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Foraging ecology of Mediterranean fin whales in a changing environment elucidated by satellite tracking and baleen plate stable isotopes

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ABSTRACT: We investigated seasonal shifts in diet and distribution of fin whales *Balaenoptera physalus* occurring in the western Mediterranean Sea. For this purpose, we combined carbon and nitrogen stable isotope ratios ($\delta^{13}\text{C}$, $\delta^{15}\text{N}$) along 10 baleen plates collected from stranded fin whales between 1975 and 2002 with satellite tag deployments on 11 fin whales during summer 2003. Baleen plate stable isotopes were compared with those of the krill *Meganyctiphanes norvegica*, the main prey of fin whales in the northwestern Mediterranean Sea. Two plates collected near Malaga, Spain, exhibited larger $\delta^{13}\text{C}$ variations, while only smaller variations could be detected in the other 8. While all mean baleen plate results were consistent with the $\delta^{13}\text{C}$ signature of Mediterranean *M. norvegica*, the most depleted $\delta^{13}\text{C}$ values were intermediate between those of Atlantic and Mediterranean *M. norvegica*, suggesting westward migrations perhaps extending to the Strait of Gibraltar but not extensive, prolonged feeding in the Northeast Atlantic. This pattern was confirmed by satellite tracking; 1 out of 8 fin whales we successfully tracked left the Mediterranean for the Atlantic. Longer-term changes in isotopic signatures of baleen plates exhibited significant depletion trends, indicating that changes due to increasing input of nutrients and anthropogenic carbon are occurring in the western Mediterranean Sea ecosystem.

KEY WORDS: *Meganyctiphanes norvegica* · Mediterranean Sea · Foraging · Migration · Satellite telemetry · Environmental change

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INTRODUCTION

Fin whales *Balaenoptera physalus* are the largest marine mammals in the Mediterranean Sea (Notarbartolo di Sciara et al. 2003), with an estimated population of 3500 individuals. *B. physalus* feed on the krill *Meganyctiphanes norvegica* (euphausiacea) as

revealed both by feces (Orsi Relini & Giordano 1992) and fatty acid (Ruchonnet et al. 2006) analyses. They gather during summer in the northern part of the western basin (Forcada et al. 1996, Monestiez et al. 2006, Cotté et al. 2009), where they are exposed to shipping collisions, a major threat for the species (Panigada et al. 2006) particularly in the Mediterra-

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nean, which accounts for 30% (in Notarbartolo di Sciarra et al. 2003) of the world's merchant shipping covering only 0.8% of the global ocean surface. Information about their population distribution outside the summer period is lacking, yet it is needed to develop mitigation measures for the conservation of the species. Mediterranean fin whales are genetically distinct from the other north Atlantic populations (Bérubé et al. 1998). The degree of contact between Mediterranean and Atlantic fin whale populations has been debated for the last 2 centuries, and the prevailing idea is that the stocks are geographically isolated (Notarbartolo di Sciarra et al. 2003). Recently, from fin whale sighting and satellite tracking, Cotté et al. (2008) suggested a regional fidelity to the northwestern Mediterranean Sea. Sightings have been recorded in shallow water around Lampedusa Island in winter (Canese et al. 2006), and Aïssi et al. (2008) suggested a probable year-round distribution along synchronic seasonal dispatching, with a feeding behaviour occurrence in the Ligurian Sea during late spring and summer and in the Lampedusa area from late winter to early spring. However, the results are still insufficient to prove year-round residency of a large portion of the fin whale Mediterranean population.

In the present study, the distribution of fin whales was investigated by monitoring movement using satellite tags deployed on whales present in the northwestern Mediterranean Sea during summer. This approach was complemented by the measurements of stable isotopes in fin whale tissues, which can be a powerful complement for investigating foraging behaviour, not only in terms of diet but also in terms of movement of marine predators (Kelly 2000, Bearhop et al. 2004, Bailleul et al. 2010). Recently developed methods of diet analysis (e.g. lipids and stable isotopes) are increasingly used to investigate the feeding habits of predator species. The stable isotope ratio for an animal's tissue is often related to that of its diet, with a difference due to the trophic isotopic enrichment factor between dietary and consumer tissue (DeNiro & Epstein 1978, 1981). Consumer tissues are enriched in ^{15}N relative to their food, and, consequently, $\delta^{15}\text{N}$ measurements serve as indicators of a consumer trophic position (McCutchan et al. 2003, Vanderklift & Ponsard 2003) integrated over months while stomach contents or feces analysis are generally only representative of the very last foraging areas visited. Pioneering studies of DeNiro & Epstein (1981), Minagawa & Wada (1984), and Peterson & Fry (1987) suggested that consumers become enriched in ^{15}N relative to their food by 3 to 4‰. However a num-

ber of studies reveal a wider range of isotopic ^{15}N enrichment between trophic levels (1.7 to 4.5 ‰; Minagawa & Wada 1984, Abend & Smith 1997, Post 2002). A mean 3.4‰ enrichment value was used by Pinnegar et al. (2003) in their exhaustive western Mediterranean fishery landing study.

In contrast, small increases of $\delta^{13}\text{C}$ between prey and predators are reported in the literature (<2 ‰, mean 1‰) (De Niro & Epstein 1978, Wada et al. 1991) providing clues about the ultimate origin of their food. $\delta^{13}\text{C}$ values of organic tissue along food chain are mainly used to assess consumer foraging areas, i.e. inshore versus offshore, pelagic versus benthic, or latitudinal variations in the contribution to food intake (Kelly 2000, Cherel & Hobson 2007). The origin of the latitudinal depletion of marine organic matter $\delta^{13}\text{C}$ with increasing latitude has been mainly attributed to changes in the concentration of aqueous CO_2 (Rau et al. 1989, Bentaleb et al. 1998). These regional and latitudinal $\delta^{13}\text{C}$ gradients have been also used as an effective way for investigating seasonal change in the foraging areas of marine predators (Cherel & Hobson 2007, Cherel et al. 2007).

The whale baleen plates are a particularly suitable tissue to investigate the temporal change of the $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ due to their continuous growth throughout lifetime (Hobson et al. 2004). The baleen plates grow down from the gums of the upper jaw and are arranged in rows that extend down each side of the mouth. The terminal end continually wears off. The baleens composed of keratinous protein are metabolically inactive after formation (Schell et al. 1989a). Thus, their isotopic composition is believed to record the diet of the whales through its synthesis. Schell et al. (1989a,b) measured stable isotope ratios in the baleen plates of bowhead whales *Balaena mysticetus* and found isotopic oscillations along the length of the baleen plates. Similar oscillations were also found in the baleen of southern right whales *Eubalaena australis* (Best & Schell 1996). The technique was also successfully applied to investigate seasonal shift in diet of Minke whales off Japan (Mitani et al. 2006) and long-term average diet (Hobson et al., 2004).

The present study focuses on the Mediterranean *Balaenoptera physalus* (hereafter fin whale). The main objectives of our study were to assess seasonal movements or dietary shift of this population and their movements inside the Mediterranean Sea by combining the information provided by: (1) satellite tracking of individual fin whales marked in the Mediterranean Sea at the end of summer, and (2) analyses of $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ signatures along baleen

plates collected from stranded fin whales in the western Mediterranean, and of krill *Meganyctiphanes norvegica* (hereafter krill) sampled in the Mediterranean sea and in the Atlantic ocean.

We assert that the isotopic composition of the krill consumed throughout the fin whale lifetime is preserved. Whenever Mediterranean fin whales migrate, or shift preys, they should exhibit corresponding changes in the isotopic signature along their plates. Stable isotope values should be different between different prey or between the Mediterranean Sea and the Atlantic Ocean. Finally, from plates collected from the 1970s to the 2000s, we tried to detect whether $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ stable isotope signatures shift over time, revealing some large-scale and long-term environmental change within the Mediterranean Sea.

MATERIALS AND METHODS

Sampling of materials

Baleen plates are made of Keratin. They grow from the gums of the upper jaw and are arranged in rows that extend down each side of the mouth. Baleen grows throughout the whale's lifetime; the terminal end continually wears off. The outer (outside-facing)

edge of each plate is smooth, while the inner edge is frayed (see Fig. 1). The frayed edge of each plate intertwines to form a mat, which functions as a strainer during filter feeding.

A total of 10 baleen plates (BP) was collected over the period 1975 to 2002 from 9 fin whales stranded along the western Mediterranean coast. Eight BPs were collected on the French coast (BP1 to BP7 and BP9) and 2 in the Malaga area and the Strait of Gibraltar, Andalusia, Spain (BPs 8 and 10). BPs 3 and 4 were collected from the same individual stranded in Port-La-Nouvelle (France). BP3 was collected from the right upper jaw, whereas BP4 was taken in the anterior part (see Fig. 1, Table 1).

Krill from the Atlantic Ocean (between 38 and 45°N and 12 and 13°W) was sampled during the POMME 2 and 3 cruises in March and September 2001 using a mid-water trawl. Stable isotopic study concerned a total of 12 adult and sub-adult individuals (6 each for spring and fall), and the 6 adults were sexed (4 males and 2 females; Table 2). Samples were immediately frozen at -80°C (see Salomon et al. 2000) until laboratory analysis. For the Mediterranean Sea, we used stable isotopic data and carbon:nitrogen ratio (C:N) of 12 adult specimens (6 each from spring and fall) of krill sampled in the Gulf of Lion (Ferraton 2007; Table 3).

