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Franco Maria Ruggeri, Roberto Delogu, Tatiana Petouchoff, Olga Tcheremenskaia, Simona de Petris, et al.. Molecular characterization of rotavirus strains from children with diarrhea in Italy, 2007-2009. Journal of Medical Virology, 2011, 83 (9), pp.1657. 10.1002/jmv.22163 . hal-00657579

HAL Id: hal-00657579 https://hal.science/hal-00657579

Submitted on 7 Jan 2012

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Journal of Medical Virology

Molecular characterization of rotavirus strains from children with diarrhea in Italy, 2007-2009

Journal:	Journal of Medical Virology
Manuscript ID:	JMV-10-2253.R2
Wiley - Manuscript type:	Research Article
Date Submitted by the Author:	17-May-2011
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Keywords:	Rotavirus, Genotyping, Molecular epidemiology, Gene reassortment, Zoonotic transmission

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	Rotavirus strain number, by year*							
Region	2007	2008	2009	Total				
Piedmont	100	133	44	277				
Liguria		22	-	22				
Lombardia		59	255	314				
Veneto	31	358	132	521				
Total Northern regions	131	572	431	1134				
Emilia-Romagna	53	155	120	328				
Marche	0	63	58	121				
Tuscany	17	176	35	228				
Umbria	53	89	55	197				
Lazio		19	31	50				
Fotal Central regions	123	502	299	924				
Calabria	17	33	40	90				
Campania		4	41	45				
Apulia	180	73	87	340				
Sicily		69	95	164				
Fotal Southern regions	197	179	263	639				
All regions	451	1253	993	2697				

Table 1. Number of rotavirus-positive stools collected in 2007-2009 in Italy, by region.

*Rotavirus year is the 12-month period containing the winter-spring infection peak, from September of the previous calendar year through August of designated year.

Table 2. Genotypes of rotavirus characterized in Italy, 2007-2009.

	Number of genotyped strains (%), by year*								
Genotype	2007		20	2008		2009		-2009	
G1P[8]	212	(47.3)	598	(48.8)	377	(38.8)	1187	(44.9)	
G2P[4]	76	(17.0)	88	(7.2)	60	(6.2)	224	(8.5)	
G3P[8]	2	(0.4)	45	(3.7)	35	(3.6)	82	(3.1)	
G4P[8]	53	(11.8)	75	(6.1)	143	(14.7)	271	(10.2)	
G9P[8]	74	(16.5)	285	(23.3)	233	(24.0)	592	(22.4)	
Common genotypes	417	(93.1)	1091	(89.1)	848	(87.2)	2356	(89.1)	
Uncommon	1	(0.2)	20	(1.6)	31	(3.2)	52	(2.0)	
Mixed types	22	(4.9)	106	(8.7)	72	(7.4)	200	(7.6)	
Untypable	8	(1.8)	8	(0.7)	21	(2.2)	37	(1.4)	
Total	448	(100.0)	1225	(100.0)	972	(100.0)	2645	(100.0)	

*Rotavirus year is the 12-month period containing the winter-spring infection peak, from September of the previous calendar year through August of designated year.

		Number of str	ains, by year*	
Genotype	2007	2008	2009	2007-2009
G1P[4]	-	6	-	6
G2P[8]	-	5	3	8
G4P[4]	-	2	1	3
G9P[4]	-	2	6	8
G1P[6]	-	-	2	2
G3P[6]	_	-	7	7
G8P[8]	- 0	-	1	1
G9P[9]	-	1	-	1
G10P[8]	-	4	11	15
G12P[8]	1	-	-	1
Total	1	20	31	52

Table 3. Uncommon genotypes or G and P combinations among rotaviruses characterized in Italy, 2007-2009.

*Rotavirus year is the 12-month period containing the winter-spring infection peak from September of the previous calendar year through August of designated year.

		Ye	ear		Pro	bable rotavirus involved
G-P detected	2007	2008	2009	Total	Cases	Types
G1,2 P[4,8]	1	5	9	15		
G1,2 P[4]		14	4	18	20	
G1,2 P[8]		3		3	38	$GIP[\delta], G2P[4]$
G1 P[4,8]		2		2		
G1 P[6,8]			1	1	1	G1P[8], G?P[6]
G1,3 P[8]		1		1	1	G1P[8], G3P[8]
G1,4 P[8]	2	6	5	13	13	G1P[8], G4P[8]
G1,9 P[8]	18	43	7	68	68	G1P[8], G9P[8]
G2,3 P[4,8]			1	1	2	
G2,3 P[4]			2	2	3	$G2P[4], G3P[\delta]$
G2 P[4,6]			1	1	1	G2P[4], G?P[6]
G2,4 P[4,8]			2	2		
G2,4 P[4]		7	1	8	11	G2P[4], G4P[8]
G2,4 P[8]			1	1		
G2,9 P[4,8]		8	12	20		
G2,9 P[4]			2	2	27	G2P[4], G9P[8]
G9 P[4,8]	1	3	1	5		
G2 P[4,8]		1		1	1	G2P[4], G?P[8]
G4,9 P[8]		5	3	8	8	G4P[8], G9P[8]
G9 P[6,8]			1	1	1	G9P[8], G?P[6]
G1,2 P[4,6]		1	1	2	2	G1P[?]°, G2P[4], G?P[6]°
G1,2,4 P[8]			3	3	3	G1P[8], G2P[4], G4P[8]
G1,2,9 P[4,8]		1		1	17	
G1,9 P[4,8]		5	11	16	17	01P[8], 02P[4], 09P[8]
G2,4 P[6,8]			1	1	1	G2P[4], G4P[8], G?P[6]
G2,9 P[4,6]			1	1	1	G2P[4], G9P[8], G?P[6]
G3 P[4,6]			1	1	1	G2P[4], G3P[?] ^{°°} , G?P[6] ^{°°}
G3,4 P[4,8]		1		1	1	G2P[4], G3P[8], G4P[8]
G4,9 P[4,8]			1	1	1	G2P[4], G4P[8], G9P[8]
Total	22	105	65	192	192	

Table 4. Mixed infections with G and P genotypes rotaviruses in Italy, 2007-2009.

*Rotavirus year is the 12-month period containing the winter-spring infection peak from September

of the previous calendar year through August of designated year.

°Possible G1P[6] reassortant strain

°°Possible G3P[6] reassortant strain

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Table 5. Mixed infections of rotaviruses of different common G and P types observed in Italy, 2008-2009 as possible source for the emergence of natural reassortants

Mixed infecti	ons detected	I		Ν	lixed infect	tions as pos	ssible sourc	e for natura	al reassort	ants detecte	ed	
Genotypes*	No	.**	G11	P[4]	G2	P[8]	G3	P[4]	G4	P[4]	G9	P[4]
	2008	2009	2008	2009	2008	2009	2008	2009	2008	2009	2008	2009
G1,2P[4,8]	5	9	5	9	5	9						
G1,2P[4]	14	4	14	4								
G1,2P[8]	3				3							
G1P[4,8]	2		2									
G2,3P[4,8]		1				1		1				
G2,3P[4]		2						2				
G2,4P[4,8]		2				2				2		
G2,4P[4]	7	1							7	1		
G2,4P[8]		1				1						
G2,9P[4,8]	8	12			8	12					8	12
G2,9P[4]		2										2
G9P[4,8]	3	1									3	1
G2P[4,8]	1				1							
G1,2P[4,6]	1	1	1	1								
G1,2,4P[8]		3				3						
G1,2,9P[4,8]	1		1		1						1	
G1,9P[4,8]	5	11	5	11							5	11
G2,4P[6,8]		1				1						
G2,9P[4,6]		1										1
G3P[4,6]		1						1				
G3,4P[4,8]	1						1		1			
G4,9P[4,8]		1								1		1
Total			28	25	18	29	1	4	8	4	17	28
No. of natural reassortants detected			6	0	5	3	0	0	2	1	2	6

* The P[6] genotypes were not considered in this context, as they are of likely animal origin and thus, whilst being found in mixed infections, they are not a factor in the natural reassortment of common human rotavirus strains.

** Values taken from Table 4.

*** Values taken from Table 3.

If total number of mixed infections/year is used as independent variable x, and the number of natural reassortants detected as dependent variable y, the following linear regression line is found: y = 0.14 x + 0.17. The correlation coefficient r is 0.66, i.e. for degree of freedom of 8 (= n-2) the relationship is significant at P<0.05.

