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## Antibacterial activity of some natural products against bacteria expressing a multidrug-resistant phenotype

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## ABSTRACT

The present study assessed the antimicrobial activities of various natural products belonging to the terpenoids, alkaloids and phenolics against a collection of Gram-negative multidrug-resistant (MDR) bacteria. The results demonstrated that most of the compounds were extruded by bacterial efflux pumps. In the presence of the efflux pump inhibitor phenylalanine arginine  $\beta$ -naphthylamide (PA $\beta$ N), the activities of laurentixanthone B (xanthone), plumbagin (naphthoquinone), 4-hydroxy lonchocarpin (flavonoid) and MAB3 (coumarin) increased significantly against all studied MDR bacteria. Laurentixanthone B, 4-hydroxy lonchocarpin and MAB3 contained the same pharmacophoric moiety as plumbagin. This study indicates that the AcrAB–TolC (Enterobacteriaceae) and MexAB–OprM (*Pseudomonas aeruginosa*) efflux pumps are involved in resistance of Gram-negative bacteria to most of the natural products.

## 1. Introduction

Multidrug resistance is a worrying cause of treatment failure in bacterial infections [1–3]. Various widely used antibiotics favour the selection of bacteria resistant to multiple antibacterial agents [4]. The expression of mutated genes, coding for resistance to a single drug (e.g. mutation in a target) or a specific resistance mechanism (e.g. enzymatic barrier), in association with the modulation of multidrug efflux pump or porin expression involved in transport, are the most common processes described in multidrug-resistant (MDR) bacteria [4–6]. A numbers of chemicals, such as phenylalanine arginine  $\beta$ -naphthylamide (PA $\beta$ N) and 1-(1-naphthylmethyl)-piperazine, quinoline derivatives, etc. [7], as well as natural products [8] have been found to inhibit bacterial efflux pumps. In addition, several natural compounds and integral components belonging to the usual pharmaceutical library have been tested to combat resistant bacteria [9,10]. The aim of the present work was to determine the antibacterial activities and potencies of several molecules, including terpenoids, alkaloids and phenolic compounds, against different bacterial strains expressing a MDR phenotype. The role of efflux pumps was investigated using pump-deleted strains and the efflux pump inhibitor (EPI) PA $\beta$ N. Furthermore, this study aimed to determine the pharmacophoric group present in the active compounds.

## 2. Material and methods

### 2.1. Chemicals for antimicrobial assays

Chloramphenicol (Sigma-Aldrich, St-Quentin-Fallavier, France) was used as a reference antibiotic molecule. *p*-Iodonitrotetrazolium chloride (INT) and PA $\beta$ N were

used as a microbial growth indicator and EPI, respectively. Natural products (Fig. 1) were obtained from the chemical bank of the Laboratory of Organic Chemistry, University of Yaoundé I, Cameroon. Isolation and identification of the following compounds have been reported previously [10–13]: crotozambefurans A, B and C; bartericin C; MAB3; mammeisin; norcassaide; norerythrosuaveolide; 1,8-dihydroxy-6-methoxy-3-methylantraquinone; 3-geranyloxy-6-methyl-1,8-dihydroxyanthraquinone; vismiaquinone and vismiaquinone C; bivismiaquinone; laurentixanthone B; kaempferol; lapachol; 2-acetylfuro-1,4-naphthoquinone; newbouldiaquinone and newbouldiaquinone A; hardwickiic acid; 3-friedelanone; betulinic acid; oleanolic acid; 7-methoxycoumarin; 7-hydroxycoumarin; 1,5-dihydroxyxanthone; smeathxanthone A and smeathxanthone B; gancaonin Q; stipulin; angusticornin B; 1,7-dihydroxyxanthone; 4-hydroxylonchocarpin; and quercetin.

## 2.2. Bacterial strains and culture media

MDR isolates and reference strains of *Escherichia coli*, *Enterobacter aerogenes*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* are summarised in Table 1. Isolates were conserved at 4 °C and were grown on Mueller–Hinton agar for 24 h before minimal inhibitory concentration (MIC) testing. Mueller–Hinton broth (MHB) was used for the susceptibility tests [20].