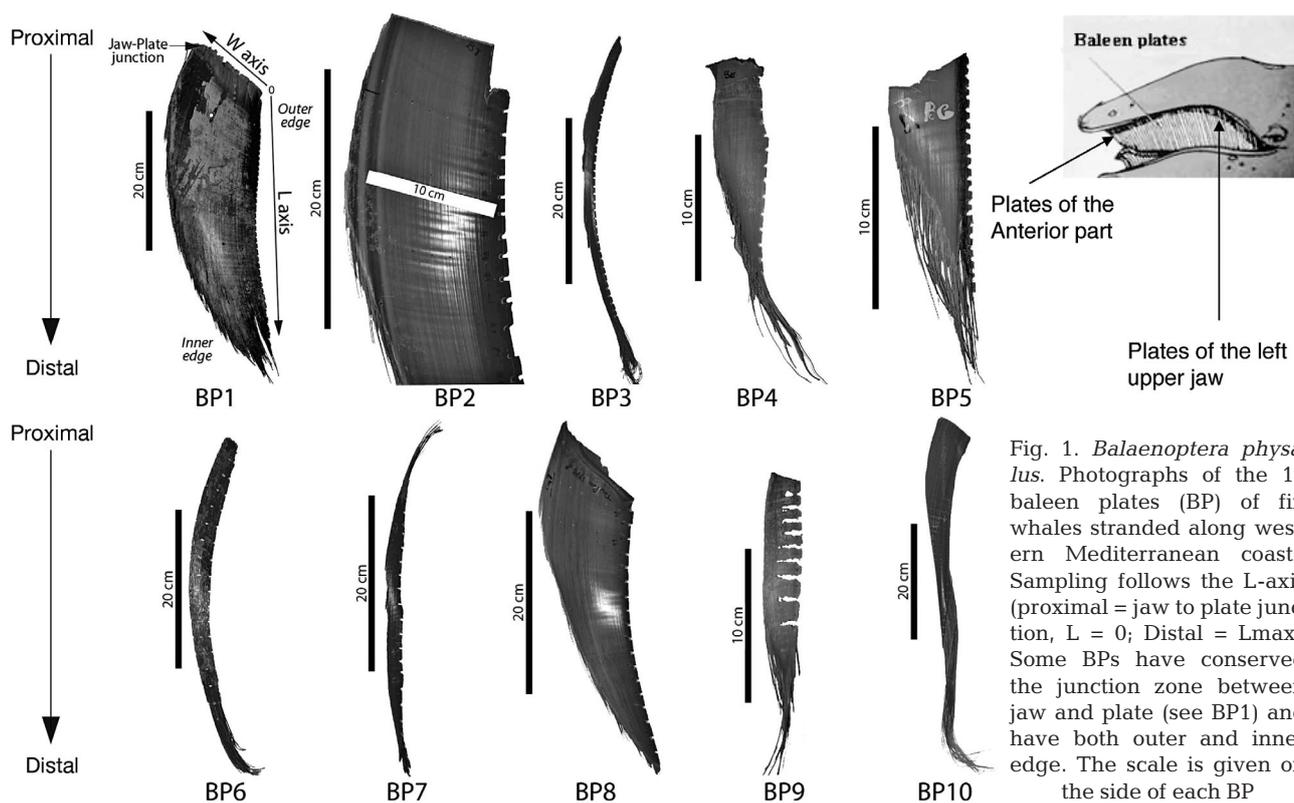


Fig. 1. *Balaenoptera physalus*. Photographs of the 10 baleen plates (BP) of fin whales stranded along western Mediterranean coasts. Sampling follows the L-axis (proximal = jaw to plate junction, L = 0; Distal = Lmax). Some BPs have conserved the junction zone between jaw and plate (see BP1) and have both outer and inner edge. The scale is given on the side of each BP

Table 1. *Balaenoptera physalus*. Sampling data, carbon (C) and nitrogen (N) stable isotope and C:N (see Fig. 1) ratios of 10 fin whale baleen plates (BP). BP3 and 4 were from the same individual but sampled at lateral and frontal sides of the jaw. All samples were measured along W = 0 axis except for BP6, which was measured as W = 2 cm. ID: identification number; SD: standard deviation; F: female; M: male; A: adult; J: juvenile; nd: no data

ID	Year	Collection Location	Sex	Age class	$\delta^{13}\text{C}$ (‰)				$\delta^{15}\text{N}$ (‰)				C:N	SD	No. of samples per BP	Length of the L-axis (cm)		
					Mean	SD	Min	Max	Range	Mean	SD	Min					Max	Range
BP1	1975	Toulon, France	nd	A	-18.0	0.2	-18.6	-17.7	0.8	9.4	0.2	8.9	10.0	1.0	3.4	0.1	17	31.5
BP2	1980	Toulon, France	nd	A	-17.4	0.4	-18.0	-17.0	1.0	8.1	0.2	7.9	8.3	0.5	3.5	0.1	7	38.0
BP3	1989	Port La Nouvelle, France	F	A	-18.0	0.3	-18.5	-17.5	1.0	7.2	0.4	6.4	7.9	1.6	3.4	0.1	30	32.2
BP4	1989	Port La Nouvelle, France	F	A	-18.1	0.4	-18.6	-17.5	1.1	7.7	0.3	7.4	8.3	0.9	3.4	0.1	9	14.1
BP5	1990	Port La Nouvelle, France	nd	J	-18.3	0.2	-18.7	-17.9	0.8	7.2	0.3	6.7	7.6	1.0	3.3	0.1	10	11.4
BP6	1992	Port La Nouvelle, France	nd	nd	-18.3	0.3	-18.7	-17.8	1.0	7.5	0.4	6.7	8.1	1.4	3.5	0.1	17	33.8
BP7	1993	Port La Nouvelle, France	nd	nd	-18.3	0.2	-18.7	-18.1	0.6	7.7	0.4	6.8	8.2	1.3	3.4	0.1	12	34.0
BP8	1996	Malaga, Spain	nd	A	-18.5	0.4	-19.3	-18.0	1.3	6.8	0.2	6.3	7.2	0.9	3.6	0.1	32	33.9
BP9	2000	Toulon, France	M	A	-18.0	0.2	-18.3	-17.5	0.8	7.0	0.4	6.4	7.8	1.5	3.4	0.0	12	9.7
BP10	2002	Malaga, Spain	nd	A	-18.3	0.4	-19.2	-17.4	1.9	7.7	0.4	6.5	8.7	2.2	3.4	0.1	44	51.5

Stable isotope analyses of fin whale baleen plates and krill

Baleen plates were rinsed with distilled water and dried in an oven at 50°C. We used a dental drill to collect the keratin powder along the L-axis from the proximal part (below the gum line) to the most distal, i.e. oldest part of the plates at an interval of ~1 cm so as to represent an approximately constant time interval. The drilled area was 2 to 3 mm in diameter. The sampling took place at the outer edge (W = 0) for most of the plates except BP6 (W = 2 cm). The plates are generally broken in different areas such as BPs1, 2, 7, and 8, for which an undetermined and probably significant top portion is missing (Fig. 1).

The powder (0.5 to 1.0 mg) was collected in a glass tube previously washed with hydrochloric acid and rinsed with distilled water, and dried at 60°C for 48 h. The stable-carbon and stable-nitrogen isotope assays and the C and N contents were performed in 2008 at the Institut des Sciences de l'Evolution de Montpellier Laboratory. About 70 to 120 µg of homogenized powder samples were loaded into tin cups and combusted at 1200°C in a Euro Vector EA3000 elemental analyzer. Resultant CO₂ and N₂ gases were analyzed by continuous flow using an interfaced GV Optima Isotope Ratio Mass Spectrometer (IRMS). $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values are expressed in δ notation relative to V-PDB belemnite and atmospheric N₂ (air), respectively. The δ notation represents the deviation from standards in parts per thousand (‰):

$$\delta^{13}\text{C} \text{ or } \delta^{15}\text{N} = (R_{\text{sample}}/R_{\text{standard}} - 1) \times 1000 \quad (1)$$

where $R = {}^{13}\text{C}/{}^{12}\text{C}$ or ${}^{15}\text{N}/{}^{14}\text{N}$. Replicate $\delta^{15}\text{N}$ measurements of both potassium nitrate ($n = 36$; IAEA-NO-3- $\delta^{15}\text{N}$: 4.7‰) and alanine (ISEM internal standard unknown origin; $n = 41$; alanine- $\delta^{15}\text{N}$: -0.5‰) indicate a reproducibility better than 0.2‰ and an accuracy of 0.1‰. Both accuracy and reproducibility of the $\delta^{13}\text{C}$ Nist-8541 graphite (also known as USGS-24 with a $\delta^{13}\text{C}$ of -16.1‰; $n = 34$) are 0.3‰. The alanine $\delta^{13}\text{C}$ accuracy was quite poor (0.5‰), but the reproducibility was 0.2‰. Using the C and N contents of the IAEA-NO-3 (N% = 13), and Alanine (C% = 40%), the baleen plate C and N contents (%) were measured with a precision of 1%.

Whole Atlantic krill individual samples were lyophilized, powdered using a mortar pestel and sieved through a 60 µm mesh. We did not remove the chitin assuming that the small content (3.7 to 5% of krill dry weight (DW) according to Raymond et al. 1971) will not significantly affect the isotopic value of the whole individual. Moreover it allowed us to compare

Table 2. *Meganyctiphanes norvegica*. $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and C:N values of 12 bulk and 12 lipid free samples of Atlantic krill. Spring, fall, and annual values are mean \pm SD. F: female, M: male; Sub: sub-adult

Sample	Collection date (mo/yr)	n	Bulk samples			Lipid-free samples		
			$\delta^{13}\text{C}$	$\delta^{15}\text{N}$	C:N	$\delta^{13}\text{C}$	$\delta^{15}\text{N}$	C:N
1753_5F	03/2001	1	-20.9	5.3	3.8	-20.9	6.3	3.5
1753_6F	03/2001	1	-21.4	4.5	3.9	-21.2	5.5	3.7
1755_1M	03/2001	1	-21.5	5.7	3.7	-21.4	6.0	3.6
1755_2M	03/2001	1	-20.7	6.8	3.5	-20.6	7.1	3.5
1755_3M	03/2001	1	-21.2	5.9	3.9	-20.8	6.5	3.5
1755_4M	03/2001	1	-20.7	6.6	3.5	-20.7	6.7	3.5
1754_SUB6	09/2001	1	-21.3	7.1	3.9	-21.0	7.1	3.6
1756_SUB1	09/2001	1	-21.1	7.8	3.6	-21.2	7.8	3.6
1756_SUB2	09/2001	1	-21.2	8.4	4.0	-20.9	8.6	3.7
1756_SUB3	09/2001	1	-21.4	7.2	3.9	-21.2	7.3	3.7
1756_SUB4	09/2001	1	-20.9	7.4	4.0	-20.9	7.3	4.0
1756_SUB5	09/2001	1	-21.7	6.9	3.8	-21.4	7.2	3.5
Spring mean		6	-21.1	5.8	3.7	-20.9	6.3	3.5
\pm SD			± 0.4	± 0.9	± 0.2	± 0.3	± 0.6	± 0.1
Fall mean		6	-21.3	7.4	3.8	-21.1	7.5	3.7
\pm SD			± 0.3	± 0.6	0.2	± 0.2	± 0.6	± 0.2
Annual mean		12	-21.2	6.6	3.8	-21.0	6.9	3.6
\pm SD			± 0.3	± 1.1	± 0.2	± 0.3	± 0.8	± 0.1