Table 6. Genotypes of rotavirus characterized in Northern, Central, and Southern Italy, 2007-2009. Number of genotyped strains (%), by Area Northern Central Southern All Genotype NT 07. NL α NL α

2006 - 7		INO.	%	NO.	%	NO.	%	NO.	%
	G1P[8]	95	(73.6)	76	(61.8)	41	(20.9)	212	(47.3)
	G2P[4]	11	(8.5)	31	(25.2)	34	(17.3)	76	(17.0)
Common	G3P[8]	-	-	-	-	2	(1.0)	2	(0.4)
	G4P[8]	3	(2.3)	4	(3.3)	46	(23.5)	53	(11.8)
	G9P[8]	13	(10.1)	8	(6.5)	53	(27.0)	74	(16.5)
Uncommon		-	-	-	-	1	(0.5)	1	(0.2)
Mixed		7	(5.4)	4	(3.3)	11	(5.6)	22	(4.9)
Untypable		-	-	-	-	8	(4.1)	8	(1.8)
Total		129	(100.0)	123	(100.0)	196	(100.0)	448	(100.0)
2007 -	· 8				1				
	G1P[8]	283	(50.8)	261	(53.0)	54	(30.7)	598	(48.8)
	G2P[4]	24	(4.3)	31	(6.3)	33	(18.8)	88	(7.2)
Common	G3P[8]	14	(2.5)	17	(3.5)	14	(8.0)	45	(3.7)
	G4P[8]	31	(5.6)	39	(7.9)	5	(2.8)	75	(6.1)
	G9P[8]	155	(27.8)	92	(18.7)	38	(21.6)	285	(23.3)
Uncommon		9	(1.6)	6	(1.2)	5	(2.8)	20	(1.6)
Mixed		41	(7.4)	46	(9.3)	19	(10.8)	106	(8.7)
Untypable	Untypable		-	-	-	8	(4.5)	8	(0.7)
Total		557	(100.0)	492	(100.0)	176	(100.0)	1225	(100.0)
2008 -	9								
	G1P[8]	188	(45.0)	123	(42.1)	66	(25.2)	377	(38.8)
	G2P[4]	20	(4.8)	15	(5.1)	25	(9.5)	60	(6.2)
Common	G3P[8]	17	(4.1)	11	(3.8)	7	(2.7)	35	(3.6)
	G4P[8]	30	(7.2)	21	(7.2)	92	(35.1)	143	(14.7)
	G9P[8]	124	(29.7)	78	(26.7)	31	(11.8)	233	(24.0)
Uncommon		10	(2.4)	11	(3.8)	10	(3.8)	31	(3.2)
Mixed		24	(5.7)	31	(10.6)	17	(6.5)	72	(7.4)
Untypable		5	(1.2)	2	(0.7)	14	(5.3)	21	(2.2)
Total		418	(100.0)	292	(100.0)	262	(100.0)	972	(100.0)
ALL YE	ARS								
	G1P[8]	566	(51.3)	460	(50.7)	161	(25.4)	1187	(44.9)
	G2P[4]	55	(5.0)	77	(8.5)	92	(14.5)	224	(8.5)
Common	G3P[8]	31	(2.8)	28	(3.1)	23	(3.6)	82	(3.1)
	G4P[8]	64	(5.8)	64	(7.1)	143	(22.6)	271	(10.2)
	G9P[8]	292	(26.4)	178	(19.6)	122	(19.2)	592	(22.4)
Uncommon		19	(1.7)	17	(1.9)	16	(2.5)	52	(2.0)
Mixed		72	(6.5)	81	(8.9)	47	(7.4)	200	(7.6)
Untypable		5	(0.5)	2	(0.2)	30	(4.7)	37	(1.4)
Total		1104	(100.0)	907	(100.0)	634	(100.0)	2645	(100.0)





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2 3	1	Molecular characterization of rotavirus strains from children with diarrhea
4 5	2	in Italy, 2007-2009
6 7	3	
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10 11	5	Franco Maria Ruggeri ¹ , Roberto Delogu ² , Tatiana Petouchoff ² , Olga Tcheremenskaia ¹ , Simona
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24 25	13	*List of RotaNet-Italy Study Group members is shown in Annex 1
26 27	14	
28 29	15	
30	16	
31 32	17	Short title:
33 34	18	Molecular epidemiology of rotavirus in Italy
35 36	19	
37 38	20	Key words
39 40	21	Rotavirus, genotyping, molecular epidemiology, gene reassortment, zoonotic transmission
40 41	22	
42 43	23	The work was designed and performed at the Istituto Superiore di Sanità, coordinating the
44 45	24	activities of regional partners of the RotaNet-Italy Study Group
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RACT

illance network RotaNet-Italia was established in 2007 in order to investigate the of co-circulating rotavirus strains in Italy, and to provide a baseline for future of possible effects of vaccine implementation in selecting novel versus common trains.

2,645 rotavirus strains from pediatric patients with acute diarrhea were collected over secutive seasons from September 2006 through August 2009, and partially ed by standardized multiplex RT-PCR. Most of strains (89.1%) belonged to G1-G4, and G9, associated with either P[8] or P[4], commonly found in humans . However, in at least 2.0% of cases, viruses exhibited either a G or P type typical of al strains, suggesting gene reassortment events between rotaviruses of different origin. ections with two or more rotavirus strains were observed frequently (7.6% of patients), ded on the frequencies of co-circulating rotaviruses of one particular genotype. The nd genotypes of likely natural reassortants of common genotype rotaviruses were be correlated with the observed numbers and genotypes of mixed infections. Large in the relative frequency of different rotavirus genotypes was observed between seasons and/or areas of Italy, suggesting independent evolution or differential on of viral strains with respect to both time and space.

INTRODUCTION

Rotaviruses cause a large proportion of acute gastroenteritis cases in infants and young children worldwide, with a death toll of more than 500,000 yearly, mostly in developing countries [Parashar et al., 2006]. Each year, rotaviruses cause approximately 111 million episodes of acute gastroenteritis requiring home care, 25 million clinic visits, and 2 million hospitalizations, altogether constituting a major economic burden for health care systems and families in both USA and European countries [Forster et al., 2009; Parashar et al., 2006; Soriano-Gabarro et al., 2006]. These figures are consistent with the need for implementing vaccination strategies in both developing and developed countries, as recommended by the World Health Organization [WHO, 2007].

Two rotavirus vaccines have been licensed in Europe, USA and other countries throughout the world, and have shown high efficacy in phase III clinical trials [Ruiz-Palacios et al., 2006; Vesikari et al., 2006b]. These are: (i) the live attenuated monovalent human rotavirus vaccine Rotarix (GlaxoSmithKline, London, England) constituted by a single G1P1A[8] strain [Ruiz-Palacios et al., 2006]; (ii) the live attenuated pentavalent human-bovine reassortant vaccine RotaTeq [Clark et al., 2006; Vesikari et al., 2006b] presenting human serotype G1 through G4, and P1A[8] specificities (Merck, Whitehouse Station, NJ). A remarkable decline of pediatric admissions for rotavirus diarrhea was reported in Philadelphia in 2008, and attributed to specific herd immunity consolidated by the increasing vaccine use during the previous two years [Clark et al., 2009]. Although this study noted that most of 2008 cases were infected with genotype G3 rotavirus, their low number allowed no inference on a possible differential efficacy of vaccination by viral serotype. Nonetheless, a high efficacy (95%) of the pentavalent vaccine towards G3P[8] infection was later demonstrated in Texas [Boom et al., 2010]. Further studies will likely generate more data, particularly in the Americas, where children immunization has been adopted on a larger scale than elsewhere [de Oliveira et al., 2008; Parashar et al., 2006].

The correlates of protection from rotavirus disease are not fully defined [Desselberger and Huppertz, 2011], but it is largely accepted by the scientific community that both outer capsid proteins of rotavirus VP7 and VP4 stimulate neutralizing antibody responses [Yuan et al., 2009], which may be important for forming complexes with the virus on the intestinal mucosa [Coffin et al., 1997; To et al., 1998] or prevent viremia and possible extra-intestinal rotavirus replication [Blutt et al., 2007].

