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4 **INFLAMMATION DYSREGULATES NOTCH SIGNALING IN ENDOTHELIAL**
5 **CELLS: IMPLICATION OF NOTCH2 AND NOTCH4 TO ENDOTHELIAL**
6 **DYSFUNCTION**
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Abstract

Although the involvement of the Notch pathway in several areas of vascular biology is now clearly established, its role in vascular inflammation at the endothelial level remains to be elucidated. In this study, we demonstrated that proinflammatory cytokines drive a specific regulation of the Notch pathway in vascular endothelial cells (ECs). In arterial ECs, $\text{TNF}\alpha$ strongly modulates the pattern of Notch expression by decreasing Notch4 expression while increasing Notch2 expression. Changes in Notch expression were associated with a reduction in *hes1* and *hey2* expression and in CBF1 reporter gene activity, suggesting that $\text{TNF}\alpha$ regulates both Notch expression and activity. Notch2 and Notch4 regulations occurred independently and were found to be mostly mediated by the $\text{NF}\kappa\text{B}$ signaling pathways and PI3-kinase signaling pathways, respectively. Functionally, TNF -mediated Notch regulation promotes caspase-dependent EC apoptosis. Finally, our findings confirmed that dysregulated Notch signaling also occurs upon inflammation *in vivo* and correlates with caspase activation and apoptosis. In conclusion, inflammatory cytokines elicit a switch in Notch expression characterized by Notch2 predominance over Notch4 leading to a reduced Notch activity and promoting apoptosis. Thus, here we provide evidence for a role of soluble mediators of inflammation (i.e. cytokines) in the regulation of Notch signaling and for the implication of a dysregulated Notch pathway to endothelial and vascular dysfunction.

(211 words)

1. Introduction

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3 Notch signaling is an evolutionarily conserved pathway that allows cell communication
4 through molecular cell/cell interactions [1]. Notch encodes a single pass transmembrane
5 protein with epidermal growth factor (EGF) repeats in the extracellular domain and ankyrin
6 repeats in the intracellular domain that can binds to two different ligands, Delta and
7 Serrate/Jagged. Vertebrates express multiple Notch receptors (Notch 1 to 4) and ligands
8 including Delta-like (DII) 1, 3 and 4, and Jagged 1 and 2. The Notch receptors undergo three
9 successive cleavages before allowing transcription of downstream targets. The first
10 proteolytic event occurs in the trans-Golgi network by a furin-like convertase and leads to the
11 cell surface presentation of a functional heterodimeric form of the receptors. The second
12 cleavage, mediated by a disintegrin and metalloprotease (ADAM) family member, occurred
13 after interaction with a ligand expressed on neighboring cells. Finally, the γ -secretase
14 complex allows the cytoplasmic release of the intracellular domain of the receptor. This
15 fragment is then translocated into the nucleus where it binds to the mammalian transcription
16 factor CBF1/RBP-J κ docked in a transcriptional repressor complex. This interaction
17 ultimately leads, through displacing the silencing complex and by the recruitment of
18 coactivator factors, to the expression of primary target genes such as the hes and herp/hey
19 genes [2]. Many studies have reported that the Notch pathway plays a fundamental role in
20 drosophila and mammal development [1]. More recently, it was shown that Notch also plays
21 major roles in the adult in several contexts involving cell plasticity, such as proliferation,
22 oncogenesis [3], immune recognition [4], and angiogenesis [5].

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48 Endothelial cells (ECs) control vascular tone, leukocyte adhesion, coagulation and
49 thrombosis by a fine-tuned regulation of many cell surface and soluble molecules [6]. EC
50 activation is considered to be an early event which subsequently leads to EC dysfunction and
51 ultimately to vascular injury, key events associated with acute and chronic inflammation,
52 including sepsis, atherosclerosis and acute vascular and chronic allograft rejection [7] [8]. EC
53 changes involve membrane damage, increased permeability, swelling, apoptosis and
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1 necrosis. The EC loss of function could be as a result of changes in hemodynamic forces
2 (shear and/or hoop stress), direct drug-induced cytotoxicity, mechanical device implant-
3 induced injury and/or immune-mediated mechanisms [9] [10]. Inflammatory signaling
4 cascades alter EC integrity by enhancing expression of cellular adhesion molecules,
5 activation of cytotoxic T cells and/or induction of antibodies directed against EC surface [7].
6 Local release of inflammatory cytokines, including $\text{TNF}\alpha$ and $\text{IL-1}\beta$, and chemokines activate
7 ECs to upregulate soluble adhesion molecules, activate neutrophils and generate reactive
8 oxygen species that amplify the initial inflammation leading to dysregulated apoptosis,
9 secondary necrosis and overt vascular injury lesions. Considering the role of the endothelium
10 in the initiation and propagation of vascular wall injury, there is a need for the discovery of
11 molecular targets to serve as inhibitors of EC activation, dysfunction and vascular injury [6].
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25 Both embryonic and adult ECs express Notch receptors and Notch ligands [2]. Notch
26 signaling has been extensively implicated in endothelial cell-fate determination along
27 vasculogenesis and angiogenesis [11]. Several studies examining the effects of activated
28 Notch signaling on EC phenotype and function have identified potential mechanisms
29 including endothelial-to-mesenchymal (EMT) transformation [12], EC proliferation [13] and
30 control of apoptosis [14]. Recent findings further suggest a potential role for deregulated
31 Notch signaling in tumor angiogenesis and metastasis [15]. It has also been reported that
32 Notch may be necessary for the establishment and/or maintenance of quiescent EC
33 phenotype [16]. However, implication of Notch signaling in activated EC phenotype and
34 function upon inflammation has not been documented.
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48 In previous studies, we investigated signaling pathways regulated by $\text{TNF}\alpha$ in vascular
49 ECs [17-20]. Of particular interest, we have shown that the desintegrin and metalloproteinase
50 known as Kuzbanian or ADAM-10 is strongly upregulated at mRNA and protein level in ECs
51 activated with $\text{TNF}\alpha$ [17]. ADAM-10 is involved in the processing of Notch receptors and
52 ligands [21], suggesting a potential crosstalk between TNF signaling and Notch pathway that
53 may contribute to changes in EC phenotype and functions. We also reported on the
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1 contribution of Notch signaling in transplant arteriosclerosis and endothelial injury [22, 23]. In
2 this study, we investigated the regulation of Notch receptors and effector molecules in human
3 vascular ECs upon stimulation with TNF α and other pro-inflammatory mediators *in vitro* and
4 *in vivo*. Moreover, the overall Notch activity and the respective involvement of TNF α -
5 mediated signaling pathways, including NF κ B, PI-3 kinase and JNK MAPK, in Notch
6 regulation was also examined.
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2. Material and Methods

2.1 Cell culture and reagents

Primary cultures of human ECs issued from segments of renal artery (HAEC) or from human umbilical veins (HUVEC) were isolated and cultured as we previously reported [19]. ECs were grown in endothelial basal growth medium (ECBM, Promocell, Heidelberg, Germany) supplemented with 10% fetal calf serum (FCS), 0.4% EC growth supplement/heparin, 0.1 ng/mL human epidermal growth factor, 1 ng/mL human basic fibroblast growth factor, 1 μ g/mL hydrocortisone, 50 μ g/mL gentamicin, and 50 ng/mL amphotericin B (Promocell). Before activation, confluent EC monolayers were growth factor and serum depleted by culture for 24 h in basal ECBM supplemented with only 2% FCS. For activation, confluent EC monolayers were cultured with 100 U/mL recombinant human gamma interferon (IFN γ) (Imukin, Boehringer Ingelheim, Germany) or human tumor necrosis factor-alpha (TNF α) (provided by Professor P. Neuman, BASF, Ludwigshafen, Germany). Human Recombinant IL1 β (R&D Systems, Abingdon, UK) was used at 5 ng/mL. For inhibition experiments, SP600125 (10 μ M), N-acetyl-cysteine (NAC, 10 mM), pyrrolidine dithiocarbamate (PDTC; 100 μ M) and wortmannin (100 nM) (all purchased from Sigma–Aldrich, Saint Quentin Fallavier, France) were added to cells 1 h before TNF α treatment.