Table 3. *Meganyctiphanes norvegica*. Spring, fall, and annual values of $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and C:N (mean \pm SD) of Atlantic Ocean (AO) and Mediterranean Sea (MS) (Ferraton 2007) bulk krill samples

Origin, year	Season	n	$\delta^{13}\text{C}$	$\delta^{15}\text{N}$	C/N
AO, 2001 (present study)	Spring	6	-21.1 ± 0.4	5.8 ± 0.9	3.7 ± 0.2
	Fall	6	-21.3 ± 0.3	7.4 ± 0.6	3.8 ± 0.2
	Annual	12	-21.2 ± 0.3	6.6 ± 1.1	3.8 ± 0.2
MS, 2004 (Ferraton 2007)	Spring	6	-19.0 ± 0.4	4.1 ± 0.5	5.6 ± 0.4
	Fall	6	-19.7 ± 0.2	5.0 ± 0.2	5.7 ± 0.1
	Annual	12	-19.3 ± 0.5	4.4 ± 0.6	5.6 ± 0.2

Atlantic krill isotopic composition values to those of the Mediterranean Sea analysed with chitin intact (Ferraton 2007). The isotopic measurements were performed in the laboratory of Littoral Environnement et Sociétés (LIENS). Precision of the acetanilide is 0.2‰ for ^{13}C and ^{15}N , and $\sim 1\%$ for the C and N elemental content. We did not proceed to a cross-calibration of standards. Instead we compared the isotopic results of the 12 bulk Atlantic krill powders analyzed at an interval time of 4 yr in 2009 at LIENS and 2005 at ISEM laboratories. Results show very consistent $\delta^{13}\text{C}$ values (differences $< 0.1\%$), while ISEM $\delta^{15}\text{N}$ values were consistently depleted by 0.6‰ compared to LIENS. This difference does not affect our interpretation.

For the Mediterranean Sea we used stable isotope and C:N ratio data for spring and fall Mediterranean krill from Ferraton (2007). We assumed that Ferraton's data were representative endmember of $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ compositions for the Mediterranean fall and spring krill. High lipid concentration can mask a large part of the $\delta^{13}\text{C}$ isotopic signature because lipids are ^{13}C depleted (6 to 8‰) compared with proteins and carbohydrates (De Niro & Epstein 1978, McConnaughey & McRoy 1979). We tested the effect of the lipid content by comparing the 12 Atlantic Ocean krill C and N isotopic compositions measured before and after the lipid extraction treatment using cyclohexane. Available Mediterranean krill isotopic data concerns only non-lipid extracted samples.

Satellite tracking

Semi-implantable tags (26 cm in length \times 1.9 cm in diameter) consisted of a Telonics Argos ST-15 transmitter in a stainless steel tube, incorporating a flexible 12.5 cm whip-antenna, a flexible 4 cm saltwater switch (SWS), and 2 solid flanges (0.9×1.5 cm) to prevent inward migration. Half of the anterior housing was coated with 2.5 g of Gentamycin sulfate antibiotic in methacrylate (Eudragrit), designed for extended time-release. The tag was applied to the fin whale using an air-powered applicator (Heide-Jørgensen et al. 2001) with 7 bar pressure from an el-

evated 1 m bow pulpit at a distance of < 3 m from a 6 m outboard-powered boat. Tags were deployed within 2 m of the dorsal fin frontal part. To conserve batteries and extend tag operation, tags transmitted for only 4 periods of 1 h daily for 90 d and then every fourth day. A saltwater switch prevented the tag from transmitting when underwater. At the surface, transmissions were programmed to coincide with optimal satellite coverage within the western Mediterranean Sea.

Statistics

Data were analyzed statistically using the R language (R Development Core Team 2010). Isotopic

ratios were means \pm SD, and statistical tests regarding potential differences in variances or means for different sub-groups were performed using Fisher-Snedecor and Student's *t*-test or Welsh's test (e.g. see Wilks 2006), respectively, at a 95% level of significance ($\alpha = 0.05$). Statistical analyses performed on BPs in the sections that follow refer always to BPs with $W = 0$, hence excluding BP6 ($W = 2$ cm).

RESULTS

Stable isotopes of krill

Bulk powders of 12 Atlantic krill individuals sampled in spring ($n = 6$; adults) and fall ($n = 6$; subadults) were analyzed for C and N stable isotopes and C and N contents (%) before and after lipid removal by chemical treatment (Table 2). Sample sizes were too small to parse out class age or seasonal effect. Hence seasonal effects cannot be independently studied for the Atlantic krill data.

Using a dependent *t*-test for paired samples, carbon stable isotope values (mean \pm SD, $n = 12$) before ($-21.2 \pm 0.3\text{‰}$) and after ($-21.0 \pm 0.3\text{‰}$) chemical treatment were statistically significantly different ($t = 3.1$, $p < 0.01$). For the nitrogen isotopic values, significant differences between none lipid extracted ($6.6 \pm 1.1\text{‰}$) and lipid free samples ($6.9 \pm 0.8\text{‰}$) were found ($t = 3.06$, $p < 0.05$). These statistical analyses suggest an effect of the chemical treatment for carbon and nitrogen isotopic composition of the krill. These delipidation treatments have been discussed in Kiljueinen et al. (2006) and Post et al. (2007).

Lipid removal does cause slight and statistically significant changes in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values for whole-body krill. However, on the absolute scale the differences in $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ between lipid extracted and bulk krill are small, by 0.32 and 0.15‰, respectively, on average. Therefore in order to allow direct comparison with the results provided by Ferraton (2007) for Mediterranean krill, bulk krill values were used, for which seasonal $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ differences were reported (Table 3). Mean $\delta^{13}\text{C}$ is higher in spring ($-19.0 \pm 0.4\text{‰}$) than during fall ($-19.7 \pm 0.2\text{‰}$), and $\delta^{15}\text{N}$ mean increases from $4.1 \pm 0.5\text{‰}$ in spring to $5.0 \pm 0.1\text{‰}$ in fall.

Significant differences were found between bulk krill sampled in the Atlantic Ocean (AO) and the Mediterranean Sea (MS) at the annual scale for both the $\delta^{13}\text{C}$ (MS: $-19.3 \pm 0.5\text{‰}$; AO: $-21.2 \pm 0.3\text{‰}$; $t = -6.53$, $p < 0.01$) and $\delta^{15}\text{N}$ (MS: $4.4 \pm 0.6\text{‰}$, AO: $6.6 \pm 1.1\text{‰}$, $t = 7.36$, $p < 0.01$).

Assuming enrichments of $\sim 1\text{‰}$ ^{13}C and 3.4‰ ^{15}N per trophic level (De Niro & Epstein 1978, 1981) the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ annual means for a fin whale feeding on krill in the MS or in the AO would be $-18.3 \pm 0.5\text{‰}$; $7.8 \pm 0.6\text{‰}$ and $-20.2 \pm 0.3\text{‰}$; $10 \pm 1.1\text{‰}$, respectively.

Stable isotopes of fin whale baleen plates

The isotopic variations in the 10 baleen plates, representing 190 pairs of $\delta^{13}\text{C}$ to $\delta^{15}\text{N}$ analyses and C:N ratios along the L-axis of the baleen plates are reported in Table 4. The mean and SD of all 190 C:N ratios for all BPs across all baleen lengths is 3.4 ± 0.1 (Table 4), which is in agreement with the theoretical C:N atomic ratio of keratin (O'Connell & Hedges 1999).

Among all 190 baleen samples and across all lengths, $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values ranged between -19.3 and -17‰ , and 6.3 and 10.0‰ , respectively (Table 1). The $\delta^{13}\text{C}$ ranges (max. $\delta^{13}\text{C}$ to min. $\delta^{13}\text{C}$ along a baleen plate) are rather small for most BPs ($< 1.1\text{‰}$; Table 1), except for BP8 and BP10 (1.3 and 1.9‰ , respectively). BPs 3, 9, and 10 show the highest $\delta^{15}\text{N}$ range values (1.6 , 1.5 , and 2.2‰ , respectively; Table 1). Both BP1 and BP2 showed a higher $\delta^{15}\text{N}$ mean ($9.4 \pm 0.2\text{‰}$ and $8.1 \pm 0.2\text{‰}$, respectively) than the other BPs (6.8 to 7.7‰ ; p always < 0.05 and even < 0.001 for most of the BPs, $W = 0$).

Considering the records along the plate, positive or negative trends in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ with respect to the distance from the top of the plates ($W = 0$) were found (Figs. 2 & 3). The relationships between $\delta^{13}\text{C}$ along the L-axis were only significant for BP5 and BP9, which both exhibit significant slopes ($p < 0.05$).

While $\delta^{13}\text{C}$ of BP3 (lateral position) and BP4 (frontal area) were sampled from the same fin whale, no relationship was found ($p = 0.92$; Fig. 2). BP4 was half the size of BP3 and probably represents a shorter time-span. Mean BP4 $\delta^{13}\text{C}$ record ($-18.1 \pm 0.4\text{‰}$) is very similar to that found for BP3 between 0 and 15.7 cm ($-18.3 \pm 0.2\text{‰}$) ($p > 0.05$). $\delta^{13}\text{C}$ of BP3 showed a change at ~ 17.6 to 20.5 cm (mean \pm SD = $-17.8 \pm 0.3\text{‰}$). The same results were observed regarding $\delta^{15}\text{N}$ with relatively similar mean values for BP4 ($7.7 \pm 0.3\text{‰}$) and BP3 (between 0 and 15.7 cm = $7.4 \pm 0.3\text{‰}$, $p > 0.05$). The mean $\delta^{15}\text{N}$ values of BP3 between 17.6 and 20.5 cm ($6.7 \pm 0.3\text{‰}$) are significantly lower than the values observed between 0 to 15.7 cm ($p < 0.05$) and 21.4 to 32.2 cm ($p < 0.05$). Significant covariation between $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ was also found in BP3 ($r^2 = 0.33$); slope = -0.45 , $p < 0.001$).

