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Author manuscript

Targeting natural killer cells and natural killer T cells in cancer

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Preface text

Natural killer (NK) and NKT cells are subsets of lymphocytes that share some phenotypic and functional similarities. Both cell types can rapidly respond to the presence of tumour cells and participate in antitumour immune responses. This has prompted interest in the development of innovative anticancer therapies that are based on the manipulation of NK and NKT cells. Recent studies have highlighted how the immune reactivity of NK and NKT cells is shaped by the environment in which they develop. The rationale use of these cells for cancer immunotherapies awaits a better understanding of their effector functions, migratory patterns and survival properties in humans.

Introduction

The immune system is classically divided into innate and adaptive branches. Adaptive immunity can be defined by the presence of cells (T cells and B cells in higher vertebrates) that can respond to many diverse environmental antigens. This is achieved by the clonal expression of a colossal repertoire of receptors with antigen specificities (T cell receptors (TCRs) and B cell receptors (BCRs)), the diversity of which results from somatic DNA rearrangements. In contrast, the recognition of various assaults by cells of the innate immune system has been described to depend so far only upon germline-encoded receptors. A recent paradigm shift in our understanding of immunity in mammals has resulted from the

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discovery of these recognition receptors used by the innate immune system, such as the Tolllike receptors (TLRs), and their specificity $^{1-3}$.

Besides conventional T and B cells, a series of innate lymphoid cells (ILCs) were recently identified ⁴. ILCs include various cells of the innate immune system, such as lymphoid tissue-inducer (LTi) cells, but also cells that produce interleukin-5 (IL-5), IL-13, IL-17 and/or IL-22 helping to initiate immune responses to pathogens. Natural killer (NK) cells are now recognized as a subset of cytotoxic ILCs that express the transcription factor E4BP4/ Nfil3. NK cells also secrete cytokines, such as interferon- γ (IFN- γ), that participate in the shaping of the adaptive immune response ⁵. An important feature of NK cells is their capacity to distinguish stressed cells (such as tumour cells, infected cells and cells which have undergone physical or chemical injuries) from normal cells. NK cells were initially identified through their ability to kill tumour cells (hence their name) $^{6-8}$. Since then, the anti-tumour effect of NK cells has been documented in many models and instances. In vitro, mouse and human NK cells can kill a broad array of tumour cells of hematopoietic and nonhematopoietic origin. In vivo, mouse NK cells can eliminate many transplantable and spontaneous tumours ^{9, 10}. Selective NK cell deficiencies are extremely rare¹¹, thus preventing the monitoring of a high incidence of cancers in these patients, but also possibly testifying of the physiological importance of NK cells. Nevertheless, an epidemiological study has linked low peripheral blood NK cell activity with increased cancer risk ¹². In addition, NK cell infiltration in tumour tissue is associated with better disease prognosis in non-small cell lung carcinomas ^{13, 14}, clear cell renal cell carcinoma ¹⁵ and colorectal cancer ¹⁶. Observations in patients with advanced gastrointestinal stromal tumour treated with imatinib mesylate support the hypothesis that NK cells exert anti-tumour effects not only through direct cytolytic activity, but also indirectly through their ability to produce cytokines such as IFN- γ ¹⁷.

Natural Killer T (NKT) cells have been classified in four different groups ¹⁸. Only type I and type II NKT cells are CD1d restricted as they respond to CD1d expressing cells and are absent in CD1d deficient mice ¹⁸ (Box 1). The most studied group, type I NKT cells or invariant NKT cells (iNKT cells), is well conserved in mammals ¹⁸. iNKT cells develop within the thymus, arising from the same common lymphoid precursor pool from which conventional T cells develop ^{19, 20}. Once the aß T cell lineage commitment is made and double positive thymocytes are generated, the iNKT cell and conventional T cell selection pathways diverge $^{21, 22}$. iNKT cell precursors are selected following α -chain rearrangement and expression of the semi-invariant T cell receptor (Va14-Ja18 in mice, Va24-Ja18 in humans) ^{21, 23–26}. In contrast to conventional T cells that are selected by antagonist/partial ligands, it is believed that iNKT cells are selected by agonist glycolipids ^{27–29}. iNKT express respond rapidly to a variety of glycolipids presented by CD1d ^{30, 31} (Box 1). Strong iNKT cell agonists, such as a-galactosylceramide (a-GalCer), induce an immediate and powerful cytokine release ^{32–35}. Similarly to NK cells, a role for iNKT cells in tumour immuno-surveillance has been documented ^{36–39} and their potential in anti-tumour therapy is beginning to be uncovered.

Box 1

CD1d-restricted T cells and activation of iNKT cells

CD1d restricted cells are well conserved in mammals and have been initially classified as type I NKT cells and type II NKT cells. Type I NKT cells (iNKT cells) express a semiinvariant TCR while type II NKT cells express a more diverse TCR. iNKT cells have been characterized using the CD1d tetramer loaded with α -GalCer or other agonist in both mouse and human as well as using the 6B11 monoclonal antibody in human. Type II NKT cell functions are unclear and have been indirectly examined by comparing the phenotype and immune response of CD1d and Ja18 deficient animals. iNKT cell frequencies is lower in humans than mice while type II NKT cell frequency is mostly unknown due to lack of tools to phenotype them. Although all iNKT cells can be identified by the CD1d tetramer, recent data indicate that there are several subsets of iNKT cells with different functions and phenotype. Notably a α-GalCer-reactive NKT cell subset that expresses a canonical Va10-Ja50 TCR was recently identified ²¹³. iNKT cells are unique as they have the ability to respond as innate cells with minimal TCR involvement or as memory like cells through engagement of their semi invariant TCR. The TCR dependent activation has been extensively studied using the strong agonist α -GalCer. In this case, the iNKT cell response is CD1d dependent and inflammatory cytokines are dispensable. In fact α -GalCer loaded CD1d coated on a plate is sufficient to activate iNKT cells. Interestingly pathogens, which are known to express iNKT cell antigens and require iNKT cells for effective protection, predominantly depend on inflammatory cytokines such as IL-12. The indirect pathway has recently been characterized during viral infection and is active in the presence of large amounts of inflammatory cytokines such as IL-12, IL-18 and IFN-a. In this case, iNKT cells behave functionally as classical NK cells and produce mostly Th1 cytokines. iNKT cells can also be stimulated by a combination of inflammatory cytokines and CD1d/TCR dependent stimulus. This has been reported with bacteria such as Salmonella typhimurium, Staphylococcus aureus and Mycobacterium tuberculosis that lack agonist glycolipids, and activate iNKT cells through recognition of presumably endogenous glycosphingolipids such as β -D-glucopyranosylceramide ²¹⁴, presented by pathogen-activated dendritic cells 215-218.

Although their developmental programmes are controlled by different transcription factors (PLZF and E4BP4/Nfil3 respectively) ^{40–43}, iNKT and NK cells have phenotypic and functional similarities. For instance, NK cells use Ly49H, a receptor that can be seen as an invariant TCR-like receptor, to recognize the mouse cytomegalovirus (MCMV) protein m157, and functionally behave as innate T cells ⁴⁴. Conversely, during certain viral infections, iNKT cells can be activated by inflammatory cytokines with minimal TCR involvement and functionally behave as NK like cells ^{45–47}. In addition, both NK and iNKT cells are poised to secrete cytokines ⁴⁸ and they depend on IL-15 and T-bet for maturation and homeostasis ^{49–51}. However, unlike NK cells, which rely mostly on perforin/granzyme-mediated mechanisms, iNKT cell cytotoxicity is mostly restricted to the CD95/CD178 pathway ⁵².

In this article, we discuss recent research on the role of NK and NKT cells in the control of tumours with a special emphasis on how the molecular dissection of their mode of recognition of tumour cells is leading to the development of innovative cancer therapies. The heterogeneity of NK and NKT cells, their roles in infections or auto-immunity, as well as their regulatory function in the shaping of adaptive immunity will not be discussed here because these topics have been extensively covered elsewhere.

How do NK cells recognize tumour cells?

NK cells express an array of receptors, notably expressed at the cell surface, that enable them to detect their cellular targets while sparing normal cells. These molecules include inhibitory, activating, adhesion and cytokine receptors. NK cells integrate these various signals that might be transduced simultaneously during the contact with the interacting cells, dictating whether NK cells will be activated or not (Figure 1). Whether and how this integration of signals leads to a commensurate or all-or-none NK cell response (such as for target cell lysis) remains to be dissected.

Missing-self and NK cell education

NK cells express MHC class I-specific inhibitory receptors (Box 2). These receptors and their ligands (H-2 in mice and HLA in humans) are highly polymorphic molecules encoded by multigenic, multiallelic families of genes that are inherited independently ^{53, 54}. NK cells have thus to discriminate self in a context where self molecules differ from individual to individual. Similarly to T cells, NK cells are educated to self versus altered-self discrimination, but the molecular strategies involved in this education are different. T cell education involves the stimulatory T cell receptor whereas NK cell education is mediated through the engagement of the MHC class I-specific inhibitory receptors. This education, also termed 'licensing' or 'arming' leads to the maturation of a NK cell functional repertoire (that is, the ensemble of signals towards which NK cells are reactive), which is adapted to self-MHC class I environment^{5, 55-60}. In MHC class I-deficient individuals, NK cells are hyporesponsive to stimulatory receptor stimulation and thereby tolerant to self ^{61, 62}. Hyporesponsiveness of NK cells grown in a MHC class I-deficient environment can nevertheless be overcome in inflammatory conditions ⁶³. Thus, 2 mechanisms of selftolerance for NK cells coexist in steady-state conditions: functionally competent NK cells, whose effector responses are inhibited by the recognition of self MHC class I molecules, and hyporesponsive NK cells that cannot detect self-MHC class I. Of course, this is a simplistic view, and NK cell education does not result in a bipolar situation, but rather in a continuum allowing fine tuning of NK cell responsiveness. For example, NK cells that express higher levels of inhibitory receptors specific for self MHC class I molecules show greater immunoreactivity that NK cells that express lower levels of inhibitory receptors specific for self MHC class I molecules 60, 64, 65. The molecular mechanisms underlying this MHCdependent NK cell education are only partially understood. Nevertheless, a functional immunoreceptor tyrosine-based inhibitory motif (ITIM) is necessary in the intracytoplasmic tail of the mouse Ly49 inhibitory receptors ⁵⁵. In addition, NK cell education via MHC class I-specific inhibitory receptors does not substantially alter the NK cell transcriptional program. Rather, it is associated with the membrane confinement of activating receptors in

nanodomains, whereas these receptors are preferentially located in actin-rich membrane meshwork in uneducated NK cells ⁶⁶. Thus, competent and hyporesponsive NK cells might switch from one state to another via this mechanism dependent on membrane confinement of activating receptors, conferring NK cells more plasticity than originally thought. Along this line, NK cells expressing inhibitory receptors revert from a hyporesponsive to a competent status upon exposure to cognate MHC class I molecules ^{67, 68}. Therefore, upon encounter with target cells expressing low MHC class I surface density, one could predict that the reactivity of NK cells will decrease upon adaptation to this altered MHC class I environment. Despite the recent confirmation of the importance of 'missing self' recognition for NK cell responsiveness to tumours in vivo in a mouse model of chronic myeloid leukaemia ⁶⁹, the kinetics of NK cells' adaptation to their surrounding milieu which are still unclear, represent a key issue to understand the reactivity of NK cells.

Box 2

NK cells and missing-self

NK cells use inhibitory receptors to detect the presence of constitutively expressed selfmolecules on susceptible target cells. In particular, NK cells express MHC class Ispecific receptors and are relieved from inhibitory signals when encountering MHC class I–deficient cells ^{219–224}. As a consequence, NK cells can recognize 'missing self' on interacting cells ²¹⁹. The MHC class I–specific inhibitory receptors include the killer cell immunoglobulin-like receptors (KIRs) in humans, the lectin-like Ly49 dimers in the mouse and the lectin-like CD94-NKG2A heterodimers in both species ^{54, 222–227}. These inhibitory receptors possess intracytoplasmic inhibitory signalling domains called immunoreceptor tyrosine-based inhibition motifs (ITIMs) that mediate their inhibitory function ^{228–230}. Other inhibitory receptors (for example, mouse NKR-P1B, human NKR-P1A and mouse 2B4) that recognize non-MHC self-molecules (for example, Clr-b, LLT-1 and CD48, respectively) also regulate NK cell activation ²³¹.

