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Running title: PTGES gene polymorphism in RA.

Microsomal PGE synthase 1 (mPGES-1) is the terminal enzyme in induced state of prostaglandin E_2 (PGE₂)synthesis and constitutes a therapeutic target for rheumatoid arthritis (RA) treatment. We examined the role of the PTGES gene polymorphism in susceptibility to and severity of RA and related variations in the gene to its function. The PTGES gene polymorphism was analyzed in 3081 RA patients and 1900 controls from two study populations: Swedish Epidemiological Investigation of Rheumatoid Arthritis (EIRA) and the Leiden Early Arthritis Clinic (Leiden EAC).Baselinedisease activity score (DAS28) was employed as disease severity measure.mPGES-1 expression was analyzed in synovial tissue from RA patients with known genotypes using immunohistochemistry.In the Swedishstudypopulation, among women a significant association with risk for RA was observed for *PTGES* SNPs in univariate analysis and for the distinct haplotype. These results were substantiated by meta-analysis of data from EIRA and Leiden EAC studies with overall OR 1.28 (95%CI 1.09-1.51). Several PTGES SNPs were associated with earlier onset of disease or with higher DAS28 in women with RA. The patients with the genotype associated with higher DAS28 exhibited significantly higher mPGES-1 expression in synovial tissue. Our data reveal a possible influence of PTGES polymorphism on the pathogenesis of RA and on disease severity through upregulation of mPGES-1 at the sites of inflammation. Genetically predisposed individuals may develop earlier and more active disease due to this mechanism.

Keywords

mPGES-1, gene polymorphism, rheumatoid arthritis, gender

INTRODUCTION

Rheumatoid arthritis (RA) is a chronic inflammatory disease characterized by persistent synovial inflammation resulting in joint destruction and severe disability. It is one of the most common complex autoimmune diseases with up to 1% prevalence, with a higher prevalence, activity and earlier disease onset in women. Genetic factors significantly contribute to the development of RA and its severity[1]. The major genetic risk factor for RA is the shared epitope (SE) alleles of HLA-DRB1 gene locus[2, 3]. Recentlyseveral common genetic variants have been discoveredoutside HLA-DRB1 locus identifying important biological pathways predisposing to RA (for review[4]).

Prostaglandin E₂ (PGE₂) isone of the key mediators of inflammation, pain and joint destruction in RA, anditsproduction is an important target ofanti-inflammatory drugs.HencePGE₂biosynthetic pathway is of considerable interest togene association and pharmacogenetic studies[5, 6]. PGE₂is produced by conversion of arachidonic acid to PGH₂ by cyclooxygenases (COX-1/COX-2) and subsequent conversion of PGH₂to PGE₂ by terminal PGE synthases. Microsomal PGE synthase 1 (mPGES-1) is the terminal enzyme in the induced PGE₂ production at the sites of inflammation [7, 8]and plays an important role in the pathogenesis of inflammatory cytokines in RA synovial fibroblasts and synovial fluid monocytes [11, 12] and markedly up-regulated in RA synovial tissue[13].In mice with experimental arthritis, genetic deletion of mPGES-1 results in significantly reduced disease incidence, severity pain[9, 10]. mPGES-1 is recognized as analternative target for anti-inflammatory treatment with improved selectivity and safety compared to COX-1/COX-2[8].

The human mPGES-1 is encoded by prostaglandin E synthase (*PTGES*) gene, which is localized at chromosome 9q34.3, spans 14.8 Kb and contains 3 exons. According to NCBI Single nucleotide polymorphisms (SNPs)database, there are 249 SNPs in *PTGES* gene region, including 4 coding SNPs (2 synonymous and 2 non-synonymous SNPs). While none of the studied variants in *PTGES* gene had a significant effect on the risk of lymphoma [14], non-fatal myocardian infarction or ischemic stroke [15], and breast cancer susceptibility [16], in a large-scale association study one SNP in 5'region of*PTGES* gene was associated with hypertension in women in the Japanese population [17]. SNPs in *PTGES* gene could modify enzyme expression levels or activity and alter inducible PGE_2 biosynthesis. Consequently *PTGES* gene polymorphisms might contribute to inter-individual differences in susceptibility to or severity of RA, as well as clinical response to future mPGES-1 inhibitors. Hence, *PTGES* genetics may have clinical importance from disease risk assessment to choice of personalized treatment in RA.

