

Heteroresistance to glycopeptides in Italian meticillin-resistant (MRSA) isolates

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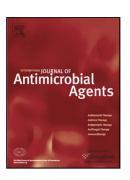
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2	Staphylococcus aureus isolates
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6	Tashia Bertuccio, Simona Purrello, Daria Nicolosi, Cristina Scuderi, and Stefania Stefani
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9	Department of Microbiology – University of Catania (I)
10	
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12 13 14	
15 16	Corresponding Author:
17	Prof Stefania Stefani
18	Department o f Microbiology
19	University of Catania
20	Via Androne 81
21	95124 Catania, Italy
22	tel +39 095 2504714
23	fax +39 095 2504733
24	email <u>stefanis@unict.it</u>

25 Abstract

The prevalence and the molecular characterization of hetereoresistant *S.aureus* strains 26 27 to vancomycin (hVISA) were determined in a large group of Italian strains, isolated from 28 2005 to mid 2007. Among the 1284 strains isolated from documented infections from hospitalized patients (BSI, pneumonia and SSTIs), 139 S.aureus with MICs to vancomycin 29 30 between 1 and 2 mg/L were screened for the presence of hVISA using three different methods, and confirmed by the population analysis profile (PAP). Thirty-six strains 31 (25.8%) were detected. Among the three screening methods used, the macroEtest 32 33 (MET) demonstrated 100% specificity and 75% sensitivity. hVISA strains were *agr*I and II and belonged to the major nosocomial clones circulating in Italy (ST8, ST239, ST247, 34 and ST228); all strains were susceptible to quinupristin/dalfopristin, linezolid, 35 36 daptomycin, tigecycline and dalbavancin. In conclusion, we have demonstrated that 37 hVISA isolates are common in MRSA isolates with MICs between 1 and 2 mg/L, in Italy. MET, with its high sensitivity and specificity should be used for an early detection of 38 hVISA, above all inpatients with serious or prolonged infections sustained by MRSA. 39 40

Finally, the most recent anti Gram-positive drugs maintained their full spectrum of *in vitro* susceptibility against these strains.

43

First reports of *Staphylococcus aureus* with reduced susceptibility to vancomycin (RSV)
date back to 1997 [1] when Hiramatsu et al described the Mu50 (VISA) and the Mu3
(hVISA) strains in Japan, possessing MICs to vancomycin respectively of 8 and 4 mg/L,
isolated from patients in which vancomycin therapy failed. After these reports, it did not
take long for this resistance phenotype to be recognized all around the world,
predominantly in methicillin-resistant strains. Although the homogeneous resistance to

50 vancomycin (VISA) continues to be rare, there are increasing reports of strains showing 51 heteroresistance (hVISA), often with vancomycin MICs $\leq 2 \text{ mg/L} [2,3,4]$. Even if 1997 is 52 the official beginning of the isolation of strains with reduced vancomycin susceptibility 53 (RVS), a retrospective analysis of stored isolates detected previously unrecognized 54 hVISA/VISA strains at least back to 1987 both in the USA and in Europe [5]. Heteroresistance to glycopeptides, in which subpopulations with reduced susceptibility 55 (approximately 10⁶ CFU/ml) coexist in a seemingly susceptible phenotype, has been 56 recently associated with clinical failure [6,7,8,9], and can be found predominantly in 57 MRSA strains, even if reports from methicillin-susceptible *S.aureus* have recently been 58 published [10]. 59

60

61 One of the major difficulties in interpreting the literature on prevalence, epidemiology and significance of hVISA itself, derives from the lack of standardized criteria for the 62 definition of hVISA and the use of different methodologies to detect them [11]. MIC 63 breakpoints are not sensitive enough to distinguish precursors of strains with reduced 64 65 susceptibility (hVISA) from the vancomycin-susceptible S.aureus isolates. For these 66 reasons, a large number of screening methods have been analyzed, and although a number of them are useful for the detection of VISA, they are not adequate for hVISA 67 screening, in which resistant subpopulations can be found only at 10⁻⁵ to 10⁻⁶ CFU/ml, at 68 the limit of the inoculum size used in the MIC testing. Methods to detect hVISA therefore 69 tend to rely on the testing of higher inoculum (Etest 2Mc-Farland) or population 70 analysis profile (PAP) [2,11]. 71

72

73 Significant controversy also exists also regarding the current and future roles of
74 vancomycin and teicoplanin in the treatment of serious MRSA infections, above all in

