

Deep-sea parasitic nematodes of the genus Rubtsov et Platonova, 1974 (Benthimermithidae) from the Equatorial Atlantic, with the descriptions of two new species

Dmitry M. Miljutin

▶ To cite this version:

D
mitry M. Miljutin. Deep-sea parasitic nematodes of the genus Rubtsov et Platonova, 1974 (Benthimer
mithidae) from the Equatorial Atlantic, with the descriptions of two new species. Helgol
and Marine Research, 2010, pp.245-256. 10.1007/s10152-010-0225-0. hal-00624426

HAL Id: hal-00624426 https://hal.science/hal-00624426

Submitted on 17 Sep 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

3 Deep-sea parasitic nematodes of the genus *Trophomera* Rubtsov et Platonova,
4 1974 (Benthimermithidae) from the Equatorial Atlantic, with the descriptions
5 of two new species

6

7 , Forschungsinstitut Senckenberg, Senckenberg am Meer, Deutsches Zentrum für Marine
8 Biodiversitätsforschung, Südstrand 44, 26382 Wilhelmshaven, Germany. E-mail:
9 dmiljutin@senckenberg.de (corresponding author)

10

11 Abstract Nematode females of the genus Trophomera (Benthimermithidae) from the 12 collection of the Smithsonian's National Museum of Natural History (Washington, DC, USA) 13 were examined. Nematodes were collected in different parts of the Western Atlantic (Hatteras 14 Abyssal Plain, Brazil Basin, and Argentina Basin) from depths of 467–5223 m. Two new species 15 are described. Body length of T. americana sp. n. is 3250-4470 µm; posterior end conical with 16 rounded tip; cephalic setae about 3-4 µm long; trophosome consisting of several longitudinal rows of large cells; ovaries reflected; mature eggs 35 µm in diameter. Body length of T. 17 18 longiovaris sp. n. is 7870–15400 µm; posterior end conical with rounded tip; cephalic sensilla 7 19 µm long; mouth opening vestigial, present as very narrow apical pore; pharynx devoid of 20 internal lumen and muscular envelope; midgut represents a trophosome without internal lumen; 21 trophosomal cells arranged in 3-4 longitudinal rows; rectum and anus vestigial; female 22 reproductive system didelphic, amphidelphic, very long, occupying about 0.8 total body length; 23 ovaries telogonic, outstretched; oviducts very long, repeatedly folded across body axis; proximal 24 parts of oviducts being than distal ones, uterus distinctly formed. New finds of two known 25 species, T. arnauidi and T. marionensis are also recorded and described.

2 Keywords Benthimermithidae, Benthimermis, deep-sea, Nematoda, parasite, taxonomy,

3 Trophomera

1 Introduction

2 Benthimermithids are marine nematodes that have a larval stage parasitizing the body 3 cavity or internal organs of a wide spectrum of mainly deep-sea invertebrates (nematodes, polychaetes, priapulids, and crustaceans). These worms possess a vestigial mouth opening and 4 5 pharynx (the latter is devoid of a muscular envelope and a functional axial internal lumen), and 6 their midgut also has no internal lumen. For this reason, it is presumed that the parasitic stages 7 absorb nutrients through the body surface. The midgut is transformed into a trophosome, which 8 consists of large cells with numerous internal lipid-containing vesicles, and serves as a depot for 9 nutrients (Chesunov 1997; Miljutin & Tchesunov 2001). Late parasitic stages can occupy the 10 entire body of their hosts, and may possible cause the death of the host. Before the terminal molt, 11 benthimermithids exit their host to the external environment, where they do not feed but do 12 reproduce. Evidently, adult benthimermithids consume the nutrients accumulated in their 13 trophosome during the larval parasitizing stage, because senile specimens are virtually lacking in 14 lipid-containing vesicles inside their trophosome, whereas the trophosomes of young, virgin 15 adults are tightly packed with such vesicles. The hosts of the majority of the described 16 benthimermithids are still unknown. Most species were described from very few free-living adult 17 specimens, either males or females, and both sexes are known in only two nominal species 18 [Trophomera australis (Petter, 1983) and T. petterae (Miljutin, 2004)] so far. Some species 19 descriptions have been based on a single specimen. The size range of known benthimermithids 20 varies from less than one millimeter to 17 centimeters; however the body length of most species 21 is several millimeters. Miljutin (2004, 2006) reviewed the anatomy and life cycle of 22 benthimermithids, and discussed the taxonomic problems within the family. Miljutin and 23 Miljutina (2009) produced an up-to-date key to the genera of the family Benthimermithidae and tabular keys to the species of the genus Trophomera.