Stress-induced self recognition

Besides using inhibitory receptors that recognize self, NK cells are also equipped with cell surface activating receptors ^{70, 71}. In addition to the recognition of microbial molecules by a variety of innate immune receptors, the so-called 'infectious non-self recognition', it has been shown that several receptors of innate immune cells can detect internal changes that occur in damaged host tissues, leading to the concept of 'stress-induced' self recognition'^{72–74}. This mode of detection relies on the recognition of self molecules that are barely detectable in steady-state conditions, but whose expression increases in various forms of stress. A prototypical example is the activation of NK cells via engagement of the activating NKG2D receptor. NKG2D interacts with self-molecules that are selectively upregulated on stressed cells, such as tumour cells ^{75–78}. *In vivo*, NKG2D was shown to be crucial for immunosurveillance of epithelial and lymphoid malignancies in two transgenic models of *de novo* tumour genesis ⁷⁹. In the transgenic Eµ-Myc mouse model of spontaneous B cell lymphoma, the tumour expression of NKG2D ligands represents an early step of tumour genesis that is associated with still unknown genetic lesions of cancer

cells ⁸⁰. NKG2D ligands are the stress-inducible MICA/B and ULBP/RAE proteins in humans ^{72, 77} and Rae1, H60, and Mult-1 proteins in mice ^{75, 76, 78}. A link between tumour genesis, DNA damage response (DDR) and the immune response has been proposed. DNA-damaging agents or DNA lesions associated with tumour genesis activate the DDR, which results in up-regulation of NKG2D ligands leading NK cells to attack the diseased cells ⁸¹. The up-regulation of NKG2D ligands depends on the PI-3 kinase-related ATM (ataxia telangiectasia, mutated) or ATR (ATM- and Rad3-related) protein kinases, which initiate the DDR pathway after exposure to DNA damage ⁷³. Treatment with proteasome inhibitors also induces NKG2D ligand expression in multiple myeloma cells via the ATM/ATR pathway ⁸².

Besides NKG2D, NK cells express an array of cell surface molecules, such as the natural cytotoxicity receptors (NCR), which were shown to be involved in the activation of NK cells by tumour cells more than a decade ago. The NCR family includes NKp46 (NCR1, CD335) 83, NKp44 (NCR2, CD336) 84 and NKp30 (NCR3, CD337) 85. NCR association with immunoreceptor tyrosine-based activation motif (ITAM)-bearing transducing polypeptides is reminiscent of the architecture of other pivotal immune receptor complexes, such as the TCR-, BCR- and Fc-receptors, and makes them very potent activating receptors ⁷¹. The tumour cell surface ligands for the NCR family have remained elusive. hindering a complete understanding of their role in tumour surveillance. An exception resides in the identification of a novel member of the B7 family of immunoreceptors, B7-H6, as a cellular ligand for NKp30^{86,87}. The expression of B7-H6 on tumour cells induces NKp30-dependent NK cell activation and cytotoxicity. Importantly, B7-H6 is absent from all normal cells tested in steady-state conditions, but is expressed by tumour cells. Considering that NK cells do not compromise the integrity of normal healthy cells and tissues, a reasonable hypothesis is that, as for NKG2D ligands, self-ligands for all activating NK cells receptors are tightly down-regulated in healthy cells and are up-regulated in stressed cells, such as tumour cells. There are many other activating receptors and adhesion molecules that are present on NK cells and may participate in the recognition of tumour cells. These receptors include DNAM-1 via its ligands poliovirus receptor (PVR) and Nectin-2⁸⁸, and the signalling lymphocytic activation molecule (SLAM)-receptors (2B4, NTB-A) as reviewed elsewhere ⁸⁸. Importantly, both human and murine NK cells also express FcyRIIIA (CD16), which recognizes antibody-coated target cells through their Fc portion and mediates antibody-dependent cellular cytotoxicity (ADCC). Growing evidence indicates that ADCC contributes to the beneficial clinical response of human cancers to several monoclonal antibody-based therapies, including treatment with rituximab ^{89, 90}, implying that NK cells are involved in the antitumour response in these settings.

Manipulation of NK cells in antitumour therapy

The remarkable conservation of their antitumour activity against many types of murine and human tumours suggests that NK cells detect common modifications in cellular metabolism and/or gene expression that are shared or induced by many oncogenic processes. This ability of NK cells to target a common mechanism present in cancers while respecting the integrity of healthy cells, has led to them being considered as promising therapeutic tools for cancer immunotherapy ⁹¹.

Allogeneic hematopoietic stem cell transplantation for cancer patients

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is an efficient form of adoptive immunotherapy in the treatment of patients with haematological malignancies ⁹², although the therapeutic effect was originally thought to derive mostly from the escalated doses of cytotoxic agents and radiation therapy used in the so-called "conditioning regimen". The reality of a graft-versus-leukaemia (GVL) effect developed by the allogeneic graft was nevertheless established more than 3 decades ago 93. Since this time, the possibility of disease control in other haematological malignancies ⁹⁴ but also in solid tumours ^{95, 96} has been confirmed. Despite these proof-of-concepts, allo-HSCT has long been carried out in a limited subset of cases as compared to the authentic extent of potential population. This is in part due to the high rate of fatal toxicities attached to the procedure. which eventually impairs patient survival despite effective disease control. These complications are mainly related to the inner mechanism of this treatment which relies on immunological effect ⁹⁷, at least when used to treat patients with various forms of advanced or poor-prognosis cancers. Indeed alongside the searched GVL effect, allogeneic immune activation is the source of a Graft-Versus-Host disease (GVHD) recognition targeting minor histocompatibility antigens on non tumour tissues. Even in the context of fully HLA identical sibling donor, this latter effect is highly effective: when fully developed, it conducts to unwanted and potentially serious injuries of normal tissue as gut, skin and liver epithelia. This has and still represents an important limitation for allo-HSCT development along with the lack of donors. Indeed, for years, allogeneic HSCT protocols have failed to manipulate the immune system to boost graft-versus-tumour (GVT) effects without inducing GVHD. In order to solve this issue, most of HSCT protocols have relied on the manipulation of T cell responses, an attempt that proved to be extremely hazardous due to the underestimated and unpredictable cross-reactivity of the TCRs. In contrast, it is now established in pre-clinical and clinical settings that NK cells have a unique capacity to exert potent GVT effects without inducing GVHD, a feature which most likely results from the differential distribution of ligands for activating NK cell receptors on hematopoietic and nonhematopoietic tissues, such as epithelial cells ⁹⁸.

Allo-HSCT has been an evolving field in the last two decades, as a consequence of the diversification of stem cell and donor sources and changes in conditioning regimen. One type of allo-HSCT is the so-called 'haplo-identical' transplantation, which was developed to overcome the problem posed by a lack of an HLA-matched donor and has been recently put in light again the potential of NK cells in cancer control. In haplo-identical transplantation the related donor and recipient share only one haplotype encoding HLA molecules. In some but not all cases, the recipient expresses MHC alleles (that is, KIR ligands) that are not expressed by the donor. In this 'KIR ligand-mismatched' situation, subsets of donor-derived NK cells that are not restrained by host MHC class I molecules develop in the recipient and have GVL potential. In classical haplo-identical HSCT protocols, the allograft is depleted of T-cells before reinfusion to reduce the incidence of severe and potentially lethal GVHD, and the patient receive highly cytotoxic and immune-suppressive conditioning regimen to prevent graft rejection ⁹⁹. In these settings, recipients with acute myeloid leukaemia (AML) who received a haplo-identical transplant from a KIR ligand-mismatched donor had a marked reduction in relapse compared with an otherwise similar group of AML patients

transplanted from a KIR ligand-matched related donor¹⁰⁰. In haplo-identical HSCT, the positive influence of KIR-incompatibility was detected in other studies with patients with AML ^{101, 102} or multiple myeloma ¹⁰³, but not in others ¹⁰⁴, indicating that other parameters in the treatment protocol - such as the dose of stem cells and the extent of T cell depletion contribute to clinical outcome. Hence, the spectrum of alloreactivity displayed by donorderived NK cells remains to be fully elucidated (Figure 2a). Clinical efficacy against residual patient tumour cells requires that fully competent NK cells of donor origin develop in the recipient following transplantation. Considering the role of MHC class I in NK cell education and the plasticity of NK cells described above, the development of alloreactive NK cells of donor origin is likely to depend upon the recipient's conditioning regimen and the dose of donor hematopoietic progenitors (that is, the likelihood and the duration of interaction with donor or recipient MHC class I molecules). The complexity and changing nature of such interactions is a likely explanation for the apparently inconsistent observations of NK cell alloreactivity in different protocols and studies. The challenge will be to harness the therapeutic potential of NK cells in this context. NK cells could also be used as part of the conditioning regimen, as it has been shown in mouse models that donor NK cells can kill host dendritic cells impairing donor T cell-mediated GVHD ¹⁰⁰. Irrespective of this possibility, NK cells are especially attractive therapeutic tools because their association with a reduction in relapse rates was not associated with increased GVHD incidence. Though limited, these observations thus suggest that NK cell manipulation can dissociate GVL and GVHD effects, an objective that has never been attained through the manipulation of T cell compartments. In addition, cyclosporin A that is administered to inhibit T cells and prevent GVHD can also induce NK cell expansion and boost NK cell functions^{105, 106}. Finally, recent work has also focused on the role of activating KIR (KIR-S) in transplantation for patients with leukaemia. KIR-S are molecular homologs of the inhibitory KIRs, with shorter cytoplasmic tails that are devoid of ITIMs and with a transmembrane domain that associates with the ITAM-bearing transducing polypeptide DAP12¹⁰⁷⁻¹⁰⁹. KIR-S are potent NK cell activators and can also be expressed by certain rare T cell subsets. The genes encoding KIR-S are absent in approximately 25% of Caucasians who are homozygous for the so-called KIR gene group A. Investigations into KIR-S genetics, showed that the presence of activating KIRs was associated with less leukaemia relapse and cytomegalovirus reactivation and improved survival in some patients ^{110–115}. Thus, several years after the clinical proof of concept was brought by a pivotal study ¹⁰⁰, the question remains of how NK cell alloreactivity can be fully exploited to produce clinical benefits in haplo-identical HSCT. In addition, these protocols of haploidentical HSCT remain used only for a minor subset of patients with poor prognosis malignancies, because the protocols are associated with a high transplant-related mortality due to profound and durable immunosuppression.

HLA-identical HSCT is still the most frequently used and continuously expanding approach in patients with cancer and has highly benefited from the reduced toxicity associated with the new approaches for conditioning regimen. Following the initial description of the first clinical success ⁹², allo-HSCT procedures were carried out with basically unmodified protocols for 20 to 25 years. It begins with a preparative phase, so called conditioning. This phase is followed by allogeneic graft cell infusion and a post infusion drug

immunosuppression in attempt to limit unwanted GVH reactions. The goals of conditioning are to treat residual tumour disease and to allow for donor engraftment through a profound immunosuppression of the host. Both goals have been carried out rather successfully by the association of high dose total body irradiation with chemotherapy or by the association of highly myeloablative anti-neoplasic drugs. However, the cytotoxicity of the conditioning generates pro-inflammatory cytokines which represent the starting point of the previously described negative effects ¹¹⁶. Later, this pro-inflammatory effect was integrated in a more complex cytokine-storm hypothesis. Indeed, these cytokines when reaching blood stream get together with the circulating immune cells of the infused graft, turning on their immune capacities and eventually generating GVH effects. This now classical scenario suggested that reducing cytokine generation could afford less toxicity. Some years later, this hypothesis was convincingly verified by the combination of distinct potent immunosuppressive agents, including notably a purine analogue compound, to carry out allogeneic engraftment, leaving the anti-tumour action of allo-HSCT mainly to the development of GVL or GVT effect ^{117–119}. Since this time initial findings have been widely confirmed and the so-called reduced intensity conditionings (RIC) are now broadly used. Indeed the dramatic diminution of procedure related mortality¹²⁰ has conducted to propose this treatment to a wider population, including patients with other diagnoses than leukaemia - lymphoid malignancies and solid tumours - and patients above the classical age of 50 years, generally not considered for transplantation because of the increased risk of fatal complication. This has conducted to a nearly doubling of the number of transplants worldwide in less than 10 years ¹²¹. Altogether these achievements justify the present attempts to further improve disease control after HLA-identical HSCT and in particular, approaches that manipulate NK cells. Indeed donor NK cells are among the first cells to arise in the recipient following allo-HSCT ¹²², with a rapid NK cell reconstitution evident as early as 1.5 months post-RIC HSCT ^{123, 124}. Various studies have shown an association between higher numbers of NK cells in the graft and lower rates of relapse ¹¹⁵. High NK cell counts on day 30 post allo-HSCT have also been associated with improved clinical outcomes^{125, 126}. More recently, an association between high NK cell counts at day 60 and reduced relapse after RIC HSCT was reported ¹²⁷. This effect was not documented after standard conditioning regimens. Altogether, these data demonstrate that NK cells are readily present at early times following HSCT in RIC and can exert their functions even during immunosuppressive treatment.