2.2. Recombinant adenovirus, SiRNAs, plasmids and transfection

The recombinant adenovirus for Notch2NICD and GFP (AdN2ICD) was generated as we previously described [23] and produced in the 293 cells by the vector core laboratory of the University Hospital of Nantes (INSERM UMR649 Gene Therapy Laboratory, Nantes, France). The recombinant adenovirus AdTrack-GFP was used as control (AdGFP). Adenoviral infection was carried out in ECGM supplemented with 1% FCS for 3h at 37°C, 5% CO $_2$ under agitation. Transduction efficiency was analyzed 24h after infection through GFP

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detection by direct microscopy imaging and Flow Cytometry using a FACScalibur® (BD Biosciences, Franklin Lakes, NJ, USA).

For gene silencing, cells were transfected according to manufacturer's recommendations with RNAiMax lipofectamine® (Invitrogen, Cergy Pontoise, France) and siRNA targeting Notch4 (#107458, 95% knockdown, #107459, 74% knockdown), or a scrambled negative control (#AM4611) (Ambion, Austin, TX, USA) at a final concentration of 10nM. Specific expression knockdown by siRNA was attested by qRT-PCR and functional assays were assessed 48h post-transfection.

2.3 Semi-quantitative RT-PCR, quantitative real-time PCR and Southern blotting

RNA was isolated using Trizol reagent (Invitrogen, Carlsbad, CA, USA) and treated with DNase (Ambion) before reverse transcription (RT). Subsequent to RT, cDNA were amplified by PCR and analyzed in agarose gels stained with ethidium bromide. For semiquantitative RT-PCR, PCR conditions were optimized for each primer set and performed for 18 to 35 cycles of amplification to allow semiquantitative analysis (**Table 1**). For Southern blotting, PCR products were purified according to the nucleospin extract II protocol (Macherey-Nagel, Hoerd, France). Purified amplicons were then cloned into the TOPO TA cloning® vector (Invitrogen) and sequenced before their use as cDNA probes. Southern blotting and hybridizations were performed as we previously described [17]. Quantitative PCRs were performed using the ABI PRISM 7700 sequence detection application program (PE Applied Biosystems, Foster City, CA, USA). For quantification, duplicates were normalized by the concomitant quantification of hypoxanthine-guanine phosphoribosyl transferase (HPRT). Normalization was made with the control samples in the human cells and with an additional reference sample for the rat study. Relative expression was calculated according to the $2^{-\Delta\Delta C_t}$ method, as previously described [24]. Custom primers were obtained from MWG (High Point, NC, USA) and used for semiquantitative PCR and qPCR (**Table 1**). Transcript levels were quantified by qRT-PCR with the following primers and probe from Applied Biosystems (Foster City, CA, USA): Notch1 (Hs00413187_m1), Notch2

1 (Hs00225747_m1; Rn00577522_m1), Notch4 (Hs00270200_m1; Rn01525737_g1), hey1
2 (Hs00232618_m1), VCAM-1(Hs00365486_m1; Rn00563627_m1) and HPRT
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4 (Rn01527838_g1).
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7 *2.4. Immunoblotting*

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10 Cells were lysed on ice in 20 mmol/L Tris-HCl (pH 7.4), 137 mmol/L NaCl, 0.05%
11 Triton X-100, 1 mmol/L phenylmethylsulfonyl fluoride supplemented with protease inhibitors
12 (PIC, Sigma–Aldrich). Lysates were resolved by sodium dodecyl sulfate–polyacrylamide gel
13 electrophoresis (6%–10%) and subjected to Western immunoblot analysis using specific
14 antibodies against Notch2 (C651.6DbHN, Developmental Studies Hybridoma Bank, IA,
15 USA), Notch4 (Santa Cruz Biotechnology, Santa Cruz, CA, USA), total and cleaved caspase
16 3 and 7 (Cell Signaling Technology, St Quentin-en-Yveline, France) and tubulin (Oncogene,
17 MERCK EuroLab, Val de Fontenay, France), and secondary horseradish peroxidase-labeled
18 anti-rabbit, anti-mouse, or anti-goat antibodies (Cell Signaling Technology). Antibody-bound
19 proteins were detected using an enhanced chemiluminescence kit (ECL kit, Amersham, Les
20 Uis, France).
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37 *2.5. Apoptosis Assays*

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39 *Measurement of caspase activity* - Caspase activity was analyzed by western blot using
40 antibodies against cleaved Caspase-3 (Asp175), cleaved Caspase-7 (Asp198), and
41 antibodies against total caspase-3 and -7 (all from CST).
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46 The cell-permeable fluorogenic substrate PhiPhiLux-G2D2 (OncoImmunin, Gaithersburg,
47 MD) containing the cleavage site DEVD was used to monitor caspase-3-like activity in intact
48 cells. ECs were incubated with the substrate solution for 1 h at 37°C in the dark, according to
49 the manufacturer's instructions. Caspase 3 activation/apoptosis was examined during 18h by
50 time lapse imaging using a microscope DMI6000B (Leica Microsystems SAS. Rueil
51 Malmaison) equipped with an objective lens X40 (HCX FL Plan), and a CCD camera
52 (Coolsnap HQ2, Photometrics Roper Scientifics SAS Evry). Caspase 3 positive cells/field
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2 were counted every 2 h between 48h and 66h post-infection with AdN2ICD and controls
3 adenovirus. Results were expressed as the percentage of caspase-positive ECs.
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5 *2.6 Animal model of vascular inflammation*

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8 The care and use of animals in this study complied with institutional guidelines. Male
9 Sprague Dawley rats (300 to 400 g body weight) purchased from Charles River (Saint-Aubin,
10 les Elbeuf, France) were injected intravenously, under anesthesia, with 10 µg/kg of
11 recombinant rat TNF α (PreproTech, Neuilly-Sur-Seine, France) or PBS as vehicle. Animals
12 were euthanized for organ collection 1, 4 or 6 h after treatment. Rat RNA and proteins were
13 isolated using TriZol® (Invitrogen) and RIPA (0.5% sodium deoxycholic acid, 0.1% SDS, 1%
14 NP40, PBS, protease inhibitors) buffers, respectively, and were then treated as reported
15 above.
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26 *2.7 Statistics*

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29 Data are represented as means \pm SE for replicates experiments (n=3 independent
30 experiments). Statistical analysis was performed on Graphpad Prism® Software (Graphpad
31 Software, San Diego, CA) with the parametric or Kruskal Wallis non parametric analysis of
32 variance test as appropriate. p<0.05 was considered statistically significant.
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3. Results

3.1. Constitutive and cytokine-regulated expression of the Notch2 and Notch4 receptors in vascular cells

To further characterize the vascular changes in expression and activity of the Notch pathway molecules upon inflammatory processes, we sought to analysis the pattern of mRNA levels for Notch receptors in resting and cytokine-activated vascular cells. To this aim, primary cultures of ECs from two different vascular beds (HAEC from arteries and HUVEC from veins) were treated for 0 to 24 h with recombinant tumor necrosis factor-alpha (TNF α) and transcript levels were analyzed by semi-quantitative RT-PCR. As shown in **Figures 1A and 1B**, transcripts for the receptor Notch4 were detected in untreated vascular ECs whereas only minimal levels of Notch2 mRNA were found, indicating that quiescent ECs differentially express Notch receptors. TNF elicits an upregulation of Notch2 and a downregulation of Notch4 in both HAEC and HUVEC.