The issue of protective immunity from rotavirus infection is related to the complexity of the
 virion, exhibiting three protein layers and six structural proteins, and of its genome consisting

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of 11 double-stranded (ds) RNA segments which encode 11 or 12 proteins [Pesavento et al., 2006]. The ability of rotavirus to exchange entire genes during multi-strain infection of cells (reassortment) is known since many years [Gombold and Ramig, 1986; Offit and Blavat, 1986], is observed during infection in vivo in humans and animals [Esona et al., 2009; Iturriza-Gomara et al., 2001; Maunula and Von Bonsdorff, 2002; Watanabe et al., 2001], and is a major cause of the exceptional genetic variability observed for clinical rotavirus strains worldwide [Santos and Hoshino, 2005]. Interspecies rotavirus reassortment may have implications for vaccine programs effectiveness because it may introduce viral serotypes typical of animals into potentially naïve human populations, independent of vaccine coverage [Gentsch et al., 2005]. The second major factor in rotavirus genetic diversity is the accumulation of point mutations generated by the error-prone RNA-dependent RNA polymerase, that can generate neutralization escape variants [Arista et al., 2005; Ciarlet et al., 1997; Iturriza-Gomara et al., 2001; Shaw et al., 1988] and misrecognition of viral genes by established genotyping primers [Parra and Espinola, 2006; Santos et al., 2003; Solberg et al., 2009].

- Despite the large diversity among rotavirus strains, a broad cross-reactive immunity between viral serotypes *in vitro* and *in vivo* is supported by several studies [Gorrell and Bishop, 1999; Vesikari et al., 2007; Yuan et al., 2005], and may explain the reported efficacy of vaccines against circulating wild-type rotaviruses of G- and P-types which are not encompassed in vaccine formulations.
- To track rotavirus genes evolution, zoonotic interchange and reassortment of group A rotaviruses [Martinez-Laso et al., 2009; Matthijnssens et al., 2008a; Song and Hao, 2009], a recently proposed new classification scheme considering all 11 genome segments [Matthijnssens et al., 2008a; Matthijnssens et al., 2008b] has been very useful. Based on extensive genome sequencing, this system is suited for very specific studies on rotavirus phylogeny or for detailed investigations on uncommon strains emerging in the population. Multiplex RT-PCRs with diverse primer sets are normally used for characterizing fecal group A rotavirus strains into G- and P- types, relative to the genes encoding capsid (Glycosylated) VP7 and (Protease-sensitive) VP4 proteins [Gentsch et al, 1992; Gouvea et al., 1990; Iturriza-Gomara et al., 2004; Simmonds et al., 2008]. By applying this dual nomenclature method [Desselberger et al., 2009; Estes and Kapikian, 2007], at least 23 G-genotypes and 32 P-genotypes were differentiated [Collins et al., 2010; Trojnar et al., 2009; Ursu et al., 2009], including 10 G- and 11 P-types in human infections, respectively [Desselberger et al., 2009; Jayaram et al., 2004]. Molecular typing methods have improved the understanding of

rotavirus strain diversity, and permitted establishment of large molecular epidemiology
programs [Gentsch et al., 2005; Iturriza-Gomara et al, 2010; Iturriza-Gomara et al., 2009;
Santos and Hoshino, 2005], which will be valuable to control the efficacy of present vaccines
towards different co-circulating wildtype rotaviruses, estimate the impact of possible vaccinedriven immune selection of wildtype strains and help shaping possible future rotavirus
vaccines development [Desselberger et al., 2006].

To gather comprehensive information on co-circulating rotavirus genotypes throughout
Europe, a large consortium was established in 2007 [Iturriza-Gomara et al., 2009], embracing
17 countries, using shared methods and viral databases.

10 The present study reports the molecular characterization of rotaviruses from hospitalized 11 children with diarrhea during a 3-year surveillance activity in Italy, and discusses temporal 12 and geographical differences in the incidence of infection with different genotypes.

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MATERIALS AND METHODS

2 RotaNet-Italy Study Group

A laboratory-based network was established by the Istituto Superiore di Sanità in 2007, in collaboration with the Ministry of Health, Regional Public Health Services, Hospitals and Universities in different Italian regions. Table 1 lists the collaborating institutions and the number of rotavirus strains contributed. Regional centers were selected to ensure a large pediatric patients catchment area across the country and to broaden the capacity of laboratory rotavirus diagnosis.

Patients and samples

Stool samples were collected from children aged less than 5 years, admitted with acute gastroenteritis to hospitals or outpatient wards between 2006 and 2009 seasons. According to the European network, a rotavirus season was defined as the 12-month period from September through the end of August of the following year. Each specimen was labeled with a unique surveillance identification number. Epidemiological and patient data (age, sex, geographical location, date of onset, date of sample collection, and symptoms) were recorded using *ad hoc* questionnaires, and entered into a database for linkage to genotyping data. Patients' privacy and confidentiality issues were managed in agreement with national legislation.

20 Rotavirus testing

Stools were collected at admission, analyzed right after collection or stored at -20°C until processing. Rotavirus screening was performed with commercial tests validated in each laboratory, including immuno-chromatographic, immuno-enzymatic and latex-agglutination formats. After rotavirus identification by local laboratories, positive stools were shipped in dry ice to the Istituto Superiore di Sanità in Rome, for strain characterization, maintaining the cold-chain until genotyping. For molecular testing, 10% suspensions in water, were clarified by micro-tube centrifugation (15,700 x g for 15 min) and used for rotavirus-specific reverse transcription (RT)-PCR and nested PCR.

Nucleic Acid Extraction and G/P Rotavirus Typing

Rotavirus dsRNA was extracted from 140 µL of the 10% fecal suspension by either QIAamp
 Viral RNA Mini Kit (Qiagen, Milan, Italy) or SV Total RNA Isolation System (Promega,
 Milan, Italy), according to the manufacturer's instructions. RNA was eluted in 40 µL of
 RNase-free water, and stored at -80°C or used immediately. G and P rotavirus genotyping

2 3	1	was	performed	by	standardized	RT-PCR	methods
4 5	2	(<u>http://w</u>	ww.who.int/nuvi/rot	tavirus/WH	O_IVB_08.17_eng.pdf;		
6 7	3	http://ww	ww.eurorota.net/docs	<u>s.php</u>). Esse	entially, for identification	on of G type, VP7	-F and VP7-
8	4	R conser	nsus primers were us	sed in first-r	ound PCR. Subsequent	ly, the VP7-R prin	mer was used
9 10	5	in a nes	sted multiplex PCR	together w	with the G1-G4, G8-G	10 and G12-spec	cific forward
11 12	6	primers.	Con2-Con3 conser	nsus primer	s were used in RT-PC	CR for P typing,	followed by
13 14	7	standard	multiplex PCR incl	uding the C	on3 primer, in combina	tion with type-spe	ecific primers
15	8	for P4, a	nd P6-P11. When n	ecessary, ro	otavirus infection was co	onfirmed by VP6	-specific RT-
17	9	PCR [G	entsch et al., 1992;	Gouvea et a	al., 1990; Iturriza-Goma	ara et al., 2004].]	For genotype
18 19	10	assignm	ent, the molecular s	ize of PCR	amplicons, separated b	y agarose gel ele	ectrophoresis,
20 21	11	was det	ermined using a M	olecular Im	nager Gel Doc XR wit	th the Quantity-O	One software
22	12	(BioRad	, Segrate, Italy). T	he perform	ance of the laboratory	v is evaluated ar	nnually by a
23	13	genotypi	ing Ring Test on blir	nd samples a	administered within the	EuroRotaNet circ	cuit.
25 26	14						
27 28	15	DNA Se	quencing and Anal	ysis			

A total of 53 rotavirus strains were confirmed by nucleotide sequencing of PCR amplicons. These included a random selection of "common" strains to control the reliability of the genotyping PCR, samples exhibiting an uncommon or ambiguous genotype, and a fraction of amplicons suggesting mixed genotype infections. Gene 4 or gene 9 DNA bands were purified using Wizard SV Gel and PCR Clean-Up System (Promega), and sequenced using the PCR primers with the BigDye Terminator Cycle Sequencing Ready Reaction Kit version 3.1 (Perkin Elmer, Applied Biosystems, Foster City, CA) in an automated sequencer (ABI Prism 310 DNA sequencer, Applied Biosystems, Foster City, CA). Sequences obtained were compared against the NCBI GenBank database (http://www.ncbi.nlm.nih.gov), using the DNASIS Max v.2 software (Hitachisoft).

Analysis of Mixed Infections and Natural Reassortants

The numbers of observed likely natural reassortants G1P[4], G2P[8], G3P[4], G4P[4], and G9P[4] of the seasons 2008 and 2009 were compared with the numbers of relevant mixed infections observed during this time period, using linear regression analysis.