Here, we studied the association between *PTGES* gene polymorphisms and risk and severity of RA. In addition, we examined whether *PTGES* gene polymorphism could contribute to altered mPGES-1 expression.

MATERIALS AND METHODS

Patients.

In this study two independent cohorts have been included: Swedish Epidemiological Investigation of Rheumatoid Arthritis (EIRA) and Leiden Early Arthritis Clinic (EAC). The first cohort consisted of 2012 RA patients and 1033 controls matched on age, sex and residential area from the EIRA, a population-based case-control study in Sweden (Table 1). A case was defined as a person who received for the first time a diagnosis of RA based on the diagnostic criteria of the American College of Rheumatology[18].Data regarding age of disease onsetwere available for all patients, data regardingbaselinedisease activity score (DAS28) for 887(44%)RApatients. Further characteristics of the EIRA study are described elsewhere[19].

At the replication stage the second cohort consisted of 1069 Dutch Caucasian individuals with RA, fulfilled theACR classification criteria for RA and 867 controls, unrelated Dutch Caucasians with no history of RAfromEAC(Table 1). More details about the EAC study population have been described elsewhere[20]. The approval was granted by the regional ethical committees and all the participants gave their informed consent to participate in the study.

Single nucleotide polymorphism (SNP) genotyping

11 SNPs were selected across the *PTGES*gene locus and flanking regions(Fig 1).The selection was based on the minor allele frequency (MAF) higher than 10% according to NCBI for all SNPs except for rs11792431 (MAF 2.5%). In the replication phase, we genotyped 3 SNPs in *PTGES* locus which showed significantly association with RA risk. Genotyping was performed using TaqManSNP genotyping assays (Applied Biosystems, Europe BV, The Netherlands). The average genotyping call rate was 96.6% varying from 95% to 98% for different SNPs.

In addition, we included in the analysis data from our previous genome-wide association scan (GWAS) [21] for 14 SNPs in the *PTGES* locus and inter-gene regions with neighboring *PRRX2* and *TOR1B* genes (Fig. 1).Genotyping was performed on the Illumina HumanHap300 (version 1.0) array[21].

Immunohistochemical analysis.

Synovial tissue biopsy specimens from 24 RA patientsfrom EIRA, carrying different *PTGES* genotypeswere obtained during arthroscopy. Synovial tissue samples were snap frozen and kept at -70°C until sectioned. Staining of cryostat sections with rabbit polyclonal anti-human mPGES-1 antiserumwas performed as previously described[13]. Irrelevant rabbit IgG was used as a negative control. Stained tissue sections were examined using a Polyvar II microscope (Reichert-Jung, Vienna, Austria) and photographed with a digital Leica camera 300F (Leica, Cambridge, UK). The positive staining was assessed quantitatively using computer-assisted image analysis and expressed as the percentage of the total area of counterstained tissue.

Statistical analysis.

Genotype and allele frequencies in the patient and the control groups were compared using the Chi-square test and/or Fisher's exact test when appropriate. Haplotype frequencies were estimated using HaploView 4.0 (http://www.broad.mit.edu/mpg/ haploview/ index.php), and the permutation test was set to 1,000,000 permutations for the single markers and for the haplotypes with P<0.05 for significance of empirical pvalue. Anodds ratio (OR) with 95% confidence interval (CI) wascalculated for the association between different genetic factors and risk of RA. Evaluation of Linkage Disequilibrium (LD) and deviations from Hardy-Weinberg equilibrium (HWE) were performed using HaploView. The distribution of genotypes for all studied SNPs within the RA group and within the control group was in agreement with HWE (p>0.05). Prior to combining for meta-analysis, the EIRA and Leiden EAC data were tested for the homogeneity in the ORs, using a Pearson χ^2 goodness-of-fit test as described elsewhere[22]. In the meta-analysis haplotype-frequency data of the RA cases versus the controls were compared in EIRA and Leiden EAC studies using Mantel-Haenszel analysis assuming random effects for the ORs[22].

In RA patients stratified by sex, the associations between *PTGES* polymorphisms and DAS28 and age at the first diagnosis of RA, as well as the results of functional study were analyzed using Mann-Whitney's U test with Bonferroni corrections for multiple comparisons. P values <0.05were considered statistically significant.