Quantitative PCRs confirmed that, in ECs, TNF α selectively modulates the mRNA steady-state levels for the Notch receptors. TNF α decreased transcript levels for Notch4 with a significant effect starting 2 h after treatment and maximal inhibition of $78 \pm 2\%$ as compared with basal levels (*p < 0.05) (**Figure 1C**). In contrast, an enhanced mRNA level for Notch2 was found in response to TNF α , corresponding to a maximal 3.3 ± 0.3 -fold increase at 24 h (*p < 0.05) as compared to the basal mRNA level. Western blotting analysis (**Figure 1D**) indicates that regulation in of Notch2, and 4 protein level paralleled changes in steady state mRNA levels for these molecules (up to 2.69 ± 1.26 and 0.28 ± 0.02 –fold the baseline for Notch 2 and Notch 4, respectively; *p<0.05), suggesting that TNF α triggers an effective and selective Notch regulation at both mRNA and protein levels in ECs.

Next, we tested whether Notch expression could be regulated by cytokines other than TNF α . To address this question, qRT-PCR was used to compare mRNA levels for Notch2 and Notch4 in ECs treated with the cytokines TNF α , interleukin-1 β (IL1 β) and interferon γ

1 (IFN γ) (**Figure 2**). A comparable regulation in both time course and magnitude was observed
2 for Notch2 in ECs activated with TNF α , IL1 β or IFN γ (up to a 2.3 ± 0.3 -fold increase for
3 Notch2 as compared with untreated cells). Notch4 exhibited the same pattern of regulation
4 upon TNF α and IL1 β stimulation, with a maximal decrease in mRNA of $65 \pm 5\%$ and $69 \pm 1\%$
5 for TNF α and IL1 β , respectively. Regulation of vascular cellular adhesion molecule-1
6 (VCAM-1) is shown as a control of EC activation. Similarly to VCAM-1, IFN γ had no
7 significant effect on the reduction of Notch4 transcripts.
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19 *3.2. TNF α -mediated regulation of Notch effectors and impact on basal Notch activity* 20 *in cultured ECs.*

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23 Expression of Hairy/Enhancer of split (hes) and Hairy-related (hey/hrt/herp)
24 transcription factors has been shown to be regulated by activation of Notch receptors [25].
25 Consequently, activity of the Notch pathway should be reflected by the expression of the hes
26 and hey transcripts. Cells were incubated with recombinant TNF α for 0 to 24 h and mRNA
27 levels for hes-1, 2, 3, 4, 5, 6, and 7 and hey 1, 2 and 3 were analyzed by RT-PCR. As shown
28 in **Figures 3A**, ECs basally express significant levels of hey1, hey1 and hes1 mRNAs. We
29 found that transcript levels of hes2, 4 and 6 were lower than hes1/hey1-2 (detection achieved
30 at >30 cycles of PCR amplification, see also Table 1). In addition, no mRNA for hes3, 5 or 7
31 or hey3 was detected by RT-PCR (at 35 PCR cycles), suggesting that these effectors
32 molecules are weakly expressed in ECs and play minor roles in Notch signal in ECs (data
33 not shown) as compared to hes1/hey1-2. Among these molecular targets of Notch activity,
34 only hey1 was found upregulated in TNF α -activated ECs. In contrast, after TNF α treatment,
35 hey2 and hes1 showed a significant decrease in their expression.
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55 In parallel, the expression of effector molecules in response to TNF α , IL1 β and IFN γ
56 was investigated. As shown in **Figure 3**, a drastic down-regulation of both hes-1 and hey-2
57 was found in response to TNF α ($79 \pm 1\%$ and $78 \pm 1\%$ inhibition at 24 h, respectively).
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1 Similarly, IL1 β induced a comparable regulation pattern with a maximal inhibition achieved at
2 6 h for hes1 and at 12 h for hey2 (52 ± 3 % and 71 ± 4 % of decrease, respectively, as
3 compared to untreated cells). In addition, the enhancement of hey1 (2.2 ± 0.3 -fold increase
4 versus control) by TNF α was further observed with IL1 β (6.5 ± 1.6 -fold increase as compared
5 to basal expression level). However, no significant regulation of these effector genes was
6 obtained after IFN γ treatment, suggesting selective Notch receptors/effectors regulations and
7 therefore functions in response to inflammatory stimuli.
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16 3.3. TNF-mediated regulation of Notch2 and Notch4: involvement of NF κ B, PI-3 17 kinase and MAP kinase signaling pathways. 18 19 20 21

22 The selective effects of inflammatory cytokines by TNF α , IL1 β and IFN γ on Notch
23 molecules also suggest that specific signaling pathways are implicated in this process. In
24 ECs, TNF α activates several signaling pathways including the phosphatidylinositol 3-kinase
25 (PI3-K), nuclear factor- κ B (NF κ B) and mitogen-activated protein kinase (MAPK) pathways
26 [26]. The respective involvement of these pathways in Notch regulation mediated by TNF α in
27 ECs was examined. For this purpose, HAECs were pretreated with or without signaling
28 pathway inhibitors (N-acetyl cysteine (NAC), PDTC, wortmannin and SP600125) for 1 h and
29 then activated with TNF α for 24 h, a time point leading to maximal regulation as shown
30 above. Transcript levels for Notch2 and 4 were determined by qRT-PCR.
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42 We found that blocking PI3-K using wortmannin does not affect Notch4
43 downregulation triggered by TNF α (**Figure 4**). In contrast, Notch2 transcriptional
44 upregulation was totally inhibited. Inhibition of the PI3K as well as inhibition of JNK also
45 significantly prevents transcriptional regulation of hey1. An efficient prevention of TNF α -
46 dependent Notch4 and hes1 downregulation was obtained after NF κ B inhibition using PDTC
47 or NAC. The blockade of c-Jun N-terminal kinase (JNK) MAPK with SP600125 has no effect
48 on Notch 2, Notch4 and hes1 suggesting that this pathway is not involved. Overall, our data
49 demonstrated for the first time that opposite regulations of Notch2 and Notch4 in activated
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1 EC require selective signaling pathways suggesting that Notch receptors exert non
2 redundant, complementary, functions upon inflammation. Moreover, our data showing that
3 hes/hey effector molecules are also selectively regulated by the NF κ B and PI3Kinase
4 pathways substantiated the hypothesis that Notch receptors control specific functions
5 through the regulation of selected effectors (i.e. Notch2/hey1, Notch4/hes1).
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10 Next, we used silencing experiments to mimic the changes in Notch4 expression mediated
11 by TNF α in vascular ECs. To this aim, silencing of Notch4 was achieved using two different
12 siRNAs targeting Notch4(SiN4#1 and SiN4#2) or a scramble non targeting SiRNA (scramble)
13 as we previously described [22]. Cells were then analyzed by qPCR for Notch2 and Notch4
14 transcript levels. Notch1 mRNA was used as a control for potential off-target effects. We
15 found that silencing Notch4 has no significant effect on Notch1 and Notch2 expression
16 indicating that downregulation of Notch4 doesn't trigger the regulation of Notch2. Conversely,
17 we also observed that silencing or overexpressing Notch2 has no regulatory effect of Notch4
18 expression in ECs (data not shown).
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31 Overall, our findings suggest that TNF α -mediated changes of Notch2 and 4 transcription
32 may occur independently and are dependent, at least partially, on the PI3K and NF κ B
33 pathways and support a major role for NF κ B in the control of Notch4 and hes1 [27].
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39 *3.4. Endothelial changes in Notch2 and Notch4 expression promote EC apoptosis.*