75	those strains in which vancomycin MICs creep towards the upper levels of susceptibility,
76	affecting the global susceptibility of the <i>S.aureus</i> population.
77	
78	The resolution of all these problems requires a detailed understanding of the
79	epidemiological and clinical impact of changes in glycopeptide susceptibility of S.aureus
80	as well as criteria for detection and a full comprehension of the mechanisms at the basis
81	of reduced susceptibility (RS).
82	
83	With this in mind, a retrospective analysis of MRSA isolates was performed in order to:
84	i) evaluate the prevalence of hVISA strains in a sample of 1284 sequential Italian MRSA
85	isolates, by using current methods for the screening of hVISA and VISA strains and
86	confirmed by PAP/AUC analysis; ii) perform the molecular characterization of these
87	strains with reduced susceptibility and their susceptibility to anti-Gram-positive drugs.
88	
89	We screened 1284 sequential MRSA strains (isolated in a Italian surveillance study
90	involving 20 centres distributed throughout the country, between 2005 and mid 2007)
91	from documented bloodstream, pneumonia and skin-structure infections [12], and 139
92	strains with vancomycin MICs between 1 and 2 mg/L were selected for further studies.
93	Mu50 (VISA) and Mu3 (hVISA) were included in the study as control strains for
94	phenotypic and genotypic assays.
95	
96	MIC determinations for glycopeptides, linezolid, dalbavancin, quinupristin/dalfopristin,
97	tigecycline and daptomycin, were performed according to CLSI [13].
98	

99 Strains were screened using a rapid screening method as follows: $BHIV_4$ (containing 4) mg/L vancomycin, originally developed by Hiramatsu et al [1]) and BHIT₅ (containing 5 100 101 mg/L teicoplanin, described by Fitzgibbon et al as the best performing method used [14]) compared with the macro Etest (MET), and confirmed by the reference PAP/AUC 102 103 method. Briefly, tests were performed as previously described [1,14], with the following 104 modifications: all isolates were screened on BHIT5 and BHIV4 using 10 µL-loop volumes 105 of bacterial suspensions with two densities, equivalent to a 0.5 and 2 McFarland 106 turbidity standard. Plates were incubated for 48 h at 37°C. S. aureus Mu3 (hVISA), Mu50 107 (VISA) and ATCC 29213 (vancomycin-susceptible *S. aureus*, VSSA) were used as control 108 strains.

109

To assay macro Etest (MET) procedures, all clinical isolates were grown overnight to a
2.0 McFarland standard in Mueller-Hinton broth; a 100µl sample was plated and
streaked onto BHI agar, and vancomycin and teicoplanin Etest strips were applied (AB
Biodisk, Sweden). Plates were incubated for 48h at 37°C and then evaluated for growth
following manufacturer's instructions (EAS003; AB Biodisk, Solna, Sweden) [15].

To perform population analysis (PAP/AUC), colonies from cultures grown overnight on
tryptic soy agar were inoculated into tryptic soy broth. After incubation for 24h,
dilutions of 10⁻³ (10⁵ CFU/ml) and 10⁻⁶ (10² CFU/ml) were prepared in saline, and 50µl
were inoculated onto BHI agar plates containing 4.0, 6.0, 8.0, 12.0, 16.0 mg of
vancomycin/liter and 4.0, 8.0, 16.0, 32.0 mg of teicoplanin/liter, respectively. After 48h
of incubation at 37°C, the colonies were counted and the log of the CFU/ml was plotted
against the vancomycin concentration by using Prism software (GraphPad Software Inc).

- 123 The ratio of AUC of the test isolates to the AUC of *S. aureus* Mu3 was calculated and
- 124 interpreted as previously published [11].
- 125

126 Molecular characterization of all the MRSA strains included in the study was conducted

- 127 by PFGE, MLST, SCCmec, and *agr*-typing; all techniques were performed as previously
- 128 described [12,16].
- 129
- 130 Prevalence of hVISA in clinical specimens

131 Among the 139 isolates with vancomycin MIC between 1 and 2 mg/L, PAP/AUC

132 identified 78 VSSA, and 36 hVISA strains, no VISA strains, as defined by CLSI

133 breakpoints, were detected. However, 9 out of the 36 hVISA, showed a population

134 profile similar to Mu50 (VISA), and these strains showed a higher number of cells with

135 MIC \ge 12 mg/L to vancomycin as determined by PAP.

136

Overall, the incidence of hVISA strains among the group of MRSA with MICs between 1
to 2 mg/L, was 25.8%, almost all isolated from blood and LRT infection specimens. As
reported in table 1, of the three screening methods used, the macro Etest detected all
strains with a specificity of 100% and a sensitivity of 75%. For the other two screening
methods BHIT₅ and BHIV₄, specificity was respectively 43.5 and 61.5%, while sensitivity
was respectively 75 and 66.6%.

