

# Model-Based Multifactor Dimensionality Reduction to detect epistasis for quantitative traits in the presence of error-free and noisy data

Kristel van Steen, Jestinah M Mahachie John, François van Lishout

# ► To cite this version:

Kristel van Steen, Jestinah M Mahachie John, François van Lishout. Model-Based Multifactor Dimensionality Reduction to detect epistasis for quantitative traits in the presence of error-free and noisy data. European Journal of Human Genetics, 2011, 10.1038/ejhg.2011.17. hal-00624157

# HAL Id: hal-00624157 https://hal.science/hal-00624157

Submitted on 16 Sep 2011

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# Model-Based Multifactor Dimensionality Reduction to detect epistasis for quantitative traits in the presence of error-free and noisy data

Jestinah M. Mahachie John<sup>1, 2</sup>, François Van Lishout<sup>1, 2</sup>, Kristel Van Steen<sup>1, 2</sup>

<sup>1</sup>Systems and Modeling Unit, Montefiore Institute, University of Liege, Grande Traverse 10, 4000 Liège, Belgium

<sup>2</sup>Bioinformatics and Modeling, GIGA-R, University of Liege, Avenue de l'Hôpital 1, 4000 Liège, Belgium

#### **Correspondence to:**

#### Kristel Van Steen, PhD

University of Liege

Systems and Modeling Unit, Montefiore Institute

Bât. B28 Bioinformatique

Grande Traverse 10

4000 Liège 1

Belgium

Telephone: +3243662692

E-mail: kristel.vansteen@ulg.ac.be

Running title: MB-MDR for quantitative traits and noisy data

#### Abstract

Detecting gene-gene interactions or epistasis in studies of human complex diseases is a big challenge in the area of epidemiology. To address this problem, several methods have been developed, mainly in the context of data dimensionality reduction. One of these methods, Model-Based Multifactor Dimensionality Reduction, has so far mainly been applied to casecontrol studies. In this study, we evaluate the power of Model-Based Multifactor Dimensionality Reduction for quantitative traits to detect gene-gene interactions (epistasis) in the presence of error-free and noisy data. Considered sources of error are genotyping errors, missing genotypes, phenotypic mixtures and genetic heterogeneity. Our simulation study encompasses a variety of settings with varying minor allele frequencies and genetic variance for different epistasis models. On each simulated data, we have performed Model-Based Multifactor Dimensionality Reduction in two ways: with and without adjustment for main effects of (known) functional SNPs. In line with binary trait counterparts, our simulations show that the power is lowest in the presence of phenotypic mixture or genetic heterogeneity compared to scenarios with missing genotypes or genotyping errors. In addition, empirical power estimates reduce even further with main effects corrections, but at the same time, false positive percentages are reduced as well. In conclusion, phenotypic mixtures and genetic heterogeneity remain challenging for epistasis detection, and careful thought must be given to the way important lower-order effects are accounted for in the analysis.

**Keywords**: Model-Based Multifactor Dimensionality Reduction, gene-gene interactions, quantitative traits, complex diseases, noisy data

#### Introduction

Understanding the effects of genes on the development of complex diseases and traits in human is a major aim of genetic epidemiology. These kinds of diseases are controlled by complex molecular mechanisms characterized by the joint action of several genes which could have different effect sizes. In this context, traditional methods within a regression paradigm involving single markers have limited use and more advanced and efficient methods are needed to identify gene-gene interactions and epistatic patterns of susceptibility. One of these methods is the Multifactor Dimensionality Reduction method (MDR)<sup>1</sup>, which nicely tackles the dimensionality problem involved in interaction detection by pooling multi-locus genotypes into two groups of risk based on some threshold value. Those cells with a case/control ratio equal to or above the threshold are labeled as High risk and the remaining cells as Low risk. Although MDR has been widely and successfully used for interaction detection (e.g., URL: http://compgen.blogspot.com/2006/05/mdrapplications.html), it suffers from some major drawbacks, including that important interactions could be missed due to pooling too many cells together or that proposed MDR analyses will only reveal at most one significant epistasis model, selection being based on computationally demanding cross-validation and permutation strategies. To overcome the aforementioned hurdles, Calle et al. (2008)<sup>2,3</sup> developed Model-Based MDR (MB-MDR) for dichotomous traits and unrelated individuals, hereby providing the basis for a flexible framework to detect gene-gene interactions. The method has been made available via an R package *mbmdr*. The principal difference between MDR and MB-MDR is that MB-MDR merges multi-locus genotypes exhibiting some significant evidence of High or Low risk, based on association testing or modeling, rather than on comparison with a threshold value. In addition, those multi-locus genotypes that either show no evidence of association or have no sufficient sample size contribute to an additional MB-MDR category, that of 'No