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41 Next, to functionally assess the impact of Notch regulation, apoptosis assays were
42 performed after modulation of Notch2 and/or Notch4 in cultured ECs. We used gene transfer
43 to mimic the changes in both Notch2 and Notch4 mediated by TNF α in vascular ECs. To this
44 aim, silencing of Notch4 was achieved as above using siRNAs while Notch2 was modulated
45 and activated using an adenoviral vector encoding Notch2-ICD and GFP (AdN2ICD) as we
46 previously described [23]. Controls include a non targeting siRNA (scramble) and a
47 recombinant adenovirus for the reporter gene GFP (AdGFP). Transduced and knock-down
48 cells were then analyzed by Western blots for caspase activation. We found that silencing
49 Notch4 and overexpression of Notch2 (NICD) similarly induce the cleavage of caspase-3
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(Asp175) and caspase-7 (Asp198) indicating that both events are pro-apoptotic in vascular ECs (**Figure 5A**). To confirm these results, caspase-3 activity was monitored in live ECs by videomicroscopy using a cell permeable substrate (PhiPhiLux ®) to detect real-time activation of caspase (Figures 5B, 5C, 5D, 5E). The PhiPhiLux probe becomes fluorescent (red) when cleaved by active caspase-3. For these experiments, ECs were silenced for Notch4, transduced using AdN2ICD or both. Higher basal caspase-3 activity in treated cells compared to control reflects the pro-apoptotic effect of Notch modulation (Figures 5B&5C). Consistent with immunoblotting, we show a time-dependent increase in caspase-3 activity in ECs with a sustained Notch2 NICD expression or with a knocked-down for Notch4. Moreover, we found that combination of both further elicits caspase-3 activity suggesting that despite partly independent regulation of Notch2 and 4, apoptosis is a common effector mechanism. These data were further confirmed by annexin V labeling and facs analysis (data not shown).

3.5. Modulation of Notch2 and Notch4 in vascular inflammation *in vivo*.

In order to establish a preliminary evidence *in vivo* for the biological relevance of our *in vitro* findings related to Notch regulation in activated ECs, we investigated Notch expression in rats treated with recombinant TNF α . EC activation was assessed by measuring VCAM-1 expression, a representative marker of EC activation *in vitro* and *in vivo* [7]. To induce vascular inflammation and EC activation, rats were treated intravenously with recombinant TNF α or vehicle as control. At 0 to 6 h postinjection, kidney, heart and lung were collected for analysis. First, basal expression of Notch2 and 4 transcripts in the different tissues from untreated rats was compared by qRT-PCR. As shown in **Figure 6A**, the transcript level of Notch2 and 4 molecules varied greatly according to tissues, with the highest expression levels consistently observed in the lung. The lower levels of Notch2 and 4 transcripts were found in the heart. Ratios of expression levels in the lung compared with the heart were 16.2 ± 3.0 -fold (*p < 0.05) for Notch2, and, 6.4 ± 1.1 -fold (*p < 0.05) for Notch4.

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Expression of Notch receptors was further examined in lung from rats treated with TNF α for 1, 4 and 6 h (**Figure 6B**). Notch2 mRNAs were significantly and transiently increased (1.8 ± 0.1 -fold increase as compared to untreated rats; $p < 0.05$ at 1 h). In contrast, Notch4 was downregulated upon TNF α treatment, with a maximal 2.2 ± 0.1 -fold decrease in mRNA level, ($p < 0.05$ versus control). Western blotting for Notch2 and Notch4 further confirmed the respective up- and down-regulation at the protein level in tissues (**Figure 6C**). We also performed immunochemistry analysis on lung sections and we confirmed the decrease at endothelial level of Notch4 in TNF-treated animals (data not shown). Unlike Notch4, Notch2 is ubiquitously expressed in cells and tissues and we were not able to appreciate a clear quantitative increase in Notch2 expression in the endothelium (data not shown). Immunoblotting experiments also associated Notch regulation with pro-apoptotic events reflected by the activation of caspase-3 and caspase-7. The 19-kDa form of cleaved caspase-3 (Asp175) and the 20-kDa form of cleaved caspase-7 (Asp198) were detected in lung from TNF-treated but not in controls (**Figure 6D**).

4. Discussion

Although the impact of the Notch pathway in several areas of vascular biology is now clearly established, its role in vascular inflammation at the endothelial level remains to be elucidated. A large number of studies demonstrated, mostly through modulation of Notch pathway activity, that Notch is involved in EC differentiation, apoptosis and proliferation [12] [16] [28]. In addition, recent studies also investigated the effect of various effectors, such as soluble mediators of cell growth (VEGF or FGF-2) [28] [29], differentiation (TGF β) [30] or activation (LPS) [31] on the Notch pathway in various cell types. In the present study, we asked whether inflammatory mediators could also modulate the Notch signaling and the pattern of Notch molecules expressed in vascular endothelial cells. To this aim, the effect of cytokines on Notch receptors expression and on Notch activity in human ECs was examined.

Here, we demonstrated that TNF α , the prototype of pro-inflammatory cytokines, drives a specific regulation of the Notch pathway in vascular ECs. In arterial ECs, TNF α strongly

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modulates the pattern of Notch molecules expression by decreasing Notch4 expression while increasing Notch2 expression. Changes in Notch levels were further observed at the protein level, and were associated with a reduction in *hes-1* and *hey-2* expression and CBF1 reporter gene activity as previously reported [23], suggesting that inflammation regulates both Notch expression and activity. Interestingly, regulation of Notch4 expression seems to be cytokine-specific since no regulatory effect was found in response to IFN γ , similar to VCAM-1 that is not affected by IFN γ . This TNF α -driven transcriptional regulation was found to be mostly mediated by the NF κ B and the PI3-kinase signaling pathways. *In vivo* analysis confirmed that in the lung, TNF α regulates Notch2 and Notch4 at both transcriptional and protein levels.

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Four distinct Notch receptors, Notch1, 2, 3, and 4, and five different Notch ligands, Jagged-1 and 2, and Delta-1, 3, and 4, have been identified and characterized in mammals. ECs express endothelium-specific Notch members, including Notch4 and Dll-4. However, whether normal, quiescent, human ECs express basal levels of other Notch receptors and ligands is not clearly established. Here we show that Notch2 is also expressed in cultured ECs and is upregulated in response to TNF α . However, the concomitant downregulation of Notch4 expression and Notch activity may suggest that Notch4 is the major Notch receptor in arterial ECs or that Notch2 partly exerts its functions by a non-canonical mechanism.

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In contrast to ECs and consistent with previous data [32], we also found that vascular SMCs express Notch2 and Notch3 but not Notch4 at mRNA level. Consistent with our results on ECs, vSMCs responded to TNF α with a significant upregulation of Notch2 (about a 4.2-fold increase as compared to untreated cells) and a strong downregulation of Notch3 expression (**data not shown**).