Donor lymphocyte infusions

Nowadays, allo-HSCT anti-tumour activity can be reinforced through single or sequential reinfusions of donor-derived immune competent cells once hematopoietic chimerism is established. Initially, in the mid 90's, donor lymphocyte infusions (DLIs) were shown to result in a high incidence of durable cytogenetic and molecular remissions when used as a treatment for chronic myeloid leukaemia (CML) that relapsed after conventional allo-HSCT ^{128, 129}. However, despite these positive effects, significant side-effects occurred, including GVHD and secondary aplasia, as a consequence of the high numbers of cytotoxic T cells in the infusions ¹³⁰. Attempts to deplete CD8⁺ T cells from DLIs produced encouraging, although incomplete results ^{131, 132}. The interest of lymphocyte-driven immunotherapy led to the infusion of allogeneic lymphocytes being explored as a therapy for cancer outside the context of allogeneic HSCT ¹³³. Short-term lymphocyte survival was

noted and this prevented long-term cancer control although clinical responses were documented in some patients. In line with these initial data, a recent study infusing HLAmismatched peripheral blood stem cells established the outcome improvement of chemotherapy for acute myeloid leukaemia in elderly patients ¹³⁴. Whether the use of appropriately selected and activated donor-derived NK cells instead of regular DLIs will better support an antitumour effect remains to be shown, but this warrants further study. Several protocols of clinical grade NK cell purification and *in vitro* expansion are now validated, and studies have shown that the infusion of allogeneic NK cells is safe in humans ^{135–149} (Figure 2b). Injections of mature HLA mismatched NK cells are also well-tolerated ¹⁵⁰. Therefore there are encouraging signs that NK cell infusion could be a useful antitumour strategy

KIR-specific monoclonal antibodies

As already mentioned, the use of allo-HSCT remains restricted to minor subsets of patients affected with poor prognosis malignancies, mainly subsets of patients presenting with acute leukaemia associated with poor-prognosis criteria (mostly cytogenetic), relapse, patients with myelodysplastic syndromes, as well as patients with lymphoid malignancies who failed or relapsed after initial therapy. Restrictions to its use include the difficulty in identifying a suitable donor as well as the physical condition of the recipient and his/her predicted ability to sustain the morbidity associated with the transplantation; as a consequence allo-HSCT can rarely be offered to elderly people, populations in which the incidence of haematological malignancies is increasing. In vivo, activation of NK cells is another avenue for medical progress, with potential applications in larger populations of patients. Fully humanized KIRspecific monoclonal antibodies have been generated to achieve such progress ^{151, 152} (Figure 2c). By blocking the interaction between all inhibitory KIR molecules that recognize HLA-C alleles, KIR-specific monoclonal antibodies are expected to boost the reactivity of NK cells against tumour cells expressing ligands for activating receptors, without inducing autoimmunity against normal cells, which do not express sufficient density of activating ligands ¹⁵³ (Figure 2c). However, the recognition of MHC class I molecules by KIRs is crucial for NK cell education. The blocking of KIRs by monoclonal antibodies may have more complex consequences than simply triggering tumour elimination. To fully evaluate this issue *in vivo*, a 'humanized' preclinical mouse model has been developed in which all NK cells are educated by a transgenic inhibitory receptor, human KIR2DL3, through the engagement with its HLA-Cw3 ligand. This approach revealed that NK cells could be reprogrammed to kill HLA-Cw3+ target cells without compromising self-tolerance and without abolishing NK cell education ¹⁵⁴. Following pre-clinical evaluation, 100 patients with AML or MM were treated in phase I or II clinical studies (N. Vey et al., in preparation). These studies showed that the infusion of KIR-specific monoclonal antibodies was safe, even in mostly elderly patients who had been heavily pre-treated by chemotherapy. Although the results of efficacy are awaited, the availability of KIR-specific monoclonal antibodies thus paves the way for the design of innovative NK cell-based antitumour therapies. For instance, new protocol might include the combination of KIR-specific monoclonal antibodies with HLA-identical HSCT, with the infusion of HLA-identical NK cells, with other monoclonal antibodies (for example, CD20-specific or HER2/Neu-specific

monoclonal antibodies), or with drugs that induce the expression of ligands for activating NK cell receptors, such as lenalinomide ¹⁵⁵.

In conclusion, the efficacy of NK cell-based antitumour therapies still remains to be firmly established. Question remain concerning the sensitivity of tumour cells to NK cell attack, on the migratory properties of endogenous ¹⁵⁶ and infused NK cells ¹⁵⁷, as well as on the survival and homeostatic proliferative capacity of donor NK cells in cancer patients in conditions of chemotherapy and/or HSCT ¹⁵⁸. Many clinical trials have been initiated which hopefully will answer some of these key issues (Box 2).

Roles of NKT cells in cancer

In earlier studies, NK and NKT cell functions in antitumour immune responses have been sometimes confounded. Recent progress in the characterization of iNKT cells allowed for a better understanding of their functions in response to tumours. We review below how iNKT cells are believed to recognize tumour cells and how these findings have led to current strategies to target these cells for tumour therapy.

How do NKT cells recognize tumour cells?

Activation of iNKT cells with strong agonists such as α -GalCer leads to strong antitumour responses in mice (Figure 3). However, in this case, the iNKT cell contribution to the antitumour response is indirect and mediated in part by downstream effectors such as NK cells (Figure 3) and by the cytokine IFN- γ , rather than directly targeting cancer cells by cytotoxic mechanisms. iNKT cells can directly respond to IL-12 in the B16 melanoma model and produce IFN- γ , yet iNKT cells are dispensable in the B16 melanoma model ¹⁵⁹. However, their contribution in the absence of NK cells using NK cell deficient mice has not been addressed in this model. A direct recognition of CD1d-expressing tumour cells by iNKT cells has been demonstrated in vitro (Figure 3a). CD1d is expressed on some myelomonocytic leukaemia cells and it has been shown that these cells are sensitive to lysis by human NKT cells ¹⁶⁰. Similarly, using a CD1d-transfected mouse B-cell lymphoma, it was shown that iNKT cells protect against tumour progression in a CD1d-dependent manner ¹⁶¹. However, it is unclear how iNKT cells distinguish CD1d expression on malignant cells from CD1d expression on normal cells. It is possible that a different set of self ligands is presented by CD1d in transformed cells, yet the evidence for this are lacking. Although CD1d expression has been demonstrated on human malignant hematopoietic cells, the majority of solid tumours and cell line models do not express CD1d or poorly express CD1d suggesting an indirect mechanism or cross-presentation to iNKT cells (Figure 3b) ¹⁶².

Role of iNKT cells in anti-tumour immuno-surveillance

There are strong evidences for a role of iNKT cells in tumour immunosurveillance at least in mice. A protective role for iNKT cells in tumour immunosurveillance has been demonstrated in various tumour models, including MCA-induced fibrosarcomas, P53 loss and in the TRAMP prostate cancer model ^{36–39}. These studies were performed in the absence of exogenous stimuli and by comparing Ja18-deficient and CD1d-deficient mice to wild-type

mice suggesting that iNKT cells were critical while type II NKT cells were dispensable. Adoptive transfer of CD4/CD8 negative iNKT cells from the liver suggest a crucial role for this subset of iNKT cells in promoting antitumour immune responses. iNKT cell IFN- γ production has been shown to be critical at least against MCA-induced sarcomas ¹⁶³ and most likely promotes NK cell activation. Overall, these studies in mice suggest that iNKT cells have an active role in tumour immunosurveillance and their absence predisposes to cancer development. However, it is not entirely clear how iNKT cells become activated in these models and which CD1d-expressing cells activate them. In addition, the contribution of endogenous lipids and inflammatory cytokines is unknown. In fact, it has been proposed that iNKT cells do not target directly the tumour but instead control CD1d-expressing tumour-associated macrophages, preventing these cells from promoting angiogenesis ¹⁶⁴ (Figure 3c). In humans, observational studies have focused mostly on iNKT cells. Overall, iNKT frequency is decreased in solid tumours including in melanoma, colon, lung, breast and head and neck squamous cell carcinoma while an increased iNKT cell number is associated with a better prognosis ^{165–167}. Therefore these observational studies in human are consistent with the mouse studies and suggest that iNKT cells may play a role in tumour immunosurveillance. Interestingly, type II NKT cells have been shown to suppress tumour immunosurveillance provided by iNKT cells potentially explaining the paradox in the role of CD1d restricted T cells in the regulation of tumour immunity ¹⁶⁸.

Manipulating iNKT cells for tumour therapy

Initial studies clearly show the antitumour effects of soluble α -GalCer in mice ^{169, 170}. In mice, α-GalCer induces a rapid iNKT cell activation that leads to downstream activation of NK cells. The cascade of activation initiated by α-GalCer-activated iNKT cells extends to cells of the adaptive immune system, although this is delayed in comparison to the rapidity at which iNKT cells promote NK cell activation. However, a-GalCer administration can also induce IL-4 production and in some cases anergy, as they become unresponsive to subsequent activation ^{171, 172}. To circumvent these issues, other delivery methods have been examined including adoptive transfer of a-GalCer pulsed monocyte-derived dendritic cells, which induce a more potent anti-tumour effect than treatment with soluble α -GalCer alone (Figure 4) due to the dendritic cell superior ability to present antigens and expression of costimulatory molecules at their cell surface. A significant amount of work has been undertaken to design glycolipids that will induce a stronger Th1-type response and target a specific subset of iNKT cells known to be biased toward Th1 responses at least in human¹⁷³. Although polarization of the iNKT cell response has been observed using a-GalCer analogues, the mechanism leading to this polarization is still under intense investigation $^{174-176}$. Regardless of the mechanism(s), it was shown that although α -GalCer analogues that are associated with either a Th1-type or a Th2-type immune response can induce the predicted systemic cytokine bias in mice (production of IFN- γ or IL-4. respectively), the immediate iNKT cell response is not polarized ¹⁷⁷. Two groups targeted murine CD1d itself and showed that injection of a CD1d-specific monoclonal antibody has the capacity to directly mature antigen-presenting cells, leading to inflammatory cytokine secretion and tumour growth prevention ^{178, 179}. However, the CD1d mediated intracellular signalling pathway has not been defined and it is not known whether human CD1d can

perform similar functions to mouse CD1d. Notably, a recent study showed that blocking antibodies specific for CD1d had opposite effects and increased tumour metastasis ¹⁸⁰. Targeting specific CD1d-expressing cell subsets has been attempted using antigen-a-GalCer conjugate particles preferentially adapted for B cells or CD169 positive macrophages ^{181, 182}. Another strategy was to expand autologous iNKT cells *in vitro* in order to compensate for the decreased iNKT cell frequency observed in cancer patients ¹⁸³¹⁸⁴. Clinical trials using a-GalCer or a-GalCer-pulsed antigen presenting cells showed that these treatments were relatively safe and well tolerated. Soluble administration of a-GalCer did not result in significant clinical benefits ^{185, 186}, although injection of a-GalCer-loaded immature dendritic cells led to modest iNKT cell activation in vivo 187, 188. However, when mature monocyte-derived dendritic cells were used, iNKT cell population expansion was observed and this led to an increase in serum levels of IL-12 and IFN- γ ¹⁸⁹. Interestingly, the combination of iNKT cell transfer and α -GalCer-pulsed Dendritic cells has been reported to induce significant antitumour immunity in patients with head and neck squamous cell carcinomas ^{184, 190} (Figure 4). Overall, there are strong evidences for iNKT cell roles in tumour immunosurveillance and anti-tumour potential of ligand activated iNKT cells. Additional studies to optimize glycolipid delivery and to target specific iNKT cell subsets warrant future investigations.