## 2.3. Determination of bacterial susceptibility

The respective MICs of the various compounds against the bacteria were determined using a rapid XTT colorimetric assay [14]. Briefly, samples were first dissolved in dimethyl sulphoxide (DMSO)/MHB. The solution obtained was then added to MHB

and was sequentially diluted two-fold in a 96-well microplate. Then, 100  $\mu\text{L}$  of fresh inoculum ( $1.5 \times 10^6$  colony-forming units/mL) prepared in MHB was added. The plates were covered with a sterile plate sealer, homogenised and incubated at 37 °C for 18 h. The concentration of DMSO used did not affect microbial growth. Wells containing MHB, 100  $\mu\text{L}$  of inoculum and DMSO was used as a negative control (systematically added). Chloramphenicol was used as a reference antibiotic. MICs were measured after 18 h incubation at 37 °C, followed by addition of 40  $\mu\text{L}$  of 0.2 mg/mL INT and incubation at 37 °C for 30 min. Viable bacteria reduced the yellow dye (INT) to pink. The MIC was defined as the lowest concentration of compound that prevented this change and exhibited complete inhibition of microbial growth. Samples were tested alone or in the presence of PA $\beta$ N at a final concentration of 20 mg/L. All assays were carried out in triplicate and were repeated three times.

### 3. Results

#### 3.1. Studied compounds

The different natural compounds investigated in the present study are shown in Fig. 1. The terpenoids included four diterpenoids (hardwickiic acid and crotozambefurans A–C) and three pentacyclic triterpenoids (3-friedelanone, betulinic acid and oleanolic acid). Two alkaloids (norcassaide and norerythrosuaveolide) were tested. Phenolic compounds, including four coumarins, five xanthonenes, nine quinones and seven flavonoids, were also investigated. The compounds were tested for their ability to prevent the growth of MDR and reference strains of Gram-negative bacteria, alone and in the presence of the EPI PA $\beta$ N. The results are summarised in Table 2.



### 3.2. Activity of the terpenoids

Both the diterpenoids and the triterpenoids exhibited very weak activities. Tested alone, none of them was active against any of the tested bacterial strains (Table 2). The presence of EPI conferred only a little activity to some of the compounds against AcrAB- or TolC-deleted strains.

### 3.3. Activity of the alkaloids

Norcassaide and norerythroaveolide showed a moderate and limited activity against the tested bacteria. An MIC value of 64 mg/L was obtained with norcassaide against *E. coli* AG100A, *E. aerogenes* EA294 and EA298 and the reference *P. aeruginosa* PA01 strain (Table 2). A corresponding value for norerythroaveolide of 32 mg/L was noted against *E. coli* AG100A. A significant decrease in MIC values was observed in the presence of the EPI PA $\beta$ N.

### 3.4. Activity of phenolics (coumarins, xanthenes, quinines and flavonoids)

Most of the coumarins presented a reduced and limited activity when tested alone. The best activity (MIC = 32 mg/L) was obtained with MAB3 against *E. coli* AG100A. Most of the xanthenes were poorly active. Laurentixanthone B showed significant activity, with its antibacterial effects being observed on all investigated bacteria including MDR strains. The quinone plumbagin, the two chalcones (flavonoids) 4-hydroxylonchocarpin and bartericin C, and MAB3 (coumarin) were found to be active against bacteria including MDR strains. MIC values <10 mg/L were obtained for plumbagin against *E. coli* AG100A and AG100A<sub>Tet</sub>. The activity of this compound was better than that of chloramphenicol (MIC = 32 mg/L) against AG100A<sub>Tet</sub>. 4-

Hydroxylonchocarpin and bartericin C showed interesting antimicrobial properties, but the two compounds had an MIC > 10 mg/L against the bacteria studied in this work.