RESULTS

PTGES polymorphisms and risk of developing RA.

In the discovery phase, we analyzed allelic and haplotype frequencies of 11 SNPs in PTGES gene in the entireSwedish cohort as well as in men and women separately. An association betweenthe SNP rs16931419 and risk of RA was observed in the allelic model (women and men combined),but the difference was not significant after permutation test (Table 2). Among womenthree SNPs, rs10988484, rs11999368 and rs16931419,from a single LD block in the gene locus downstream the *PTGES* gene showed higher minor allele frequencies in female RA patients compared to that in female controls (Table 2). The association of the SNP rs16931419 with the risk of RA was still apparent after permutation test (OR 1.38; 95%CI 1.14-1.67, empirical p value=0.0082). Among men, a trend towards an association between the SNP rs 3844048 and risk of RA was observed, but this associationwas not statistically significant after permutation test (Table 2).

To exclude possible correlation with neighboring genes due to extended linkage disequilibrium (LD), we expanded the analysis by including 14 SNPs in the *PTGES* and intergenic areas with the*PRPX2* or the *TOR1B* genes using data from our previous GWAS [21] (Fig.1). There was no evidence for an association

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between SNPs in proximity to the *PRPX2* gene or the *TOR1B* gene and susceptibility to RA (data are not shown).

Further, we examined the frequency of haplotypes (only with the frequency above 0.01) based on the three SNPs significantly associated with the risk of RA. The frequency of the haplotype TTC (susceptible haplotype) was significantly higher in women with RA than in women without RA (OR 1.34; 95%CI 1.11-1.63, empirical p value=0.0077) (Table 3). The frequency of the opposite haplotype CCT (protective haplotype) was significantly lower in women with RA compared to controls (OR 0.78; 95%CI 0.66-0.93; empirical p value=0.0172). We did not findsignificant associations between thesehaplotypes and RA risk amongmen (Table 3).

In the replication phase, we analyzed the association of the three SNPs, rs10988484, rs11999368 and rs16931419, with RA susceptibility in the Dutch cohort. Similarly tothe Swedish cohort, these SNPs showed higher minor allelic frequencies in female patients with RAcompared to the frequencies in female controls, though the difference was not statistically significant (Table 2). In the same way, the frequency of the susceptible haplotype TTC was higher in women with RA than in women without RA, though the difference was not significant (Table 3). The allelic frequencies amongfemale controls in the two populations were identical, while the haplotype frequencies were highly similar.

Genetic variations with low or moderate effects on disease susceptibility may not be evident in an individual study due to low statistical power. Thus, we performed a metaanalysis combining our results from the Swedish and the Dutch cohorts to verify the significance of *PTGES* polymorphisms for susceptibility to RA. Test for the heterogeneity between EIRA and EAC showed no significant differences (p>0.05). The overall statistical power in the combined analysis was >80% (in two-tail test) to

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detect associations with $OR \ge 1.3$, given the prevalence of studied genetic factors. The meta-analysis demonstrated a statistically significant association between the susceptible haplotype TTC and risk of developing RA in women with an overall OR of 1.31 (95%CI 1.10-1.56) and p=0.002 (Fig.2).

PTGES polymorphisms and age of onset and activity of RA.

RA is a heterogeneous syndrome with relatively broad variations in the disease onset and activity. Therefore, we tested the correlation between these parameters and *PTGES* genetic markers in patients from EIRA. Baseline DAS28 and mPGES-1 expression in RA synovial tissue were used as disease activity measures.

There was no association of RA risk haplotype TTC with baseline DAS28 scores or age of disease onset in women or men. The minor allele of the SNP rs10739757 corresponded to 3 years earlier onset of RA in female patients (median 51, range 19-74 *vs.* median 54, range 16-82; p= 0.0065) (Table 4).

Furthermore, we observed that the homozygous major allele in the SNP rs4837404 was associated with higher DAS28 scores in women with RA(median 5.57, range 1.74-8.89, *vs* median 5.28, range 1.57-8.29, p=0.0080), but not in men. In addition, the minor allele in the SNP rs2302821 showed a trend towards associationwith higher DAS28 scores in women with RA (median 5.57, range 2.39-8.89 *vs*. median 5.31, range 1.57-8.37, p=0.0321). Also, the female RA patients carrying the minor allele of the SNP rs11792431 showed a trend towards higher DAS28 scores (median 5.63, range 3.82-8.89 *vs*median 5.32, range 1.57-8.66, p=0.0369) (Table 4).