Table 4. *Balaenoptera physalus*. Carbon (C) and nitrogen (N) stable isotope and C:N ratios measured along fin whale baleen plate length (L-axis, see Fig. 1) by plate reference number (collection year). All samples were measured along W = 0 axis except for BP6, which was measured as W = 2 cm. nd: lost sample

Reference number (collection year)	L-axis (cm)	W-axis (cm)	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)	C:N	Reference number (collection year)	L-axis (cm)	W-axis (cm)	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)	C:N	
BP1(1975)	4	0	-17.84	8.90	3.4	BP4 (1989)	1.7	0	-17.54	7.44	3.4	
	5.7	0	-17.95	9.36	3.4		3.8	0	-17.99	7.54	3.4	
	7.3	0	-17.98	9.30	3.4		4.7	0	-18.12	8.34	3.4	
	9.1	0	-18.55	9.21	3.5		6.4	0	-18.62	7.92	3.4	
	10.5	0	-18.49	9.61	3.4		7.4	0	-18.53	7.46	3.4	
	12	0	-18.18	9.95	3.4		9.3	0	-18.44	7.59	3.4	
	13.7	0	-18.13	9.61	3.4		10.4	0	nd	7.78	nd	
	15.2	0	-17.99	9.53	3.5		12.4	0	-17.65	7.70	3.4	
	16.7	0	-17.87	9.47	3.5		13.3	0	-17.78	7.61	3.3	
	18.2	0	-17.87	9.51	3.5		BP5 (1990)	0.65	0	-18.11	7.64	3.4
	19.7	0	-17.92	9.40	3.4			1.75	0	-17.91	7.33	3.3
	21.2	0	-18.07	9.22	3.5			2.85	0	-18.25	7.27	3.3
	22.7	0	-18.04	9.15	3.4			3.90	0	-18.45	7.09	3.4
	24.2	0	-18.12	9.36	3.5			5.10	0	-18.19	6.69	3.3
	25.6	0	-17.75	9.75	3.5			6.20	0	-18.24	7.15	3.3
	29	0	-18.03	9.38	nd			7.25	0	-18.75	7.11	3.4
	30.7	0	-17.87	9.60	3.5			8.30	0	-18.56	7.31	3.3
BP2 (1980)	7.1	0	-17.41	8.16	3.6	9.40		0	-18.51	7.56	3.3	
	12.2	0	-17.97	7.99	3.5	10.40		0	-18.30	7.25	3.3	
	16.7	0	-17.74	8.26	3.5	BP6 (1992)	0	2	-17.98	6.71	3.7	
	21.3	0	-16.98	7.99	3.5		2	2	-18.36	7.48	3.5	
	26.8	0	-17.16	8.20	3.5		3.8	2	-18.72	7.38	3.6	
	31.1	0	-17.61	8.35	3.5		5.8	2	-17.98	6.82	3.6	
36.4	0	-17.09	7.86	3.5	7.8		2	-18.48	7.96	3.4		
BP3 (1989)	0	0	-18.15	7.62	3.4		9.8	2	-18.47	7.60	3.4	
	0.8	0	-18.05	7.60	3.4		12	2	-18.29	7.47	3.7	
	1.5	0	-18.45	7.03	3.4		16.2	2	-18.40	7.75	3.4	
	2.4	0	-18.12	6.72	3.4		18.3	2	-17.76	7.39	3.6	
	3.3	0	-17.85	7.35	3.4		20.2	2	-18.52	7.80	3.4	
	4.3	0	-18.35	7.93	3.4	22.2	2	-18.54	7.84	3.4		
	6.3	0	-18.24	7.77	3.4	23.9	2	-17.91	6.68	3.6		
	7.2	0	-18.38	7.70	3.4	25.8	2	-18.33	7.39	3.4		
	8.9	0	-18.37	7.28	3.4	27.7	2	-18.14	7.90	3.4		
	9.9	0	-18.35	7.31	3.4	29.8	2	-17.84	7.36	3.7		
	11.7	0	-18.36	7.73	3.4	31.6	2	-18.68	7.86	3.6		
	14.8	0	-18.53	7.47	3.5	33.8	2	-18.23	8.06	3.4		
	15.7	0	-18.18	7.21	3.4	BP7 (1993)	5.2	0	-18.37	7.38	3.4	
	17.6	0	-18.12	7.14	3.4		6.9	0	-18.71	7.45	3.4	
	18.5	0	-17.69	6.63	3.4		10.5	0	-18.45	8.18	3.4	
	19.5	0	-17.54	6.36	3.4		11.7	0	-18.42	7.71	3.4	
	20.5	0	-17.67	6.70	3.4		14.8	0	-18.27	7.74	3.5	
	21.4	0	-17.89	7.27	3.3		16.5	0	-18.28	7.96	3.5	
	22.4	0	-17.82	6.86	3.4		20.1	0	-18.20	7.93	3.4	
	23.3	0	-17.96	7.08	3.4		21.6	0	-18.29	7.95	3.4	
	24.5	0	-18.10	6.65	3.3		26.2	0	-18.24	7.46	3.4	
	25.4	0	-17.79	7.28	3.4		28.4	0	-18.32	7.77	3.4	
	26.4	0	-17.72	7.23	3.4	30	0	nd	7.74	nd		
27.8	0	-17.95	7.49	3.4	34	0	-18.09	6.84	3.3			
28.2	0	-17.98	7.06	3.4	BP8 (1996)	4	0	-18.41	6.87	3.5		
29	0	-17.64	7.15	3.4		5.4	0	-18.90	6.69	3.7		
29.9	0	-17.62	7.10	3.4		5.8	0	-18.28	7.08	3.5		
30.8	0	-17.71	7.01	3.4		6.2	0	-19.32	6.69	3.9		
31.6	0	-17.83	7.10	3.4		6.9	0	-18.78	6.96	3.5		
32.2	0	-17.57	7.02	3.3		8.3	0	-18.82	6.74	3.6		

Table continued on next page

Table 4. (continued)

Reference number (collection year)	L-axis (cm)	W-axis (cm)	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)	C:N	Reference number (collection year)	L-axis (cm)	W-axis (cm)	$\delta^{13}\text{C}$ (‰)	$\delta^{15}\text{N}$ (‰)	C:N	
BP (1996)	9.1	0	-19.25	6.70	4.0	BP10 (2002) (continued)	6.2	0	-18.33	7.14	3.4	
	9.7	0	-18.76	6.57	3.7		7.25	0	-18.83	7.31	3.8	
	10.5	0	-18.78	6.44	3.6		8.15	0	-18.46	7.15	3.3	
	11.2	0	-18.66	6.58	3.6		9.25	0	-18.70	7.56	3.4	
	12	0	-18.02	6.79	3.4		10.35	0	-18.41	7.68	3.4	
	12.7	0	-18.23	6.93	3.5		10.35	0	-18.34	7.96	3.4	
	13.5	0	-18.05	6.61	3.6		11.45	0	-18.24	7.95	3.4	
	15.5	0	-18.20	6.79	3.6		12.55	0	-18.16	7.90	3.3	
	16.4	0	-18.05	7.01	3.5		13	0	-18.66	8.38	3.6	
	17.2	0	-17.97	7.07	3.6		14.75	0	-18.12	8.35	3.3	
	18.1	0	-18.64	6.77	3.7		16	0	-18.11	8.00	3.4	
	18.9	0	-18.26	7.03	3.6		17.2	0	-18.69	8.49	3.5	
	19.7	0	-18.62	7.00	3.7		18.35	0	-18.50	8.68	3.3	
	20.5	0	-18.21	7.04	3.6		19.5	0	-18.53	8.23	3.3	
	22.4	0	-18.46	6.32	3.5		20.65	0	-18.84	7.17	3.4	
	23.4	0	-18.29	6.64	3.5		21.75	0	-18.94	6.50	3.4	
	24.4	0	-18.73	7.02	3.6		23.25	0	-18.83	8.14	3.4	
	25.4	0	-18.28	6.66	3.5		24.55	0	-19.25	7.95	3.5	
	26.2	0	-18.80	7.23	3.7		27.15	0	-18.26	7.82	3.4	
	27.1	0	-18.39	6.99	3.5		28.55	0	-18.05	8.09	3.4	
	28.2	0	-18.52	6.93	3.6		29.65	0	-17.94	7.23	3.3	
	29.5	0	-18.36	6.69	3.6		30.9	0	-18.14	7.46	3.3	
	30.6	0	-17.99	6.97	3.5		32.1	0	-18.50	7.92	3.5	
	31.8	0	-18.14	6.69	3.5		33.2	0	-18.34	7.97	3.4	
	32.9	0	-18.30	6.88	3.5		34.3	0	-18.15	7.87	3.4	
	33.9	0	-18.08	7.16	3.5		35.5	0	-18.08	7.60	3.5	
	BP9 (2000)	0.35	0	-17.57	7.29		3.5	36.7	0	-18.19	7.29	3.5
		1.25	0	-18.05	6.84		3.4	37.9	0	-18.24	7.31	3.4
2.15		0	-17.53	7.16	3.4	39	0	-18.29	7.34	3.4		
3		0	-17.79	7.11	3.4	40.2	0	-17.95	7.23	3.4		
3.85		0	-18.02	7.83	3.4	42.4	0	-18.10	7.38	3.3		
4.75		0	-18.16	7.55	3.4	43.4	0	-19.21	7.08	3.7		
5.8		0	-17.86	6.58	3.4	44.5	0	-17.79	7.64	3.3		
6.85		0	-18.18	6.40	3.4	45.6	0	-18.07	7.36	3.3		
7.25		0	-18.32	6.36	3.3	46.5	0	-17.88	8.10	3.3		
8.05		0	-17.90	7.10	3.4	47.5	0	-17.95	7.83	3.3		
8.7		0	-18.18	7.09	3.3	48.5	0	-18.50	7.81	3.5		
9.7	0	-18.06	6.83	3.4	49.55	0	-17.73	7.47	3.3			
BP10 (2002)	0.9	0	-18.02	7.61	3.3	50.65	0	-17.37	7.91	3.3		
	3	0	-18.15	8.12	3.4	51.55	0	-17.75	7.25	3.3		
	4.1	0	-18.08	8.08	3.4							
	5.1	0	-18.19	7.73	3.3							

Stable isotopes signature of krill and fin whale baleen, and trophic level of fin whales

The mean level of $\delta^{15}\text{N}$ signature of baleen plates ($7.6 \pm 0.7\text{‰}$) was 3.2 and 1.0‰ higher than that of bulk (non-lipid-extracted) *Meganyctiphanes norvegica* sampled in the Mediterranean Sea or in the Atlantic Ocean, respectively. The mean level of $\delta^{13}\text{C}$ signature of baleen plates ($-18.2 \pm 0.4\text{‰}$) was 1.1 ‰ and 3.0‰ higher than that of annual mean bulk krill sampled in the Mediterranean Sea or in the Atlantic Ocean, respectively (Tables 2 & 3).