### 3.5. Role of efflux pumps in the susceptibility of tested bacteria: identification of PA $\beta$ N-sensitive efflux in addition to the AcrAB–TolC pump

To assess the involvement of efflux in the activity of the tested compounds, the compounds were combined with the well known EPI PA $\beta$ N. The results are summarised in parentheses in Table 2. When tested alone none of the terpenoids were active against *K. pneumoniae* or *P. aeruginosa* (Table 2), whereas a reduced and limited activity was noted with crotozambefuran B and C against *E. aerogenes* EA294 and EA298. Despite this moderate activity, it is important to note that an antibacterial effect of the tested terpenoids was detected against the efflux-deleted strain *E. coli* AG100A, except for crotozambefuran A. However, the enhancing effect of the EPI was limited during combination with terpenoids. PA $\beta$ N significantly increased the activity of most of the tested phenolic compounds. For the coumarins, MICs < 10 mg/L were recorded with MAB3 against four *E. coli* strains and *E. aerogenes* EA298. Interestingly, PA $\beta$ N increased the activity of laurentixanthone B, and MIC values < 10 mg/L were noted for two *E. coli* strains (Table 2). Amongst the quinones, plumbagin (naphthoquinone), which was active against all studied bacteria when tested alone, showed a significant increase in activity in the presence of the EPI. MICs < 10 mg/L were obtained with studied bacteria including MDR and reference strains. Furthermore, this activity was quite similar to or better than that of chloramphenicol in several cases. It should also be noted that one of the most active flavonoids (4-hydroxylonchocarpin) also exhibited MIC values < 10 mg/L against

several MDR bacteria (Table 2). It is important to note the effect of the EPI on the susceptibility of *E. coli* and *E. aerogenes* strains deleted of AcrAB and TolC components (Table 2). This suggests the involvement of PA $\beta$ N-sensitive efflux in addition to the AcrAB–TolC efflux pump. When tested in the presence of the EPI, compounds were active in ca. 46% (94/204), 54% (37/68), 64% (65/102) and 75% (127/170) of the cases against *E. aerogenes*, *P. aeruginosa*, *K. pneumoniae* and *E. coli*, respectively, whilst they were only active in 29%, 41%, 48% and 46% of the cases when tested alone.

### 3.6. Structure–activity relationship study

When exploring the structure–activity relationship, it can be noted that terpenoids (both diterpenoids and triterpenoids) are poor antimicrobial compounds. A careful examination of the activities of the two tested alkaloids (norcassaide and norerythrosuaveolide with a terpenoid-like ring) suggests that the presence of nitrogen in the structure may enhance the antimicrobial activity of terpenoids. Regarding the activity of coumarins, it could be noted that 7-methoxycoumarin is generally more active than 7-hydroxycoumarin against MDR Gram-negative bacteria. 1,5-Dihydroxyxanthone and 1,7-dihydroxyxanthone exhibited similar activities, indicating that the location of the hydroxyl group in the structure does not significantly modulate their activities. It is also important to note that a simple naphthoquinone (plumbagin) exhibited better activity compared with 4-hydroxyionochocarpin (flavonoid), laurentixanthone B (xanthone) and MAB3 (coumarin). From a molecular structure point of view, the natural products exhibiting the best antibacterial activities have the same pharmacophore (Fig. 2). As plumbagin is the minimal scaffold required to have significant antibacterial activity in the absence of an EPI, it could be

suggested that this pharmacophore is involved in the antibacterial activity. Other functional groups and their position (Fig. 2), in addition to those of the pharmacophoric moiety, may affect the antibacterial properties and modulate the susceptibility of the molecule to bacterial resistance mechanisms.

In the presence of the EPI, MIC values < 10 mg/L were obtained in 2 (12.5%), 5 (31.3%), 9 (56.3%) and 16 (100%) of the 16 tested bacteria, respectively, for laurentixanthone B, MAB3, 4-hydroxyonchocarpin and plumbagin (Table 2).