Perspectives on NK and NKT cells anti-tumour properties

Recent studies indicate that NK cell activation could lead to the generation of 'memory' NK cells, a feature only ascribed to B and T cells so far, and recognized as the hallmark of the adaptive immune system. In the model of MCMV infection, memory-like NK cells present increased reactivity not only to molecules that activate their MCVM receptor Ly49H but also to those that activate NK1.1 ⁴⁴. Similarly cytokine (IL-12/IL-18)-pre-activated NK cells respond better to NK1.1 or Ly49H stimulation ¹⁹¹. In addition, hapten- and virus-specific memory NK cells have been described ^{192, 193}. As a consequence of the expression of activating receptors involved in tumour elimination on most if not all NK cells (e.g. NKp46, NKp30, NKG2D), NK cells that have experienced a non-tumour driven activation (e.g. infection, adjuvants) might display broad cross-reactivity and thus exert better anti-tumour function as shown many years ago ¹⁹⁴. The capacity to produce primed/memory NK cells to fight against cancer, as well as the possibility to drive the expansion of tumour-specific memory NK cells thus represents an attractive avenue to explore.

 α -GalCer has been tested as a potential adjuvant due to its ability to induce the activation of a variety of immune cells, including NK cells, in both mice and humans ^{195–202}. These and other findings led to the development of α -GalCer analogs and strategies to better exploit iNKT properties in the design of adjuvants or vaccines ^{203–206}. Therefore, anti tumour therapy taking advantage of the adjuvant potential of iNKT cell ligands and effector functions of both NK and iNKT cells warrant future investigations.

Finally, detrimental roles of NK cells have recently emerged in conditions of inflammation, where NK cells can aggravate sepsis conditions ²⁰⁷, in conditions of auto-immunity as they might contribute to the onset of pathology and to tissue damage ²⁰⁸, as well as in conditions of microbial infections where they can dampen subsequent T cell responses ^{209, 210}. As

inflammation is now recognized as a key element in tumour development ²¹¹ and antitumour T cell functions have been shown to increase patient survival ²¹², these issues should be considered for the establishment of robust immunotherapy protocols. It is also mandatory to improve blood and tissue immune-monitoring to explore in large cohorts of patient the role of NK and NKT cells during cancer.

The knowledge of antitumour immune control has recently rapidly progressed, and a new vision of immunotherapy has emerged from new concepts, medical strategies, medications and medical devices. It is likely that in the coming years the reciprocal movement from bench to bed and from bed to bench will continue to accelerate feeding both scientific knowledge and innovative treatments meeting medical needs. In this perspective, NK/NKT cells undoubtedly represent very exciting players of antitumour immunotherapy.

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Glossary terms

Toll-like receptors (TLRs)

A family of evolutionarily conserved pattern-recognition receptors. These molecules are located intracellularly and at the cell surface of macrophages, dendritic cells, B cells and intestinal epithelial cells. Their natural ligands are molecules that are found in bacteria, viruses and fungi.

Innate lymphoid cells (ILCs)

A group of cells of lymphoid origin that includes NK cells, LTis and other non-T, non-B cells that produce distinct cytokines such as IL-5, IL-13 or IL-17.

Natural killer cells (NK cells)

Non-T, non-B lymphocytes and can mediate natural killing against prototypical NK-cellsensitive targets — K562 (human) and YAC-1 (mouse) and/or produce IFN- γ . In humans, NK cells are typically NKp46⁺CD56⁺CD3⁻, and they are NK1.1⁺CD3⁻ in the C57BL/6 mouse strain and generally NKp46⁺CD3⁻ in all mouse strains.

imatinib mesylate (Gleevec®)

First of its class Tyrosine Kinase Inhibitor (TKI), with clinical activity against Chronic Myeloid Leukaemia (CML) associated with the t(9;22) reciprocal translocation. Introduction of imatinib mesylate in clinical practice at the end of the 20th century, induced a rapid shift in medical practices, leading to abandon alloHSCT for what previously was a standard indication.

CD1d

A MHC-like molecule that associates with β 2-microglobulin and present lipids.

T-bet

A member of the T-box family of transcription factors. It is a master switch in the development of T helper 1 (T_H 1)-cell responses, through its ability to regulate expression of the interleukin-12 receptor, inhibit signals that promote T_H 2-cell development and promote the production of interferon- γ .

perforin/granzyme-mediated mechanisms

Granzymes are serine proteinases that are found primarily in the cytoplasmic granules of cytotoxic T lymphocytes and natural killer cells. They enter target cells through perform pores, then cleave and activate intracellular caspases and lead to target-cell apoptosis.

CD95/CD178 pathway

CD178 (also known as FAS ligand) binds to CD95 (also known as FAS). This results in the formation of the death-inducing signalling complex and subsequent activation of caspases, which leads to apoptosis in the CD95-expressing target cell.

Immunoreceptor tyrosine-based inhibitory motif (ITIM)

This motif is present in the cytoplasmic domain of several inhibitory receptors. After ligand binding, ITIMs are tyrosine phosphorylated and recruit lipid- or tyrosine-phosphatases.

Immunoreceptor tyrosine-based activation motif (ITAM)

Activating receptors often have ITAMs consisting of a consensus amino-acid sequence with paired tyrosines and leucines ($YxxI/Lx_{(6-12)}YxxI/L$). These are normally located in the cytoplasmic domains of ligand-binding transmembrane receptors (such as FceRI and TCR) and they mediate interaction between the transmembrane receptor complex and protein tyrosine kinase activity, which is required to initiate early and late signalling events.

DNA damage response (DDR)

A cell response triggered by DNA damage, such as single or double strand breaks. DDR stops cell cycle progression to enable repair before the damage is transmitted to progeny cells. Checkpoints in the mammalian DDR are the phosphatidylinositol 3-kinases ATM and ATR.

Antibody-dependent cellular cytotoxicity (ADCC)

The capacity of NK cells to lyse antibody-coated target cells via their Fc receptor, CD16.

Rituximab

A chimeric monoclonal antibody that is specific for the CD20 molecule, which is primarily expressed by B cells. Rituximab is the most frequently used antibody therapy for patients with cancer

Graft-versus-leukaemia (GVL) effect

The anti-tumour activity of donor T cells against residual leukaemic cells of the graft recipient following (allogeneic) bone marrow transplantation.

Graft-versus-host disease (GVHD)

Tissue damage in a recipient of allogeneic transplanted tissue (usually a bone-marrow transplant) that results from the activity of donor CTLs that recognize the recipient's tissue

as foreign. GVHD varies markedly in severity, but can be life threatening in severe cases. Typically, damage to the skin and gut mucosa leads to clinical manifestations.

Reduced-intensity conditioning (RIC) regimen

A regimen that uses less chemotherapy and radiation than that normally used for myeloablation.

cyclosporin A

A commonly used immunosuppressive drug that blocks calcineurin A and thereby inhibits T cell activation. It is used to prevent the rejection of transplanted organs and to treat some inflammatory diseases.

References

- Janeway CA Jr, Medzhitov R. Innate immune recognition. Annu Rev Immunol. 2002; 20:197–216. [PubMed: 11861602]
- 2. Beutler B. Innate immunity: an overview. Mol Immunol. 2004; 40:845–59. [PubMed: 14698223]
- Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. Nat Immunol. 2001; 2:675–80. [PubMed: 11477402]
- 4. Spits H, Di Santo JP. The expanding family of innate lymphoid cells: regulators and effectors of immunity and tissue remodeling. Nat Immunol. 2011; 12:21–7. [PubMed: 21113163]
- 5. Vivier E, et al. Innate or adaptive immunity? The example of natural killer cells. Science. 2011; 331:44–9. [PubMed: 21212348]
- Oldham RK, Herberman RB. Evaluation of cell-mediated cytotoxic reactivity against tumor associated antigens with 125I-iododeoxyuridine labeled target cells. J Immunol. 1973; 111:862–71. [PubMed: 4750872]
- Herberman RB, Nunn ME, Lavrin DH. Natural cytotoxic reactivity of mouse lymphoid cells against syngeneic acid allogeneic tumors. I. Distribution of reactivity and specificity. Int J Cancer. 1975; 16:216–29. [PubMed: 50294]
- Kiessling R, Klein E, Wigzell H. "Natural" killer cells in the mouse. I. Cytotoxic cells with specificity for mouse Moloney leukemia cells. Specificity and distribution according to genotype. Eur J Immunol. 1975; 5:112–117. [PubMed: 1234049]
- 9. Smyth MJ, Hayakawa Y, Takeda K, Yagita H. New aspects of natural-killer-cell surveillance and therapy of cancer. Nat Rev Cancer. 2002; 2:850–61. [PubMed: 12415255]
- Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. Annu Rev Immunol. 2011; 29:235–71. [PubMed: 21219185]
- Orange JS, Ballas ZK. Natural killer cells in human health and disease. Clin Immunol. 2006; 118:1–10. [PubMed: 16337194]
- Imai K, Matsuyama S, Miyake S, Suga K, Nakachi K. Natural cytotoxic activity of peripheralblood lymphocytes and cancer incidence: an 11-year follow-up study of a general population. Lancet. 2000; 356:1795–9. [PubMed: 11117911]
- Carrega P, et al. Natural killer cells infiltrating human nonsmall-cell lung cancer are enriched in CD56 bright CD16(-) cells and display an impaired capability to kill tumor cells. Cancer. 2008; 112:863–75. [PubMed: 18203207]
- Platonova S, et al. Profound coordinated alterations of intratumoral NK cell phenotype and function in lung carcinoma. Cancer Res. 2011; 71:5412–22. [PubMed: 21708957]
- Eckl J, et al. Transcript signature predicts tissue NK cell content and defines renal cell carcinoma subgroups independent of TNM staging. J Mol Med (Berl). 2011
- Halama N, et al. Natural killer cells are scarce in colorectal carcinoma tissue despite high levels of chemokines and cytokines. Clin Cancer Res. 2011; 17:678–89. [PubMed: 21325295]

- Menard C, et al. Natural killer cell IFN-gamma levels predict long-term survival with imatinib mesylate therapy in gastrointestinal stromal tumor-bearing patients. Cancer Res. 2009; 69:3563–9. [PubMed: 19351841]
- Kronenberg M, Gapin L. The unconventional lifestyle of NKT cells. Nat Rev Immunol. 2002; 2:557–568. [PubMed: 12154375]
- Tilloy F, et al. An invariant T cell receptor alpha chain defines a novel TAP-independent major histocompatibility complex class Ib-restricted alpha/beta T cell subpopulation in mammals. J Exp Med. 1999; 189:1907–21. [PubMed: 10377186]
- Benlagha K, Kyin T, Beavis A, Teyton L, Bendelac A. A thymic precursor to the NK T cell lineage. Science. 2002; 296:553–5. [PubMed: 11968185]
- 21. Gapin L, Matsuda JL, Surh CD, Kronenberg M. NKT cells derive from double-positive thymocytes that are positively selected by CD1d. Nat Immunol. 2001; 2:971–8. [PubMed: 11550008]
- 22. Egawa T, et al. Genetic evidence supporting selection of the Valpha14i NKT cell lineage from double-positive thymocyte precursors. Immunity. 2005; 22:705–16. [PubMed: 15963785]
- Lantz O, Bendelac A. An invariant T cell receptor alpha chain is used by a unique subset of major histocompatibility complex class I-specific CD4+ and CD4-8- T cells in mice and humans. J Exp Med. 1994; 180:1097–106. [PubMed: 7520467]
- 24. Porcelli S, Yockey CE, Brenner MB, Balk SP. Analysis of T cell antigen receptor (TCR) expression by human peripheral blood CD4-8- alpha/beta T cells demonstrates preferential use of several V beta genes and an invariant TCR alpha chain. J Exp Med. 1993; 178:1–16. [PubMed: 8391057]
- Schumann J, Voyle RB, Wei BY, MacDonald HR. Cutting edge: influence of the TCR V beta domain on the avidity of CD1d:alpha-galactosylceramide binding by invariant V alpha 14 NKT cells. J Immunol. 2003; 170:5815–9. [PubMed: 12794105]
- Wei DG, Curran SA, Savage PB, Teyton L, Bendelac A. Mechanisms imposing the Vbeta bias of Valpha14 natural killer T cells and consequences for microbial glycolipid recognition. J Exp Med. 2006; 203:1197–207. [PubMed: 16651387]
- 27. Sidobre S, et al. The V alpha 14 NKT cell TCR exhibits high-affinity binding to a glycolipid/CD1d complex. J Immunol. 2002; 169:1340–8. [PubMed: 12133957]
- Pellicci DG, et al. A natural killer T (NKT) cell developmental pathway involving a thymusdependent NK1.1(-)CD4(+) CD1d-dependent precursor stage. J Exp Med. 2002; 195:835–44. [PubMed: 11927628]
- Cantu C 3rd, Benlagha K, Savage PB, Bendelac A, Teyton L. The paradox of immune molecular recognition of alpha-galactosylceramide: low affinity, low specificity for CD1d, high affinity for alpha beta TCRs. J Immunol. 2003; 170:4673–82. [PubMed: 12707346]
- 30. Kawano T, et al. CD1d-restricted and TCR-mediated activation of valpha14 NKT cells by glycosylceramides. Science. 1997; 278:1626–1629. [PubMed: 9374463]
- Burdin N, et al. Selective ability of mouse CD1 to present glycolipids: a-galactosylceramide specifically stimulates Va14⁺ NK T lymphocytes. J Immunol. 1998; 161:3271–3281. [PubMed: 9759842]
- Brigl M, Brenner MB. CD1: Antigen Presentation and T Cell Function. Annu Rev Immunol. 2004; 22:817–890. [PubMed: 15032598]
- Kronenberg M. Toward an Understanding of NKT Cell Biology: Progress and Paradoxes. Annu Rev Immunol. 2004; 23:877–900.
- Godfrey DI, Pellicci DG, Smyth MJ. Immunology. The elusive NKT cell antigen--is the search over? Science. 2004; 306:1687–9. [PubMed: 15576595]
- Bendelac A, Savage PB, Teyton L. The Biology of NKT Cells. Annu Rev Immunol. 2006; 25:297– 336.
- 36. Smyth MJ, et al. Differential Tumor Surveillance by Natural Killer (NK) and NKT Cells. Journal Of Experimental Medicine. 2000; 191:661–668. [PubMed: 10684858]
- Swann JB, et al. Type I natural killer T cells suppress tumors caused by p53 loss in mice. Blood. 2009; 113:6382–5. [PubMed: 19234138]
- Nowak M, et al. Defective NKT cell activation by CD1d+ TRAMP prostate tumor cells is corrected by interleukin-12 with alpha-galactosylceramide. PLoS One. 2010; 5:e11311. [PubMed: 20593019]