Evidence for risk'. Note that despite the fact that Lou *et al.* $(2007)^4$  recognized in part the necessity to adjust for covariates and to extend MDR to quantitative traits, issues related to significance assessment remain, as explained in detail by Cattaert *et al.*(2010).<sup>5</sup>

Although a beta-version of MB-MDR for quantitative traits has already been applied by Mahachie John *et al.* (2010)<sup>6</sup>, its power under several conditions, including the presence of error sources or noise (e.g., genotyping errors, missing genotypes, phenotypic mixtures and genetic heterogeneity) has never been investigated. The aim of the current study is to evaluate the power of MB-MDR for quantitative traits to detect gene-gene interactions, for a variety of simulated scenarios. We will restrict attention to 2-order interactions, although MB-MDR can be used to highlight gene-gene interactions of any order.

#### Methods

#### **MB-MDR:** Model-Based Multifactor Dimensionality Reduction

The three steps of the MB-MDR strategy used for this simulation study are summarized below and visualized in Figure 1. For more general details, we refer to Cattaert *et al.* (2010).<sup>5</sup>

#### **MB-MDR Step 1: Multi-locus cell prioritization**

For every pair of markers, data is organized in a 2-way table, with 9 genotype cells. Each 2locus genotype cell,  $c_j$ , in such a table, is assigned to one of three risk categories, High risk (H), Low risk (L) or No Evidence for risk (O), as a result of association tests on each of the individual 2-locus genotype cells with the response variable Y. Cell-dependent testing for H, L, O labeling is done with a Student's t test, at the liberal significance level of 0.10, since the power to detect association using individual cells is likely to be limited. If for a 2-locus cell, the Student's t-test, comparing the cell's mean with the mean of the remaining 8 cells is not significant at 0.10, the cell is labeled as O. The sign of the Student's t-test statistic is used to distinguish between H and L: a positive (negative) sign refers to risk H(L).

#### MB-MDR Step 2: Association test on lower-dimensional construct

The result of the first step is thus a new categorical variable X with values H, L and O, which captures information about the importance of the pair of markers with respect to the trait. A new association test is subsequently performed now for the new construct X on Y. In particular, we consider the maximum of  $W_H$  and  $W_L$ , which are Student's t-tests for comparing H versus  $\{L, O\}$  means or L versus  $\{H, O\}$  means, respectively.

#### **MB-MDR Step 3: Significance assessment**

Once the dimensionality reduction procedure has been implemented and tests for association have been performed, for every pair of markers in the data, a single test result  $\{W_H, W_L\}$  per marker pair is obtained. Because the test statistics are obtained after combining cells according to X, using information about the trait  $Y, W_H$  and  $W_L$  will no longer be t-distributed. In fact, these tests are expected to generate inflated type I errors. We therefore assess significance of max  $\{W_H, W_L\}$  per marker-pair, by adopting a permutation-based strategy (999 replicates) that corrects for multiple testing (over all marker pairs) and adequately controls FWER at  $\alpha$ =5%. In particular, we implement the step-down maxT adjusted p-values approach, as outlined Westfall and Young (1993).<sup>7</sup>

#### FIGURE 1: ABOUT HERE

#### Adjustment for main effects

Some interactions can be identified simply because of highly significant lower-order effects, and are therefore not genuine. That is why we also consider MB-MDR adjusted analyses in the following way: with and without adjustment for main effects of functional SNPs. Main effects are adjusted for in MB-MDR by first regressing them out in a data preparation step and then considering the residuals from the regression model as new traits. Two extreme ways of correcting are considered: the additive model and the co-dominant model. When adjusting for main effects in the presence of genetic heterogeneity, we take into account that different functional pairs are relevant for heterogeneous sub-populations.