#### 4. Discussion

Tripartite drug efflux pumps, mainly those clinically reported as AcrAB–TolC in Enterobacteriaceae or as MexAB–OprM in *P. aeruginosa*, play a key role in multidrug resistance of pathogenic Gram-negative bacteria [4,6]. PA $\beta$ N has been reported as a potent inhibitor of the resistance–nodulation–cell division (RND) efflux systems and is particularly active on AcrAB–TolC and MexAB–OprM [7,21]. To determine the involvement of efflux pumps, the concentration of PA $\beta$ N used in this study (20 mg/L) had no intrinsic effect on the bacteria as previously determined [19], i.e. no significant variation in the MIC of chloramphenicol was observed for AG100A (AcrAB-deleted derivative of AG100). In contrast, under these conditions a significant increase in the antibacterial activity of several compounds was noted, showing that a number of natural products are substrate of efflux pumps acting in resistant strains of *E. coli*, *E. aerogenes*, *K. pneumoniae* and *P. aeruginosa*. However, this was not the general case with terpenoids, indicating that either this class of secondary metabolites is not highly active or that RND efflux pumps are not the main mechanism of resistance involved against such compounds in these MDR bacteria. Terpenoids could also

have some difficulties in passing through the bacterial membrane barrier. The enhancing effect of PA $\beta$ N observed against the studied MDR bacteria with various compounds such as laurentixanthone B, plumbagin, 4-hydroxylonchocarpin, bartericin C and MAB3 suggests that PA $\beta$ N-sensitive efflux could be a major means of resistance. Regarding the involvement of MDR bacteria in treatment failures and the re-emergence of infectious diseases [1,2,22], the activity of some compounds (in the presence of PA $\beta$ N) could be considered very promising. *Pseudomonas aeruginosa* is an important nosocomial pathogen highly resistant to clinically used antibiotics, causing a wide spectrum of infections and leading to substantial morbidity and mortality [23]. MDR Enterobacteriaceae, including *K. pneumoniae*, *E. aerogenes* and *E. coli*, have also been classified as antimicrobial-resistant organisms of concern in healthcare facilities [22]. Therefore, laurentixanthone B, plumbagin and 4-hydroxylonchocarpin could be attractive candidates for the development of a new antimicrobial family against MDR strains in combination with EPIs. The antimicrobial activities of bartericin C and crotozambefurans A–C are being reported and the results of the present study provide a clear explanation of the low bacterial susceptibility previously observed for other compounds [12,24]. The studied compounds included terpenoids, phenolics and alkaloids. Within the phenolic compounds, some of them, such as laurentixanthone B, MAB3 and 4-hydroxylonchocarpin, exhibited significant activity and contained the same pharmacophore as observed in plumbagin.

By analysing the role of efflux mechanisms in the antimicrobial activity of the various compounds, it can be observed that compounds were more active against the tested bacteria in the presence of an EPI. These data clearly draw attention to the

involvement of efflux pumps AcrAB–TolC or MexAB–OprM [7,10,19,25] in the resistance of Gram-negative bacteria to natural compounds. This study highlights the key role of efflux mechanisms against several phenolic and alkaloid compounds and, consequently, indicates that inhibition of such efflux pumps could significantly improve their antimicrobial potencies.

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### **Competing interests**

None declared.

### **Ethical approval**

Not required.

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**Fig. 1.** Chemical structures of the natural compounds tested.