143

144 Molecular characterization of hVISA strains

145 All our hVISA strains belonged to the *agr*I and *agr*II types and derived from the major

- 146 nosocomial pathogens circulating in Italy, i.e. MRSA ST8 (formerly the Archaic clone),
- 147 ST239 (formerly the Brazilian clone), ST247 (formerly the Rome clone), and ST228

148	(formerly the Iberian clone) (table 2). The strains were potently inhibited by all
149	antibiotics tested, i.e. linezolid, daptomycin, tigecycline, quinupristin/dalfopristin and
150	dalbavancin, with one or two-dilution variations considering the different clones.
151	
152	Over the last few years, there has been significant interest regarding the changing
153	patterns of vancomycin MICs within the S.aureus population. This has been driven partly
154	by studies demonstrating poorer outcomes for vancomycin treatment of MRSA
155	infections with higher vancomycin MICs but still in the range of susceptibility
156	[7,8,17,18]. This change has directly impacted the proportion of hVISA, as has been
157	clearly demonstrated by many authors [19,20]. This "MIC creep" has been reported by
158	many centres [12,19,21,22,23,], while some others did not find any change over time
159	[24]. Therefore, changes in <i>S.aureus</i> vancomycin MICs can occur over time within
160	specific institutions.
161	
162	The fact that the "MIC creep" obscures the presence of hVISA with subpopulations of
163	cells resistant to vancomycin, but also to teicoplanin [12], requires an accurate
164	assessment of the clinical significance of hVISA with reliable screening and confirmatory
165	tests.
166	
167	In this study we surveyed nosocomial MRSA isolates sequentially collected from
168	different Italian hospitals during a period of 36 months. PAP/AUC analysis
169	demonstrated that, among strains with MICs between 1 and 2 mg/L, hVISA were found
170	in 25.8% of cases and these strains belonged to the major nosocomial clones circulating

172 hVISA has varied significantly. Some of these differences can be explained by differences

173	in laboratory definitions and testing strategies, however it appears that rates of hVISA
174	vary globally, which could be related to the amount of subpopulations present. Studies
175	reported rates ranging from less than 1% to as high as 50% [6,9,25].
176	
177	In the present study, using MET at the manufacturer's criteria of \geq 8 mg/L for
178	vancomycin and teicoplanin or $\geq 12~mg/L$ for teicoplanin alone, we found a good
179	correlation with the PAP/AUC, with a good sensitivity and a high specificity. BHIT5, as
180	demonstrated by other authors [14] is a useful method with a sensitivity comparable to
181	MET, but with a lower specificity. BHIV4, in this respect, is less useful.
182	
183	Thirty-three out of the 36 hVISA strains emerged from the most diffused nosocomial
184	pandemic MDR MRSA clones. In Italy they belong to <i>agr</i> I and II type strains, as already
185	observed [26,27] and there is evidence of a potential association between a non-
186	functional <i>agr</i> locus and a reduced response to vancomycin therapy.
187	
188	hVISA is susceptible to many antimicrobials currently available. Despite their MDR
189	phenotype, including resistance to fluoroquinolones, aminoglycosides, macrolides, these
190	strains were fully susceptible to linezolid, quinopristin/dalfopristin, tigecycline,
191	daptomycin and to the new investigational drug dalbavancin. There has been an
192	association between hVISA, VISA and reduced susceptibility to daptomycin [28,29,30],
193	however this appears to be strain specific and may be unstable [31]. Data from <i>in vitro</i>
194	studies suggest that daptomycin may have a lower rate on <i>in vitro</i> killing, still retaining
195	its potent bactericidal activity [32,33,34]. RVS strains maintain susceptibility to
196	linezolid, with no changes in the MIC distribution with respect to the VSSA. This drug is

197 essentially bacteriostatic, but it was used with success in the treatment of infections

sustained by MRSA, hVISA and VISA strains [35,36,37].

199 Tigecycline, the first glycylcycline to be available for clinical use, has a very good activity

200 against MSSA and MRSA: it has a very good *in vitro* activity against hVISA strains with

201 very low MIC values.

202

203 With regards to new drugs under development, the potent activity of dalbavancin was

204 demonstrated in our study. Data from the literature reported higher MICs of this drug in

205 hVISA or VISA strains, and an *in vitro* model [38] shows a need for the highest

206 concentrations of drug to achieve bactericidal activity, however, the clinical implications

207 of these findings are unclear.

208

209 In conclusion, we have demonstrated that hVISA isolates are common in MRSA isolates

210 with MICs between 1 and 2 mg/L, in Italy. Detection of hVISA requires specific and often

211 cumbersome laboratory tests, among them the optimal one has not yet been

212 determined. MET, with its high sensitivity and specificity should be used for an early

213 detection of hVISA, above all in patients with serious or prolonged infections sustained

214 by MRSA. Finally, the most recent anti Gram-positive drugs maintained their full

215 spectrum of *in vitro* susceptibility against these strains.