Allmentioned variations related to disease course were in low LD ($r2 \le 0.34$) with SNPs included in the susceptibility haplotype TTC.

Using immunohistochemical analysis we evaluated the expression of mPGES-1 in RA synovial tissue in relation to *PTGES* genotypes. There was no difference in mPGES-1 expression in relation to RA risk haplotype TTC. mPGES-1 expression was significantly higher in RA synovial tissue from patients (mean \pm SD age, 52.8 \pm 14.4 years) carrying the minor allele C in SNP rs2302821 compared to the patients (mean \pm SD age, 54.4 \pm 14.1 years)that did not carry that allele (median 12.9 % of stained area, range 8.1-23.6 *vs* median 7.1 %, range 0.9-10.5, p<0.001, n=9 and 15 respectively) (Fig.3). No differences were observed in mPGES-1 expression in relation to gender or treatment of patients with NSAIDs, MTX or prednisolone.

DISCUSSION

The present study demonstrates the association of variations in the*PTGES* gene, encoding mPGES-1, with susceptibility to RA in women. The data are consistent for single SNP and haplotypes in the large study from Sweden. Moreover, thefinding was substantiated by meta-analysis of data from the EIRA and Leiden EAC studies. We also found that the *PTGES* genotypes are likely to be related to earlier disease onset and disease severity, as they were associated with higher baseline disease activity scores and related to higher expression of mPGES-1 in synovial tissue biopsies from RA patients.

PGE₂ is actively involved in the regulation of immune response, inflammation and pain, and polymorphisms in the genes in the PGE₂ biosynthetic pathway are important risk factors forcardiovascular, neoplastic and inflammatory diseases[6, 23]. The role of gene polymorphisms in the PGE₂ pathway in relation to RA risk and severity is less investigated. Two case-control studies have revealed a possible protective influence of COX-2 -765C allele on the susceptibility to RA and disease activity[24, 25]. To our

knowledge, the present study is the first one to examine an association between PTGES polymorphisms and the risk of developing RA.

We performed an association study in two independent populations. In a large material from Sweden, we found the association between three SNPs from the same recombination block, which spread approximately within 9 Kb downstream to *PTGES*, and RA risk in women. This significant association was apparent for the SNP rs16931419 (3 Kb down-stream to *PTGES*) after 1,000,000 permutations. Moreover, the TTC haplotype based on the SNPs from this recombination block demonstrated a significant association with RA risk in women in the Swedish population.

We expanded the analysis by including 14 SNPs in the *PTGES* locus and intergenic areas with the *PRPX2* or the *TOR1B* genes to exclude possible association with neighboring genes. No evidence for the association between the SNPs in proximity to the *PRPX2* and the *TOR1B* genes and RA risk was seen in our study population.

In order to replicate our finding, we analyzed the association between three*PTGESSNPs* included in the haplotype and risk of RA in a Dutch cohort. Though the respective case-control frequencies had a similar pattern in RA females from Leiden EAC as in EIRA, there was no significant association between single SNPs or haplotype and susceptibility to RA in the Dutch cohort. The power of the replication study was though limited by the small size of female control group. To increase the power of the study we performed a meta-analysis of the data from EIRA and Leiden EAC studies and confirmed the association between TTC haplotype and increased risk to RA in women. This finding substantiates the evidence that *PTGES* polymorphisms might be important for susceptibility to RA in women.

The revealed genetic association was consistent for women but not detected for men. The higher prevalence of RA in women suggests that genetic risk of RA

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might vary with sex. Indeed, there is a number of associations that show genetic effects in only one sex, or reciprocal effects on risk of RA in males and females[26, 27]. Thus, there is strong association of SNP rs2900180 in the *TRAF-C5* locus with RA without obvious evidence of an effect in males [27]. Also, SNPrs11761231 has been reported in a large study to be associated with RA in females, but has no effect on disease status in males [26].On the other hand, several SNPs in *NCF4* (rs729749), *NCF2* (rs789181) and *RAC2* (rs1476002) genes were found to be mildly associated with RA only in men[28].