**Fig. 2.** Representation of pharmacophoric groups detected in the tested compounds.

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**Table 1**

## Bacterial strains and features

Bacterial strain	Relevant features	Reference
<i>Escherichia coli</i>		
ATCC 8739 and ATCC 10536	Reference strains	
AG100	Wild-type <i>E. coli</i> K-12	[14]
AG100A	AG100 $\Delta$ <i>acrAB</i> ::KAN <sup>R</sup>	[14]
AG100A <sub>Tet</sub>	$\Delta$ <i>acrAB</i> mutant of AG100; TET <sup>R</sup> owing to <i>acrF</i> gene markedly overexpressed	[14]
AG102	AG100 overexpressing AcrAB pump	[15]
<i>Enterobacter aerogenes</i>		
ATCC 13048	Reference strain	
EA-CM64	CHL <sup>R</sup> variant obtained from ATCC 13048 overexpressing AcrAB pump	[16]
EA27	Clinical MDR isolate exhibiting energy-dependent NOR and CHL efflux; KAN <sup>R</sup> AMP <sup>R</sup> NAL <sup>R</sup> STR <sup>R</sup> TET <sup>R</sup>	[17]
EA289	KAN-sensitive derivative of EA27	[17]
EA294	EA289 <i>acrA</i> ::KAN <sup>R</sup>	[17]
EA298	EA289 <i>tolC</i> ::KAN <sup>R</sup>	[17]
<i>Klebsiella pneumoniae</i>		
ATCC 11296	Reference strain	
Kp55	Clinical MDR isolate, TET <sup>R</sup> AMP <sup>R</sup> ATM <sup>R</sup> CEF <sup>R</sup>	[18]
Kp63	Clinical MDR isolate, TET <sup>R</sup> CHL <sup>R</sup> AMP <sup>R</sup> ATM <sup>R</sup>	[18]
<i>Pseudomonas aeruginosa</i>		
PA01	Reference strain	
PA124	MDR clinical isolate	[19]

KAN, kanamycin; TET, tetracycline; CHL, chloramphenicol; NOR, norfloxacin; AMP, ampicillin; NAL, nalidixic acid; STR, streptomycin; ATM, aztreonam; CEF, cefalothin; <sup>R</sup>, resistant; MDR, multidrug-resistant.

Table 2

Minimal inhibitory concentrations (MICs) of the studied components against the tested bacterial species

Sample	MIC (mg/L) (with 20 mg/L PAβN <sup>a</sup> )															
	<i>Escherichia coli</i>					<i>Enterobacter aerogenes</i>						<i>Klebsiella pneumoniae</i>			<i>Pseudomonas aeruginosa</i>	
	ATCC 8739	AG100	AG100A	AG100A <sub>Tet</sub>	AG102	ATCC 13048	EA-CM64	EA289	EA294	EA298	EA27	ATCC 11296	Kp55	Kp63	PA01	PA124
<b>Terpenoids</b>																
Hardwickiic acid	i	i	i (16)	i	i	i	i	i	i	i	i	i	i	i	i	i
Crotozambefuran A	i	i	i	i	i	i	i	i	i	i	i	i	i	i	i	i
Crotozambefuran B	i	i	i (256)	i	i	i	i	i	i (128)	i (128)	i	i	i	i	i	i
Crotozambefuran C	i (128)	i	i (128)	i	i	i	i	i	i (128)	i (128)	i	i	i	i	i	i
3-Friedelanone	i	i	i (128)	i	i	i	i	i	i	i	i	i	i	i	i	i
Betulinic acid	i	i	i (256)	i	i	i	i	i	i	i	i	i	i	i	i	i
Oleanolic acid	i (128)	i	i (128)	i	i	i	i	i	i	i	i	i	i	i	i	i
<b>Coumarins</b>																
MAB3	i (32)	128 (8)	32 (4)	i (8)	i (8)	256 (64)	i	i (32)	i (16)	i (8)	i	256 (32)	i (16)	i (16)	i (128)	i (256)
Mammeisin	i (16)	i (32)	i (32)	i (32)	i (64)	i	i	i	i (128)	i (16)	i	i (64)	i (16)	i (128)	i	i
7-Hydroxycoumarin	i	i	256 (128)	i (128)	i	i	i	i	256 (64)	256 (32)	i	i (128)	i	i	i	i (256)
7-Methoxycoumarin	i (128)	256 (128)	128 (32)	256 (128)	256 (64)	i (128)	128 (32)	i	128 (32)	128 (16)	256 (64)	256 (64)	i (128)	i (128)	256 (128)	256 (64)
<b>Xanthenes</b>																
1,5-Dihydroxyxanthone	i (16)	i (128)	i (16)	i (16)	256 (32)	256 (128)	i	i	128 (8)	256 (8)	i	256 (32)	i (32)	i (32)	i (256)	256 (128)
1,7-Dihydroxyxanthone	i (32)	i (32)	i (16)	i (32)	i (32)	i (256)	i	i	64 (4)	128 (4)	i	256 (64)	i (32)	i (32)	i (128)	i (128)
Laurentixanthone B	128 (8)	256 (32)	32 (8)	256 (16)	256 (32)	256 (16)	256 (32)	128 (64)	256 (32)	256 (32)	256 (16)	64 (32)	256 (64)	256 (16)	256 (64)	256 (16)
Smeathxanthone A	i	i	256 (256)	i	256 (256)	i	i	i	i	i	i	i	i	i	i	i
Smeathxanthone B	i	i	256 (256)	i	256 (256)	i	i	i (128)	i	i	i	i	i	i	i	i
<b>Quinones</b>																