As a rule, benthimermithids are quite rare, and comprise a fraction of one percent of nematode assemblages (Bussau 1993; Miljutin and Miljutina 2009). Thirty-eight species of benthimermithids belonging to three genera have been described to date (Miljutin and Miljutina 2009). They have been found from the Arctic to Antarctic and throughout the Atlantic and the Indian oceans (Miljutin 2004). It is obvious that benthimermithids are widespread in the World Ocean, and their known habitat will be extended as new deep-sea areas are studied.

7 This contribution presents the results of a study of females belonging to the genus
8 *Trophomera* from the collection of the Smithsonian's National Museum of Natural History
9 (NMNH, Washington, DC, USA).

10 Material and methods

Adult free-living nematodes were collected from sediment samples in several survey cruises in the western and southwestern parts of the Atlantic Ocean. All the cruises mentioned below were organized and carried out by the Woods Hole Oceanographic Institution (USA).

All examined specimens were selected from samples and deposited in the NMNH collection by Dr. D. G. Murphy (Smithsonian Institution, Washington, DC, USA). The USNM collection numbers are provided. Specimens were initially fixed on board with 4% formaldehyde in seawater. Nematodes were then rinsed in distilled water, transferred to anhydrous glycerol according to the progressive evaporation method (Seinhorst 1959), and mounted on a glass slide.

For the present study, nematode specimens were examined using a LEICA DMR
interference microscope fitted with the 1.5x and 2x magnifier and equipped with a drawing tube.

- 21 *Abbreviations:*
- 22 a: ratio of 'body length / maximum body width';

23 amph.dist.: distance from anterior apex to amphid;

c: ratio of 'body length / tail length';

c': ratio of 'tail length / anal body diameter';

1	c.b.d:	corresponding body diameter;
2	d:	ratio of 'body length / distance from anterior apex to amphid';
3	L:	total body length;
4	V:	distance from anterior end to vulva / total body length', %.
5	Taxonomic descri	ptions
6	Trophomera amer	<i>icana</i> sp. n.
7	Figs 1, 2; T	able 1
8	Type mater	ial. Holotype: gravid female, USNM 1000098, station No. 1036 (RV "Chain",
9	cruise 58, 39°43.3	66' N, 70°37.24' W, depth 2022 m, 04.05.1966). Paratype: virgin female,
10	USNM 1000084, s	tation No. 119 (RV "Atlantis II", cruise 24, 32°16' N, 64°32' W, depth 2095
11	m, 19.08.1966).	
12	Location. H	atteras Abyssal Plain, Western Atlantic.
13	Etymology.	Latinized form of "American".
14	Description	
15	Measureme	nts: L = 3250–4470 μ m; a = 40.6–42.6; V = 50.5–55.0% (Table 1).
16	Females. B	ody cylindrical, slightly narrowed at posterior end. Anterior end rounded.
17	Posterior end conic	cal with rounded tip. Cuticle about 3 μ m thick in anterior and midbody, and
18	about 8 µm thick in	n posterior body. Four mediolateral cephalic setae about 3–4 μ m long inserted
19	in small pits. So	matic sensilla in shape of small setae 1.0-1.5 µm long arranged along
20	hypodermal chords	s and present also anterior to amphids. Amphids non-spiral, situated about 1
21	c.b.d. from cephal	ic apex. Amphidial apertures oval, transversely oriented, about 4 μm long.
22	Amphidial fovea 7	$-10 \ \mu m$ in diameter. Mouth opening reduced to small pit in cuticle of apex.
23	Pharynx devoid o	f internal lumen and appearing as a short irregular bar. Gravid female
24	possessing several	large, optically transparent cells without intracellular inclusions anteriorly to
25	beginning of trop	hosome. Trophosome without internal lumen and consisting of several

longitudinal rows of large cells. Cellular borders in trophosome distinct. Rectum and anus vestigial. Female reproductive system amphidelphic and occupying about half of body length in gravid female. Ovaries reflected. Oviduct long, at first directed away from vulva from proximal end of ovary, and then turning towards vulva, filled with eggs. Most mature eggs about 35 µm in diameter, possessing large light vacuoles. Some small cells (possibly spermatozoa) present in narrow middle part of oviduct. Uterus morphologically undifferentiated. Vagina short, with thick cuticular walls. Vulvar glands absent. Male unknown. Males and host unknown.