- Bellone M, et al. iNKT cells control mouse spontaneous carcinoma independently of tumorspecific cytotoxic T cells. PLoS One. 2010; 5:e8646. [PubMed: 20072624]
- 40. Savage AK, et al. The transcription factor PLZF directs the effector program of the NKT cell lineage. Immunity. 2008; 29:391–403. [PubMed: 18703361]
- 41. Kovalovsky D, et al. The BTB-zinc finger transcriptional regulator PLZF controls the development of invariant natural killer T cell effector functions. Nat Immunol. 2008
- 42. Gascoyne DM, et al. The basic leucine zipper transcription factor E4BP4 is essential for natural killer cell development. Nat Immunol. 2009; 10:1118–24. [PubMed: 19749763]
- 43. Kamizono S, et al. Nfil3/E4bp4 is required for the development and maturation of NK cells in vivo. The Journal of Experimental Medicine. 2009; 206:2977–2986. [PubMed: 19995955]
- 44. Sun JC, Beilke JN, Lanier LL. Adaptive immune features of natural killer cells. Nature. 2009; 457:557–561. [PubMed: 19136945]
- Wesley JD, Tessmer MS, Chaukos D, Brossay L. NK cell-like behavior of Valpha14i NK T cells during MCMV infection. PLoS Pathog. 2008; 4:e1000106. [PubMed: 18636102]
- 46. Tyznik AJ, et al. Cutting edge: the mechanism of invariant NKT cell responses to viral danger signals. J Immunol. 2008; 181:4452–6. [PubMed: 18802047]
- 47. Brigl M, et al. Innate and cytokine-driven signals, rather than microbial antigens, dominate in natural killer T cell activation during microbial infection. J Exp Med. 2011; 208:1163–77.
 [PubMed: 21555485]
- 48. Stetson DB, et al. Constitutive Cytokine mRNAs Mark Natural Killer (NK) and NK T Cells Poised for Rapid Effector Function. J Exp Med. 2003; 198:1069–76. [PubMed: 14530376]
- 49. Matsuda JL, et al. Homeostasis of V alpha 14i NKT cells. Nat Immunol. 2002; 3:966–74. [PubMed: 12244311]
- 50. Ranson T, et al. IL-15 availability conditions homeostasis of peripheral natural killer T cells. Proc Natl Acad Sci U S A. 2003; 100:2663–8. [PubMed: 12598649]
- 51. Townsend MJ, et al. T-bet Regulates the Terminal Maturation and Homeostasis of NK and Valpha14i NKT Cells. Immunity. 2004; 20:477–94. [PubMed: 15084276]
- Wingender G, Krebs P, Beutler B, Kronenberg M. Antigen-Specific Cytotoxicity by Invariant NKT Cells In Vivo Is CD95/CD178-Dependent and Is Correlated with Antigenic Potency. The Journal of Immunology. 2010; 185:2721–2729. [PubMed: 20660713]
- Carlyle JR, et al. Evolution of the Ly49 and Nkrp1 recognition systems. Semin Immunol. 2008; 20:321–30. [PubMed: 18595730]
- Parham P. MHC class I molecules and KIRs in human history, health and survival. Nat Rev Immunol. 2005; 5:201–14. [PubMed: 15719024]
- 55. Kim S, et al. Licensing of natural killer cells by host major histocompatibility complex class I molecules. Nature. 2005; 436:709–13. [PubMed: 16079848]
- Fernandez NC, et al. A subset of natural killer cells achieves self-tolerance without expressing inhibitory receptors specific for self-MHC molecules. Blood. 2005; 105:4416–23. [PubMed: 15728129]
- 57. Yokoyama WM, Kim S. How Do Natural Killer Cells Find Self to Achieve Tolerance? Immunity. 2006; 24:249–257. [PubMed: 16546094]
- Raulet DH, Vance RE. Self-tolerance of natural killer cells. Nat Rev Immunol. 2006; 6:520–31. [PubMed: 16799471]
- 59. Brodin P, Karre K, Hoglund P. NK cell education: not an on-off switch but a tunable rheostat. Trends Immunol. 2009; 30:143–149. [PubMed: 19282243]
- 60. Orr MT, Lanier LL. Natural killer cell education and tolerance. Cell. 2010; 142:847–56. [PubMed: 20850008]
- 61. Hoglund P, et al. Recognition of beta 2-microglobulin-negative (beta 2m-) T-cell blasts by natural killer cells from normal but not from beta 2m-mice: nonresponsiveness controlled by beta 2m-bone marrow in chimeric mice. Proc Natl Acad Sci U S A. 1991; 88:10332–6. [PubMed: 1946452]
- 62. Liao NS, Bix M, Zilstra M, Jaenish R, Raulet D. MHC class I deficiency: susceptibility to natural killer (NK) cells and impaired NK activity. Science. 1991; 253:199–202. [PubMed: 1853205]

- 63. Orr MT, Murphy WJ, Lanier LL. 'Unlicensed' natural killer cells dominate the response to cytomegalovirus infection. Nat Immunol. 2010; 11:321–7. [PubMed: 20190757]
- 64. Joncker NT, Fernandez NC, Treiner E, Vivier E, Raulet DH. NK cell responsiveness is tuned commensurate with the number of inhibitory receptors for self-MHC class I: the rheostat model. J Immunol. 2009; 182:4572–80. [PubMed: 19342631]
- Brodin P, Lakshmikanth T, Johansson S, Karre K, Hoglund P. The strength of inhibitory input during education quantitatively tunes the functional responsiveness of individual natural killer cells. Blood. 2009; 113:2434–41. [PubMed: 18974374]
- 66. Guia S, et al. Confinement of activating receptors at the plasma membrane controls natural killer cell tolerance. Sci Signal. 2011; 4:ra21. [PubMed: 21467299]
- Joncker NT, Shifrin N, Delebecque F, Raulet DH. Mature natural killer cells reset their responsiveness when exposed to an altered MHC environment. J Exp Med. 2010; 207:2065–2072. [PubMed: 20819928]
- Elliott JM, Wahle JA, Yokoyama WM. MHC class I-deficient natural killer cells acquire a licensed phenotype after transfer into an MHC class I-sufficient environment. J Exp Med. 2010; 207:2073– 2079. [PubMed: 20819924]
- Kijima M, Gardiol N, Held W. Natural Killer Cell Mediated Missing-Self Recognition Can Protect Mice from Primary Chronic Myeloid Leukemia In Vivo. PLoS ONE. 2011; 6:e27639. [PubMed: 22132120]
- Bottino C, Moretta L, Moretta A. NK cell activating receptors and tumor recognition in humans. Curr Top Microbiol Immunol. 2006; 298:175–82. [PubMed: 16323416]
- Moretta A, et al. Activating receptors and coreceptors involved in human natural killer cellmediated cytolysis. Ann Rev Immunol. 2001; 19:197–223. [PubMed: 11244035]
- Bauer S, et al. Activation of NK cells and T cells by NKG2D, a receptor for stress-inducible MICA. Science. 1999; 285:727–729. [PubMed: 10426993]
- 73. Raulet DH, Guerra N. Oncogenic stress sensed by the immune system: role of natural killer cell receptors. Nat Rev Immunol. 2009; 9:568–80. [PubMed: 19629084]
- 74. Raulet DH. Roles of the NKG2D immunoreceptor and its ligands. Nat Rev Immunol. 2003; 3:781– 90. [PubMed: 14523385]
- Diefenbach A, Jamieson AM, Liu SD, Shastri N, Raulet DH. Ligands for the murine NKG2D receptor: expression by tumor cells and activation of NK cells and macrophages. Nat Immunol. 2000; 1:119–26. [PubMed: 11248803]
- 76. Cerwenka A, et al. Retinoic Acid early Inducible genes define a Ligand family for the Activating NKG2D receptor in Mice. Immunity. 2000; 12:721–727. [PubMed: 10894171]
- Cosman D, et al. ULBPs, Novel MHC Class I-Related Molecules, Bind to CMV Glycoprotein UL16 and Stimulate NK Cytotoxicity through the NKG2D Receptor. Immunity. 2001; 14:123– 133. [PubMed: 11239445]
- Carayannopoulos LN, et al. Ligands for murine NKG2D display heterogenous binding behavior. Eur J Immunol. 2002; 32:597–605. [PubMed: 11857333]
- Guerra N, et al. NKG2D-deficient mice are defective in tumor surveillance in models of spontaneous malignancy. Immunity. 2008; 28:571–80. [PubMed: 18394936]
- Unni AM, Bondar T, Medzhitov R. Intrinsic sensor of oncogenic transformation induces a signal for innate immunosurveillance. Proc Natl Acad Sci U S A. 2008; 105:1686–91. [PubMed: 18223157]
- Gasser S, Orsulic S, Brown EJ, Raulet DH. The DNA damage pathway regulates innate immune system ligands of the NKG2D receptor. Nature. 2005; 436:1186–90. [PubMed: 15995699]
- 82. Soriani A, et al. ATM-ATR dependent up-regulation of DNAM-1 and NKG2D ligands on multiple myeloma cells by therapeutic agents results in enhanced NK cell susceptibility and is associated with a senescent phenotype. Blood. 2008
- 83. Sivori S, et al. p46, a novel natural killer cell-specific surface molecule that mediates cell activation. J Exp Med. 1997; 186:1129–1136. [PubMed: 9314561]
- Vitale M, et al. NKp44, a novel triggering surface molecule specifically expressed by activated natural killer cells, is involved in non-major histocompatibility complex-restricted tumor cell lysis. J ExpMed. 1998; 187:2065–2072.