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221

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224	
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231	
232	
233	
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238	invitations from many pharmaceutical companies. SS has received partial funding from
239	Pfizer Pharma to perform studies on linezolid (who also supplied the drug).
240	All other Authors have none to declare.
241	Ethical Approval: Not necessary
242	
243 244	
245	

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Table 1. Sensitivity and specificity values for the three screening methods used, with respectto PAP/AUC (at 48h).

METHODS	PAP hVISA N°36		% SENS	PAP VSSA	N°78	% SPEC	
	VSSA	hVISA		VSSA	hVISA		
BHI T5 2MF	9	9 27 75%		34	44	43.5%	
BHI V4 2MF	12	24	66.6%	48	30	61.5%	
Macro E-test 2MF	9	27	75%	78	0	100%	

- 397
- Table 2. Genetic (ST, SCCmec, PFGE, and *agr* types) and phenotypic features of the 36
- 399 hVISA strains in study and MICs distribution (mg/L) of the major anti-Gram positive drugs
- 400 tested.
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	0.03	0.06	0.12	0.25	0.50	1	2	4
Archaic (3 hVISA) ST8-HA-MRSA-I agr-type I/IV								
	0	0	0	0	0	3	0	0
Teicoplanin	0	0	0	0	0	3	0	0
Daptomycin	0	0	0	1	2	0	0	0
Linezolid	0	0	0	0	0	2	1	0
Quinupristin/Dalfopristin	0	0	0	0	1	2	0	0
Tigecycline	0	0	0	1	2	0	0	0
Dalbavancin	1	2	0	0	0	0	0	0
Iberian (3 hVISA)								
ST247-HA-MRSA-IA agr-type I		-						
Vancomycin	0	0	0	0	0	3	0	0
Teicoplanin Daptomycin	0 0	0 0	0 0	0 0	0 3	0	1	2 0
Linezolid	0	0	0	0	3	2	1	0
Quinupristin/Dalfopristin	0	0	0	2	1	0	0	0
Tigecycline	0	0	0	0	3	0	0	0
Dalbavancin	1	2	0	0	0	0	0	0
Rome (6 hVISA)		_	-	-			-	-
ST247-HA-MRSA-I/IA agr-type I	_	-	<i>.</i>				_	
Vancomycin	0	0	0	0	0	1	5	0
Teicoplanin Daptomycin	0 0	0 0	0 0	0 0	0	1 4	1 0	4 0
Linezolid	0	0	0	0	2	4 3	0	0
Quinupristin/Dalfopristin	0	0	0	0	0 4	3 0	3	0
Tigecycline	0	0	3	1	2	0	0	0
Dalbavancin	1	1	4	0	0	0	0	0
Rome (6 hVISA) ST247-HA-MRSA-I/IA agr-type I			\sim		Ū	Ū	Ū	Ū
Vancomycin	0	0	0	0	0	0	6	0
Teicoplanin	0	0	0	0	0	0	4	2
Daptomycin	0	0	0	0	2	4	0	0
Linezolid	0	0	0	0	0	4	2	0
Quinupristin/Dalfopristin	0	0	0	2	4	0	0	0
Tigecycline	0	0	1	5	0	0	0	0
Dalbavancin Brasilian (3 hVISA)	2	2	2	0	0	0	0	0
ST239-HA-MRSA-IIIA agr-type I								
Vancomycin	0	0	0	0	0	3	0	0
Teicoplanin	0	0 0	Õ	0	0	3	0 0	Õ
Daptomycin	0	0	0 0	1	2	0	0	0 0
Linezolid	0	0	0	1	2	0	0	0
Quinupristin/Dalfopristin	0	0	0	1	2	0	0	0
Tigecycline	0	0	0	1	2	0	0	0
Dalbavancin	0	3	0	0	0	0	0	0
Italian (12 hVISA) ST228-HA-MRSA-I agr-type II								
Vancomycin	0	0	0	0	0	12	0	0
Teicoplanin	0	0	0	0	0	12	0	0
Daptomycin	0	0	0	0	9	3	0	0
Linezolid	0	0	0	0	0	6	3	3
Quinupristin/Dalfopristin	0	0 0	0	0 0	5 10	7	0 0	0 0
Tigecycline Dalbavancin	0 0	0 12	2 0	0	10 0	0 0	0	0
Sporadic (3 hVISA)	U	12	U	0	U	0	0	U
Vancomycin	0	0	0	0	0	3	0	0
Teicoplanin	0	0	0	0	0	1	0	2
Daptomycin	0	0	0	1	2	0	0	0
Linezolid	0	0	0	0	0	2	1	0
Quinupristin/Dalfopristin	0	0	0	1	1	1	0	0
Tigecycline	0	0	0	2	1	0	0	0
Dalbavancin	0	3	0	0	0	0	0	0