Associated with the constitutive expression of Notch receptors, we found a basal expression of a selective pattern of effector molecules of the *Hairy/Enhancer of split* (*Hes*)

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2 and Hairy-related transcription factors (Hey, also known as Hrt, Hesr, Hey, CHF, grl, and
3 Herp) family. Previous studies showed basal transcript levels for hes1, hey1 (herp2, hrt1,
4 hesr1) and hey2 (herp1, hrt2, hesr2) in ECs [25, 33]. Consistent with these results, we
5 reported significant expression of hes-1 and 2 and hey-1 and 2 associated with a basal
6 CBF1/luciferase activity (data not shown), confirming that endogenous Notch activity occurs
7 in quiescent ECs and is probably implicated in the maintenance of endothelium quiescence
8 [34]. A microarray comparison of large series of human EC lines confirmed arterial-specific
9 expression for hey2 [35]. Further, those authors showed that ectopic expression of hey2 in
10 HUVECs specifically induces expression of a series of genes that are characteristic of
11 arterial endothelia, implicating hey2 as a key regulator of the arterial phenotype. Consistent
12 with our results, Espinosa et al. provided evidence that $TNF\alpha$ triggers an important decrease
13 in the level of hes1 mRNA, while a lower effect was found on hey1 [36]. Our findings further
14 indicate that, consistent with an overall decreased expression for the major effector
15 molecules hes1 and hey1, $TNF\alpha$ reduces basal CBF1 reporter activity in activated ECs.
16 Considering that CBF1 activity reflects canonical Notch pathway activity, we may extrapolate
17 that $TNF\alpha$ decreases Notch activity in ECs.
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36 The functional consequences of Notch modulation mediated by $TNF\alpha$ in the endothelium
37 appear to promote EC apoptosis. Notch4 has been implicated in the control of proliferation,
38 apoptosis and migration of SMCs and ECs [14, 16, 37]. Notch2 has mostly been involved in
39 monocyte and T lymphocyte maturation and differentiation [38-40]. Its role in EC biology is
40 still unclear. We recently demonstrated that Notch2 signaling sensitizes EC to apoptosis [23].
41 TNF elicits a broad array of cellular effects via two receptors TNFR1 and TNFR2. TNFR1
42 mediates inflammation and cell death while TNFR2 serves to enhance TNFR1-induced
43 apoptosis or to promote cell activation, migration, growth or proliferation in a cell-specific
44 manner [41]. Here, our results suggest that concomitant changes in Notch2 and Notch4
45 expression elicited by $TNF\alpha$ may have an additive pro-apoptotic effect that triggers
46 endothelial injury and vascular damage.
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1 To conclude, inflammatory cytokines trigger a selective expression pattern of Notch
2 receptors in the endothelium associated with a reduced canonical Notch activity. Consistent
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4 with previous models where growth factors modulate the expression of Notch receptors and
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6 ligands at both qualitative and quantitative levels, our findings suggest that inflammation may
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8 provide additional control of Notch signaling.
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10 11 12 13 14 **Contributors**

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17 T.Q. contribute most of the experimental work and participated in the design or the
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19 study, data analysis and drafting the manuscript. J.D. contributed significantly to the in vitro
20
21 experiments and apoptosis assays. S.C. performed initial in vivo experiments and analysis.
22
23 B.C. performed conception and design of the study, contributed substantially to interpretation
24
25 of the data and drafting of the manuscript. All authors read and approved the final
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27 manuscript.
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46 Fondation pour la Recherche Médicale.
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57 **Abbreviations**

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ECs: endothelial cells; HAECs: human arterial ECs; HUVECs: human umbilical vein

ECs; ICD: intracellular domain; SMCs: smooth muscle cells.

Figure Legends

Figure 1.

Expression of Notch receptors in resting and TNF α -activated endothelial cells.

Confluent cells monolayers were incubated with 100 U/mL TNF α for the indicated periods.

Cells were lysed in parallel experiments to allow RNA and protein analysis. Semiquantitative

RT-PCR analysis of mRNA levels for Notch2 and 4 in vascular cells: HAEC (A), HUVEC (B).

PCR products were separated on 1.2% agarose gels, blotted onto nylon membranes and

hybridized with specific radiolabeled PCR probes. β -Actin mRNAs were amplified as

normalization controls. Representative autoradiographs of three independent experiments

are shown. (C) Real-time quantitative PCR for Notch2 Notch4, VCAM-1 in HAECs. Results

shown are the mean \pm SEM from three independent experiments performed in duplicates

and are expressed as relative expression, calculated according to the $2^{-\Delta\Delta Ct}$ method (*p <

0.05 versus control). (D) A representative analysis of Notch protein expression in HAEC by

Western blotting. Blots were reprobed with an anti-tubulin antibody to ensure equal loading.

(E) Quantification of western blot analysis for Notch2 and Notch4 from at least 4 independent

experiments.

Figure 2.

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Comparative effects of various cytokines on Notch signaling. HAECs were treated for 0, 6, 12 and 24 h with TNF α (100 U/mL), IL1 β (5 ng/mL) or IFN γ (100 U/mL). Transcriptional regulation was analyzed by Real-time quantitative RT-PCR for Notch2, Notch4 and VCAM-1. Results shown are the mean \pm SEM from three independent experiments performed in duplicates and are expressed as relative expression, calculated according to the $2^{-\Delta\Delta Ct}$ method, after normalization to HPRT levels (*p < 0.05 versus control).

Figure 3.

Regulatory effects of TNF on Notch effectors and Notch activity. HAECs were treated for 0, 2, 6, 12 and 24 h with TNF α (100 U/mL), IL1 β (5 ng/mL) or IFN γ (100 U/mL). (A) Expression pattern of Notch effectors hes and hey was determined by semiquantitative RT-PCR. PCR products were separated on 1.2% agarose gels and stained with ethidium bromide. β -actin mRNA was amplified as a control. Results are representative of three experiment performed. Transcriptional regulation was analyzed by real-time quantitative PCR for hes1 (B), hey1 (C) and hey2 (D). Results shown are the mean \pm SEM from three independent experiments performed in duplicates and are expressed as relative expression, calculated according to the $2^{-\Delta\Delta Ct}$ method, after normalization to HPRT levels (*p < 0.05 versus control).

Figure 4.

Signaling pathways involved in TNF α -dependent Notch regulation. HAECs were preincubated with PDTC (100 μ M), SP600125 (10 μ M) or wortmannin (100 nM) for 1 h before 24 h treatment with TNF α (100 U/mL). Cells were lysed to allow RNA analysis by quantitative RT-PCR for Notch2 (A), hey1 (B), Notch4 (C) and hes1 (D). Results shown are the mean \pm SEM from three independent experiments and are expressed as relative expression, calculated according to the $2^{-\Delta\Delta Ct}$ method, after normalization to β -actin levels. *p<0.05 versus TNF α -untreated cells (ctrl) and between non pre-treated and pre-treated TNF α -activated cells with inhibitors. (E) ECs were transfected with siRNAs targeting Notch4 (SiN4#1 and

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SiN4#2) or a non targeting scramble siRNA. Notch1, Notch2 and Notch4 mRNA steady states were analyzed by qRT-PCR 48h post-transfection. Results shown are means \pm SEM from 3 independent experiments and are express as a percentage of control expression (medium).

Figure 5.