8

Differential diagnosis

9 Females of *T. americana* sp. n. resemble the females of *T. iturupiensis* (Rubtzov et
10 Platonova, 1974), *T. hureaui* (Petter, 1983), *T. conicauda* (Petter, 1987), *T. tchesunovi* (Miljutin,
11 2004), *T. elegantis* Miljutin et Miljutina, 2009, and *T. senckenbergi* Miljutin et Miljutina, 2009,
12 in the shape of the tail (conical with rounded tip).

T. americana sp. n. differs from *T. elegantis* and *T. senckenbergi* by its longer body (3.2–
4.5 mm versus 1.5 mm and 1.6 mm respectively); by its larger mature eggs (35 μm in diameter
versus 24 μm and 15x9 μm respectively); by the construction of the trophosome (consisting of
several rows of cells, versus one row of cells in both compared species).

T. americana sp. n. differs from *T. iturupiensis* by its much shorter body (3.2–4.5 mm
versus 26–38 mm); and by its smaller "a" index (40–43 versus 98–104).

T. americana sp. n. differs from *T. conicauda* by the shape of the head (rounded versus
conical with rounded tip in *T. conicauda*); by its smaller body size (3.2–4.5 mm versus 5.2–6.8
mm), by the relatively thicker body (a = 41-43 versus 49-58); and by the smaller "d" index (61–
69 versus 108–148).

T. americana sp. n. differs from *T. hureaui* by the relatively thinner body (a = 41–43
versus 25–32), by its relatively short reproductive system in gravid specimens (occupying ca. 0.5

body length versus ca. 0.25 body length), and by its larger eggs (35 μm in diameter versus 25
μm).

- 3 Trophomera arnaudi (Petter, 1983)
- 4 Fig. 3; Table 2
- 5 Petter 1983: 12-14, Fig. 7 (*Benthimermis arnaudi*); Chesunov 1988: 13-15, Figs 1, 2
 6 (*Benthimermis arnaudi*); Miljutin 2006 (*Trophomera arnaudi* comb. n.)
- *Material examined.* One gravid and three virgin females found free-living. Specimen No.
 1 (USNM 1000080) and specimen No. 2 (USNM 1000105), station No. 64 (RV "Atlantis II",
 cruise 12, 38°46.0′N, 70°06.0′W, depth 2886 m, 21.08.1964, NW Atlantic); specimen No. 3
 (USNM 1000107), station D1 (RV "Atlantis II", cruise 277, 39°54.5′ N, 70°35.46′ W, depth 467
 m, 23.05.1962, NW Atlantic); specimen No. 4 (USNM 1000113), station No. 125 (RV "Atlantis
 II", cruise 24, 39°37′ N, 66°44–47′ W, depth 3806 m, 23.08.1966, NW Atlantic).
- 13 *Location*. Western Atlantic, off Cox Ledge.
- 14 *Description*

15 Measurements: $L = 3386-8240 \mu m$; a = 25.3-47.1; c = 67.7-183.0; V = 49-54% (Table

16 2).