Plumbagin	16 (2)	32 (1)	2 (0.12)	8 (0.25)	16 (0.5)	16 (4)	64 (8)	32 (4)	16 (2)	16 (2)	32 (4)	16 (1)	32 (4)	32 (4)	16 (1)	32 (0.5)
Lapachol	128 (32)	256 (64)	64 (16)	i (64)	i (64)	128 (32)	i	i	i (128)	i (128)	i	256 (128)	i (256)	i (256)	256 (32)	i (64)
2-Acetylfuro-1,4-naphthoquinone	256 (64)	i (64)	128 (32)	i (128)	i (256)	i (256)	i	i	i	i (256)	i	i (256)	i	i	256	i
															(256)	
Newbouldiaquinone	128 (16)	256 (32)	32 (16)	128 (32)	256 (64)	64 (32)	i (128)	256	128	256	i (256)	64 (16)	256	256	128 (16)	256 (16)
								(64)	(32)	(32)			(64)	(64)		
Newbouldiaquinone A	64 (16)	128 (32)	64 (32)	256 (32)	256 (64)	128 (16)	256	128	128	256	i	64 (32)	128	128	128 (16)	128 (32)
							(128)	(128)	(64)	(32)			(32)	(64)		
Vismiaquinone	i	i	i (128)	i	i	i	i	i	i	i	i	256 (256)	256	128	i	i
												(256)	(128)			
Vismiaquinone C	128	256	64 (4)	256 (128)	256	i	i	256	128	256	i	256 (256)	64 (64)	64 (64)	i	i (256)
	(128)	(128)			(256)			(128)	(32)	(64)						
3-Geranyloxy-6-methyl-1,8-dihydroxyanthraquinone	N/T	N/T	N/T	N/T	N/T	i	i (128)	i (128)	i (64)	i (64)	i (256)	256 (256)	128	256	i	i
												(128)	(64)			
Bivismiaquinone	i	i	i	i	i	i	i	i	i	i	i	i	i	256	i	i
														(256)		
Flavonoids																
Quercetin	256	i	i (256)	256 (256)	i	i	i	i	i	i (128)	i	256 (256)	256	256	256	i
	(256)												(256)	(256)	(256)	
Kaempferol	256	i	128 (64)	256 (256)	i	256 (256)	i	i	i (256)	i (128)	i	256 (256)	256	256	256	i
	(256)												(128)	(256)	(256)	
Gancaonin Q	i	i	256	i	i	i	i	i	i	i	i	256 (256)	i	i	256	256
			(128)												(256)	(256)
Stipulin	128 (64)	i (128)	64 (32)	i (256)	i (256)	256 (64)	i	i	i	i (256)	i	128 (64)	i	i	i (256)	256
																(128)
Angusticornin B	128 (32)	256 (64)	64 (16)	256 (64)	i (64)	256 (128)	i (128)	i (256)	i (256)	i (256)	i	128 (64)	i (256)	256	128 (64)	128 (32)
														(64)		
4-Hydroxylonchocarpin	128 (4)	128 (8)	32 (4)	128 (8)	256 (32)	256 (4)	256	128 (8)	256 (8)	256 (8)	256 (8)	64 (16)	256	256	128 (32)	256 (8)
							(64)						(32)	(32)		
Bartericin C	64 (8)	128 (16)	32 (4)	64 (16)	128 (32)	128 (128)	256	128	128	128	256	128 (128)	128	128	128	256
							(256)	(16)	(16)	(16)	(256)		(128)	(128)	(128)	(256)
Alkaloids																
Norcassaide	128 (32)	256 (64)	64 (8)	128 (16)	256 (16)	256 (64)	i (128)	256	64 (32)	64 (16)	i (128)	128 (16)	256	128	64 (8)	128 (32)
								(128)					(32)	(32)		