TNF-mediated Notch regulation and EC apoptosis

(A) A representative Western blot analysis showing caspase-3 and caspase-7 cleavage in ECs silenced for Notch4 or overexpressing Notch2NICD. Immunoblotting was performed using specific anti-cleaved or total form of caspases antibodies. Blots were reprobred with anti-GAPDH antibodies (B, C, D, E) Caspase-3 activity in EC transduced with AdN2ICD or AdGFP and/or transfected with either non targeting (scramble) or Notch4 siRNAs. Caspase-3-like activity was visualized in individual, live ECs by time lapse fluorescence videomicroscopy. Cultures were incubated with cell-permeable PhiPhiLux-G₂D₂ substrate at 37°C, 5% CO₂. The quenched fluorescence PhiPhiLuxG2D2 substrate is cleaved intracellularly by caspase-3-like proteases, greatly enhancing red fluorescence. Non apoptotic ECs expressing Notch2NICD-GFP or GFP alone appeared in green while apoptotic ECs are round red fluorescent cells. ECs were examined under a $\times 20$ objective and the total number of apoptotic cells determined by counting. (B, C,D) Results are expressed as the percentage of caspase-3 positive EC (*p<0.05). (E) Representative pictures of fields analyzed.

Figure 6.

TNF α -dependent Notch regulation upon inflammation *in vivo*. Rats (n = 3) were treated intravenously with rat TNF α (10 μ g/kg) for 0, 1, 4 or 6 h. Lungs, heart and kidneys were collected and frozen for mRNA and protein analysis. Real-time quantitative PCR was used for mRNA analysis. Results shown are the mean \pm SEM of three independent experiments and are expressed as relative expression, calculated according to the 2^{- $\Delta\Delta$ Ct} method, after

1 normalization with β -actin levels (*p < 0.05 versus control). **(A)** Basal levels of Notch2 and
2 Notch4 transcripts in heart, kidney and lung from untreated rats;*p<0.05 versus transcript
3 level in heart. **(B)** Time-course analysis of transcript levels in lung in response to TNF α
4 (*p<0.05 versus untreated rats). **(C, D)** Western blotting for Notch2, Notch4, cleaved
5 (Asp175) and total caspase-3, cleaved (Asp198) and total caspase-7 and tubulin in lung in
6 response to TNF α (4h). A representative experiment out of 3 independent experiments is
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Table 1.

Oligonucleotide primer pairs and PCR conditions for semi-quantitative analysis

Target	Forward (Sequence 5'-3')	Reverse (Sequence 5'-3')	Number of PCR cycles
Notch 2	GCAGGAGGTGGATGTGTTAG	CCAGGATCAGGGGTGTAGAG	21
Notch 4	TGTTTGATGGCTACGACTGT	TCCTTACCCAGAGTCCTACC	27
hes 1	AGAGGCGGCTAAGGTGTTTG	GAGAGGTGGGTTGGGGAGTT	25
hes 2	TCATCCTGCCGCTGCTGGG	TACCCTGGAGCTGCTGAAG	30
hes 3	TCCTCCTCCCCGAAAGTCTC	CACGACCAGAACGGACGACT	35
hes 4	CTCAGCTCAAACCCCTCATC	GCGGTACTTGCCCAGAACGG	30
hes 5	TGGGGTTGTTCTGTGTTTGC	CAGACCACCAGGCACACTCA	35
hes 6	CCCTGAGGCTGAACTGAGTC	CTACCCACCCACATCTGAAC	30
hes 7	TAGGGGTGGGGTAGAGACTC	AGACAGAAGGGAAGGGAAAG	35
hey 1	CAGGCAACAGGGGGTAAAGG	GTGGAGCGGATGATGGTGGT	27
hey 2	GTCGCCTCTCCACAACCTTCA	CTGGACGTGGCTGATACTGA	27
hey 3	TGGGACAGGATTCTTTGATG	GGTAAGCAGGAGAGGAGACA	35
VCAM-1	AATGTTGCCCCAGAGATAC	TCTCCTGTCCTCGCTTTTTT	27
β -actin	TCTGGCACACACCTTCTAC	CAGCTTCTCCTTAATGTCAC	18