17 Females. Body cylindrical. Anterior end conical with rounded apex, posterior end 18 rounded. Cuticle with fine annulation visible in light microscope, or lacking annulation. Cuticle 19 about 5 µm thick along entire body. Four cephalic mediolateral papilloid sensilla, from 2 to 6 µm 20 long. Each cephalic sensillum located in small pit. Ring of several tiny, very narrow labial setae 21 ca. 1 μ m long seen anterior to cephalic papillae in specimen No. 4. Somatic setae 1–2 μ m long, 22 arranged irregularly over entire body and also anteriorly to amphid. Amphidial aperture oval, 23 transversely oriented, about 3 µm wide, situated 0.5–0.85 c.b.d. from cephalic apex. Amphidial 24 fovea not seen. Mouth opening reduced, present as narrow intracuticular canal at cephalic apex. 25 Pharynx devoid of internal lumen and muscular envelope. Pharynx present as narrow bar in 1 specimen No. 3, and body cavity filled by strongly protruded hypodermal chords. Midgut a multicellular trophosome without visible internal lumen. Cellular borders of trophosomal cells 2 3 indistinct. Beginning of trophosome sometimes anterior to mphid in virgin specimens. Rectum a 4 cytoplasmic bar connecting with narrow anal opening. Female reproductive system didelphic, 5 amphidelphic, occupying from 0.3 to 0.5 body length. Ovaries reflected. Vagina short, with thin 6 walls. Diameter of mature eggs in specimen No. 4 about 24 µm. Uterus morphologically 7 undifferentiated and with widened proximal parts of ovaries near vulva. Vulvar glands absent in 8 gravid specimen and present in virgin specimens. Neither morphologically differentiated 9 spermatheca nor spermatozoa discovered.

10 Remarks

T. arnaudi was initially described by Petter (1983) as *Benthimermis arnaudi*, based on
two females from the southern Indian Ocean (46°03'S, 49°47'E, depth 267 m). Subsequently,
Chesunov (1988) reported a new find of *B. arnaudi* in the South Atlantic (52°15′S, 56°51′ W,
depth 1105 m). Afterwards, Miljutin (2006) reduced *Benthimermis* to a junior synonym of *Trophomera*. The new finds of *T. arnaudi* were made far from the previous ones, in the
northwest Atlantic at depths from 467 to 3806 m. Thus, *T. arnaudi* is known from two oceans
(Fig. 4).

The new specimens show great similarity to those described earlier, in their general appearance; in indices "a" (25–47 versus 24–42), "c" (77–183 versus 80–335), "d" (87–140 versus 97–134), and V (49–54% versus 48–56%); in the ratio "length of genital system / body length" (0.3–0.5 versus 0.5); and in the diameter of the mature eggs (24 µm versus 20–30 µm).
The main differences are in the body length: specimen No. 4 is shorter than specimens described previously (3390 µm versus 5600-6800 µm), whereas specimens No. 1 and 2 are longer (7170-8240 µm). In their body length, shape of posterior end, and indices, the longer specimens

resemble *T. edouardensis* (Petter, 1983), but their female reproductive system occupies no more
 than 0.5 body length, versus 0.8–0.9 in *T. edouardensis* (even in virgin females).

3 Large differences in body length are known for some other species of Trophomera. For 4 example, the differences in body length of specimens of *T. turpicauda* (Miljutin, 2004) are about 5 twofold (Miljutin 2004). The same occurs with T. megala (Petter, 1987) (Miljuitin & Tchesunov 6 2001), and in the type series of T. acuticauda (Petter, 1981) (Petter, 1981a), T. diploptera (Petter, 7 1981) (Petter, 1981b), T. litoralis Miljutin, 2006 (Miljutin 2006), and T. longiovaris sp. n. 8 (present paper). Such a wide range of body sizes within the same species may be caused by 9 differences in the host body sizes, the number of parasitic larvae inside the same host individual 10 (several larvae may somehow inhibit each other's growth or compete with each other for food 11 resources), or by the different locations of larvae in different hosts. For example, parasitizing a 12 crustacean's gonad may provide a larva with better food or allow it more space for growing, than 13 when a larva infects an internal cavity of a crustacean's coxal plate. Parasitizing benthimermithid 14 larvae were found both in gonads of amphipods and in internal cavities of amphipods' coxal 15 plates (author's unpublished data).

In its general appearance and body parameters, the females of *T. arnaudi* closely resemble *T. bathycola* (Rubtzov, 1980). However, according to the original description, the former species possess reflected ovaries, in contrast to the outstretched ones in *T. bathycola*. The original description was brief and lacked detailed illustrations. It is possible that the specimens described by Rubtzov (1980) do possess reflected ovaries, and if this is the case, *T. bathycola* should be merged with *T. arnaudi*.

22 Trophomera longiovaris sp. n.

23 Figs 5–7; Table 3

Type material. Holotype: virgin female, USNM 1000112, station No. 85(2) (RV "Chain",
cruise 50, 37°59.2′N, 69°26.2′W, depth 3834 m, 05.06.1965, NW Atlantic). Paratype: virgin

1 female, USNM 1000102, station No. 95 (RV "Atlantis II", cruise 38°33' N, 68°32' W, depth

2 3753 m, 12.12.1965, NW Atlantic).