- Pende D, et al. Identification and Molecular Characterization of NKp30, a Novel Triggering Receptor Involved in Natural Cytotoxicity Mediated by Human Natural Killer Cells. J Exp Med. 1999; 190:1505–1516. [PubMed: 10562324]
- 86. Brandt CS, et al. The B7 family member B7-H6 is a tumor cell ligand for the activating natural killer cell receptor NKp30 in humans. J Exp Med. 2009; 206:1495–503. [PubMed: 19528259]
- 87. Li Y, Wang Q, Mariuzza RA. Structure of the human activating natural cytotoxicity receptor NKp30 bound to its tumor cell ligand B7-H6. J Exp Med. 2011
- Bottino C, et al. Identification of PVR (CD155) and Nectin-2 (CD112) as cell surface ligands for the human DNAM-1 (CD226) activating molecule. J Exp Med. 2003; 198:557–67. [PubMed: 12913096]
- Cartron G, et al. Therapeutic activity of humanized anti-CD20 monoclonal antibody and polymorphism in IgG Fc receptor FcgammaRIIIa gene. Blood. 2002; 99:754–8. [PubMed: 11806974]
- Veeramani S, et al. Rituximab infusion induces NK activation in lymphoma patients with the highaffinity CD16 polymorphism. Blood. 2011; 118:3347–9. [PubMed: 21768303]
- 91. Terme M, Ullrich E, Delahaye NF, Chaput N, Zitvogel L. Natural killer cell-directed therapies: moving from unexpected results to successful strategies. Nat Immunol. 2008; 9:486–94. [PubMed: 18425105]
- Thomas E, et al. Bone Marrow Transplantation. The New England Journal of Medicine. 1975; 292:832–843. 895–902. [PubMed: 234595]
- Weiden PLF, Thomas NED, et al. Antileukemic effect of graft-versus-host disease in human recipients of allogeneic-marrow grafts. New England Journal of Medicine. 1979; 300:1068–1073. [PubMed: 34792]
- 94. Storb R. Allogeneic hematopoietic stem cell transplantation--yesterday, today, and tomorrow. Exp Hematol. 2003; 31:1–10. [PubMed: 12543102]
- 95. Childs R, et al. Regression of metastatic renal-cell carcinoma after nonmyeloablative allogeneic peripheral-blood stem-cell transplantation. N Engl J Med. 2000; 343:750–8. [PubMed: 10984562]
- 96. Blaise D, et al. Reduced-intensity preparative regimen and allogeneic stem cell transplantation for advanced solid tumors. Blood. 2004; 103:435–41. [PubMed: 12946991]
- 97. Ferrara JL, Levine JE, Reddy P, Holler E. Graft-versus-host disease. Lancet. 2009; 373:1550–61. [PubMed: 19282026]
- Dong Z, et al. Essential function for SAP family adaptors in the surveillance of hematopoietic cells by natural killer cells. Nat Immunol. 2009; 10:973–80. [PubMed: 19648922]
- Aversa F, et al. Treatment of high-risk acute leukemia with T-cell-depleted stem cells from related donors with one fully mismatched HLA haplotype. N Engl J Med. 1998; 339:1186–93. [PubMed: 9780338]
- 100. Ruggeri L, et al. Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants. Science. 2002; 295:2097–100. [PubMed: 11896281]
- 101. Giebel S, et al. Survival advantage with KIR ligand incompatibility in hematopoietic stem cell transplantation from unrelated donors. Blood. 2003; 102:814–819. [PubMed: 12689936]
- 102. Ruggeri L, et al. Donor natural killer cell allorecognition of missing self in haploidentical hematopoietic transplantation for acute myeloid leukemia: challenging its predictive value. Blood. 2007; 110:433–40. [PubMed: 17371948]
- 103. Kroger N, et al. Comparison between antithymocyte globulin and alemtuzumab and the possible impact of KIR-ligand mismatch after dose-reduced conditioning and unrelated stem cell transplantation in patients with multiple myeloma. Br J Haematol. 2005; 129:631–43. [PubMed: 15916686]
- 104. Bignon JD, Gagne K. KIR matching in hematopoietic stem cell transplantation. Curr Opin Immunol. 2005; 17:553–9. [PubMed: 16085405]
- 105. Poggi A, Zocchi MR. Cyclosporin A regulates human NK cell apoptosis induced by soluble HLA-I or by target cells. Autoimmun Rev. 2005; 4:532–6. [PubMed: 16214091]
- 106. Wang H, et al. The unexpected effect of cyclosporin A on CD56+CD16 and CD56+CD16+ natural killer cell subpopulations. Blood. 2007; 110:1530–1539. [PubMed: 17495133]

- 107. Moretta A, et al. Existence of both inhibitory (p58) and activatory (p50) receptors for HLA-C molecules in human natural killer cells. J Exp Med. 1995; 182:875–884. [PubMed: 7650491]
- 108. Olcese L, et al. Killer-cell activatory receptors for MHC Class I molecules are included in a multimeric complex expressed by human killer cells. J Immunol. 1997; 158:5083–5086. [PubMed: 9164921]
- 109. Lanier LL, Corliss BC, Wu J, Leong C, Phillips JH. Immunoreceptor DAP12 bearing a tyrosinebased activation motif is involved in activating NK cells. Nature. 1998; 391:703–707. [PubMed: 9490415]
- 110. Verheyden S, Schots R, Duquet W, Demanet C. A defined donor activating natural killer cell receptor genotype protects against leukemic relapse after related HLA-identical hematopoietic stem cell transplantation. Leukemia. 2005; 19:1446–51. [PubMed: 15973456]
- 111. De Santis D, et al. Natural killer cell HLA-C epitopes and killer cell immunoglobulin-like receptors both influence outcome of mismatched unrelated donor bone marrow transplants. Tissue Antigens. 2005; 65:519–28. [PubMed: 15896199]
- 112. Cook M, et al. Donor KIR genotype has a major influence on the rate of cytomegalovirus reactivation following T-cell replete stem cell transplantation. Blood. 2006; 107:1230–2. [PubMed: 16239436]
- 113. Chen C, et al. Activating KIR genes are associated with CMV reactivation and survival after non-T-cell depleted HLA-identical sibling bone marrow transplantation for malignant disorders. Bone Marrow Transplant. 2006; 38:437–44. [PubMed: 16892071]
- 114. McQueen KL, et al. Donor-recipient combinations of group A and B KIR haplotypes and HLA class I ligand affect the outcome of HLA-matched, sibling donor hematopoietic cell transplantation. Hum Immunol. 2007; 68:309–23. [PubMed: 17462498]
- 115. Clausen J, et al. Impact of natural killer cell dose and donor killer-cell immunoglobulin-like receptor (KIR) genotype on outcome following human leucocyte antigen-identical haematopoietic stem cell transplantation. Clin Exp Immunol. 2007; 148:520–8. [PubMed: 17493020]
- 116. Holler E, et al. Increased serum levels of tumor necrosis factor alpha precede major complications of bone marrow transplantation. Blood. 1990; 75:1011–6. [PubMed: 2405918]
- 117. Storb R, et al. Stable mixed hematopoietic chimerism in DLA-identical littermate dogs given sublethal total body irradiation before and pharmacological immunosuppression after marrow transplantation. Blood. 1997; 89:3048–54. [PubMed: 9108426]
- 118. Giralt S, et al. Engrafment of allogeneic hematopoietic progenitor cells ith purine analogcontaining chemotherapy: harnessing graft-versus-leukemia without myeloablative therapy. Blood. 1997; 89:4531–4536. [PubMed: 9192777]
- 119. Slavin S, et al. Nonmyeloablative stem cell transplantation and cell therapy as an alternative to conventional bone marrow transplantation with lethal cytoreduction for the treatment of malignant and nonmalignant hematologic diseases. Blood. 1998; 91:756–763. [PubMed: 9446633]
- Blaise D, Vey N, Faucher C, Mohty M. Current status of reduced-intensity-conditioning allogeneic stem cell transplantation for acute myeloid leukemia. Haematologica. 2007; 92:533– 41. [PubMed: 17488664]
- 121. Gratwohl A, et al. The EBMT activity survey 2007 with focus on allogeneic HSCT for AML and novel cellular therapies. Bone Marrow Transplant. 2009; 43:275–91. [PubMed: 19169283]
- 122. Pende D, et al. Anti-leukemia activity of alloreactive NK cells in KIR ligand-mismatched haploidentical HSCT for pediatric patients: evaluation of the functional role of activating KIR and redefinition of inhibitory KIR specificity. Blood. 2009; 113:3119–29. [PubMed: 18945967]
- 123. Mohty M, et al. Recovery of lymphocyte and dendritic cell subsets following reduced intensity allogeneic bone marrow transplantation. Hematology. 2002; 7:157–64. [PubMed: 12243978]
- 124. Larosa F, et al. Peripheral T-cell expansion and low infection rate after reduced-intensity conditioning and allogeneic blood stem cell transplantation. Bone Marrow Transplant. 2005; 35:859–68. [PubMed: 15765116]
- 125. Kim DH, et al. Non-CD34+ cells, especially CD8+ cytotoxic T cells and CD56+ natural killer cells, rather than CD34 cells, predict early engraftment and better transplantation outcomes in

patients with hematologic malignancies after allogeneic peripheral stem cell transplantation. Biol Blood Marrow Transplant. 2006; 12:719–28. [PubMed: 16785061]

- 126. Savani BN, et al. Rapid natural killer cell recovery determines outcome after T-cell-depleted HLA-identical stem cell transplantation in patients with myeloid leukemias but not with acute lymphoblastic leukemia. Leukemia. 2007; 21:2145–52. [PubMed: 17673900]
- 127. Dunbar EM, et al. The relationship between circulating natural killer cells after reduced intensity conditioning hematopoietic stem cell transplantation and relapse-free survival and graft-versus-host disease. Haematologica. 2008; 93:1852–8. [PubMed: 18945751]
- 128. Kolb HJ, et al. Graft-versus-leukemia effect of donor lymphocyte transfusions in marrow grafted patients. European Group for Blood and Marrow Transplantation Working Party Chronic Leukemia. Blood. 1995; 86:2041–50. [PubMed: 7655033]
- 129. Dazzi F, et al. Durability of responses following donor lymphocyte infusions for patients who relapse after allogeneic stem cell transplantation for chronic myeloid leukemia. Blood. 2000; 96:2712–6. [PubMed: 11023502]
- Marks DI, et al. The toxicity and efficacy of donor lymphocyte infusions given after reducedintensity conditioning allogeneic stem cell transplantation. Blood. 2002; 100:3108–3114. [PubMed: 12384406]
- 131. Shimoni A, et al. Long-Term follow-up of recipients of CD8-depleted donor lymphocyte infusions for the treatment of chronic myelogenous leukemia relapsing after allogeneic progenitor cell transplantation. Biol Blood Marrow Transplant. 2001; 7:568–75. [PubMed: 11760089]
- 132. Alyea EP, et al. CD8+ cell depletion of donor lymphocyte infusions using cd8 monoclonal antibody-coated high-density microparticles (CD8-HDM) after allogeneic hematopoietic stem cell transplantation: a pilot study. Bone Marrow Transplant. 2004; 34:123–8. [PubMed: 15133487]
- 133. Porter DL, et al. Graft-versus-tumor induction with donor leukocyte infusions as primary therapy for patients with malignancies. J Clin Oncol. 1999; 17:1234. [PubMed: 10561184]
- 134. Guo M, et al. Infusion of HLA-mismatched peripheral blood stem cells improves the outcome of chemotherapy for acute myeloid leukemia in elderly patients. Blood. 2011; 117:936–41.
 [PubMed: 20966170]
- 135. Miller JS, et al. Large scale ex vivo expansion and activation of human natural killer cells for autologous therapy. Bone Marrow Transplant. 1994; 14:555–62. [PubMed: 7532064]
- 136. Passweg JR, et al. Purified donor NK-lymphocyte infusion to consolidate engraftment after haploidentical stem cell transplantation. Leukemia. 2004; 18:1835–8. [PubMed: 15457184]
- 137. McKenna DH Jr, et al. Good manufacturing practices production of natural killer cells for immunotherapy: a six-year single-institution experience. Transfusion. 2007; 47:520–8. [PubMed: 17319835]
- Lundqvist A, McCoy JP, Samsel L, Childs R. Reduction of GVHD and enhanced antitumor effects after adoptive infusion of alloreactive Ly49-mismatched NK cells from MHC-matched donors. Blood. 2007; 109:3603–6. [PubMed: 17179231]
- Castriconi R, et al. Human NK cell infusions prolong survival of metastatic human neuroblastoma-bearing NOD/scid mice. Cancer Immunol Immunother. 2007; 56:1733–42. [PubMed: 17426969]
- 140. Shi J, et al. Infusion of haplo-identical killer immunoglobulin-like receptor ligand mismatched NK cells for relapsed myeloma in the setting of autologous stem cell transplantation. Br J Haematol. 2008; 143:641–53. [PubMed: 18950462]
- 141. Arai S, et al. Infusion of the allogeneic cell line NK-92 in patients with advanced renal cell cancer or melanoma: a phase I trial. Cytotherapy. 2008; 10:625–32. [PubMed: 18836917]
- 142. Cho D, Campana D. Expansion and activation of natural killer cells for cancer immunotherapy. Korean J Lab Med. 2009; 29:89–96. [PubMed: 19411773]
- 143. Barkholt L, et al. Safety analysis of ex vivo-expanded NK and NK-like T cells administered to cancer patients: a phase I clinical study. Immunotherapy. 2009; 1:753–64. [PubMed: 20636021]
- 144. Bachanova V, et al. Allogeneic natural killer cells for refractory lymphoma. Cancer Immunol Immunother. 2010; 59:1739–44. [PubMed: 20680271]