Figure 1

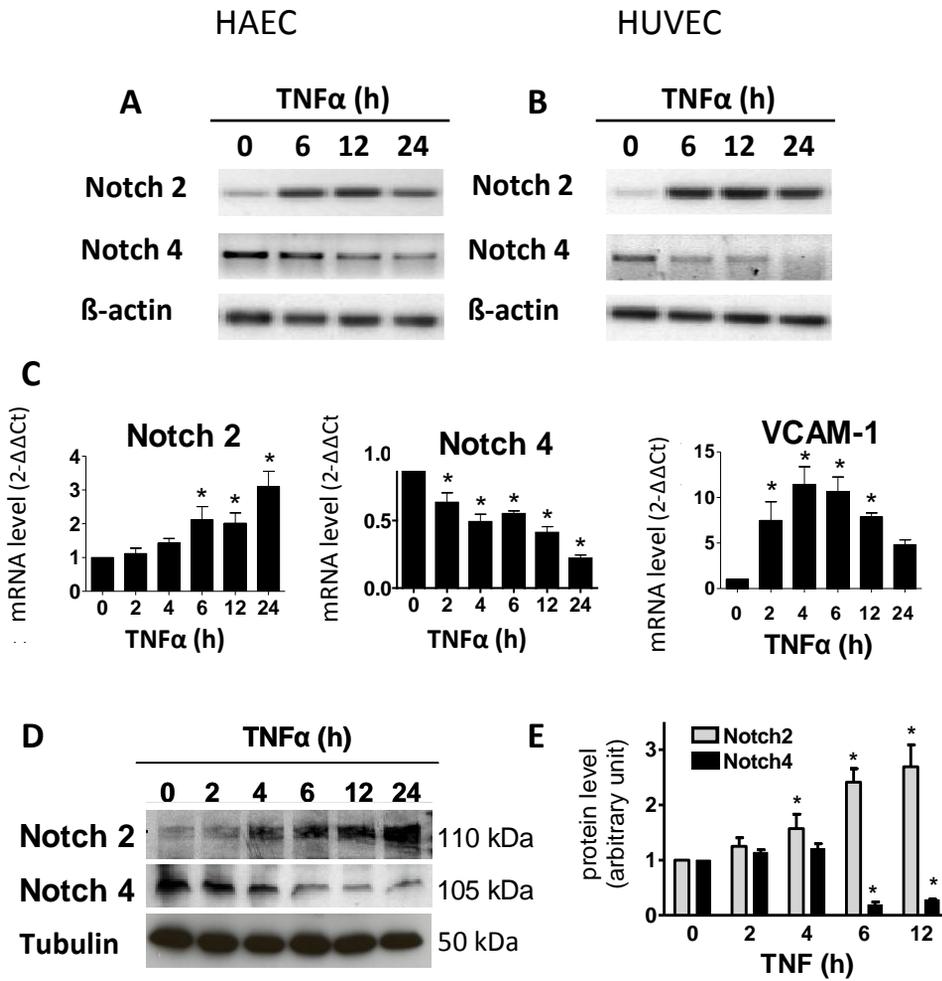


Figure 2

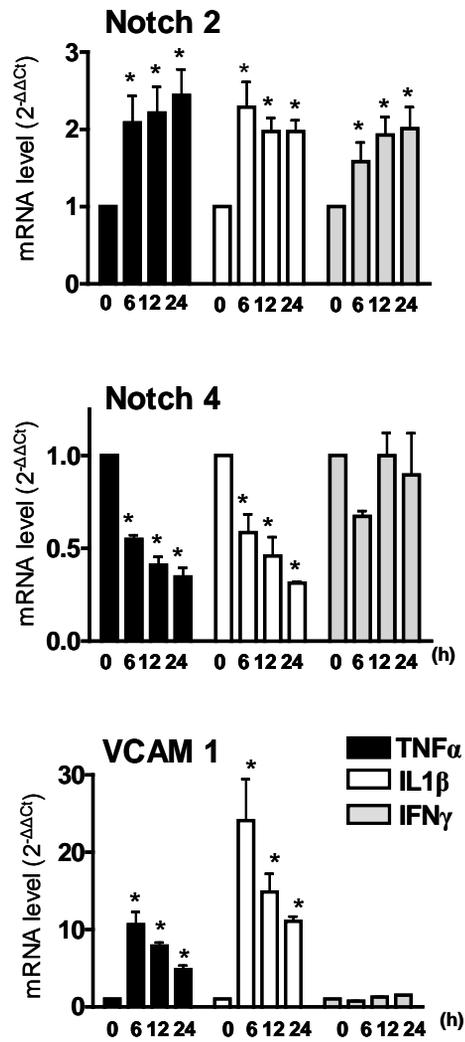


Figure 3

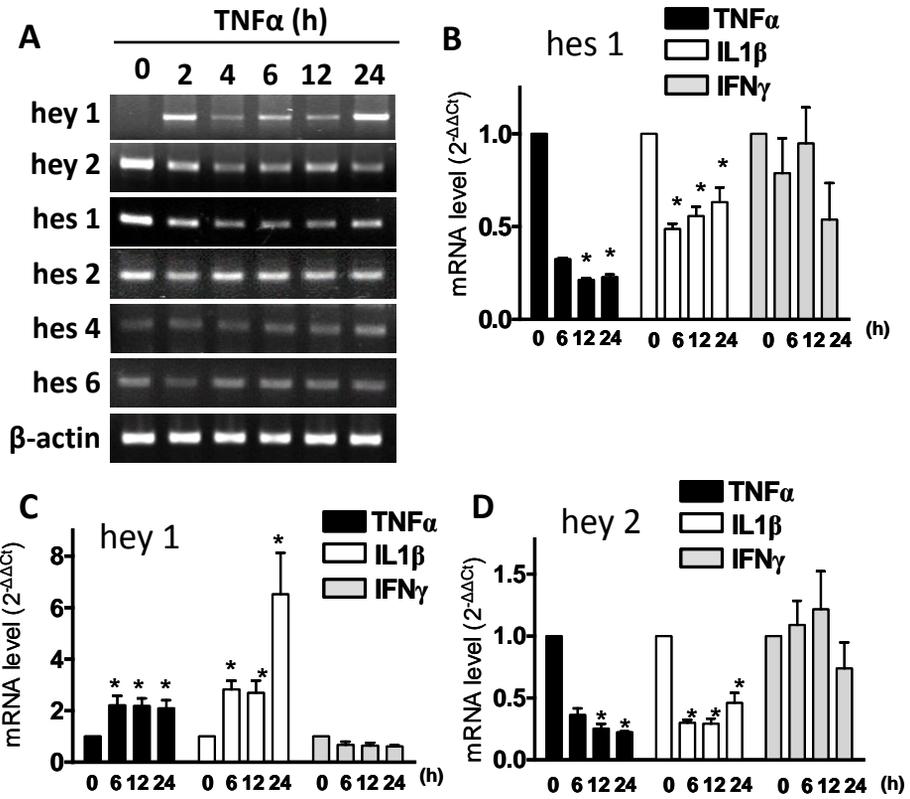


Figure 4

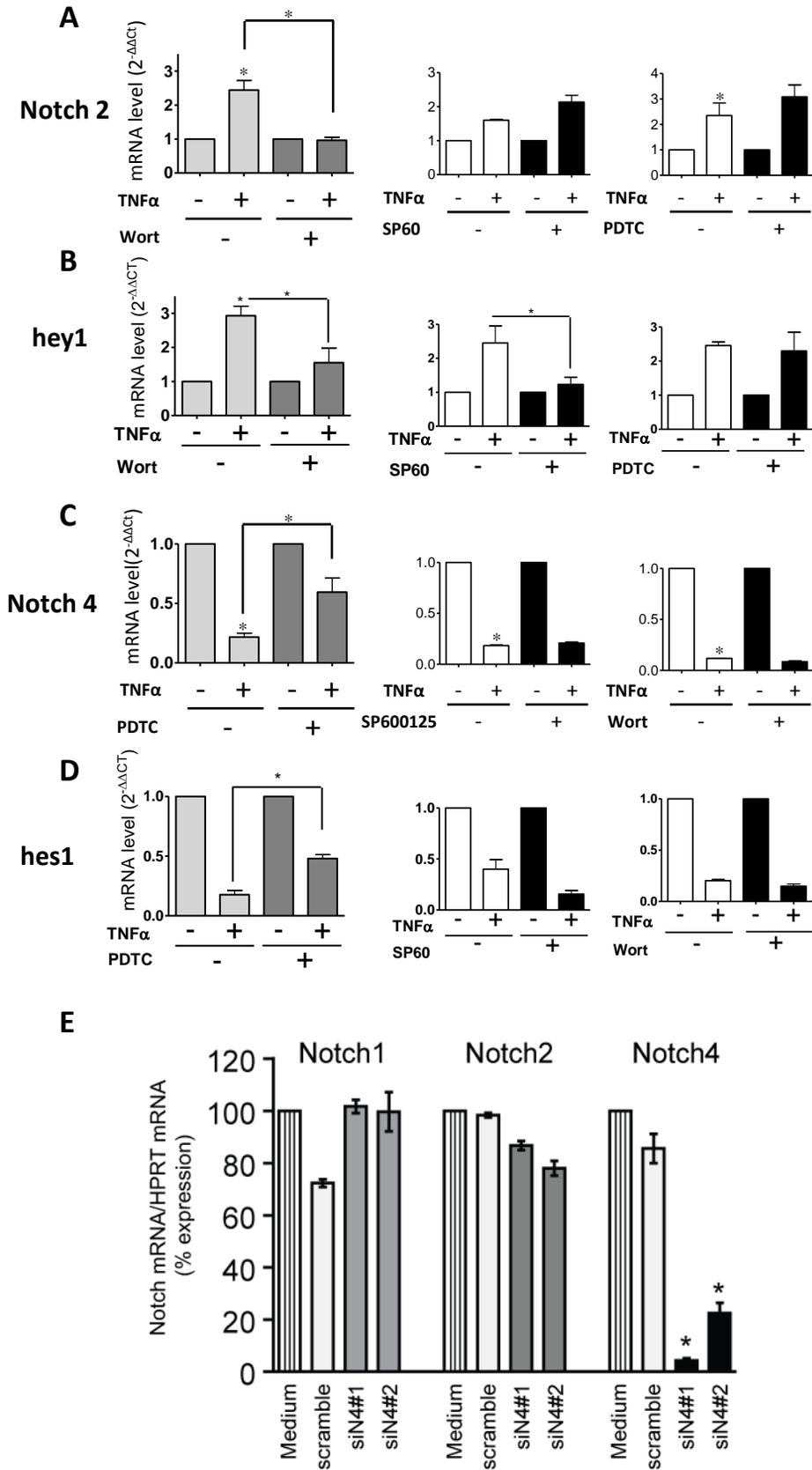


Figure 5