3 *Location*. Hatteras Abyssal Plain, Western Atlantic.

- 4 *Etymology*. From Latin *longa* (long) and *ovarium*.
- 5 *Description*

6 Measurements: L = 7870–15400 μ m; a = 52.5–71.0; c = 115.7–115.8; V = 46.8–53.2%

7 (Table 3).

8 Females. Body fusiform, narrowing slightly toward anterior and posterior ends. Anterior 9 end rounded-conical, posterior end conical with rounded apex. Body cuticle gradually thickening 10 from head to tail. Cuticle ca. 2 µm thick at anterior end, slightly thickened posteriorly up to 5–7 11 um at caudal part. Cuticle of tail forming large annular rugae. Four mediolateral cephalic sensilla 12 ca. 7 µm long. Somatic sensilla present as short conical papillae about 0.5 µm long, and arranged 13 along lateral hypodermal chords. Amphids non-spiral, situated 0.5–0.6 c.b.d. from cephalic apex. 14 Amphidial aperture round, about 1.5–2.0 µm in diameter. Amphidial fovea not seen. Mouth 15 opening vestigial, present as very narrow apical pore; cuticle at cephalic apex about half as thick 16 as cuticle of other cephalic parts. Pharynx devoid of internal lumen and muscular envelope. Anterior part of pharynx appearing as an indistinctly formed bar, posterior part an unformed 17 18 cellular mass. Midgut being a trophosome without internal lumen. Trophosomal cells arranged in 19 3-4 longitudinal rows anterior to and posterior to reproductive system) in an optical cross 20 section, with distinct borders and granular contents. Rectum appearing as a short bar without 21 cuticular walls. Anus not clearly visible, appearing as a small transverse line in cuticle. Female 22 reproductive system didelphic, amphidelphic, very long, occupying about 0.8 total body length. 23 Ovaries telogonic, outstretched. Oviducts very long, folded repeatedly across body axis. 24 Proximal parts of oviducts wider than distal ones, uterus distinctly formed. Vagina short, with thin walls. Vulva a transverse slot. Vulvar glands not developed. Neither morphologically
 differentiated spermatheca nor spermatozoa discovered. Males and host unknown.

3 Differential diagnosis

Only two known species of *Trophomera* possess outstretched ovaries: *T. laubieri* (Petter,
1987) and *T. pseudominuta* (Miljutin, 2004). *T. senckenbergi* Miljutin et Miljutina, 2009 also
possesses ovaries that may be outstretched.

The female of *T. longiovaris* sp. n. differs from the female of *T. laubieri* by its longer
body (7.9–15.4 mm versus 5.2–6.7 mm), "c" index (116 versus 58-80), length of cephalic
papillae (7.0 μm versus 3.0–3.5 μm), and the ratio "gonad length / body length" (ca. 0.8 versus
ca 0.5). The cephalic papillae of *T. longiovaris* sp. n. are not inserted in small pits, in contrast to
the cephalic papillae of *T. laubieri*.

The female of *T. longiovaris* sp. n. differs from the female of *T. pseudominuta* by its much longer body (7.9–15.4 mm versus 2.8 mm), the "a" index (53–71 versus 23), the ratio "gonad length / body length" (0.8 versus 0.2), and the shape of the tail (conical versus rounded).

The female of *T. longiovaris* sp. n. also differs from the female of *T. senckenbergi* by its
much longer body (7.9–15.4 mm versus 1.6 mm); and by the de Man indices "a" and "c"
(respectively 53–71 versus 23 and 116 versus 45).

18 Very long ovaries were described also in *T. edouardensis* (Petter, 1983) and in *T. megala*19 (Petter, 1987). However, the former species possesses reflected ovaries, and the latter has
20 hologonic ovaries (the germinal zone extends throughout the length of the ovary) (Miljutin and
21 Tchesunov 2001).

22

Remarks

The holotype is two times longer than the paratype, although both specimens are at a similar stage of maturity. However, judging by their body shape, body proportions, and the construction of the reproductive system, these specimens undoubtedly belong to the same species. Such differences in the body length have been described in other species of *Trophomera* (Petter 1981a,b; Miljutn & Tchesunov 2001; Miljutin 2004; present paper).

