIP Ŀ. CC

1	Foundation, TN, Italy) for their technical collaboration. Dr William Dundon, Dr
2	Samantha Kasloff and Francesca Ellero (IZSVe) are gratefully acknowledged for the
3	revision of the manuscript.
4	
5	
6	References
7	Appel MJ. 1987. Canine distemper virus. In: Appel MJ editor. Virus infection of
8	carnivores. Amsterdam: Elsevier. p.133-59.
9	Artois M, Delahay R, Guberti V, Cheeseman C. 2001. Control of infectious diseases of
10	wildlife in Europe. 162(2):141-52.
11	Daoust PY, McBurney SR, Godson DL, van de Bildt MW, Osterhaus AD. 2009. Canine
12	distemper virus-associated encephalitis in free-living lynx (Lynx canadensis) and
13	bobcats (Lynx rufus) of eastern Canada. J Wildl Dis.;45(3):611-24.
14	Demeter Z, Lakatos B, Palade EA, Kozma T, Forgách P, Rusvai M. 2007. Genetic
15	diversity of Hungarian canine distemper virus strains. Vet Microbiol.;122(3-4):258-69.
16	Drummond, A. J. and A. Rambaut. 2007. BEAST: Bayesian evolutionary analysis by
17	sampling trees. BMC Evol. Biol. 7:214.
18	Frölich K, Czupalla O, Haas L, Hentschke J, Dedek J, Fickel J. 2000. Epizootiological
19	investigations of canine distemper virus in free-ranging carnivores from Germany Vet
20	Microbiol.;74(4):283-92.
21	Kennedy, S., Kuiken T., Jepson P. D., Deaville R., Forsyth M., Barrett T., Van de Bildt
22	M. W. G., Osterhaus A., Eybatov T., Duck C., Kydyr-Manov A., Mitrofanov I., Wilson
23	S 2000. Mass die-off of Caspian seals caused by canine distemper virus. Emerging
24	Infectious Diseases 6: 637–639

Linnell, J.D.C., Salvatori, V. & Boitani, L. 2007. Guidelines for population level 25

Page 13 of 20

1 management plans for large carnivore in Europe. LCIE report prepared for the European

2 Commission.

Loeffler IK, Howard J, Montali RJ, Hayek LA, Dubovi E, Zhang Z, Yan Q, Guo W,
Wildt DE. 2007. Serosurvey of ex situ giant pandas (Ailuropoda melanoleuca) and red
pandas (Ailurus fulgens) in China with implications for species conservation. J Zoo
Wildl Med.;38(4):559-66.

7 Lucherini M and LovariS 1996. Habitat richness affects home range size in the red fox
8 Vulpes vulpes. Behavioural Processes 36: 103-105

9 Maddison, W. P. and D. R. Maddison. 1989. Interactive analysis of phylogeny and
10 character evolution using the computer program MacClade. Folia. Primatol. (Basel)
11 53:190-202.

12 Mamaev LV, Denikina NN, Belikov SI, Volchkov VE, Visser IK, Fleming M, Kai C,

13 Harder TC, Liess B, Osterhaus AD, Barrett, T. 1995. Characterisation of morbilliviruses

14 isolated from Lake Baikal seals (Phoca sibirica). Vet Microbiol. May;44(2-4):251-9.

15 Marsilio F, Tiscar PG, Gentile L, Roth HU, Boscagli G, Tempesta M, Gatti A. 1997.

Serologic survey for selected viral pathogens in brown bears from Italy. J WildlDis.;33(2):304-7.

Martella V, Pratelli A, Cirone F, Zizzo N, Decaro N, Tinelli A, Foti M, Buonavoglia C.
2002. Detection and genetic characterization of canine distemper virus (CDV) from
free-ranging red foxes in Italy. Mol Cell Probes.;16(1):77-83.

Martella V, Cirone F, Elia G, Lorusso E, Decaro N, Campolo M, Desario C, Lucente
MS, Bellacicco AL, Blixenkrone-Møller M, Carmichael LE, Buonavoglia C. 2006.
Heterogeneity within the hemagglutinin genes of canine distemper virus (CDV) strains
detected in Italy. Vet Microbiol.;116(4):301-9.