1 2 3	1	
4 5		
6 7	2	RESULTS
8 9	3	Sampling of rotavirus strains in Italy
10	4	Rotavirus cases, stool and data reporting to the central laboratory were implemented in Italy
11	5	since late 2006, under a surveillance project (2007) coordinated by the Istituto Superiore di
13 14	6	Sanità and supported by the Ministry of Health. The project is linked to the EuroRotaNet
15 16	7	consortium, and enrolls hospitals and laboratories spread across Italy.
17	8	Rotaviruses were identified in 2,697 stools of children, requiring hospitalization or laboratory
18 19	9	testing for acute gastroenteritis, during three consecutive years, including winter-spring peaks
20 21	10	of 2007-2009. The numbers of rotavirus strains collected by region is reported in Table 1, and
22 23	11	overall increased from 451 in 2007 (7 of 20 Regions) to 1,253 and 993 in 2008 and 2009 (13
24	12	Regions), respectively.
25 26	13	
27 28	14	Prevalence of rotavirus G and P types
29 30	15	Out of 2,697 rotaviruses detected by routine diagnostic, 2,645 were confirmed by RT-PCR
31 22	16	genotyping (Table 2). Among the VP7 genotypes identified, G1 predominated in Italy during
33	17	the entire period accounting for approximately 45% (range 38.8 - 48.8%) of strains
34 35	18	investigated, followed by G9 (22.4%), G4 and G2 (10.2 and 8.5%, respectively). G3
36 37	19	Rotaviruses were observed in 3.1% of cases, being rare in 2007. Major variation between
38 30	20	years was also observed for G2P[4] strains, which ranged from 17.0% in 2007 to 6.2 % in
40	21	2009, and G4P[8] (11.8% in 2007; 6.1% in 2008; 14.7% in 2009). In most specimens,
41 42	22	common G-types, (G1, G3, G4, and G9, and G2) were associated with common P[8] and
43 44	23	P[4], respectively, comprising on average 89.1 % of all fully G- and P-typed specimens.
45 46	24	Unconventional G and P associations, particularly G1, G4 or G9 with P[4] (6, 3, and 8 strains,
47	25	respectively), and G2P[8] (8 cases), or uncommon rotavirus genotypes, such as G10P[8] (15
40 49	26	strains) and G12P[8] (1 case), were observed in 2.0% of cases (Table 3). In further 10 cases,
50 51	27	other unusual human G or P alleles were detected, such as G8, P[6], and P[9], in combination
52 53	28	with common human P or G types.
54 55	29	Altogether, mono- (Table 3) or mixed (Table 4) infections with a P[6] rotavirus were detected
56	30	in 17 of 2645 patients (0.7%). All 7 G3P[6] strains (Table 3) were found in Apulia (Southern
57 58	31	Italy) in 2008-2009, suggesting a geographically limited circulation of this strain. The two
59 60	32	G1P[6] strains identified (Table 3) originated from Sicily (Southern Italy) and Emilia-

Romagna (North-central), in 2009. The eight mixed P[6] infections were recorded in Emilia-Romagna mostly in 2009 (Table 4).

4 Mixed rotavirus infections

Infections with more than one type of rotavirus were suggested by co-amplification of
multiple bands. Of 200 mixed infection cases (200/2645; 7.6%), 170 and 22 involved two or
three of the common (G1-G4, G9) strains (Table 4). Simultaneous presence of uncommon
P[6] with P[4] (4 cases) or P[8] (2 cases) types, or both (2 cases), and one or two of G1-G4, or
G9 G-types (Table 4) occurred in 8 cases.

18
1910Among the 170 double rotavirus infections (Table 4), the G1+G9P[8] combination occurred20
21
2111most commonly (68 stools; 40.0%). Detection of G1 and/or G2, and P[4] and/or P[8],22
2312suggested co-infection with G2P[4] and G1P[8] rotaviruses in 38 (22.4%) cases.

Similarly, presence of specific combinations of amplified DNA bands (including all or only part of the parental strain G- and P- bands; see Table 4) identified G2P[4] and G9P[8] co-infections in 27 cases (15.9%), and G2P[4] and G4P[8] in 11 (6.5%). Double infections with G1P[8]-G4P[8] (13 cases; 7.6%), and G4P[8]-G9P[8] strains (8 cases; 4.7 %) were also frequent.

3318Triple infection with common rotavirus genotypes (Table 4), involved mostly G2P[4],343519G1P[8], and G9P[8] strains (17/22; 77.3%).

The frequencies of individual "common" rotavirus infections were compared to the frequencies of observed infections with specific "mixed" genotypes. Table 5 lists the numbers of mixed infections with the five "common" genotypes observed during the years 2007-8 and 2008-9, for which the numbers of mixed infections were reasonably high. A correlation of low significance (P<0.05) was found between the numbers and genotypes of mixed infections and the numbers of emerged likely natural reassortants. However, given the relatively small numbers of natural reassortants in this clinical specimens collection, this may not necessarily be the case.

29 N

Nucleotide sequencing of untypable rotavirus strains

In a total of 72 cases, the result of genotyping PCR was ambiguous because a clearly defined DNA band was not detected after the multiplex PCR. However, 35 of these rotavirus strains were eventually characterized by sequencing the first PCR amplicon. Except one case where a G10P[8] was present, the other 34 cases belonged to common G and P genotypes. In further 37 cases, strains could not be typed, because no definite sequence could be generated and Page 17 of 31

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analyzed, as a consequence of inefficient DNA yield at PCR. Scarce virus concentration in
these stool samples is the most likely explanation for genotyping failure, because in 19 of
these cases both G and P typing failed. However, it cannot be excluded that some samples
might contain either unusual G or P types or mutations in the sequences targeted by PCR
primers. In three cases with partial genotyping, the only gene determined belonged to a type
uncommon in humans, i.e. G10, P[9], or P[10].

8 Geographical diversity in rotavirus genotype distribution in Italy

The variation in RV genotypes distribution by year of collection and geographical area is shown in Table 6, clustering data according to Northern, Central and Southern groups of Italian regions (see region grouping details in Table 1). Overall, G1P[8] rotavirus strains were constantly less prevalent in Southern Italy (from 20.9 in 2006-7 to 30.7% in 2007-8), than in the remaining Regions (from 73.6 in the North during 2006-7 to 42.1% in Central Italy, 2008-9). G2P[4] strains showed higher prevalence by both year (Table 2) and area (Table 6) in the South. In fact, compared to an overall 8.5%, this genotype represented 17.3 and 25.2% of all strains in Southern and Central regions (vs. 8.5% in the North) in 2006-7, 18.8% of strains from Southern Regions vs. 4.3 and 6.3% in North and Center in 2007-8, whereas in 2008-9 it ranged from 4.8 through 9.5% North-to-South. G3P[8] strains were relatively more constantly found in all regions (mean 3.1%), but were virtually absent in Italy in 2006-7 (0.2%), while increasing in Southern Regions in 2007-8 (8.0%). G4P[8] was generally infrequent during the central year of study (2.8 through 7.9% in the South and Center, respectively, vs. overall 10.2%), as was in 2006-7 in North-Central Italy (2.3-3.3%). Conversely, it was the first or second most prevalent genotype in Southern Regions in both 2006-7 (23.5%) and 2008-9 (35.1%), dropping to 2.8% in the South during the middle year. In 2006-7, G9P[8] rotaviruses showed a trend (10.1, 6.5 and 27.0% in Northern, Central and Southern Italy, respectively) similar to G4P[8], occurring more constantly in the other years. Uncommon rotavirus genotypes occurred rarely in 2006-7 (1 case, 0.2%), subsequently ranging from 1.2 through 3.8%. Uncommon and untypable (Table 6) strains were relatively more common in Southern Italy.

Temporal distribution of rotavirus diarrhea cases

The monthly distribution of 2,377 rotavirus patients with available date of onset is reported in Figure 1, according to Northern, Central, and Southern groups of Regions. Monthly data for the 3 study years were cumulated. Overall, most cases of rotavirus disease (85.0%) clustered between December-May, with minor variation between years (not shown). In Southern Italy, cases peaked in March-April, and in North-Central Regions the rotavirus peak was apparently broader starting in January with a tail in April (Figure 1). Although infrequent in the summer months, rotavirus admissions were recorded throughout the year.