The ovaries of *T. longiovaris* sp. n. are outstretched. Both examined specimens are virgin females without fertilized eggs in their oviducts and uteri. Two other species of *Trophomera* species possess outstretched ovaries: *T. laubieri* and *T. pseudominuta*, and both species were described from gravid females with eggs in their oviducts. Thus outstretched ovaries are not just a stage in the female's ontogenesis, and their structure can be used in benthimermithid taxonomy. Indeed, the structure of benthimermithid ovaries (telogonic reflected, telogonic outstretched, hologonic) is usually recognizable in virgin females.

10 Trophomera marionensis (Petter, 1983)

1	1	Fig. 3	8
			~

12 Petter 1983: p. 9–10, Fig. 5 (Benthimermis marionensis); Chesunov 1988: p. 15–17, Figs

13 3, 4 (B. marionensis); Miljutin 2004: p. 32–34, Fig. 7; Miljutin 2006 (Trophomera marionensis

14 comb. n.); Miljutin and Miljutina 2009: p. 181–182, Fig. 4 (*T. marionensis*).

- Material examined. One virgin female, USNM 1000078, station No. 156 (RV "Atlantis
 II", cruise 31, 14.02.1967, 06°46'S, 29°24–28'W, depth 3459 m).
- 17 *Location*. Brazil Basin, Western Atlantic.
- 18 *Description*

Measurements: L = 9030 μm; a = 48.3; c = 39.0; c'=2.3; d = 139; V = 57.7%. Maximum
body diameter = 187 μm. Diameter at level of: cephalic sensilla = 25 μm; amphids = 70 μm;
midbody = 185 μm; anus = 100 μm. Ratio of 'body length / length of caudal terminal spine' =
65.

Body long, cylindrical. Anterior end rounded-conical. Posterior end conical, possessing
elongated terminal spine 140 µm long. This terminal spine consists of distinctly visible axial core
with granular inclusions (spherical granules or vesicles) and thinner external layer without

1 inclusions. Body cuticle ca. 5 µm at level of midbody and ca. 7 µm at anterior apex. Fine 2 annulation of cuticle visible in light microscope. Mouth opening vestigial, present as narrow 3 intracuticular channel. Four thick, cylindrical, mediolateral papilloid cephalic setae 5 µm long 4 inserted in pits. Amphidial aperture round, very small (ca. 1.5 µm in diameter), situated 65 µm (ca. 1 c.b.d.) from cephalic apex. Amphidial fovea ca. 8 µm in diameter; canalis amphidialis 5 6 directed posteriorly and ventrally. Somatic sensilla ca. 1.5 µm long arranged along hypodermal chords in cervical part of body. Cuticular pores connected with hypodermal glands seen along 7 8 lateral hypodermal chords at level of midgut. Pharynx resembling a non-muscular string without 9 an internal lumen and muscular envelope. Body cavity at level of pharynx filled by strongly 10 protruded hypodermal chords. Midgut a multicellular trophosome without visible internal lumen. 11 Rectum and anus present. Female reproductive system not developed. Vulvar glands present.

12

Remarks

The features of the new-found female agree very well with the description of the type by Petter (1983) in regard to the body length, the length and shape of the cephalic papillae, the shape and core structure of the caudal terminal spine, and in the De Man indices a = 48.3 and V = 57.7% (versus 46.5-74.0 and 36.9-56.8% in the type specimens).

17 *T. marionensis* was initially described from eight gravid females from the southern Indian 18 Ocean, 46°52'S, 37°51'E, 31-110 m depth (Petter 1983). Subsequent finds were reported by Chesunov (1988) in the South Atlantic (57°09'S, 26°09'W, depth 1729 m); by Bussau (1993) in 19 20 the Tropical SE Pacific (7°04'S, 88°27'W, depth 4154 m); by Miljutin (2004) in the Western 21 Atlantic (8°07.88'N, 49°03.71'W, depth 4440 m); and by Miljutin and Miljutina (2009) in the 22 Tropical NE Pacific (13°55.63'N, 130°12.2'W, depth 4800 m). The new specimen was found quite far from all previous locations (Fig. 9). Therefore, *T. marionensis* is now known from three 23 24 oceans and may be considered as a cosmopolitan species (at least, until methods of molecular 25 taxonomy are applied).