- 145. Nguyen S, et al. Infusion of allogeneic natural killer cells in a patient with acute myeloid leukemia in relapse after haploidentical hematopoietic stem cell transplantation. Transfusion. 2011; 51:1769–78. [PubMed: 21332732]
- 146. Geller MA, et al. A phase II study of allogeneic natural killer cell therapy to treat patients with recurrent ovarian and breast cancer. Cytotherapy. 2011; 13:98–107. [PubMed: 20849361]
- 147. De Somer L, et al. Recipient lymphocyte infusion in MHC-matched bone marrow chimeras induces a limited lymphohematopoietic host-versus-graft reactivity but a significant antileukemic effect mediated by CD8+ T cells and natural killer cells. Haematologica. 2011; 96:424–31. [PubMed: 21109687]
- 148. Curti A, et al. Successful transfer of alloreactive haploidentical KIR ligand-mismatched natural killer cells after infusion in elderly high risk acute myeloid leukemia patients. Blood. 2011; 118:3273–9. [PubMed: 21791425]
- 149. Brehm C, et al. IL-2 Stimulated but Not Unstimulated NK Cells Induce Selective Disappearance of Peripheral Blood Cells: Concomitant Results to a Phase I/II Study. PLoS One. 2011; 6:e27351. [PubMed: 22096557]
- 150. Miller JS, et al. Successful adoptive transfer and in vivo expansion of human haploidentical NK cells in patients with cancer. Blood. 2005; 105:3051–7. [PubMed: 15632206]
- 151. Alici E. IPH-2101, a fully human anti-NK-cell inhibitory receptor mAb for the potential treatment of hematological cancers. Curr Opin Mol Ther. 2010; 12:724–33. [PubMed: 21154164]
- 152. Romagne F, et al. Pre-clinical characterization of 1-7F9, a novel human anti-KIR therapeutic antibody that augments NK-mediated killing of tumor cells. Blood. 2009; 114:2667–2677. [PubMed: 19553639]
- 153. Romagne F, Vivier E. Natural killer cell-based therapies. F1000 Med Rep. 2011; 3:9. [PubMed: 21654924]
- 154. Sola C, et al. Genetic and antibody-mediated reprogramming of natural killer cell missing-self recognition in vivo. Proc Natl Acad Sci U S A. 2009; 106:12879–84. [PubMed: 19561305]
- 155. Benson DM Jr, et al. IPH2101, a novel anti-inhibitory KIR antibody, and lenalidomide combine to enhance the natural killer cell versus multiple myeloma effect. Blood. 2011; 118:6387–91. [PubMed: 22031859]
- 156. Walzer T, Vivier E. G-protein-coupled receptors in control of natural killer cell migration. Trends Immunol. 2011; 32:486–92. [PubMed: 21664184]
- 157. Brand JM, et al. Kinetics and organ distribution of allogeneic natural killer lymphocytes transfused into patients suffering from renal cell carcinoma. Stem Cells Dev. 2004; 13:307–14. [PubMed: 15186726]
- 158. Sun JC, Lanier LL. NK cell development, homeostasis and function: parallels with CD8 T cells. Nat Rev Immunol. 2011; 11:645–57. [PubMed: 21869816]
- Park SH, Kyin T, Bendelac A, Carnaud C. The contribution of NKT cells, NK cells, and other gamma-chain-dependent non-T non-B cells to IL-12-mediated rejection of tumors. J Immunol. 2003; 170:1197–201. [PubMed: 12538676]
- 160. Metelitsa LS, Weinberg KI, Emanuel PD, Seeger RC. Expression of CD1d by myelomonocytic leukemias provides a target for cytotoxic NKT cells. Leukemia. 2003; 17:1068–77. [PubMed: 12764370]
- 161. Renukaradhya GJ, et al. Type I NKT cells protect (and type II NKT cells suppress) the host's innate antitumor immune response to a B-cell lymphoma. Blood. 2008; 111:5637–45. [PubMed: 18417738]
- 162. Wu DY, Segal NH, Sidobre S, Kronenberg M, Chapman PB. Cross-presentation of disialoganglioside GD3 to natural killer T cells. J Exp Med. 2003; 198:173–81. [PubMed: 12847141]
- 163. Crowe NY, Smyth MJ, Godfrey DI. A Critical Role for Natural Killer T Cells in Immunosurveillance of Methylcholanthrene-induced Sarcomas. J Exp Med. 2002; 196:119–27. [PubMed: 12093876]
- 164. Song L, et al. Valpha24-invariant NKT cells mediate antitumor activity via killing of tumorassociated macrophages. J Clin Invest. 2009; 119:1524–36. [PubMed: 19411762]

- 165. Tahir SM, et al. Loss of IFN-gamma production by invariant NK T cells in advanced cancer. J Immunol. 2001; 167:4046–50. [PubMed: 11564825]
- 166. Yanagisawa K, et al. Impaired Proliferative Response of Vα24 NKT Cells from Cancer Patients Against α-Galactosylceramide. The Journal of Immunology. 2002; 168:6494–6499. [PubMed: 12055270]
- 167. Tachibana T, et al. Increased intratumor Valpha24-positive natural killer T cells: a prognostic factor for primary colorectal carcinomas. Clin Cancer Res. 2005; 11:7322–7. [PubMed: 16243803]
- 168. Terabe M, et al. A nonclassical non-Valpha14Jalpha18 CD1d-restricted (type II) NKT cell is sufficient for down-regulation of tumor immunosurveillance. J Exp Med. 2005; 202:1627–33. [PubMed: 16365146]
- 169. Kawano T, et al. Natural killer-like nonspecific tumor cell lysis mediated by specific ligandactivated valpha14 NKT cells [In Process Citation]. ProcNatl AcadSciUS A. 1998; 95:5690– 5693.
- 170. Swann JB, Coquet JM, Smyth MJ, Godfrey DI. CD1-restricted T cells and tumor immunity. Curr Top Microbiol Immunol. 2007; 314:293–323. [PubMed: 17593666]
- 171. Parekh VV, et al. Glycolipid antigen induces long-term natural killer T cell anergy in mice. J Clin Invest. 2005; 115:2572–83. [PubMed: 16138194]
- 172. Kim S, et al. Impact of bacteria on the phenotype, functions, and therapeutic activities of invariant NKT cells in mice. J Clin Invest. 2008; 118:2301–15. [PubMed: 18451996]
- 173. Cerundolo V, Barral P, Batista FD. Synthetic iNKT cell-agonists as vaccine adjuvants--finding the balance. Curr Opin Immunol. 2010; 22:417–24. [PubMed: 20471232]
- 174. Stanic AK, et al. Another view of T cell antigen recognition: cooperative engagement of glycolipid antigens by Va14Ja18 natural T(iNKT) cell receptor [corrected]. J Immunol. 2003; 171:4539–51. [PubMed: 14568927]
- 175. Oki S, Tomi C, Yamamura T, Miyake S. Preferential T(h)2 polarization by OCH is supported by incompetent NKT cell induction of CD40L and following production of inflammatory cytokines by bystander cells in vivo. Int Immunol. 2005; 17:1619–29. [PubMed: 16291659]
- 176. Im JS, et al. Kinetics and cellular site of glycolipid loading control the outcome of natural killer T cell activation. Immunity. 2009; 30:888–98. [PubMed: 19538930]
- 177. Sullivan BA, et al. Mechanisms for glycolipid antigen-driven cytokine polarization by Valpha14i NKT cells. J Immunol. 2010; 184:141–53. [PubMed: 19949076]
- 178. Yue SC, et al. Direct CD1d-mediated stimulation of APC IL-12 production and protective immune response to virus infection in vivo. J Immunol. 2010; 184:268–76. [PubMed: 19949077]
- 179. Teng MW, Yue S, Sharkey J, Exley MA, Smyth MJ. CD1d activation and blockade: a new antitumor strategy. J Immunol. 2009; 182:3366–71. [PubMed: 19265113]
- 180. Hix LM, et al. CD1d-expressing breast cancer cells modulate NKT cell-mediated antitumor immunity in a murine model of breast cancer metastasis. PLoS One. 2011; 6:e20702. [PubMed: 21695190]
- 181. Barral P, et al. B cell receptor-mediated uptake of CD1d-restricted antigen augments antibody responses by recruiting invariant NKT cell help in vivo. Proc Natl Acad Sci U S A. 2008; 105:8345–50. [PubMed: 18550831]
- 182. Barral P, et al. CD169(+) macrophages present lipid antigens to mediate early activation of iNKT cells in lymph nodes. Nat Immunol. 2010; 11:303–12. [PubMed: 20228797]
- 183. Motohashi S, et al. A phase I study of in vitro expanded natural killer T cells in patients with advanced and recurrent non-small cell lung cancer. Clin Cancer Res. 2006; 12:6079–86. [PubMed: 17028247]
- 184. Yamasaki K, et al. Induction of NKT cell-specific immune responses in cancer tissues after NKT cell-targeted adoptive immunotherapy. Clin Immunol. 2011; 138:255–65. [PubMed: 21185787]
- 185. Giaccone G, et al. A phase I study of the natural killer T-cell ligand alpha-galactosylceramide (KRN7000) in patients with solid tumors. Clin Cancer Res. 2002; 8:3702–9. [PubMed: 12473579]

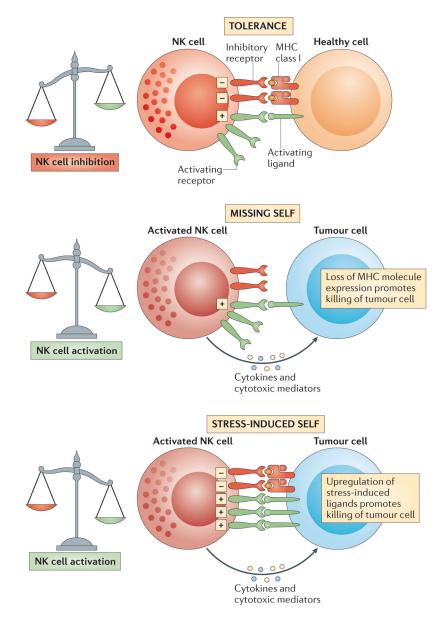
- 186. Schneiders FL, et al. Clinical experience with alpha-galactosylceramide (KRN7000) in patients with advanced cancer and chronic hepatitis B/C infection. Clin Immunol. 2011; 140:130–41. [PubMed: 21169066]
- 187. Nieda M, et al. Therapeutic activation of Valpha24+Vbeta11+ NKT cells in human subjects results in highly coordinated secondary activation of acquired and innate immunity. Blood. 2004; 103:383–9. [PubMed: 14512316]
- 188. Ishikawa A, et al. A phase I study of alpha-galactosylceramide (KRN7000)-pulsed dendritic cells in patients with advanced and recurrent non-small cell lung cancer. Clin Cancer Res. 2005; 11:1910–7. [PubMed: 15756017]
- Chang DH, et al. Sustained expansion of NKT cells and antigen-specific T cells after injection of alpha-galactosyl-ceramide loaded mature dendritic cells in cancer patients. J Exp Med. 2005; 201:1503–17. [PubMed: 15867097]
- 190. Kunii N, et al. Combination therapy of in vitro-expanded natural killer T cells and alphagalactosylceramide-pulsed antigen-presenting cells in patients with recurrent head and neck carcinoma. Cancer Sci. 2009; 100:1092–8. [PubMed: 19302288]
- 191. Cooper MA, et al. Cytokine-induced memory-like natural killer cells. Proc Natl Acad Sci U S A. 2009; 106:1915–9. [PubMed: 19181844]
- 192. Paust S, et al. Critical role for the chemokine receptor CXCR6 in NK cell-mediated antigenspecific memory of haptens and viruses. Nat Immunol. 2010; 11:1127–35. [PubMed: 20972432]
- 193. O'Leary JG, Goodarzi M, Drayton DL, von Andrian UH. T cell- and B cell-independent adaptive immunity mediated by natural killer cells. Nat Immunol. 2006; 7:507–516. [PubMed: 16617337]
- 194. Bukowski JF, Biron CA, Welsh RM. Elevated natural killer cell-mediated cytotoxicity, plasma interferon, and tumor cell rejection in mice persistently infected with lymphocytic choriomeningitis virus. J Immunol. 1983; 131:991–6. [PubMed: 6190947]
- 195. Carnaud C, et al. Cutting edge: cross-talk between cells of the innate immune system: NKT cells rapidly activate NK cells. J Immunol. 1999; 163:4647–4650. [PubMed: 10528160]
- 196. Eberl G, MacDonald HR. Selective induction of NK cell proliferation and cytotoxicity by activated NKT cells. Eur J Immunol. 2000; 30:985–992. [PubMed: 10760785]
- 197. Hermans IF, et al. NKT cells enhance CD4+ and CD8+ T cell responses to soluble antigen in vivo through direct interaction with dendritic cells. J Immunol. 2003; 171:5140–7. [PubMed: 14607913]
- 198. Wesley JD, et al. Cutting edge: IFN-gamma signaling to macrophages is required for optimal Valpha14i NK T/NK cell cross-talk. J Immunol. 2005; 174:3864–8. [PubMed: 15778340]
- 199. Gonzalez-Aseguinolaza G, et al. Natural killer T cell ligand alpha-galactosylceramide enhances protective immunity induced by malaria vaccines. J Exp Med. 2002; 195:617–24. [PubMed: 11877484]
- 200. Huang Y, et al. Enhancement of HIV DNA vaccine immunogenicity by the NKT cell ligand, alpha-galactosylceramide. Vaccine. 2008; 26:1807–16. [PubMed: 18329757]
- 201. Galli G, et al. Invariant NKT cells sustain specific B cell responses and memory. Proc Natl Acad Sci U S A. 2007; 104:3984–9. [PubMed: 17360464]
- 202. Kamijuku. Mechanism of NKT cell activation by intranasal coadministration of alphagalactosylceramide, which can induce cross-protection against influenza viruses. Mucosal Immunology. 2008
- 203. Miyamoto K, Miyake S, Yamamura T. A synthetic glycolipid prevents autoimmune encephalomyelitis by inducing TH2 bias of natural killer T cells. Nature. 2001; 413:531–534. [PubMed: 11586362]
- 204. Oki S, Chiba A, Yamamura T, Miyake S. The clinical implication and molecular mechanism of preferential IL-4 production by modified glycolipid-stimulated NKT cells. J Clin Invest. 2004; 113:1631–40. [PubMed: 15173890]
- 205. Yu KO, et al. Modulation of CD1d-restricted NKT cell responses by using N-acyl variants of alpha-galactosylceramides. Proc Natl Acad Sci U S A. 2005; 102:3383–8. [PubMed: 15722411]
- 206. Schmieg J, Yang G, Franck RW, Tsuji M. Superior protection against malaria and melanoma metastases by a C-glycoside analogue of the natural killer T cell ligand alpha-Galactosylceramide. J Exp Med. 2003; 198:1631–41. [PubMed: 14657217]