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1 DISCUSSION

The Rota-Net Italy Study Group is the Italian component of the 16 country-wide EuroRotaNet [Iturriza-Gomara et al., 2009], and besides providing rotavirus genotyping data from pediatric gastroenteritis cases into the European dataset, it was established to further analyze the diversity of co-circulating rotavirus strains in Italy at a Regional level. An overview of EuroRotaNet development and its results has been described elsewhere [Iturriza-Gomara et al., 2010; Iturriza-Gomara et al., 2009] indicating the increasing impact of this initiative. A parallel advancement was experienced in the interest and efficiency of Regional diagnostic partners in Italy, where the number of Regions involved expanded from 7 in 2006/2007 to 13 out of total 20 forming Italy, since 2009. The number of strains genotyped by year has increased from 451 during the first season to 1253 and 993 in 2008 and 2009, respectively.

Some variation in the number of rotavirus diarrhea cases analyzed was observed between years and Regions, due to changes in resources allocated locally in different years according to regional disease prioritization plans and in response to public health emergencies. This was probably the case for almost all collaborating partners in 2009, when the H1N1 influenza pandemic threat arose and attracted most epidemiological attention, funds for laboratory diagnostics and hospital admissions. The number of 701 samples per year recommended in the EuroRotaNet sampling scheme was not reached in 2006-7, due to the fewer regions enrolled during the starting year of the study. Nonetheless, as many as 2,697 rotavirus cases in 3 consecutive years were collected, beyond the total sample size (2,103) designed for early detection of emerging rotavirus genotypes with a prevalence of less than 1% in Italy [Iturriza-Gomara et al., 2009].

Since the total number of acute gastroenteritis cases tested in Regional Hospitals is undefined, this study cannot supply prevalence data on rotavirus in childhood. However, other specific studies conducted in limited areas of Italy confirmed rotaviruses as the major cause of severe pediatric diarrhea, as it was in the '80s and '90s [Forster et al., 2009; Giaquinto et al., 2010; Medici et al., 2006; Ruggeri and Declich, 1999]. Clear epidemic waves of rotavirus acute gastroenteritis were found during late winter-spring in the whole Italy, although a few patients with rotavirus infection were detected all-year round. In the South of the country, cases seem to peak with approximately one-month delay with respect to both Northern and Central Regions. Although this difference is small, it might however reflect a delayed winter in the South. This observation seems to be in loose agreement with recent findings that the rotavirus winter epidemic wave in Europe [Iturriza-Gomara et al., 2010] starts in the South-west heading on the North-east, similar to the experience in the US [Turcios et al., 2006].

The large genotype diversity (>40 G and P combinations) observed in all European countries [Iturriza-Gomara et al., 2010] is only partially reflected in Italy, suggesting that minor rotavirus strains may circulate in individual countries only. Conversely, the present data confirmed that in Italy apparent infections with "common" genotypes (G1P[8], G2P[4], G3P[8], G4P[8], and G9P[8]) occurred at an incidence rate very similar to the whole of Europe (89.1% compared to 89.5%). The same specific rates of "common" genotypes in Italy resembled closely those found for the whole European network, being 44.9% vs. 48.4%, 8.5% vs. 10.1%, 3.1% vs. 4.3%, and 10.2% vs. 15.1% for G1P[8], G2P[4], G3P[8], and G4P[8] viruses, respectively. Only G9P[8] strains were considerably more prevalent in Italy, being found in 22.4% of cases compared to an average of 11.6% in Europe [Iturriza-Gomara et al., 2010]. A similarly higher circulation of G9P[8] strains (25.1%) has also been reported in the bordering France during the same period [de Rougemont et al., 2011]. Compared to the results for Italy, the French investigation showed a higher mean prevalence of infections with G1P[8], i.e. 59.2%, and remarkably fewer G4P[8] cases (2.2%). However, also in the case of France, the vast majority of rotaviruses typed belonged to the common types, and no obvious emergence of human-animal reassortants or zoonotic strain was detected [de Rougemont et al., 2011].

Differences in the prevalence of circulating genotypes described among European countries [Iturriza-Gomara et al., 2010] were also observed between the different Regions of Italy, and between years. To minimize fluctuations due to the low sample size of specific Regions, data were cumulated according to Northern, Central, and Southern groups of Regions (for details of grouping of the regions, see Table 1). This grouping also somehow reflects differences along a North-to-South gradient in both socio-economic (industrialized/metropolitan territory towards more rural/smaller cities) and climatic (cooler/humid towards warmer/dry) parameters. Southern Regions are also more exposed to immigration particularly from Africa and other Mediterranean countries. This might partially explain the occurrence of "untypable" rotaviruses mostly in the South (4.1-5.3%), throughout the study. Accordingly, Southern Italy showed a lower (by 5-7%) frequency of "common" genotypes, and a significant decrease in G1P[8] rotavirus prevalence throughout the study period, particularly in 2006-7 (20.9% vs. 73.6 and 61.8% in Northern and Central Regions, respectively). This was compensated by more G2P[4] and G4P[8] strains in the South (14.5% and 22.6% vs. national means of 8.5% and 10.2%, respectively). The only peculiarity of Northern Italy was a higher circulation of G9P[8] rotavirus compared to the other Regions in 2007-9, whereas this genotype was more frequent in the South in 2006-7. This large variability of co-circulating rotavirus genotypes by

time and space in Italy remains unexplained, as it is in other countries [Santos and Hoshino, 2005]. The possibility of a differential selective pressure of vaccines towards emergence of individual genotypes cannot be invoked at present because rotavirus vaccination has progressed very slowly in all Regions of Italy. It is possible that strains which become predominant in a particular area and year would subsequently re-emerge depending on both their residual persistence in healthy carriers, animal reservoirs or the environment and/or on the immune pressure previously established in the population. It is also possible that novel strains may be locally introduced from other areas displacing formerly predominant strains based on limited antigenic differences and/or fitness to replicate in humans. As a matter of fact, it is not clear to what extent genotype diversity truly reflects diversity in host (immune system)-pathogen relationships, and the basis of rotavirus attenuation is still controversial, although it is tempting to believe that lower replication *in vivo* of vaccine or animal-derived strains infecting man might be involved [Anderson, 2008; Feng et al., 2011; Vesikari et al., 2006a].

Large vaccination trials and first analyses of national campaign efficacy indicate that both commercial vaccines adopted in Europe including Italy are highly efficacious against disease caused by the major rotavirus genotypes [Perez-Schael et al., 2007; Phua et al., 2009; Ruiz-Palacios et al., 2006; Vesikari et al., 2009; Vesikari et al., 2007; Vesikari et al., 2006b; Zaman et al., 2010]. Therefore, the finding that common rotavirus genotypes represent the large majority of circulating strains in Italy suggests that mass vaccination may be expected to have a significant impact on rotavirus acute gastroenteritis also in this country, as reported elsewhere. Whether vaccine efficacy will be different in different areas of Italy as a consequence of the local genotype diversities observed in this study cannot be predicted at present.

The persistence of the diversity of genotypes observed suggests *per se* that the situation may change easily. Novel strains carrying uncommon G and/or P genes as well as other genomic differences may emerge, as increasing exchange of people, food and animals between continents might eventually lead to importation of strains with new antigenic and virulence characteristics and particular fitness to generate epidemic or pandemic outburst of disease. Novel rotaviruses with reassorted gene segments may arise from co-infection of human or animal hosts [Gentsch et al., 2005; Martella et al., 2010]. In fact in the present study mixed infections were found in 7.6% of rotavirus patients, that may at least partially explain the 2.0% of "uncommon" rotavirus strains detected. Half of these may be derived from reassortment of "common" G and P types [Iturriza-Gomara et al., 2001], whereas the rest of them involved gene exchange from animal rotavirus strains, particularly P[6] or G[10] as
 recently shown by other authors [Banyai et al., 2009a; Banyai et al., 2009b; Banyai et al.,
 2010; Mukherjee et al., 2010; Steyer et al., 2010; Steyer et al., 2008].

The probability and outcome of mixed infections with two different rotavirus strains may depend on both the frequency of each strain co-circulating in the population, and possible interference phenomena during dual replication in the gut [Ward et al., 1996]. To explore these events, the observed numbers and genotypes of mixed infections involving common rotaviruses in Italy during the 2 seasons of 2007/08 and 2008/09 were compared with the numbers and genotypes of detected strains likely to have arisen by natural reassortment. Different types of reassortant rotaviruses were found to be grossly proportional with the numbers of observed relevant mixed infections. However, in this assessment the geographical or temporal incidence of mixed infections or natural reassortants was not considered, and thus, although logical, the correlation of low significance (P<0.05) found is not one which will be discovered with necessity. In this context it should be noted that Muhsen and colleagues [Muhsen et al., 2009] recently reported data from Israel where a significant correlation between the numbers of relevant mixed rotavirus infections and of natural reassortants likely to have emerged from them has been observed...