Moreover, the association of COX-2 polymorphisms with disease risk and response to NSAID varies between sexes[29, 30],most likelyas aresult ofsexual dimorphism in PGE₂ metabolism. Systemic biosynthesis of PGE₂differs markedly between males and females in human, as in mice[31, 32]. In rodents, variations in COX-2 and mPGES-1 expression at basal and pathological conditions [33, 34]and different patterns of EP receptor expression[35]have been reported in females and males. Thus, sex dependent association of *PTGES* polymorphisms with RA susceptibility may reflect different response to this genetic trigger in females and males due to differences in PGE₂ metabolism.However, the absence of an association between *PTGES* polymorphism and RA risk in men might also reflect the small number of individuals and low study power and needs further validation.

The SNPs associated with RA are in a single LD block and located down-stream to 3' untranslated region (3'UTR) of *PTGES* gene. 3'UTRis involved in post-transcriptional gene regulation via mRNA stability, the mechanism which is revealed in the regulation of mouse *Ptges*gene expression[36]. TheseSNPs may be regulatory orin LDwith other regulatory polymorphisms in 3'UTR that similarly could modulate the human mPGES-1 expression and predispose individuals to RA.

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In our study, we found that gene polymorphisms in *PTGES* might have not only disease-predisposing impact but also disease-modifying effect, since two SNPs within PTGES gene were related to early diseases onset or baseline DAS28 scores. Revealing relations between genetic variations and certain function or phenotype is currently the most challenging task for genetics of complex diseases. We investigated whether gene variations associated with disease activity could relate to mPGES-1 expression. We previously demonstrated the expression of mPGES-1 in synovial biopsies as a reliable parameter of inflammatory activity in RA[12]. Our data from the current study indicate that mPGES-1 expression seems to be dependent on the genotype, and this was also reflected by genetic association with the clinical measure of baseline DAS28. Individuals with the minor allele C in PTGES rs2302821 that showed a trend towards association with higher DAS28 had significantly higher mPGES-1 expression in RA synovial tissue, which represents additional phenotypic replication for this association. Although the total patient number in our experiment is small, it represents a unique collection of tissue samples from RA patients with distinct genotypes.

In conclusion, we provide the first evidence that *PTGES* gene polymorphisms are associated with susceptibility to RA in women. In addition, *PTGES* polymorphisms were related to earlier disease onset, higher baseline disease activity scores and might contribute to higher expression of mPGES-1 in synovial tissue biopsies from RA patients. These data suggest the importance of *PTGES* gene variants in the assessment of RA risk and severity.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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Characteristics	EIRA	Leiden EAC		
No. of patients	2012	1069		
No. of controls	1033	867		
Country of origin	Sweden	Netherlands		
Female sex, patients (%)	70.4	64		
Female sex, controls (%)	74.6	21.9		
Age of onset, years	51.7±12.7	56.4±15.4		
(mean± SD)				
Disease duration, years	<1	<2		
ACPA positive(%)	(1265/2012) 62.8	(375/625 ^a) 60		
HLA-DRB1 SE (patients)	(1486/2012) 73.8	(590/842 ^b)70.1		
(%)				

Table 1.Clinical characteristics of EIRA and Leiden EAC studies

Abbreviations: ACPA, anti-citrullinated peptide antibody; SE, shared epitope.

Details on the EIRA study[19], the Leiden EAC study[20] has been reported elsewhere.^athe number of patients with defined ACPA status; ^bthe number of patients with defined HLA-DRB1 SE status.

Table 2. Minor allele frequencies of SNPs at *PTGES* locus in the Swedish and the

Dutch populations.