1 Acknowledgements

The author thanks Dr. W. D. Hope (National Museum of Natural History, Smithsonian
Institution, Washington, DC, USA), who kindly made available the museum's collection of
benthimermithids for investigation, and Dr. A. V. Tchesunov (Moscow State University, Russia),
who examined this collection and selected the specimens to be described.

6 References

7 Bussau C (1993) Taxonomische und ökologische Untersuchungen an Nematoden des Peru8 Beckens. Dissertation, Christian-Albrechts-Universität zu Kiel, 621 pp.

9 Chesunov AV (1988) [New finds of deep-sea nematodes of the family Benthimermithidae in
10 South Atlantic with description of a new species.] Vestnik Zoologii 5: 12–22. In Russian.
11 Chesunov AV (1997) Marimermithids (Nematoda): anatomy, taxonomy, and phylogeny. Russ J

12 Zool 1(4): 440–455.

- Miljutin DM (2004) New findings of deep-sea nematodes of the family Benthimermithidae
 Petter, 1980 with descriptions of seven new species. Zoosystema 26(1): 21–48.
- Miljutin DM (2006) The genus *Trophomera* Rubtzov & Platonova, 1974 with description of *T*. *litoralis* sp. n. (Nematoda: Benthimermithidae) from the tidal zone of the Kuril
 Archipelago and proposal of *Benthimermis* Petter, 1980 as a junior synonym.
 Nematology 8(3): 411–423.
- Miljutin DM, Tchesunov AV (2001) On the histological anatomy of *Benthimermis megala*Petter, 1987, a giant nematode from the Norwegian deep-sea (Nematoda:
 Benthimermithidae). Nematology 3(6): 491–502.
- Miljutin DM, Miljutina MA (2009) Description of *Bathynema nodinautensis* gen. n., sp. n. and
 four new *Trophomera* species (Nematoda: Benthimermithidae) from the Clarion Clipperton Fracture Zone (Eastern Tropic Pacific), supplemented with the keys to genera

1	and species. In: Brökeland W, George KH (eds) Deep-sea taxonomy - a contribution to
2	our knowledge of biodiversity. Zootaxa 2096: 173–196.
3	Petter AJ (1981a) Description des mâles d'une nouvelle espèce de nématode marin de la famille
4	des Benthimermithidae. Ann Parasitol Hum Comp 56: 285–295.
5	Petter AJ (1981b) Description des mâles de trois nouvelles espèces de nématodes de la famille
6	des Benthimermithidae. Bull Mus Natl Hist Nat, Paris, 4-e Série, 3, Section A 2: 455-
7	465.
8	Petter A-J (1983) Quelques nouvelles espèces du genre Benthimermis Petter, 1980
9	(Benthimermithidae: Nematoda) du Sud de l'Océan Indien. Syst Parasitol 5: 1–15.
10	Petter A-J (1987) Quelques nouvelles espèces de femelles du genre Benthimermis Petter, 1980
11	(Benthimermithidae: Nematoda) des grands fonds de la mer de Norvège. Bull Mus Nat
12	Hist Natl Paris (3) 9(A3): 565–578.
13	Rubtzov IA (1980) [The new marine parasitic nematode, Abos bathycola, from priapulids and a
14	taxonomic position of the family Marimermithidae in the class Nematoda]. Parazitologiya
15	14: 177–181. In Russian with English summary.
16	Rubtzov IA, Platonova TA (1974) [A new family of marine parasitic nematodes.] Zool Zh 53,
17	1445–1458. In Russian.
18	Seinhorst JW (1959) A rapid method for the transfer of nematodes from fixative to anhydrous
19	glycerin. Nematologica 4: 67–69.
20	

1 Tables

Specimen status	Holotype	Paratype
Maturity	gravid	virgin
L, µm	4470	3250
Amph.dist., µm	73	47
Tail length, μm	90	103
Diameter at level of cephalic sensilla, μm	34	24
Diameter at amphidial level, µm	79	47
Diameter at level of midbody, μm	105	80
Maximal body diameter, µm	105	80
Diameter at anal level, µm	48	48
a	42.6	40.6
c	55.0	50.5
c'	1.4	2.6
d	61	69
V, %	50.5	55.0