- 207. Chiche L, et al. The role of natural killer cells in sepsis. J Biomed Biotechnol. 2011; 2011:986491. [PubMed: 21629707]
- 208. Schleinitz N, Vely F, Harle JR, Vivier E. Natural killer cells in human autoimmune diseases. Immunology. 2010; 131:451–8. [PubMed: 21039469]
- 209. Andrews DM, et al. Innate immunity defines the capacity of antiviral T cells to limit persistent infection. J Exp Med. 2010; 207:1333–43. [PubMed: 20513749]
- 210. Waggoner SN, Cornberg M, Selin LK, Welsh RM. Natural killer cells act as rheostats modulating antiviral T cells. Nature. 2011
- 211. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011; 144:646–74. [PubMed: 21376230]
- 212. Ogino S, Galon J, Fuchs CS, Dranoff G. Cancer immunology-analysis of host and tumor factors for personalized medicine. Nat Rev Clin Oncol. 2011; 8:711–9. [PubMed: 21826083]
- 213. Pellicci DG, et al. Recognition of beta-linked self glycolipids mediated by natural killer T cell antigen receptors. Nat Immunol. 2011; 12:827–33. [PubMed: 21804559]
- 214. Brennan PJ, et al. Invariant natural killer T cells recognize lipid self antigen induced by microbial danger signals. Nat Immunol. 2011; 12:1202–11. [PubMed: 22037601]
- 215. Brigl M, Bry L, Kent SC, Gumperz JE, Brenner MB. Mechanism of CD1d-restricted natural killer T cell activation during microbial infection. Nat Immunol. 2003; 26:26.
- Mattner J, et al. Exogenous and endogenous glycolipid antigens activate NKT cells during microbial infections. Nature. 2005; 434:525–9. [PubMed: 15791258]
- 217. Salio M, et al. Modulation of human natural killer T cell ligands on TLR-mediated antigenpresenting cell activation. Proc Natl Acad Sci U S A. 2007; 104:20490–5. [PubMed: 18077358]
- 218. Paget C, et al. Activation of invariant NKT cells by toll-like receptor 9-stimulated dendritic cells requires type I interferon and charged glycosphingolipids. Immunity. 2007; 27:597–609. [PubMed: 17950005]
- Kärre K, Ljunggren HG, Piontek G, Kiessling R. Selective rejection of H-2-deficient lymphoma variants suggests alternative immune defense strategy. Nature. 1986; 319:675–678. [PubMed: 3951539]
- 220. Ljunggren HG, Karre K. In search of the 'missing self': MHC molecules and NK cell recognition. Immunol Today. 1990; 11:237–44. [PubMed: 2201309]
- 221. Moretta L, et al. Allorecognition by NK cells nonself or no self? Immunol. 1992; 13:300.
- 222. Karlhofer FM, Ribaudo RK, Yokoyama WM. MHC class I alloantigen specificity of Ly-49+ IL-2activated natural killer cells. Nature. 1992; 358:66–70. [PubMed: 1614533]
- 223. Moretta A, et al. Receptors for HLA class-I molecules in human natural killer cells. Annu Rev Immunol. 1996; 14:619–48. [PubMed: 8717527]
- 224. Kärre K. How to recognize a foreign submarine. ImmunolRev. 1997; 155:5-9.
- 225. Braud VM, et al. HLA-E binds to natural killer cell receptors CD94/NKG2A, B and C. Nature. 1998; 391:795–799. [PubMed: 9486650]
- 226. Vance RE, Kraft JR, Altman JD, Jensen PE, Raulet DH. Mouse CD94/NKG2A Is a Natural Killer Cell Receptor for the Nonclassical Major Histocompatibility Complex (MHC) Class I Molecule Qa-1(b). J Exp Med. 1998; 188:1841–1848. [PubMed: 9815261]
- 227. Vilches C, Parham P. KIR: diverse, rapidly evolving receptors of innate and adaptive immunity. Annu Rev Immunol. 2002; 20:217–51. [PubMed: 11861603]
- 228. Olcese L, et al. Human and mouse killer-cell inhibitory receptors recruit PTP1C and PTP1D protein tyrosine phosphatases. J Immunol. 1996; 156:4531–4534. [PubMed: 8648092]
- 229. Burshtyn DN, et al. Recruitment of tyrosine phosphatase HCP by the killer cell inhibitory receptor. Immunity. 1996; 4:77–85. [PubMed: 8574854]
- 230. Vivier E, Nunes JA, Vely F. Natural killer cell signaling pathways. Science. 2004; 306:1517–9. [PubMed: 15567854]
- 231. Kumar V, McNerney ME. A new self: MHC-class-I-independent Natural-killer-cell self-tolerance. Nat Rev Immunol. 2005; 5:363–74. [PubMed: 15841099]

Box 3

NK cell clinical trials

A survey on clinicaltrials.gov database searching for 'clinical trials and NK cells' indicates that more than 200 clinical trials have been registered since 2003. Of these, ≈ 150 are observational studies and ≈ 50 are interventional clinical interventions with NK cells (phase I: 20; Phase I/II: 11 Phase II: 17; Phase III: 3). Various selection and expansion (if any) procedures and clinical situations are represented, demonstrating the current lack of consensus in the field. Most protocols are designed for patients with haematological malignancies; and only 5 (10% of interventional clinical interventions with NK cells) recruit patients with solid tumours. Identically only 7 (14%) are proposed to childhood. Interestingly 28 trials are designed and conducted in the context of allo-HSCT setting (18 after partially compatible transplant); the 22 other protocols evaluate the use of allogeneic (mainly from a mismatched related donor) NK cells outside of the context of regular HSCT. Altogether, if completed they anticipate to recruit a large number of 1863 patients. To date, seven clinical trials have been completed for an initial estimated enrolment of 103 patients and only two of those (NCT00274846 and NCT00354172) have reported results.

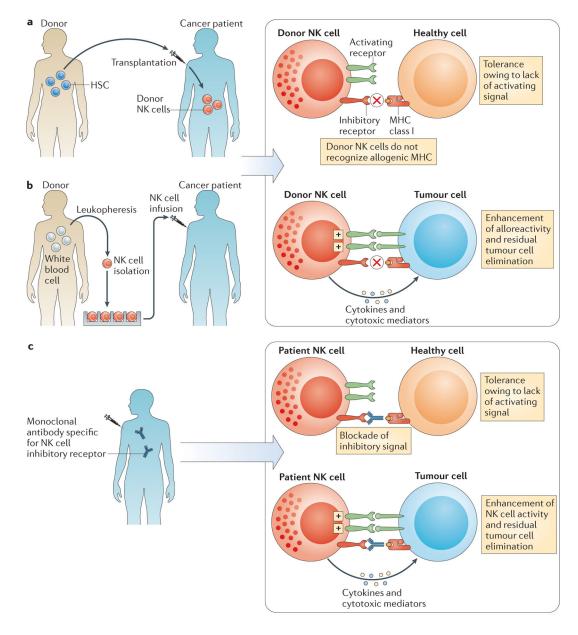


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Figure 1.

NK cell recognition of tumour cells. a | NK cells are tolerant to normal self cells as the 'strength' of activating signals is dampened by engagement of inhibitory receptors. b | NK cells are selectively activated by stressed cells as they express a density of cell surface ligands for activating receptors which overcomes signalling via inhibitory receptors. c | this NK cell activation leads to tumour elimination directly (cytotoxicity) or indirectly (production of cytokines such as IFN- γ).



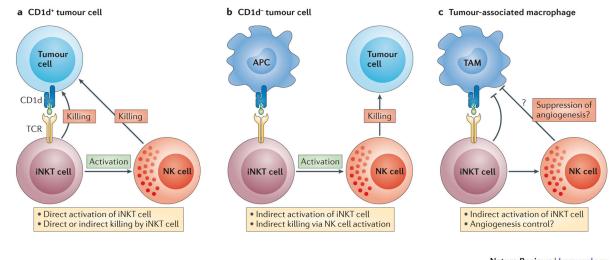


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Figure 2.

NK cellular therapy. a | In haplo-identical or MHC-matched HSCT, NK cells of healthy donor origin will develop into the cancer patient. b | Alternatively, NK cells can be isolated from healthy donors, activated and/or expanded in vitro prior to infusion into the cancer patient. In both cases (allo-HSCT and NK cell infusions), the aim is to promote the anti-tumour function of donor NK cells in the cancer patient; indeed a fraction of donor NK cells will be not be inhibited by the MHC class I molecules of the cancer patient as the KIR expressed by these subsets of NK cells will not interact with by the MHC class I molecules of the cancer patient (ELIMINATION). Most normal cells of the patients will not activate donor NK cells as, in contrast to cancer cells of the patient, they lack the cell surface density

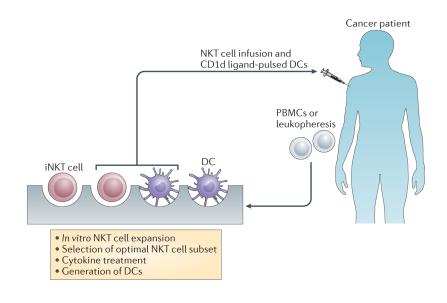
of ligands required to activate donor NK cells (IGNORANCE). c | Anti-KIR monoclonal antibody therapy. Fully human anti-KIR mAbs can be injected in cancer patient. Anti-KIR mAbs are designed to boost the antitumour activity of NK cells (ELIMINATION) without inducing auto-immunity (IGNORANCE).



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Figure 3.

iNKT cell recognition of tumours. a | normal and direct recognition of tumour cells by iNKT cells. b | Indirect activation and indirect killing mediated by iNKT cells. c | Control of angiogenesis by iNKT cells



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Figure 4.

iNKT cellular therapy. Patient Specific iNKT cell subset expanded in vitro prior to infusion together with CD1d ligand pulsed Dendritic cells.