According to the methods described by De Niro & Epstein (1978, 1981) we corrected the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of the Mediterranean Sea and the Atlantic Ocean krill by 1.0 and 3.4‰, respectively to take into account the trophic level enrichment factor for carbon and nitrogen. When compared to the corrected spring and fall $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of the bulk Mediterranean and to the corrected annual $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of the bulk Atlantic krill samples, the $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values of the 10 BPs are more closely related to the annualized Mediterranean krill values (Fig. 4A). All but one (BP2) $\delta^{13}\text{C}$ mean value fell

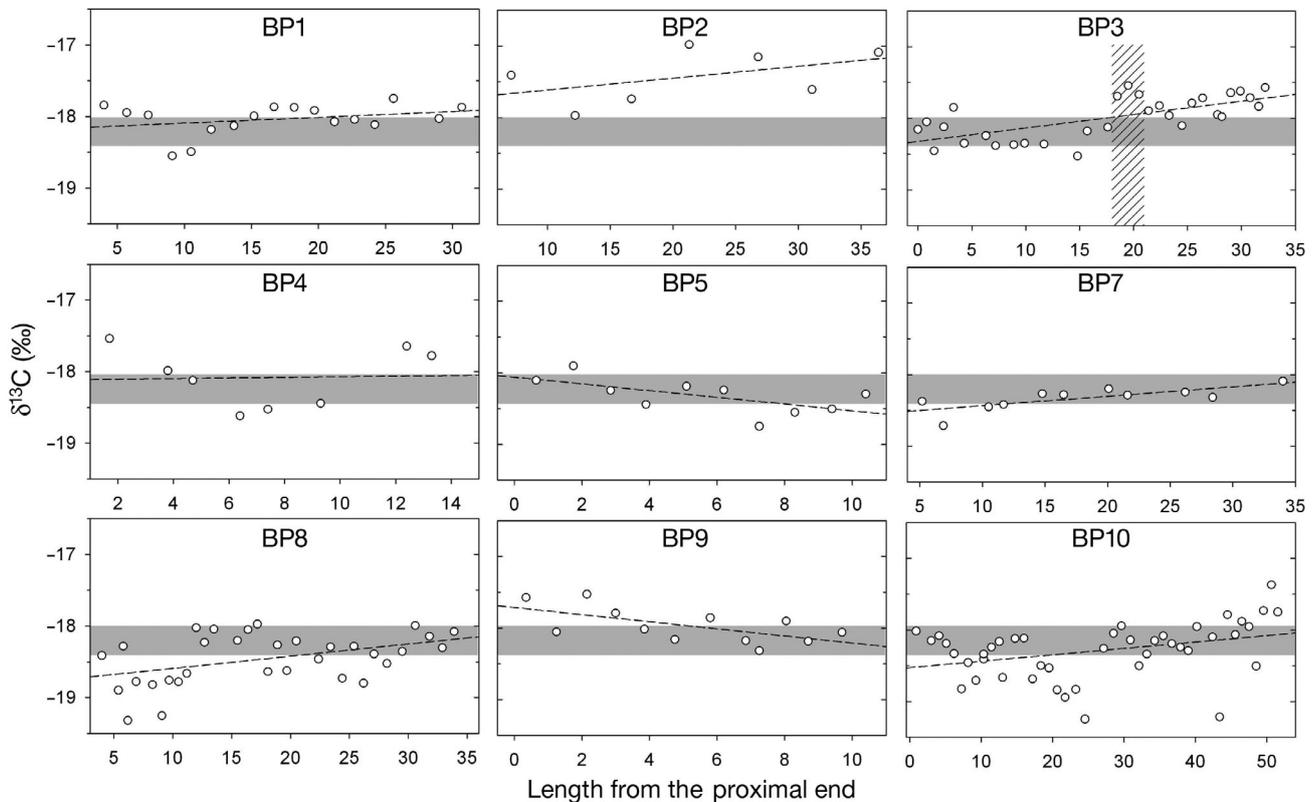


Fig. 2. *Balaenoptera physalus*. Intra-individual stable carbon isotope values (expressed per ml) along the length of baleen plates of fin whales stranded at western Mediterranean coasts between 1975 and 2002. All plots represent $W = 0$ sampled whales. Grey areas represent the range of values comprising $>50\%$ of the isotopic ratios of the whole dataset. The dashed vertical band in Panel BP3 suggests the weaning period. BP3 & BP4 were from the same individual but sampled at lateral and frontal sides of the jaw. BP6 is not included because it was sampled at $W = 2$ cm

within the reference range corresponding to the outer-limits of the ± 1 SD range of spring and fall values combined of the $\delta^{13}\text{C}_{\text{corrected}}$ values of the Mediterranean sea krill (-18.9 to -17.5 ‰).

When excluding BP2, which is well outside the range of $\delta^{13}\text{C}_{\text{corrected}}$ krill values, all but BP8 are centered on the \pm SD range of the Mediterranean spring krill mean value, and all but BP9 have few consistent values with fall Mediterranean krill $\delta^{13}\text{C}$. Fig. 4B shows that some individuals have $\delta^{13}\text{C}$ values isotopic values exceeding the range expected from the spring and fall Mediterranean krill $\delta^{13}\text{C}$ values. However, even the most depleted values (-18.9 to -19.3 ‰) of the BPs 8 and 10 plates samples were not consistent with the $\delta^{13}\text{C}_{\text{corrected}}$ signature of krill sampled in the Atlantic Ocean (-20.6 to -19.1 ‰).

Most $\delta^{15}\text{N}$ plate values are also consistent with spring Mediterranean krill $\delta^{15}\text{N}$ mean value (Fig. 4A). The main exception being the BP1 with a $\delta^{15}\text{N}$ exceeding the $\delta^{15}\text{N}$ signature expected from the fall and spring Mediterranean krill isotopic signature. BP2 has intermediate $\delta^{15}\text{N}$ values between fall and

spring Mediterranean krill $\delta^{15}\text{N}$. BPs 4 and 10 are the only plates showing consistent $\delta^{15}\text{N}$ values with both fall and spring Mediterranean krill $\delta^{15}\text{N}$ mean value. All plates except BPs 1, 2, and 4 are characterized by $\delta^{15}\text{N}$ values lower than expected from the spring and fall Mediterranean krill $\delta^{15}\text{N}$ signatures (Fig. 4C).

Assuming that the euphausiid is a primary consumer (i.e. mostly herbivorous and omnivorous), a trophic level of 2 can be assigned to this species (e.g. Vander Zanden & Rasmussen 2001). Using mean values for the whales and their euphausiid prey in the formula proposed by Vander Zanden and Rasmussen (2001) for the trophic level value (TL):

$$\text{TL} = 2 + (\delta^{15}\text{N}_{\text{baleen}} - \delta^{15}\text{N}_{\text{uncorrected } M. \text{ norvegica}}) / 3.4 \quad (2)$$

we calculate a TL of 2.9 ± 0.3 (mean \pm SD) for fin whales based on the annual mean $\delta^{15}\text{N}$ Mediterranean euphausiids. The TL varies between 2.8 in fall and 3.0 in spring due to the effect of the seasonal $\delta^{15}\text{N}$ variability observed in the euphausiids. Using the Atlantic euphausiids $\delta^{15}\text{N}$ values, a mean TL of 2.3 ± 0.4 (2.0 in fall and 2.5 for spring) can be esti-

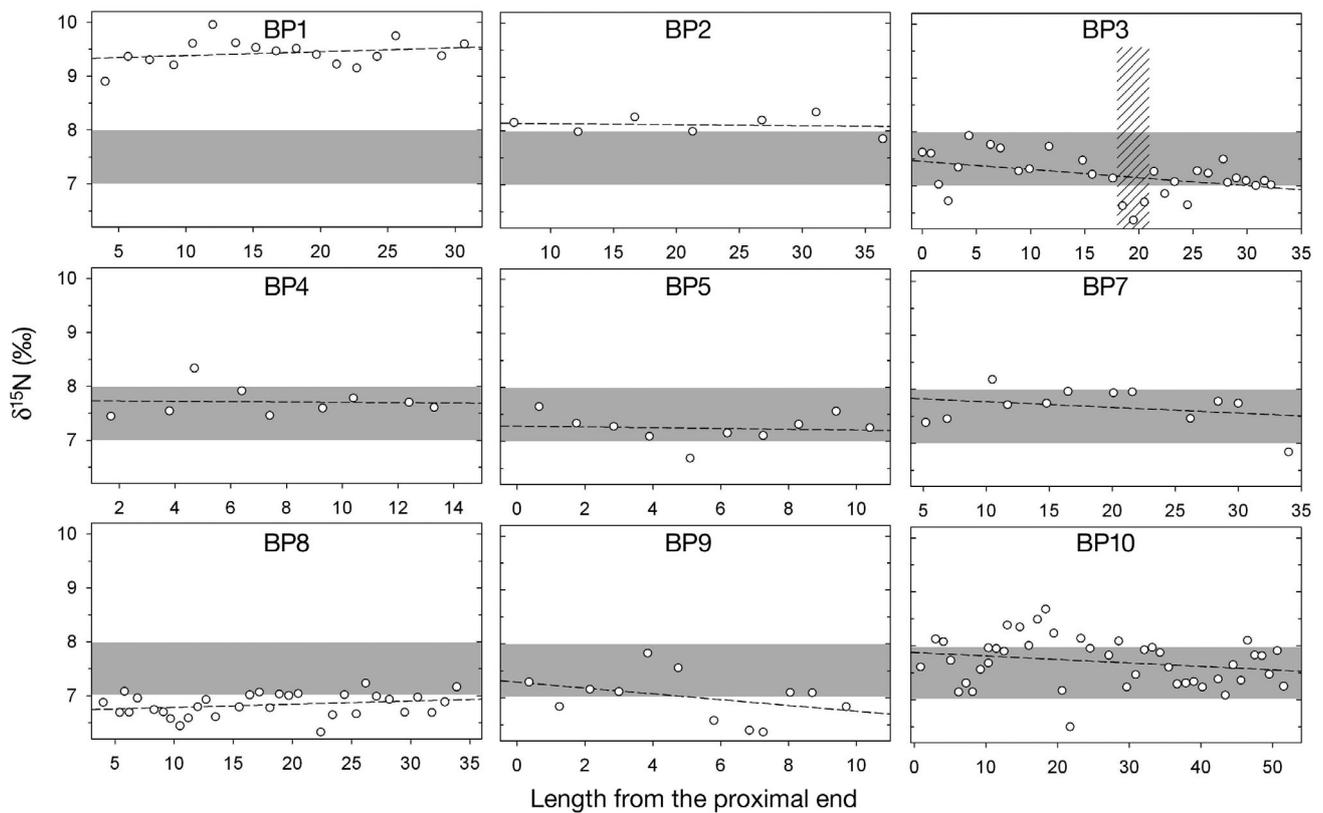


Fig. 3. *Balaenoptera physalus*. Intra-individual stable nitrogen isotope values (expressed per ml) along the length of baleen plates of fin whales stranded at western Mediterranean coasts between 1975 and 2002. For details see legend of Fig. 2

mated. The $TL = 2.9 \pm 0.3$ calculated with the Mediterranean $\delta^{15}N_{\text{uncorrected } M. norvegica}$ value is closer to the $TL = 3.2$ estimated from the dietary analysis of Mediterranean fin whale by Kaschner et al. (2004).