2 Table 1. Main measurements of females of *T. americana* sp. n.

Specimen, No.	1	2	3	4
Maturity	virgin	virgin	virgin	gravid
L, µm	7170	8240	5170	3390
Amph.dist., µm	56	95	37	36
Tail length, μm	44	45	34	44
Length of cephalic sensilla, μm	5	6	3	3
Diameter at level of cephalic sensilla, μm	53	61	43	27
Diameter at amphidial level, μm	89	112	75	54
Diameter at level of midbody, μm	160	175	187	134
Maximal body diameter, µm	164	177	187	134
Diameter at anal level, µm	130	149	100	77
a	43.7	47.1	27.7	25.3
c	163	183	152	77
c'	0.34	0.30	0.34	0.57
d	128	87	140	94
V, %	52.4	52.0	49.2	54.0

1 Table 2. Main measurements of females of *T. arnaudi* (Petter, 1983).

Collection status	holotype	paratype
Maturity	virgin	virgin
L, µm	15400	7870
Amph.dist., μm	40	37
Tail length, μm	134	88
Diameter at level of cephalic sensilla, µm	35	27
Diameter at amphidial level, μm	65	37
Diameter at level of midbody, μm	212	150
Maximal body diameter, µm	216	150
Diameter at anal level, µm	127	80
a	71.0	52.5
c	115.8	115.7
c'	1.1	1.1
d	385	213
V, %	46.8	53.2

1 Table 3. Main measurements of females of *T. longiovaris* sp. n.

1 Figure legends

2

3 Fig. 1. Trophomera americana sp. n., females. A: holotype, general appearance; B,C: holotype, 4 anterior end; D: holotype, reproductive system; E: paratype, anterior end; F: holotype, 5 body fragment at level of trophosome; G: holotype, posterior end. Scale bars: $A_{,D} = 500$ 6 μ m; B,C,E,F,G = 50 μ m. 7 Fig. 2. Trophomera americana sp. n., micrographs, female, holotype. A, B: head; C: fragment of 8 trophosome before genital system. Scale bars: $A.B = 50 \mu m$; $C = 200 \mu m$. 9 Fig. 3. Trophomera arnaudi (Petter 1983), females. A – specimen No. 4, general appearence; 10 B,C: specimen No. 4, anterior end; D: specimen No. 4, reproductive system; E: specimen 11 No. 3, posterior end; F: specimen No. 3, anterior end; G specimen No. 4, posterior end. 12 Scale bars: A = 500 μ m; B,C,F,G = 50 μ m; D = 200 μ m; E = 100 μ m. Abbreviations: a = 13 amphid; h = head; l.ch. = lateral chord; t = trophosome. 14 Fig. 4. Map of findings of *T. arnaudi*. 1: Petter 1983; 2: Chesunov 1988; 3: present work. 15 Fig. 5. Trophomera longiovaris sp. n., holotype, female. A: general appearance; B: posterior end. 16 Scale bars: A = 2 mm; $B = 300 \mu \text{m}$. 17 Fig. 6. Trophomera longiovaris sp. n., females. A,B: holotype, general appearance; C: holotype, 18 central part of reproductive system; D: holotype, posterior end; E: holotype, beginning of 19 anterior ovary; F: paratype, anterior end; G: paratype, posterior end. Scale bars: A, B, F, G 20 $= 50 \ \mu m; D, E = 100 \ \mu m; C = 500 \ \mu m.$ 21 Fig. 7. Trophomera longiovaris sp. n., micrographs, females. A: holotype, head; B: paratype, 22 tail; C: holotype, fragment of posterior branch of genital system; D: holotype, tail. 23 Abbreviations: an = anus; $g_z = germinal$ zone of ovary; d = oviduct; ov = ovary; t = ovary24 trophosome, v = vulva. Scale bar = 200 μ m.

2 hypodermal chord with glands. Scale bars: A, $C = 40 \ \mu m$; $B = 100 \ \mu m$.

- **3** Fig. 9. Map of findings of *T. marionensis*. 1: Petter 1983; 2: Chesunov 1988; 3: Bussau 1993; 4:
 - Miljutin 2004; 5: Miljutin and Miljutina 2009; 6: present work.
- 5

4