Norerythroavéolide	128 (16)	128 (32)	32 (8)	128 (16)	128 (16)	128 (32)	i (64)	256 (64)	128 (64)	128 (32)	256 (64)	128 (32)	256 (128)	128 (128)	128 (32)	128 (32)
Chloramphenicol (reference antibiotic)	4 (1)	4 (0.5)	0.5 (0.25)	32 (4)	32 (2)	4 (1)	256 (8)	i (128)	64 (16)	64 (16)	i (128)	4 (1)	32 (4)	i (128)	128 (8)	256 (8)

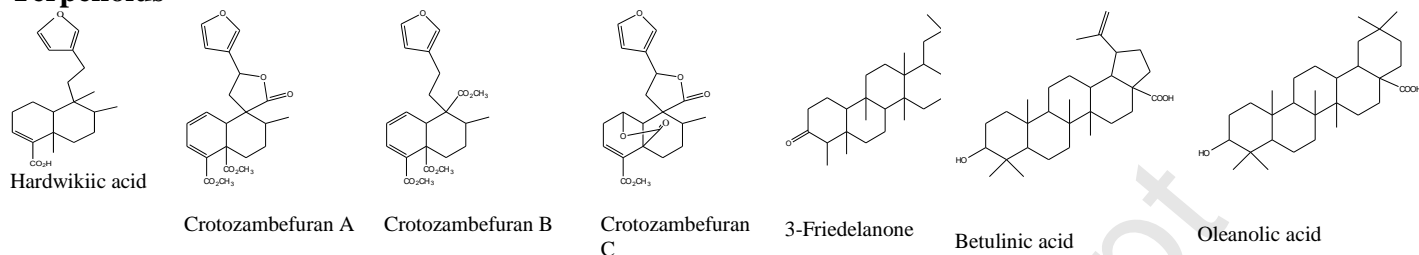
PA $\beta$ N, phenylalanine arginine  $\beta$ -naphthylamide; N/T, not tested.

<sup>a</sup> The MIC of PA $\beta$ N was 64 mg/L for AG100A and >256 mg/L for other *E. coli*, *E. aerogenes*, *K. pneumoniae* and *P. aerogenes* strains.

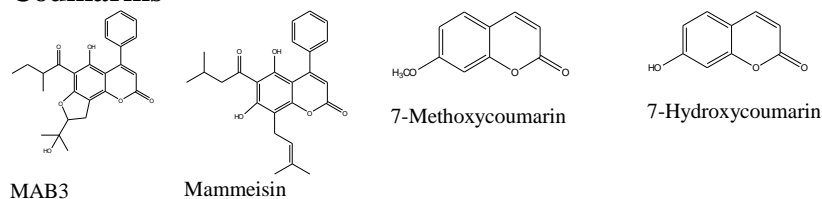
i, sample not active up to 256 mg/L.

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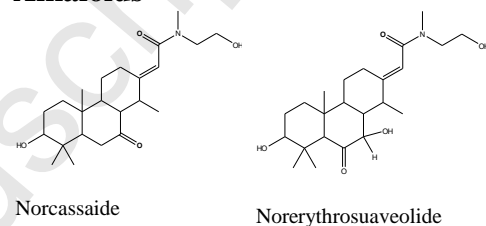
**Terpenoids**