Decadal trends in stable isotope signature

The evolution of the keratin $\delta^{13}C$ and $\delta^{15}N$ from the 10 BPs of fin whales stranded along the Western Mediterranean coasts over a 27 yr span is presented in Fig. 5A,B. $\delta^{13}C$ and $\delta^{15}N$ mean values show important changes between 1975 and 2002. Whereas $\delta^{15}N$ variances statistically significantly increase with time, the slight positive increase in $\delta^{13}C$ variances is not statistically significant ($p > 0.68$) from linear regression. We calculated the regression equation using the mean values of all BPs (Table 1). The linear fit shows a strong correlation between $\delta^{15}N$ means and years of collection ($r^2 = 0.58$, $n = 10$, $p = 0.01$) and a smaller one for $\delta^{13}C$ ($r^2 = 0.29$, $n = 10$). A polynomial fit (third order) applied to both $\delta^{13}C$ and $\delta^{15}N$ means increased the correlation between stable isotope evolution and years. The $\delta^{13}C$ versus year fit shows a

sinusoidal curve and an isotopic shift in 1980 (Fig. 5A). The $\delta^{15}N$ versus year 'abruptly' decreases between 1975 and 1980 but shows less variation between 1990 and 2002 (Fig. 5B).

Satellite tracking

Eleven fin whales were tagged with satellite-monitored radio transmitters off the Provençal coast from 1 August to 20 August 2003. Argos positions were obtained from 8 individuals over the course of 9 mo (Table 5). Seven individuals remained in the north-western part of the Mediterranean Sea through fall and winter, and 1 individual (tag 10842) moved into the Atlantic Ocean (Fig. 6).

DISCUSSION

Satellite telemetry provides evidence, that some fin whales migrate from the Mediterranean Sea to the Atlantic Ocean; however, most individuals remain within the western basin during fall and winter.

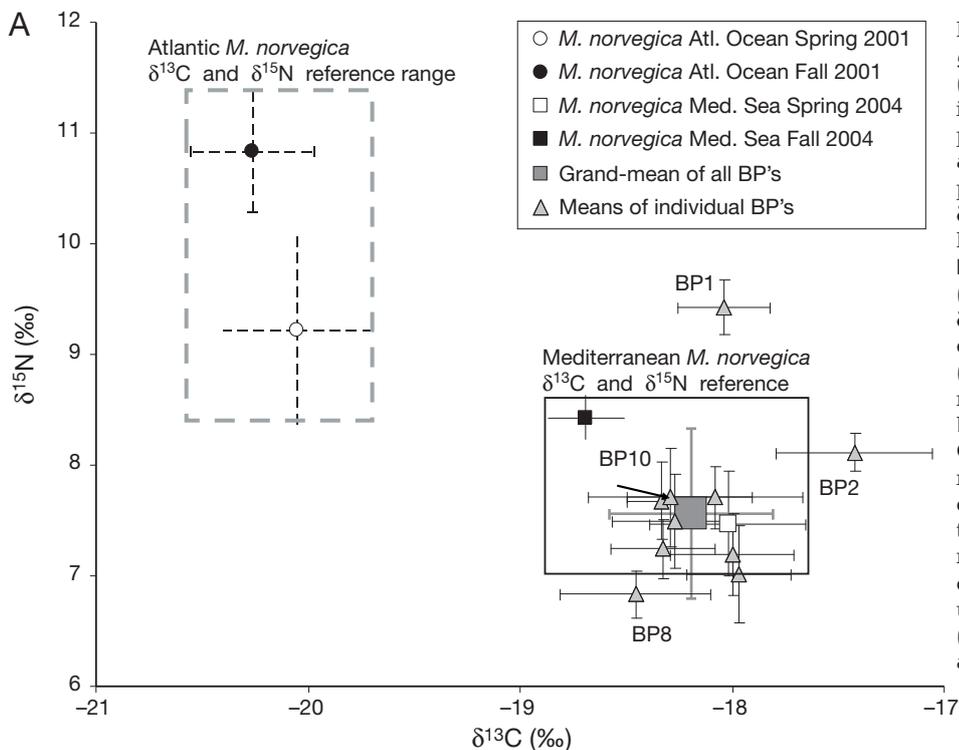
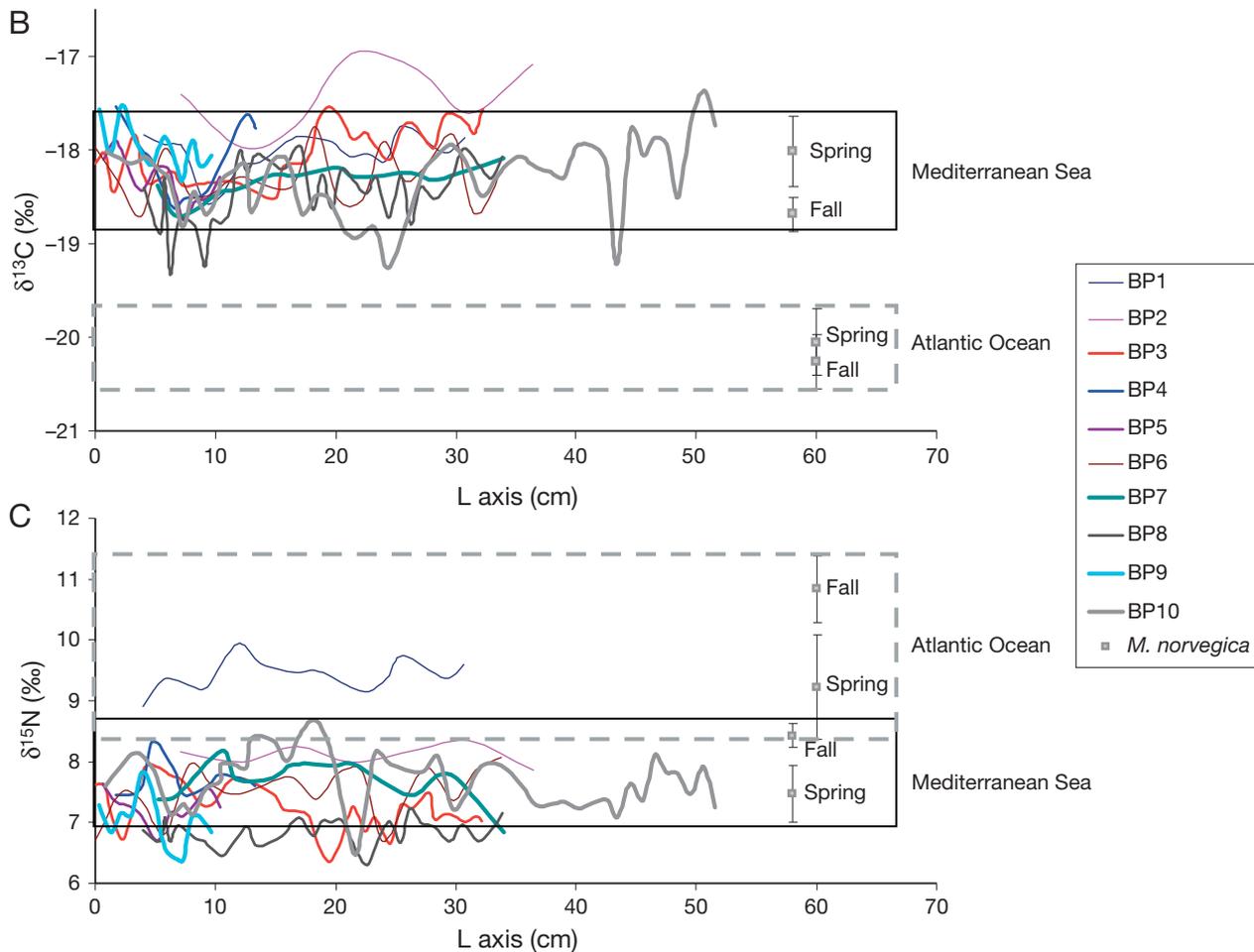


Fig. 4. *Meganyctiphanes norvegica* and *Balaenoptera physalus*. (A) Nitrogen and carbon stable isotope ratios of the 10 baleen plates of stranded fin whales (■) and of each individual (△) compared with *M. norvegica* (krill) $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values from the Mediterranean Sea (□: spring; ■: fall) and Atlantic Ocean (○: spring; ●: fall). $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ krill data represent trophic enrichment corrected values (1 and 3.4‰, respectively) and non-lipid-extracted values. The boxes represent the Atlantic Ocean (---) and the Mediterranean Sea (—) krill reference ranges that correspond to the outer-limits of the ± 1 SD range of spring and fall values combined of the $\delta^{13}\text{C}_{\text{corrected}}$ values of the krill. Intra-individual (B) $\delta^{13}\text{C}$ and (C) $\delta^{15}\text{N}$ variations along the length of the plates