The occurrence of 37 cases where genotyping was unsuccessful might be explained by limited virus concentration in some samples and/or minor variation in the nucleotide sequence targeted by the PCR primers. Nonetheless, it cannot be excluded that some of these viruses, otherwise confirmed as rotaviruses by amplification of the VP6 encoding gene, might carry significantly different G and/or P specificities, and represent uncommon genotypes either deriving from animals or imported from abroad. In fact, the multiplex PCR used for human rotavirus detection does not include all primers needed to cover the complete genotype spectrum of group A rotavirus affecting animals, and is optimized for human strains [Iturriza-Gomara et al., 2010]. Supplementary study will be needed to characterize these untypable rotaviruses further, possibly by designing different primers for G or P gene amplification and/or by sequence analysis of other genes [Matthijnssens et al., 2008a; Matthijnssens et al., 2008b].

Complex gene evolution and reassortment is likely required before animal rotavirus strains transmitted zoonotically can adapt to efficient replication and transmission in humans [Martinez-Laso et al., 2009; Matthijnssens et al., 2008a]. This is possibly the reason why strains with an animal G or P type, such as G8, G12 or P[6] in the present study, did not appear to spread significantly following their first appearance. Rotaviruses with both G and P

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type of animal origin were not detected at all. For this reason, it is also hard to address the question whether humans are the host where "humanization" of animal rotavirus strains first occurs, or an intermediate animal species other than the one of origin is required for strain adaptation to humans. Some underestimation of emerging animal rotaviruses may have occurred in this study, due to either lower viral load of animal viruses compared to homologous strains during dual infection, and/or to technical issues, such as unsuitable diagnostic or genotyping primers used in normal typing routine. This is one of the reasons of the importance of international laboratory networking, such as EuroRotaNet, in facilitating implementation, optimization and sharing of state-of-the-art detection methods and construction of large virus and typing data repositories fit to identify novel rotaviruses at an early stage of expansion in the human population. ion in u.e.

ACKNOWLEDGEMENTS

This study was supported by grants from the Ministry of Health, Italy: (i) to LF: CCM "Epidemiologia molecolare di rotavirus in età pediatrica in Italia. Creazione di una rete di sorveglianza per monitorare la diffusione e l'evoluzione di genotipi virali", and (ii) to FMR: Strategic Program: "Preparedness and response to emerging zoonoses and exotic viral infection through an integrated medical and veterinary approach"; and from ISS/NIH: to FMR: "Molecular and antigenic evolution of rotavirus strains of human and animal origin". The study was also supported by and performed within the framework of EuroRotaNet (http://www.eurorota.net).

REFERENCES

- Anderson EJ. 2008. Rotavirus vaccines: viral shedding and risk of transmission. Lancet Infect
 Dis 8:642-649.
- Arista S, Giammanco GM, De Grazia S, Colomba C, Martella V, Cascio A, Iturriza-Gomara M.
 2005. G2 rotavirus infections in an infantile population of the South of Italy: variability
 of viral strains over time. J Med Virol 77:587-594.
- Banyai K, Esona MD, Mijatovic S, Kerin TK, Pedreira C, Mercado J, Balmaseda A, Perez MC,
 Patel MM, Gentsch JR. 2009a. Zoonotic bovine rotavirus strain in a diarrheic child,
 Nicaragua. J Clin Virol 46:391-393.
 - Banyai K, Martella V, Molnar P, Mihaly I, Van Ranst M, Matthijnssens J. 2009b. Genetic
 heterogeneity in human G6P[14] rotavirus strains detected in Hungary suggests
 independent zoonotic origin. J Infect 59:213-215.
- Banyai K, Papp H, Dandar E, Molnar P, Mihaly I, Van Ranst M, Martella V, Matthijnssens J.
 2010. Whole genome sequencing and phylogenetic analysis of a zoonotic human
 G8P[14] rotavirus strain. Infect Genet Evol 10:1140-1144.
- Blutt SE, Matson DO, Crawford SE, Staat MA, Azimi P, Bennett BL, Piedra PA, Conner ME.
 2007. Rotavirus antigenemia in children is associated with viremia. PLoS Med 4:e121.
 - Boom JA, Tate JE, Sahni LC, Rench MA, Hull JJ, Gentsch JR, Patel MM, Baker CJ, Parashar
 UD. 2010. Effectiveness of pentavalent rotavirus vaccine in a large urban population in
 the United States. Pediatrics 125:e199-207.
- Ciarlet M, Hoshino Y, Liprandi F. 1997. Single point mutations may affect the serotype
 reactivity of serotype G11 porcine rotavirus strains: a widening spectrum? J Virol
 71:8213-8220.
 - Clark HF, Lawley D, Mallette LA, DiNubile MJ, Hodinka RL. 2009. Decline in cases of
 rotavirus gastroenteritis presenting to The Children's Hospital of Philadelphia after
 introduction of a pentavalent rotavirus vaccine. Clin Vaccine Immunol 16:382-386.
- Clark HF, Offit PA, Plotkin SA, Heaton PM. 2006. The new pentavalent rotavirus vaccine
 composed of bovine (strain WC3)-human rotavirus reassortants. Pediatr Infect Dis J
 25:577-583.
 - Coffin SE, Moser CA, Cohen S, Clark HF, Offit PA. 1997. Immunologic correlates of protection
 against rotavirus challenge after intramuscular immunization of mice. J Virol 71:7851 7856.
 - Collins PJ, Martella V, Buonavoglia C, O'Shea H. 2010. Identification of a G2-like porcine
 rotavirus bearing a novel VP4 type, P[32]. Vet Res 41:73.
- de Oliveira LH, Danovaro-Holliday MC, Matus CR, Andrus JK. 2008. Rotavirus vaccine
 introduction in the Americas: progress and lessons learned. Expert Rev Vaccines 7:345 353.
 - de Rougemont A, Kaplon J, Pillet S, Mory O, Gagneur A, Minoui-Tran A, Meritet JF, Mollat C,
 Lorrot M, Foulongne V, Gillet Y, Nguyen-Bourgain C, Alain S, Agius G, Lazrek M,
 Colimon R, Fontana C, Gendrel D, Pothier P. 2011. Molecular and clinical
 characterization of rotavirus from diarrheal infants admitted to pediatric emergency units
 in france. Pediatr Infect Dis J 30:118-124.
- 5443Desselberger U, Huppertz HI. 2011. Immune responses to rotavirus infection and vaccination5544and associated correlates of protection. J Infect Dis 203:188-195.
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 49
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 49
 49
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 49
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 49
 49
 49
 49
 49
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 4
- 47 Desselberger U, Wolleswinkel-van den Bosch J, Mrukowicz J, Rodrigo C, Giaquinto C, Vesikari
 48 T. 2006. Rotavirus types in Europe and their significance for vaccination. Pediatr Infect
 49 Dis J 25 (Suppl 1):S30-41.

Esona MD, Geyer A, Page N, Trabelsi A, Fodha I, Aminu M, Agbaya VA, Tsion B, Kerin TK,
 Armah GE, Steele AD, Glass RI, Gentsch JR. 2009. Genomic characterization of human
 rotavirus G8 strains from the African rotavirus network: relationship to animal
 rotaviruses. J Med Virol 81:937-951.