The Swedish population									
					MAF,	MAF,			
No	SNP	position	MA		cases	controls	OR(95%CI)	Р	P^*
1	rs10988484	3'near gene	Т	all	0.154	0.138	1.14(0.98-1.33)	0.0966	NS
				F	0.160	0.129	1.28(1.07-1.54)	0.0076	0.0591
				Μ	0.141	0.163	0.84(0.63-1.12)	0.23	NS
2	rs11999368	3'near gene	Т	all	0.148	0.134	1.12(0.96-1.31)	0.15	NS
		-		F	0.152	0.124	1.26(1.05-1.52)	0.0138	0.1035
				Μ	0.138	0.162	0.83(0.62-1.10)	0.19	NS
3	rs16931419	3'near gene	С	all	0.149	0.127	1.2(1.02-1.41)	0.02	NS
		-		F	0.153	0.116	1.38(1.14-1.67)	0.0010	0.0082
				М	0.139	0.160	0.84(0.63-1.14)	0.26	NS
4	rs2302821	3'-UNR	С	all	0.091	0.091	1.0(0.83-1.21)	0.95	NS
				F	0.093	1.0	0.93(0.75-1.15)	0.49	NS
				Μ	0.087	0.065	1.36(0.91-2.04)	0.13	NS
5	rs4837404	intron 2	G	all	0.333	0.325	1.04(0.92-1.16)	0.54	NS
				F	0.334	0.323	1.05(0.92-1.20)	0.44	NS
				Μ	0.330	0.331	0.99(0.80-1.24)	0.93	NS
6	rs10739757	intron 2	С	all	0.095	0.101	0.93(0.78-1.12)	0.44	NS
				F	0.092	0.102	0.89(0.72-1.1)	0.27	NS
				Μ	0.103	0.099	1.01(0.91-1.12)	0.82	NS
7	rs2241270	intron 1	Т	all	0.096	0.097	1.00(0.94-1.06)	0.93	NS
				F	0.096	0.096	1.00(0.83-1.23)	0.95	NS
				Μ	0.098	0.100	0.99(0.70-1.39)	0.93	NS
8	rs11792431	exon 1	А	all	0.056	0.054	1.04(0.82-1.32)	0.74	NS
				F	0.053	0.052	1.00(0.76-1.33)	0.94	NS
				Μ	0.063	0.058	1.09(0.71-1.69)	0.69	NS
9	rs10988496	5'near gene	G	all	0.104	0.104	1.00(0.84-1.20)	0.96	NS
		-		F	0.101	0.103	0.97(0.79-1.20)	0.80	NS
				Μ	0.101	0.107	1.00(0.75-1.45)	0.81	NS
10	rs3844048	5'near gene	G	all	0.301	0.308	0.97(0.86-1.09)	0.58	NS
		-		F	0.303	0.291	1.05(0.92-1.22)	0.40	NS
				Μ	0.295	0.358	0.75(0.60-0.93)	0.0114	NS
11	rs6478915	5'near gene	G	all	0.250	0.247	1.01(0.89-1.15)	0.82	NS
		-		F	0.252	0.232	1.11(0.96-1.29)	0.16	NS
				Μ	0.244	0.289	0.79(0.63-1.01)	0.05	NS
The	Dutch populat	ion					· · · · ·		
1	rs10988484	3'near gene	Т	all	0.152	0.152	1.00(0.83-1.19)	0.95	NS
		-		F	0.151	0.129	1.20(0.85-1.70)	0.28	NS
				М	0.152	0.159	0.95(0.74-1.22)	0.72	NS
2	rs11999368	3'near gene	Т	all	0.145	0.141	1.03(0.86-1.24)	0.74	NS
		e		F	0.138	0.124	1.14(0.82-1.62)	0.46	NS
				М	0.157	0.146	1.08(0.85-1.39)	0.49	NS
3	rs16931419	3'near gene	С	all	0.139	0.138	1.01(0.84-1.22)	0.93	NS
		0		F	0.131	0.116	1.16(0.80-1.67)	0.42	NS
				М	0.153	0.145	1.06(0.83-1.36)	0.59	NS

Abbreviations:MAF, minor allele frequencies; NS, not significant; F, females; M,

males. * *P*, value after 1,000,000 permutations.