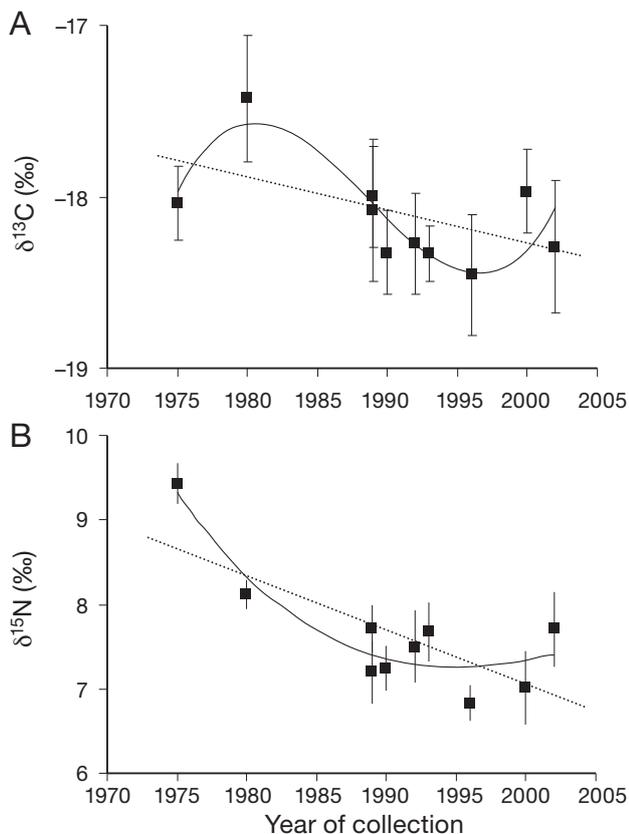


Fig. 5. *Balaenoptera physalus*. Variations of (A) $\delta^{13}\text{C}$ and (B) $\delta^{15}\text{N}$ values of 10 fin whale baleen plates versus year of collection, means \pm SD. The line curves fitting the data are linear ($\delta^{13}\text{C} = -0.02 \times \text{Years} + 20.14$; $r^2 = 0.29$; $\delta^{15}\text{N} = -0.07 \times \text{Years} + 142.5$; $r^2 = 0.58$) and cubic polynomial regressions ($\delta^{13}\text{C} = 0.0004 \times \text{Years}^3 - 2.6145 \times \text{Years}^2 + 5199 \times \text{Years} - 3 \times 10^6$; $r^2 = 0.70$; $\delta^{15}\text{N} = -8 \times 10^{-05} \times \text{Years}^3 + 0.4886 \times \text{Years}^2 - 981.82 \times \text{Years} + 657624$; $r^2 = 0.84$)

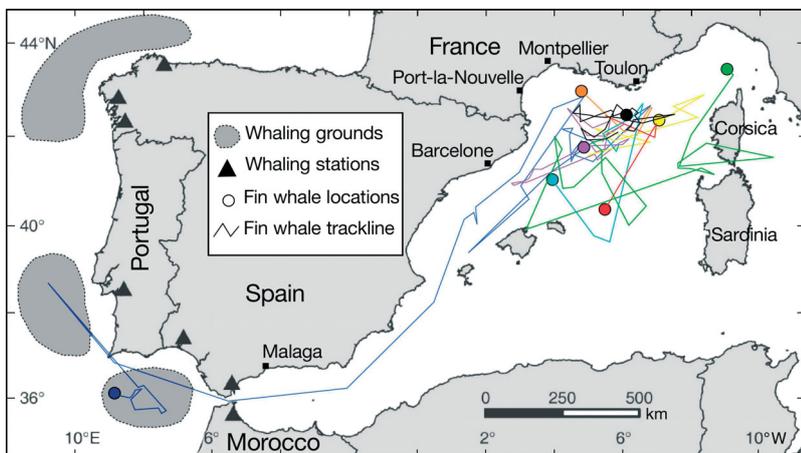


Fig. 6. *Balaenoptera physalus*. Argos locations of 7 individuals tagged with satellite-monitored radio transmitters off the Provençal coast from 1 August to 20 August 2003. Tags (see Table 5): ● = 10838, ● = 23029, ● = 10838, ● = 23032, ● = 23042, ● = 10836, ● = 10842, ● = 23041. The tag 23032 is not shown, as only 2 locations were obtained from this individual over a 43.3 d period and within initial tagging area

Table 5. *Balaenoptera physalus*. Tag identification number (PTT), deployment dates, number of days following tagging with locations, duration of operation (message days), and minimum distances travelled by 8 fin whales tagged in the northwestern Mediterranean Sea during August 2003. TAG 10842: individual moved into the Atlantic Ocean

PTT	Date deployed	Location (d)	Message (d)	Distance (km)
10836	8/12/03	145.1	192.7	913
10838	8/18/03	278.0	296.0	4260
10842	8/16/03	138.6	138.6	4057
23029	8/18/03	45.1	57.6	1079
23032	8/18/03	43.3	44.3	405
23033	8/4/03	140.4	140.4	761
23041	8/8/03	175.3	228.8	419
23042	8/16/03	301.0	394.0	1461
Averages		158.3	186.6	1669

Therefore this population is exposed, year-round, to heavy shipping traffic, as the 3 largest Mediterranean ports—Barcelona, Marseille, and Genova—are contiguous to the main fin whale summer foraging grounds (Forcada et al. 1996, Monestiez et al. 2006, Cotté et al. 2009).

The stable isotope analyses of baleen plates reveal that most stranded fin whales in the northwestern Mediterranean Sea did not present $\delta^{13}\text{C}$ oscillations. Assuming a +3.4‰ trophic enrichment factor for $\delta^{15}\text{N}$ and +1‰ for $\delta^{13}\text{C}$, these results suggest year-round residency within the western Mediterranean Sea. Few $\delta^{13}\text{C}$ values were either enriched (BP2) or depleted (BP8, BP10) compared to the $\delta^{13}\text{C}$ Mediterranean krill reference range (Fig. 4B).

Both $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ variations along the length of BP2 may be cyclical, suggesting that BP2 fin whale may regularly visit different foraging grounds, possibly by alternating between the northwestern Mediterranean basin (where this whale was ultimately killed in a ship collision) during spring–summer seasons (31 and 12 cm; Fig. 4B) and unidentified waters (where it fed at the same high trophic level compared to the other fin whales) (~21 cm). However, the high $\delta^{13}\text{C}$ values of BP2 suggest that this whale was unlikely to forage in the Atlantic Ocean. $\delta^{13}\text{C}$ of BP10 and to a lesser extent BP8, which were both sampled in the Malaga region, also exhibit 'cyclic' patterns (Fig. 4B). The high $\delta^{13}\text{C}$ ranges mea-

sured along the length of BP8 and BP10 (1.3 and 1.9‰, respectively) suggest a broader foraging area for these individuals than for the other ones (range: 0.6 to 1.1‰). However, their $\delta^{13}\text{C}$ suggest that these individuals spent most of their time in the Mediterranean basin feeding alternatively on spring and fall resources and exhibited short shifts marked by significant $\delta^{13}\text{C}$ depletion. These depleted $\delta^{13}\text{C}$ values are intermediate between the Mediterranean and Atlantic krill $\delta^{13}\text{C}$ values, suggesting possible short excursions in the Atlantic Ocean or the nearby Strait of Gibraltar. The less-depleted $\delta^{13}\text{C}$ values observed in the baleen plates compared to the expected Atlantic Ocean $\delta^{13}\text{C}$ values can be explained either by the fact that those excursions were not long enough to allow the plate to equilibrate with the Atlantic prey $\delta^{13}\text{C}$ values or that the fin whales restricted their Atlantic excursions to the Atlantic side of the Strait of Gibraltar, an historical Spanish whaling ground in the 1920s and 1950s and consistent with the data provided by satellite tracking (Sanpera & Aguilar 1992; Fig. 6). In this hypothesis, the 'Gibraltar-Atlantic' prey $\delta^{13}\text{C}$ values would be intermediate between Mediterranean and Atlantic values. As fin whales display maternally directed site fidelity, we can question the effect of the past intensive whaling on the migration patterns of this population. In the absence of such intensive whaling we would expect much higher variations in stable isotope values with time in the baleen of these whales.

As only 2 (BP10, and, to a lesser extent, BP8) out of the 10 baleen plates exhibited such 'cyclic' patterns in $\delta^{13}\text{C}$, this behavior does not seem to prevail in this fin whale population. Interestingly both fin whales from the Malaga area show indications of such migratory pattern, suggesting that fin whales encountered in this region may be in transit between the Mediterranean Sea and the Atlantic Ocean. Indeed the satellite tracking data of the individual that left the Mediterranean Sea (TAG 10842) revealed that this fin whale migrated to the Atlantic Ocean along the Spanish coastline (Fig. 6). In addition to satellite tracking that allows precise location of individual fin whales over relatively short periods of time (mo), the stable isotopes measured along the baleen plates provide an insight on the main foraging regions over long-term periods (Hobson et al. 2004). However, precise reconstructions depend on BP growth rate data. A first attempt at estimating growth rate on Mediterranean fin whale is made using BP3. The concomitant changes of both $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ between 20.5 and 17.6 cm on BP3 (Figs. 2 & 3) could result from a shift in trophic level due to prey change

or correspond to the weaning transition period from maternal milk to prey (Hobson & Schell 1998, Hobson & Sease 1998). Though small isotopic changes are observed ($\sim 1\%$), our $\delta^{13}\text{C}$ shifts and depleted $\delta^{15}\text{N}$ observed at ~ 20.5 cm might suggest weaning age (7 mo in fin whales; International Union for Conservation of Nature (IUCN) 1991) was reached, the juvenile shifting to nutritional independence and feeding at a lower trophic level than adults (Jenkins et al. 2001). However there is a very real possibility that some substantial, unknown amount of wear may have occurred for BP3, suggesting that the calculated growth rate of ~ 20.1 cm yr^{-1} ($32.2 - 20.5$ cm = 11.7 cm / 7 mo $\times 12$ mo yr^{-1}) correspond to a lower-limit growth rate for this female BP3–BP4. Interestingly, BP3 growth rate (~ 20 cm yr^{-1}) is within the range of those observed for adult Minke whales (12.9 cm yr^{-1} ; Mitani et al. 2006); bowhead whales (20 cm yr^{-1} or less; Schell et al. 1989a,b), and southern right whales (2.7 cm yr^{-1} ; Best & Schell 1996). Applying our minimum growth rate estimate, BP3 whale would be ~ 1.6 yr old. Because fin whales are believed to reach sexual maturity at ~ 6 to 12 yr, it is likely that the baleen of the adult female BP3 (Table 1) did in fact experience significant erosion at the distal end; otherwise the length of the baleen for a whale of at least 6 yr of age should have been ~ 120 cm at a minimum. In conclusion, it is likely that erosion would cause the underestimation of BP3 baleen growth rates, and hence this estimate is to be viewed as a minimum growth rate value. Using the minimum growth rate value (20 cm yr^{-1}), the multi $\delta^{13}\text{C}$ cycles observed on BP10 and BP8 suggest that these individuals were regularly alternating between different foraging grounds, with individuals traveling once or twice a year from the Mediterranean Sea in the direction of the Atlantic Ocean, and we speculate these whales remained in the Gibraltar Strait–Atlantic continuum.