- 5 Estes MK, Kapikian AZ. 2007. Rotaviruses. In: Knipe DM, Howley PM, Griffin DE, Lamb RA,
 6 Martin MA, Roizman B, Straus SE, editors. Fields Virology. 5th ed. Philadelphia:
 7 Lippincott Williams & Wilkins. p 1917-1974.
- Feng N, Sen A, Wolf M, Vo P, Hoshino Y, Greenberg HB. 2011. Roles of VP4 and NSP1 in
 determining the distinctive replication capacities of simian rotavirus RRV and bovine
 rotavirus UK in the mouse biliary tract. J Virol 85:2686-2694.
- Forster J, Guarino A, Parez N, Moraga F, Roman E, Mory O, Tozzi AE, de Aguileta AL, Wahn
 U, Graham C, Berner R, Ninan T, Barberousse C, Meyer N, Soriano-Gabarro M. 2009.
 Hospital-based surveillance to estimate the burden of rotavirus gastroenteritis among
 European children younger than 5 years of age. Pediatrics 123:e393-400.
- Gentsch JR, Glass RI, Woods P, Gouvea V, Gorziglia M, Flores J, Das BK, Bhan MK. 1992.
 Identification of group A rotavirus gene 4 types by polymerase chain reaction. J Clin Microbiol 30:1365-1373.
- Gentsch JR, Laird AR, Bielfelt B, Griffin DD, Banyai K, Ramachandran M, Jain V, Cunliffe
 NA, Nakagomi O, Kirkwood CD, Fischer TK, Parashar UD, Bresee JS, Jiang B, Glass
 RI. 2005. Serotype diversity and reassortment between human and animal rotavirus
 strains: implications for rotavirus vaccine programs. J Infect Dis 192 Suppl 1:S146-159.
- Giaquinto C, van Damme P, REVEAL Study Group. 2010. Age distribution of paediatric
 rotavirus gastroenteritis cases in Europe: the REVEAL study. Scand J Infect Dis 42:142 147.
- Gombold JL, Ramig RF. 1986. Analysis of reassortment of genome segments in mice mixedly
 infected with rotaviruses SA11 and RRV. J Virol 57:110-116.
- Gorrell RJ, Bishop RF. 1999. Homotypic and heterotypic serum neutralizing antibody response
 to rotavirus proteins following natural primary infection and reinfection in children. J
 Med Virol 57:204-211.
- Gouvea V, Glass RI, Woods P, Taniguchi K, Clark HF, Forrester B, Fang ZY. 1990. Polymerase
 chain reaction amplification and typing of rotavirus nucleic acid from stool specimens. J
 Clin Microbiol 28:276-282.
- Iturriza-Gomara M, Dallman T, Banyai K, Bottiger B, Buesa J, Diedrich S, Fiore L, Johansen K, Koopmans M, Korsun N, Koukou D, Kroneman A, Laszlo B, Lappalainen M, Maunula L, Marques AM, Matthijnssens J, Midgley S, Mladenova Z, Nawaz S, Poljsak-Prijatelj M, Pothier P, Ruggeri FM, Sanchez-Fauquier A, Stever A, Sidaraviciute-Ivaskeviciene I, Syriopoulou V, Tran AN, Usonis V, van Ranst M, de Rougemont A, Gray J. 2010. Rotavirus genotypes co-circulating in Europe between 2006 and 2009 as determined by EuroRotaNet, a pan-European collaborative strain surveillance network. Epidemiol Infect: 139:895-909.
- Iturriza-Gomara M, Dallman T, Banyai K, Bottiger B, Buesa J, Diedrich S, Fiore L, Johansen K, Korsun N, Kroneman A, Lappalainen M, Laszlo B, Maunula L, Matthinjnssens J, Midgley S, Mladenova Z, Poljsak-Prijatelj M, Pothier P, Ruggeri FM, Sanchez-Fauquier A, Schreier E, Stever A, Sidaraviciute I, Tran AN, Usonis V, Van Ranst M, de Rougemont A, Gray J. 2009. Rotavirus surveillance in europe, 2005-2008: web-enabled reporting and real-time analysis of genotyping and epidemiological data. J Infect Dis 200 (Suppl 1):S215-221.
- Iturriza-Gomara M, Isherwood B, Desselberger U, Gray J. 2001. Reassortment in vivo: driving force for diversity of human rotavirus strains isolated in the United Kingdom between 1995 and 1999. J Virol 75:3696-3705.

1		
2	1	Iturriza-Gomara M Kang G Grav I 2004 Rotavirus genotyping: keeping up with an evolving
3	1	nonulation of human rotaviruses. I Clin Virol 31:250 265
4 5	2	Journey II Estes MK Drosod DV 2004 Emerging themas in notavirus call entry conome
6	3	Jayarani H, Estes MK, Prasad DV. 2004. Emerging themes in fotavirus cent entry, genome
7	4	organization, transcription and replication. Virus Res 101:67-81.
8	5	Martella V, Banyai K, Matthijnssens J, Buonavoglia C, Ciarlet M. 2010. Zoonotic aspects of
9	6	rotaviruses. Vet Microbiol 140:246-255.
10	7	Martinez-Laso J, Roman A, Head J, Cervera I, Rodriguez M, Rodriguez-Avial I, Picazo JJ. 2009.
11	8	Phylogeny of G9 rotavirus genotype: a possible explanation of its origin and evolution. J
12	9	Clin Virol 44:52-57.
13	10	Matthijnssens J, Ciarlet M, Heiman E, Arijs I, Delbeke T, McDonald SM, Palombo EA, Iturriza-
14	11	Gomara M. Maes P. Patton JT. Rahman M. Van Ranst M. 2008a. Full genome-based
15	12	classification of rotaviruses reveals a common origin between human Wa-Like and
10	13	porcine rotavirus strains and human DS-1-like and hovine rotavirus strains. I Virol
18	17	82.3201 3210
19	14	02.5204-5217. Metthiingeong I. Ciarlet M. Dehmen M. Atteui H. Denvei K. Estes MK. Contech ID. Iturrize
20	15	Matunijissens J, Clanet M, Kalinan M, Atlour H, Danyar K, Estes MK, Gentsch JK, Ituniza-
21	10	Gomara M, Kirkwood CD, Martella V, Mertens PP, Nakagomi O, Patton JI, Ruggeri
22	17	FM, Saif LJ, Santos N, Steyer A, Taniguchi K, Desselberger U, Van Ranst M. 2008b.
23	18	Recommendations for the classification of group A rotaviruses using all 11 genomic
24	19	RNA segments. Arch Virol 153:1621-1629.
25	20	Maunula L, Von Bonsdorff CH. 2002. Frequent reassortments may explain the genetic
26	21	heterogeneity of rotaviruses: analysis of Finnish rotavirus strains. J Virol 76:11793-
27	22	11800.
20 20	23	Medici MC, Martinelli M, Abelli LA, Ruggeri FM, Di Bartolo I, Arcangeletti MC, Pinardi F, De
30	24	Conto F. Izzi G. Bernasconi S. Chezzi C. Dettori G. 2006. Molecular epidemiology of
31	25	norovirus infections in sporadic cases of viral gastroenteritis among children in Northern
32	25	Italy I Med Virol 78:1486 1402
33	20	Muhaan K. Shulman I. Dubinatain II. Kasam E. Kramar A. Garan S. Zilharstain I. Chadiak G.
34	27	Falses M. Calar D. 2000. Incidence about victime and account hunder of actuations
35	28	Ephros M, Conen D. 2009. Incidence, characteristics, and economic burden of rotavirus
36	29	gastroenteritis associated with hospitalization of israeli children <5 years of age, 2007-
37	30	2008. J Infect Dis 200 (Suppl 1):S254-263.
38	31	Mukherjee A, Chattopadhyay S, Bagchi P, Dutta D, Singh NB, Arora R, Parashar UD, Gentsch
39	32	JR, Chawla-Sarkar M. 2010. Surveillance and molecular characterization of rotavirus
40 41	33	strains circulating in Manipur, north-eastern India: increasing prevalence of emerging
42	34	G12 strains. Infect Genet Evol 10:311-320.
43	35	Offit PA, Blavat G. 1986. Identification of the two rotavirus genes determining neutralization
44	36	specificities. J Virol 57:376-378.
45	37	Parashar UD, Alexander JP, Glass RL 2006, Prevention of rotavirus gastroenteritis among
46	38	infants and children Recommendations of the Advisory Committee on Immunization
47	30	Practices (ACIP) MMWR Recomm Rep 55:1-13
48	40	Derre CL Espinole EE 2006 Nucleotide migratches between the VD7 gone and the primer are
49 50	40	rana OI, Espinola EE. 2000. Nucleotide mismatches between the vr7 gene and the printer are
50 51	41	associated with genotyping failure of a specific fineage from G1 rotavirus strains. Viroi J
52	42	3:35.
53	43	Perez-Schael I, Salinas B, Tomat M, Linhares AC, Guerrero ML, Ruiz-Palacios GM,
54	44	Bouckenooghe A, Yarzabal JP. 2007. Efficacy of the human rotavirus vaccine RIX4414
55	45	in malnourished children. J Infect Dis 196:537-540.
56	46	Pesavento JB, Crawford SE, Estes MK, Prasad BV. 2006. Rotavirus proteins: structure and
57	47	assembly. Curr Top Microbiol Immunol 309:189-219.
58	48	Phua KB, Lim FS, Lau YL, Nelson EA, Huang LM, Quak SH, Lee BW, Teoh YL, Tang H,
59	49	Boudville I, Oostvogels LC, Survakiran PV, Smolenov IV. Han HH. Bock HL. 2009.
60	50	Safety and efficacy of human rotavirus vaccine during the first 2 years of life in Asian
	51	infants: randomised, double-blind, controlled study. Vaccine 27:5936-5941
		· ····································

Ruggeri FM, Declich S. 1999. Rotavirus infection among children with diarrhoea in Italy. Acta
 Paediatr Suppl 88:66-71.