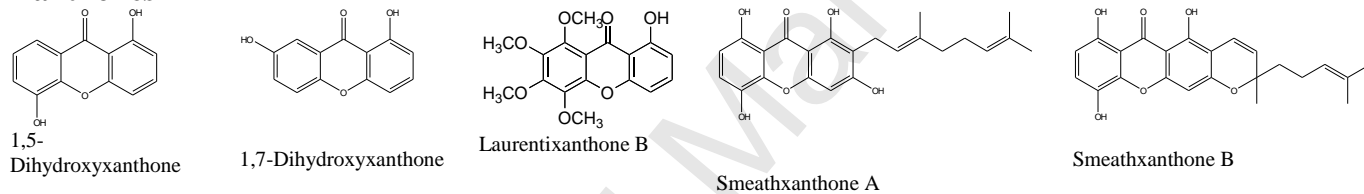
**Coumarins**



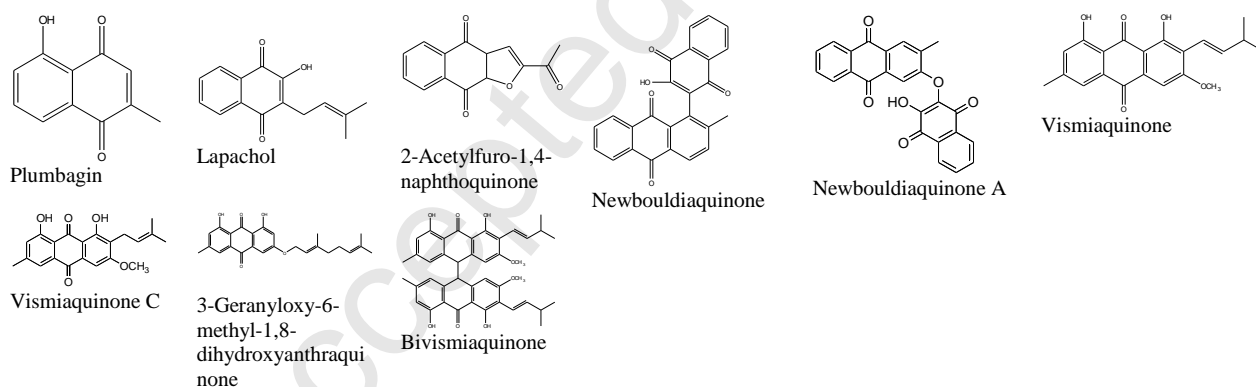
**Alkaloids**



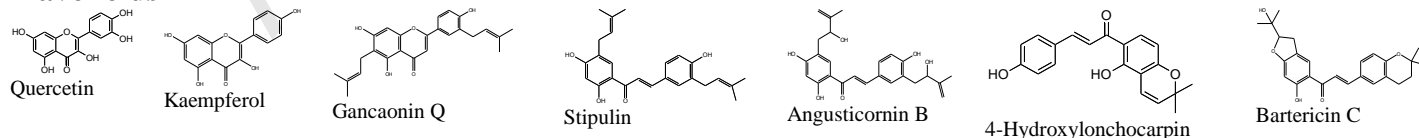
**Xanthenes**



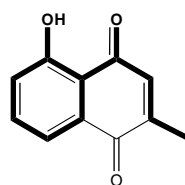
**Naphthoquinones and anthraquinones**



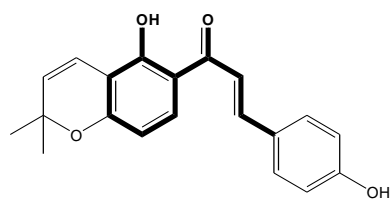
**Flavonoids**



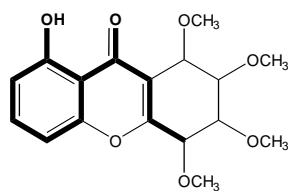




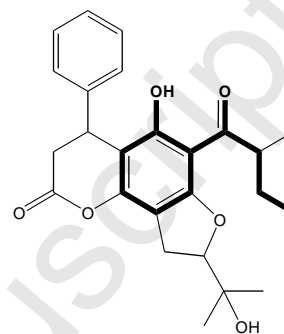
Plumbagin



4-Hydroxylonchocarpin



Laurentixanthone B



MAB3

— Pharmacophore