Table 3.Haplotype frequencies of PTGES variations in female patients with RA and

female controls from the Swedish and the Dutch pop	ulations.
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Haplo			HF,	HF,				
type		HF	cases	controls	OR(95% CI)	Р	P^*	
CCT	all	0.843	0.837	0.854	0.88(0.76-1.02)	0.09	NS	
	F	0.842	0.831	0.862	0.78(0.66-0.93)	0.0066	0.0172	
	Μ	0.844	0.851	0.828	1.19(0.90-1.57)	0.22	NS	
TTC	all	0.130	0.137	0.117	1.19(1.01-1.40)	0.03	NS	
	F	0.128	0.139	0.107	1.34(1.11-1.63)	0.0030	0.0077	
	Μ	0.136	0.131	0.146	0.88(0.65-1.18)	0.39	NS	
TCT	all	0.011	0.011	0.011	0.93(0.56-1.57)	0.75	NS	
	F	0.012	0.012	0.013	0.97(0.55-1.71)	0.86	NS	
	Μ	0.005	0.004	0.007	0.54(0.16-1.8)	0.30	NS	
The Dutch populations								
	-	•						
Haplo			HF,	HF,		Р		
type		HF	cases	controls	OR (95% CI)		P^*	
CCT	all	0.835	0.836	0.834	1.01(0.85-1.20)	0.83	NS	
	F	0.844	0.841	0.855	0.89(0.65-1.24)	0.50	NS	
	Μ	0.827	0.826	0.827	0.99(0.78-1.25)	0.97	NS	
TTC	all	0.126	0.128	0.125	1.01(0.84-1.23)	0.82	NS	
	F	0.12	0.125	0.105	1.21(0.84-1.75)	0.29	NS	
	Μ	0.131	0.132	0.131	1.00(0.77-1.30)	0.97	NS	
TCT	all	0.02	0.018	0.025	0.16(0.46-1.13)	0.13	NS	
	F	0.02	0.019	0.021	0.90(0.40-2.00)	0.80	NS	
	Μ	0.02	0.015	0.025	0.61(0.31-1.19)	0.14	NS	

The Swedish population

Abbreviations:NS, not significant; HF, haplotype frequency; F, females; M, males. Haplotypes are based on *PTGES* SNPs rs10988484, rs11999368 and rs16931419 and are considered only with the frequency above 0.01.* *P*,value after 1,000,000 permutations; Table 4.Association of PTGES genotypeswith disease onset and activity in female patients with RA.

Disease	SNP	genotype	Median (range)	counts	P value
parameters					
Disease	rs10739757	С	51 (19-74)	246	0.0065
onset, years		TT	54 (16-82)	1136	
DAS28,	rs4837404	AA	5.57 (1.74-8.89)	266	0.0080
score		G	5.28 (1.57-8.29)	321	
DAS28,	rs2302821	С	5.57 (2.39-8.89)	114	0.0321
score		AA	5.31 (1.57-8.37)	491	
DAS 28,	rs11792431	А	5.63 (3.82-8.89)	63	0.0369
score		GG	5.32 (1.57-8.66)	536	

Abbreviations:DAS28, disease activity score.

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TITLES AND LEGENDS TO FIGURES

Figure 1.Linkage disequilibrium map for the *PTGES* locus (r2). The SNPs marked with numberswere analyzed in EIRA case-control study. The SNPs marked with * were analyzed using data from our previous genome-wide association scan (GWAS). Figure 2. The association of haplotype TTC with susceptibility to rheumatoid arthritis in women in two study populations and the meta-analysis. The squares represent odds ratios (ORs) and lines represent confidence intervals (95%CI).

Figure 3.The expression of mPGES-1 in synovial tissue of RA patients in relation to *PTGES* genotypes.Photographs illustrating brown immunoperoxidase staining of mPGES-1positive cells in representative synovial tissue sections from RA patients (A)carrying the minor allele C in SNP rs2302821 and (B) did not carry that allele, counterstained with hematoxylin(original magnification x250).

Fig.1



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	Cases	¢.		Contr	ols	Odds Ratio			
Study population	TTC	Total	Haplotype frequency	TTC	Total	Haplotype frequency	Weight	M-H, Fixed, (95%CI)	
Sweden	388	2792	0.139	163	1520	0.107	76.8%	1.34 (1.11 - 1.63)	
The Netherlands	171	1368	0.125	40	380	0.105	23.2%	1.21 (0.84-1.75)	
Total (95%CI)	559	4160	0.134	203	1900	0.107	100.0%	1.31 (1.11 - 1.56)	

Test for heterogeneity: $\chi^2 = 0.23$, df = 1 (P = 0.63), I² = 0% Test for overall effect: Z = 3.11 (P = 0.002)



Fig.3