- Ruiz-Palacios GM, Perez-Schael I, Velazquez FR, Abate H, Breuer T, Clemens SC, Cheuvart B,
 Espinoza F, Gillard P, Innis BL, Cervantes Y, Linhares AC, Lopez P, Macias-Parra M,
 Ortega-Barria E, Richardson V, Rivera-Medina DM, Rivera L, Salinas B, Pavia-Ruz N,
 Salmeron J, Ruttimann R, Tinoco JC, Rubio P, Nunez E, Guerrero ML, Yarzabal JP,
 Damaso S, Tornieporth N, Saez-Llorens X, Vergara RF, Vesikari T, Bouckenooghe A,
 Clemens R, De Vos B, O'Ryan M. 2006. Safety and efficacy of an attenuated vaccine
 against severe rotavirus gastroenteritis. N Engl J Med 354:11-22.
 - Santos N, Hoshino Y. 2005. Global distribution of rotavirus serotypes/genotypes and its
 implication for the development and implementation of an effective rotavirus vaccine.
 Rev Med Virol 15:29-56.
 - Santos N, Volotao EM, Soares CC, Albuquerque MC, da Silva FM, Chizhikov V, Hoshino Y.
 2003. VP7 gene polymorphism of serotype G9 rotavirus strains and its impact on G
 genotype determination by PCR. Virus Res 93:127-138.
- Shaw RD, Mackow ER, Dyall-Smith ML, Lazdins I, Holmes IH, Greenberg HB. 1988. Serotypic
 analysis of VP3 and VP7 neutralization escape mutants of rhesus rotavirus. J Virol
 62:3509-3512.
 - Simmonds MK, Armah G, Asmah R, Banerjee I, Damanka S, Esona M, Gentsch JR, Gray JJ,
 Kirkwood C, Page N, Iturriza-Gomara M. 2008. New oligonucleotide primers for P typing of rotavirus strains: Strategies for typing previously untypeable strains. J Clin
 Virol 42:368-373.
 - Solberg OD, Hasing ME, Trueba G, Eisenberg JN. 2009. Characterization of novel VP7, VP4,
 and VP6 genotypes of a previously untypeable group A rotavirus. Virology 385:58-67.
 - Song XF, Hao Y. 2009. Adaptive evolution of rotavirus VP7 and NSP4 genes in different
 species. Comput Biol Chem 33:344-349.
 - Soriano-Gabarro M, Mrukowicz J, Vesikari T, Verstraeten T. 2006. Burden of rotavirus disease
 in European Union countries. Pediatr Infect Dis J 25 (Suppl 1):S7-S11.
- Steyer A, Bajzelj M, Iturriza-Gomara M, Mladenova Z, Korsun N, Poljsak-Prijatelj M. 2010.
 Molecular analysis of human group A rotavirus G10P[14] genotype in Slovenia. J Clin
 Virol 49:121-125.
 - Steyer A, Poljsak-Prijatelj M, Barlic-Maganja D, Marin J. 2008. Human, porcine and bovine
 rotaviruses in Slovenia: evidence of interspecies transmission and genome reassortment.
 J Gen Virol 89:1690-1698.
- To TL, Ward LA, Yuan L, Saif LJ. 1998. Serum and intestinal isotype antibody responses and
 correlates of protective immunity to human rotavirus in a gnotobiotic pig model of
 disease. J Gen Virol 79:2661-2672.
- Trojnar E, Otto P, Johne R. 2009. The first complete genome sequence of a chicken group A
 rotavirus indicates independent evolution of mammalian and avian strains. Virology
 386:325-333.
- Turcios RM, Curns AT, Holman RC, Pandya-Smith I, LaMonte A, Bresee JS, Glass RI. 2006.
 Temporal and geographic trends of rotavirus activity in the United States, 1997-2004.
 Pediatr Infect Dis J 25:451-454.
- Vesikari T, Clark HF, Offit PA, Dallas MJ, DiStefano DJ, Goveia MG, Ward RL, Schodel F,
 Karvonen A, Drummond JE, DiNubile MJ, Heaton PM. 2006a. Effects of the potency
 and composition of the multivalent human-bovine (WC3) reassortant rotavirus vaccine
 on efficacy, safety and immunogenicity in healthy infants. Vaccine 24:4821-4829.

Journal of Medical Virology

- Vesikari T, Itzler R, Karvonen A, Korhonen T, Van Damme P, Behre U, Bona G, Gothefors L,
 Heaton PM, Dallas M, Goveia MG. 2009. RotaTeq, a pentavalent rotavirus vaccine:
 efficacy and safety among infants in Europe. Vaccine 28:345-351.
- Vesikari T, Karvonen A, Prymula R, Schuster V, Tejedor JC, Cohen R, Meurice F, Han HH,
 Damaso S, Bouckenooghe A. 2007. Efficacy of human rotavirus vaccine against
 rotavirus gastroenteritis during the first 2 years of life in European infants: randomised,
 double-blind controlled study. Lancet 370:1757-1763.
- Vesikari T, Matson DO, Dennehy P, Van Damme P, Santosham M, Rodriguez Z, Dallas MJ, Heyse JF, Goveia MG, Black SB, Shinefield HR, Christie CD, Ylitalo S, Itzler RF, Coia ML, Onorato MT, Adeyi BA, Marshall GS, Gothefors L, Campens D, Karvonen A, Watt JP, O'Brien KL, DiNubile MJ, Clark HF, Boslego JW, Offit PA, Heaton PM. 2006b. Safety and efficacy of a pentavalent human-bovine (WC3) reassortant rotavirus vaccine. N Engl J Med 354:23-33.
 - Ward RL, Jin Q, Nakagomi O, Sander DS, Gentsch JR. 1996. Isolation of a human rotavirus
 containing a bovine rotavirus VP4 gene that suppresses replication of other rotaviruses in
 coinfected cells. Arch Virol 141:615-633.
 - Watanabe M, Nakagomi T, Koshimura Y, Nakagomi O. 2001. Direct evidence for genome
 segment reassortment between concurrently-circulating human rotavirus strains. Arch
 Virol 146:557-570.
 - World Health Organization. 2007. Rotavirus vaccines. WHO position paper. Wkly Epidemiol
 Rec 82:285-295.
 - Yuan L, Azevedo MS, Gonzalez AM, Jeong KI, Van Nguyen T, Lewis P, Iosef C, Herrmann JE,
 Saif LJ. 2005. Mucosal and systemic antibody responses and protection induced by a
 prime/boost rotavirus-DNA vaccine in a gnotobiotic pig model. Vaccine 23:3925-3936.
 - Yuan L, Honma S, Kim I, Kapikian AZ, Hoshino Y. 2009. Resistance to rotavirus infection in
 adult volunteers challenged with a virulent G1P1A[8] virus correlated with serum
 immunoglobulin G antibodies to homotypic viral proteins 7 and 4. J Infect Dis 200:1443 1451.
 - Zaman K, Dang DA, Victor JC, Shin S, Yunus M, Dallas MJ, Podder G, Vu DT, Le TP, Luby SP, Le HT, Coia ML, Lewis K, Rivers SB, Sack DA, Schodel F, Steele AD, Neuzil KM, Ciarlet M. 2010. Efficacy of pentavalent rotavirus vaccine against severe rotavirus gastroenteritis in infants in developing countries in Asia: a randomised, double-blind, placebo-controlled trial. Lancet 376:615-623.

LEGEND TO FIGURES

Figure 1. Distribution of rotavirus acute gastroenteritis cases in the North, Center and South of Italy, by month of admission. Regions enclosed in groups are indicated. Date of onset of rotavirus diarrhea was available for a 2,377 subjects. Columns indicate the sum of cases for all 3 years of investigation.

Annex 1:

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