- 1 Martella V, Elia G, Buonavoglia C. 2008. Canine distemper virus. Vet Clin North Am
- 2 Small Anim Pract.;38(4):787-97, vii-viii.
- McCarthy AJ, Shaw MA, Goodman SJ. 2007. Pathogen evolution and disease
 emergence in carnivores. Proc Biol Sci. 274(1629):3165-74.
- 5 Murray D.L., Kapke C.A., Everman J.J. and Fuller T.K. 1999. Infectious disease and the
- 6 conservation of free-ranging large carnivores, Animal Conservation 2 (1999), pp. 241-

7 254.

- Pomeroy LW, Bjørnstad ON, Holmes EC 2008. The evolutionary and epidemiological
 dynamics of the paramyxoviridae. J Mol Evol.;66(2):98-106.
- Pond SL, Frost SD 2005. Datamonkey: rapid detection of selective pressure on
 individual sites of codon alignments. Bioinformatics.21(10):2531-3.
- 12 Posada, D. and K. A. Crandall. 1998. MODELTEST: testing the model of DNA
- 13 substitution. Bioinformatics 14:817-818.
- 14 Rambaut, A. and A. J. Drummond. 2007. Tracer v1.4.
- 15 Rizzoli, A.P., Heidi, H.C., Tagliapietra, V. Neteler, M., Rosà, R. 2009. Forest structure
- 16 and roe deer abundance predict Tick-Borne Encephalitis risk in Italy. PlosOne 4(2).
- 17 Seki, F., Ono, N., Yamaguchi, R., Yanagi, Y., 2003. Efficient isolation of wild strains of
- 18 canine distemper virus in Vero cells expressing canine SLAM (CD150) and their
- adaptability to marmoset B95a cells. J Virol. 77,9943-50.
- 20 Sekulin K, Hafner-Marx A, Kolodziejek J, Janik D, Schmidt P, Nowotny N. 2010.
- 21 Emergence of canine distemper in Bavarian wildlife associated with a specific amino
- 22 acid exchange in the haemagglutinin protein. Vet J. In press.
- 23 Thulke HH, Eisinger D, Freuling C, Fröhlich A, Globig A, Grimm V, Müller T, Selhorst
- 24 T, Staubach C, Zips S. 2009. Situation-based surveillance: adapting investigations to
- 25 actual epidemic situations. J Wildl Dis.;45(4):1089-103.

- 1 von Messling V, Springfeld C, Devaux P, Cattaneo R. 2003. A ferret model of canine
- 2 distemper virus virulence and immunosuppression. J Virol. Dec;77(23):12579-91.
- 3 von Messling, V., Oezguen, N., Zheng, Q., Vongpunsawad, S., Braun, W., Cattaneo, R.,
- 4 2005. Nearby clusters of hemagglutinin residues sustain SLAM-dependent canine
- 5 distemper virus entry in peripheral blood mononuclear cells. J Virol. 79, 5857-62.
- 6 Wilgenbusch, J. C. and D. Swofford. 2003. Inferring evolutionary trees with PAUP*.
- 7 Curr. Protoc. Bioinformatics Chapter 6:Unit 6.4.
- 8
- 9

Figure 1. Geographical distribution of the 96 CDV strains from wildlife species analysed in the present study.

Figure 2. Maximum-likelihood (ML) phylogenetic tree for the Haemagglutinin gene of selected CDV strains. Sequences of CDV viruses analyzed in this study are highlighted in yellow. The seven main CDV clusters are identified by distinct colours. The numbers at each branch point represent bootstrap values (black). GenBank accession numbers of the CDV sequences downloaded from the public database are included.

	SLAC positive	ly selected sites	FEL positively selected sites			
	(codon position)		(codon position)			
	p < 0.1	p < 0.05	p < 0.1	p < 0.05		
			266			
			314			
			401	266		
CDV H gene	401		412	401		
	412	549	417	412		
	549		460	549		
			549	583		
			583			
			590			

Table 1.	Summarv	of the se	lection	pressures	acting in	the H	gene of	CDV	analvzed
10010 11	~ mininar j	01 010 00		p10000100			8	· · ·	

Click here to download high resolution image