A 3‰ order enrichment is expected between the $\delta^{15}\text{N}$ signature of *Meganyctiphanes norvegica* and fin whales if the whales are feeding exclusively on this prey. We found an enrichment of 2.6 to 3.5‰ (mean \pm SD = 3.2 ± 0.8) between Mediterranean krill and baleen plates, and 0.2 to 1.8‰ between Atlantic krill and baleen plates. If *M. norvegica* was the year-round predominant prey of our study of fin whale populations, the observed $\delta^{15}\text{N}$ oscillations should primarily reflect migratory patterns or seasonal change in krill $\delta^{15}\text{N}$ signature. Furthermore, our krill data show an increase of 0.9 to 1.6‰ during spring to late summer transition in the Mediterranean Sea and Atlantic Ocean, respectively. Similarly Bode &

Alvarez-Ossorio (2004) reported a zooplankton $\delta^{15}\text{N}$ increase by more than 3‰ during the spring–summer transition, partly due to the nitrate depletion during phytoplankton growth. According to that hypothesis, and considering a 3.4‰ $\delta^{15}\text{N}$ enrichment per trophic level, most fin whales in the present study could have only fed on krill in Atlantic in winter when krill $\delta^{15}\text{N}$ is depleted. However the geographical source given by the $\delta^{13}\text{C}$ does not support that hypothesis (see also the following 2 subsections).

However, an enrichment of <3.4‰ was suggested in several studies (Ostrom et al. 1993, Abend & Smith 1997). An 2.4‰ enrichment factor for the $\delta^{15}\text{N}$ would lead to different interpretations of the migration pattern. For instance BP1 and BP2 $\delta^{15}\text{N}$ values would suggest the ingestion of only Atlantic krill and many values of the other BPs would indicate that whales feed on both Mediterranean Sea and Atlantic Ocean. Again the geographical source given by the $\delta^{13}\text{C}$ does not corroborate such a scenario.

Hence potential mechanisms of decoupling carbon and nitrogen isotope signatures should be suggested. According to Hobson et al. (2004), migration and fasting could be such a mechanism. If whales migrate in winter and fast in Atlantic waters, highest $\delta^{15}\text{N}$ are expected, and carbon from stored fats is used for baleen formation. It might show that Mediterranean summer diet $\delta^{13}\text{C}$ values determine the long-term substrate values for baleen formation. These hypotheses imply that *Balaenoptera physalus* has a well-identified fasting period during winter. To our knowledge, no pattern of seasonal period of fasting has been reported in the literature.

Minagawa & Wada (1984) found a 3.4‰ enrichment factor for $\delta^{15}\text{N}$ when using whole crustacean, as was the case in the present study. Furthermore, Hobson et al (1996) found a mean value of 3‰ enrichment factor for $\delta^{15}\text{N}$ between the prey and keratinous tissues such as whiskers, nails, and hair. We might expect a similar enrichment factor between the whole krill prey and keratinous baleen plates of fin whales. Hence we suggest that a 3.4‰ enrichment factor is close to the actual number and believe that *Meganyctiphanes norvegica* is the predominant prey of this population of fin whales; therefore it follows that the observed stable isotope oscillations of BP10 and 8 primarily reflect migratory patterns.

We cannot exclude that Mediterranean fin whales are also feeding on other unknown prey species. According to Canese et al. (2006), zooplankton sampling collected in winter near Lampedusa, where fin whales fed, indicates the presence of *Nyctiphanes couchi*. However we can rule out that Mediterranean

fin whales mainly feed on small mesopelagic fishes such as sardines or anchovies, as the whales occupy the same mean trophic level as these fish species (mean $\delta^{15}\text{N}$ ranging from 7.69 to 7.87; Pinnegar et al. 2003).

Nonetheless, higher $\delta^{15}\text{N}$ values were exhibited in BP1, suggesting that part of this whale's diet could be composed of prey from higher trophic levels over extended periods of time. However, taking into account the year of stranding (1975), the BP1 $\delta^{15}\text{N}$ values can also suggest that the nitrogen isotopic signature at the base of the food chain was higher 27 yr ago.

Pinnegar et al. (2003) and Pauly et al. (1998) reported a decline in the mean trophic level of landings for the western Mediterranean Sea and interpreted it as 'fishing down the marine food web', mostly through diversification and (or) substitution in the species being targeted (marketed), as might be expected where the supply of the preferred target species has begun to dwindle (Pinnegar et al. 2002). In addition to this decline, the temporal decrease in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ may also provide some evidence of significant environmental changes affecting the Mediterranean Sea.

Schell (2000) reported a depletion of $\delta^{13}\text{C}$ in *Balaena mysticetus* plates from the Bering Sea between 1947 and 1997. The $\delta^{13}\text{C}$ depletion of the Bering Sea whales (-0.05‰ yr^{-1}) during the last 30 yr is about twice as much as the one previously reported for the Indian Ocean (-0.02‰ yr^{-1} ; Gruber et al. 1999) and the one found in the plates from the Mediterranean fin whales in the present study (-0.02‰ yr^{-1}) between 1975 and 2002. The $\delta^{13}\text{C}$ signature of the baleen plates over the study period suggests increasing $\delta^{13}\text{C}$ values between 1975 and 1980 and 1996 and 2002, and decreasing $\delta^{13}\text{C}$ between 1980 and 1996 (Fig. 5). Interestingly, Demirov & Pinardi's (2002) simulations of the interannual surface Mediterranean circulation from 1979 to 1993 identify 2 periods, 1981 to 1987 and 1988 to 1993, which differ in precipitation and winter wind regimes. Moreover, Conversi et al (2010) show important changes of the plankton at the end of the 1980s in the Mediterranean basins and provide analyses linking local, regional, and basin scale hydrological properties with 2 major indicators of large scale climate, the North Atlantic Oscillation (NAO) index and the Northern Hemisphere Temperature index, suggesting that the Mediterranean shift is part of a larger scale change affecting the northern hemisphere.

The driving force explaining the general carbon isotopic depletion patterns may reflect global hemispheric changes. Schell (2000), assuming constant

dissolved CO₂ concentration, attributed the δ¹³C depletion in the Bering Sea to a 30 to 40 % decrease in the marine productivity. An alternative hypothesis was proposed by Cullen et al. (2001) who suggested that increased CO₂ levels could also explain the δ¹³C depletion. Indeed, the atmospheric CO₂ levels increased significantly due to fossil fuel burning, land changes, and cement manufacture (IPCC 2007) all resulting in a significant decrease of the δ¹³C_{atmosphericCO₂} (Keeling et al. 2001). However, in a recent study, Williams et al. (2011) interpreted the δ¹³C shift of the Bering Sea as a response to the increased upwelling intensity linked to the Aleutian low intensification.

In region off the upwelling the ¹³C depleted atmospheric anthropogenic CO₂ is partially absorbed by the ocean surface waters, thus decreasing the isotopic composition of the marine inorganic carbon pool. Known as the Suess effect (Gruber et al. 1999, Tanaka et al. 2003), this anthropogenic CO₂ input affects the isotopic signature of primary producers and consequently of marine food web, thus possibly explaining in part the observed baleen plate δ¹³C shift noted herein.

Higher marine CO₂ content may favour the photosynthetic rate, but such conditions would lead to a δ¹³C decrease and δ¹⁵N increase (predominance of assimilatory nitrate reduction increases phytoplankton δ¹⁵N). This phenomenon has a direct effect on the isotopic signature of primary producers and consequently on the marine food web, explaining the baleen plate δ¹³C shift evidenced on a global scale (Cullen et al. 2001). However, the present δ¹⁵N values (−0.07 ‰ yr^{−1}) were as significantly depleted as those observed in the Bering Sea (−0.06 ‰ yr^{−1}; see Schell 2000 and comment by Cullen et al. 2001). This decrease in δ¹⁵N is intriguing, but poorly documented as an ecosystem indicator (Lee et al. 2005).

Atmospheric N₂ would be used as a nitrogen source for primary production (diazotrophy), otherwise the microbial loop system may become more active, recycling isotopically light NH₄⁺ used by phytoplankton. However, the most likely mechanism to explain such a rapid change in the Mediterranean Sea would be a major shift of the nutrient balance in response to increased anthropogenic input by coastal run off, river discharge and atmospheric nitrogen compounds. Though additional nitrate may stimulate the biological pump, especially for the Mediterranean Sea, which is often considered as a nitrate depleted sea, this anthropogenic forcing can also significantly decrease the productivity of the ecosystem by altering the Redfield ratios (increase of the N:P and decrease of the Si:N ra-

tios; Béthoux et al. 2002) and consequently modify the marine community (Turner et al. 2003). Data from the Gulf of Lions shows an N enrichment with respect to P due to the Rhone River influence (Diaz et al. 2001). The changes in N:P:Si ratios observed since the early 1960s (Béthoux et al. 2002) could lead to a decreased photosynthetic rate or a phytoplankton shift from a diatom-dominated ecosystem towards a non-siliceous one and a more 'regeneration-dominated' community (Marty et al. 2002).

Superimposed on this declining trend, the potential impact of the decadal changes of the NAO cannot be ruled out, causing drier (normal) Mediterranean conditions during the negative (positive) phase. Since 1980, except in 1996, the NAO has been positive. As a result, the isotopic decline between 1980 and 2002 cannot be attributed to the NAO alone. A complex combination of circulation alteration, temperature increase (Goffart et al. 2002, Conversi et al. 2010) and nutrient availability, anthropogenic impact are likely affecting marine communities of the Mediterranean sub-basins.

The combination of satellite tracking and stable isotope signature methods reveals some major aspects of Mediterranean fin whale foraging, indicating that most individuals might be year-round residents of the northwestern Mediterranean Sea and that the largest predator of the Mediterranean Sea has a trophic level equivalent to that of anchovies and sardines.

The observed decrease in δ¹⁵N is an important ecosystem indicator supporting the assumption of a changing nitrogen cycle with still unclear effects (Duce et al. 2008). More efforts need to be directed toward understanding the nitrogen biogeochemical cycle. Conversely, the change in δ¹³C reported is consistent with previously well-described studies. The decrease in primary production level or shifts in the primary producers are likely to influence the Mediterranean food web and consequently fin whales. However, the long term consequences remain unknown.

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