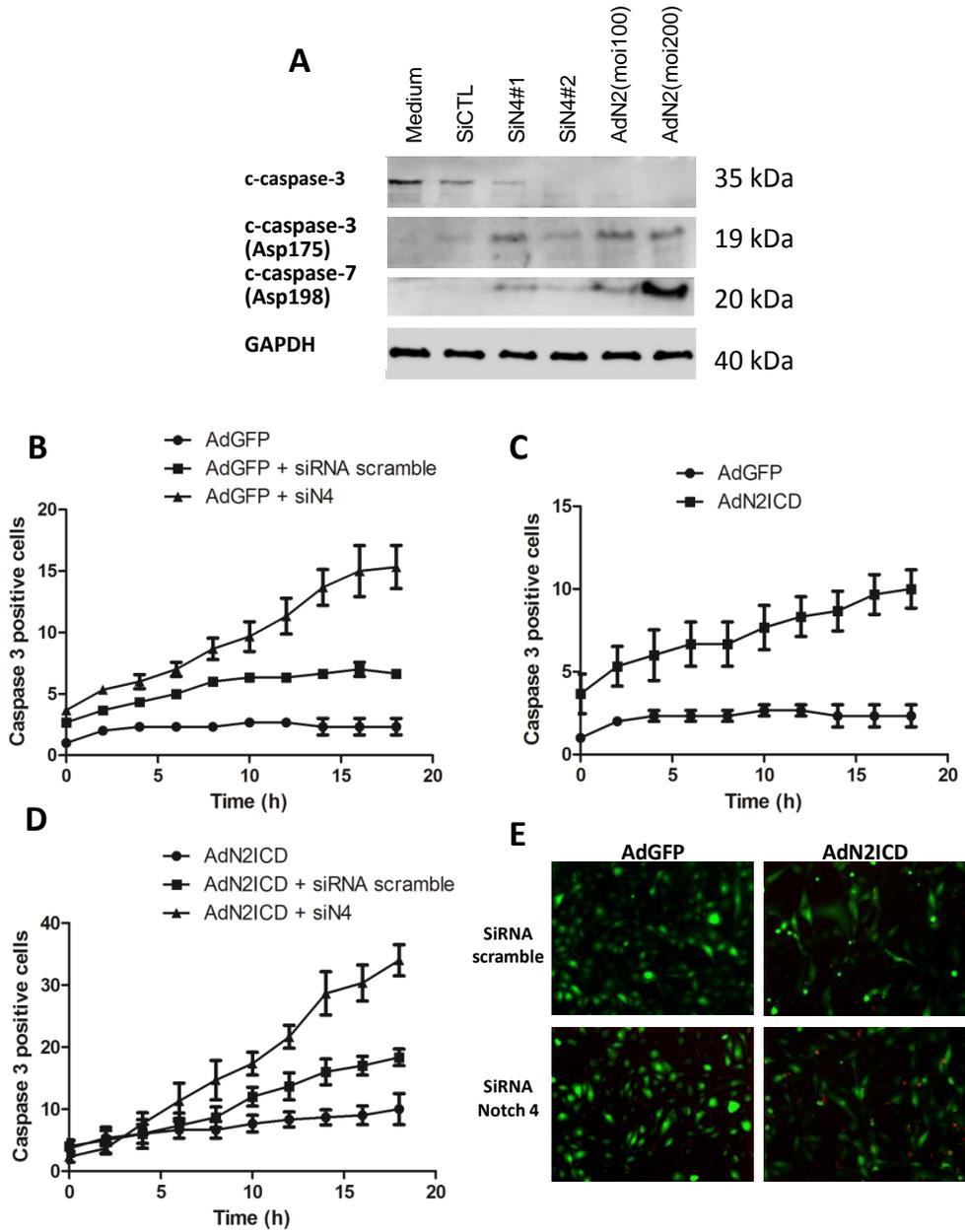
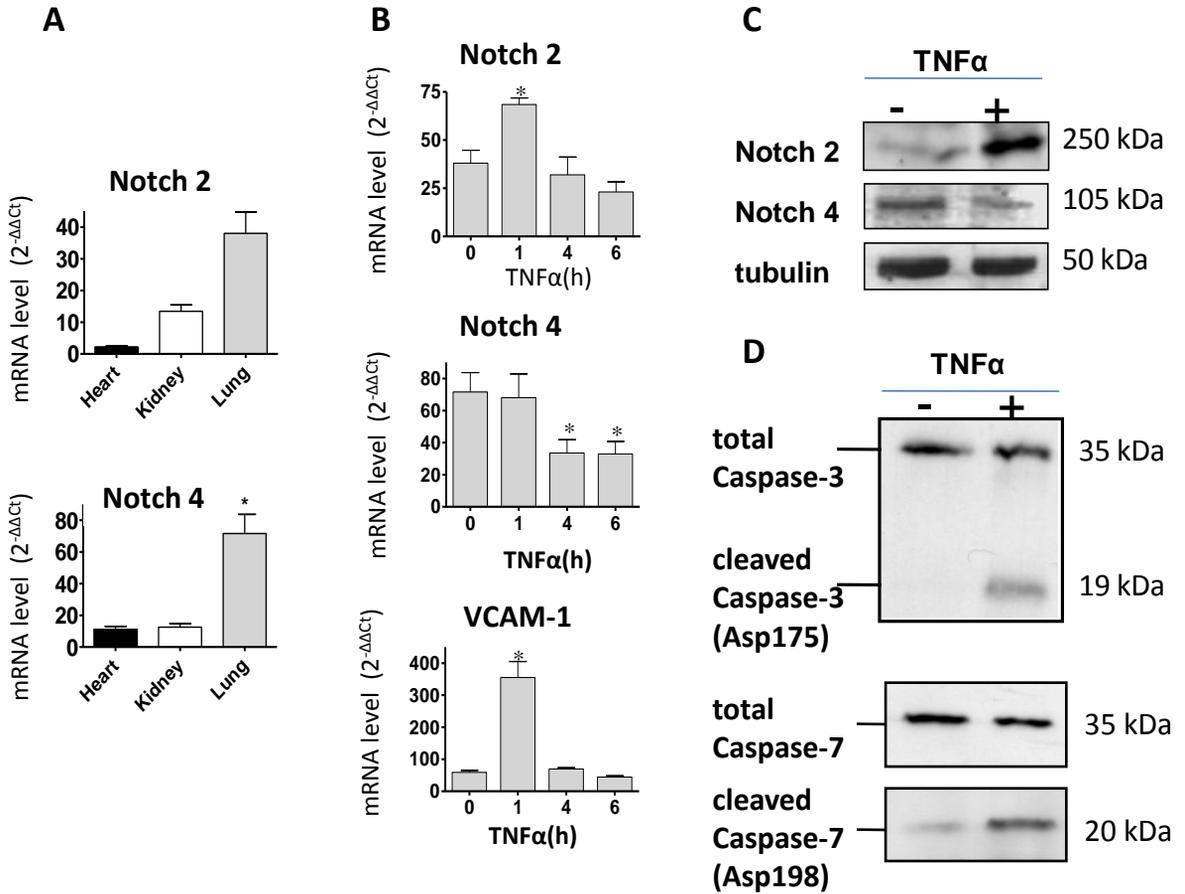


Figure 6



Graphical Abstract

TNF promotes apoptosis in endothelial cells through a downregulation of Notch activity and a phenotypic switch where Notch4 is replaced by Notch2