#### **Data Simulation**

Each of 500 data sets in a simulation setting consists of 1500 unrelated individuals with 10 SNPs (in linkage equilibrium), 2 of which are functional. The minor allele frequencies of a non-functional marker SNP<sub>j</sub> are fixed at  $p_j = 0.1 + (j-3) * 0.05$ , j = 3,...,10, whereas the minor allele frequencies of the functional SNPs (SNP<sub>1</sub>, SNP<sub>2</sub>) are taken to be equal, and varying as  $(p_1, p_2) = (p, p), p \in \{0.1, 0.25, 0.5\}$ . All SNPs are assumed to be in Hardy-Weinberg Equilibrium.

Two epistasis models that incorporate varying degrees of epistasis are considered: Model 27 and Model 170 of Evans *et al.* (2006)<sup>8</sup>, hereafter referred to as M27 and M170, respectively. In order to increase the phenotypic mean, M27 requires an individual to have at least one copy of the increaser allele at both loci whereas M170 requires an individual to be heterozygous at one locus and homozygous at the other. As *p* increases, the contribution to the total genetic variance of epistasis variance relative to main effects variances increases for M170 (decreases for M27) (Cattaert *et al.* 2010-Table 1).<sup>5</sup> The phenotypic means for these epistasis models only take two values,  $\mu_L$  (Low phenotypic mean) and  $\mu_H$  (High phenotypic mean). The total phenotypic variance  $\sigma_{not}^2$ , i.e. the sum of genetic variance at both loci  $2\sigma_1^2 = \sigma_{main}^2$  (the minor allele frequencies for the functional SNPs are taken to be the same), epistasis variance  $\sigma_{epi}^2$ , and environmental variance  $\sigma_{env}^2$ , is fixed at 1. As a consequence, the total genetic variance, defined as  $2\sigma_1^2 + \sigma_{epi}^2$ , has an interpretation of a broad heritability measure. Throughout this document it will further be referred to as  $g^2$ , to clearly indicate that the interpretation as a heritability is due to the imposed normalization constraints. The parameter  $g^2$  is varied as  $g^2 \in \{0.01, 0.02, 0.03, 0.05, 0.1\}$ . Explicit formulae for these variance components can be obtained from Evans *et al* (2006).<sup>8</sup> In addition, 1000 null data sets are generated under the most general null hypothesis of no association between any of the 10 SNPs and the trait (i.e.,  $g^2=0$ , no main effects and no

epistasis).

Typo 1

#### Introducing noise

Apart from simulating error-free data, we also simulate different error-sources to investigate their impact on the performance of MB-MDR. These involve introducing 5% and 10% missing genotypes (MG5 and MG10), 5% and 10% genotyping error (GE5 and GE10), 25% and 50% phenotypic mixtures (PM25 and PM50) and 50% genetic heterogeneity (GH). It is important to realize that the foregoing derivations of variance decomposition relate to a population as whole. When generating sources of error, estimates of variability will no longer tend to the estimates at the population level. In other words, the actual genotypic variance will no longer equal  $g^2$ . Missing genotypes (MG5 and MG10) and genotyping errors (GE5 and GE10) are also introduced in the null data, leading to a total of 255 simulation settings, so as to be able to assess the impact of these on MB-MDR's type I error control in the presence of noise.

In particular, scenarios MG5 and MG10 are generated by selecting genotypes completely at random from the original data and by setting them to missing. This introduces different perindividual and per-SNP percentages of missingness, reducing the effective sample size, yet maintaining the validity of the variance components estimates. As in Ritchie *et al.* (2003)<sup>9</sup>, genotyping error is simulated using a directed-error model.<sup>10</sup> This model postulates that there is a larger probability for the minor allele to be consistently mis-genotyped (over-represented). In this study, either 5% (GE5) or 10% (GE10) of the available genotypes in the original data set are sampled. From these, homozygous genotypes for the common allele become heterozygous and heterozygous genotypes for the rare allele become homozygous. The effect of adding genotyping errors to the original data is that the actual genetic contribution  $\sigma_{gen}^2$  to the trait variance is reduced compared to the assumed genetic variance  $g^2$ , of the simulation setting due to the additional variability (noise) introduced into the system (Table 1).

#### TABLE 1: ABOUT HERE

Genetic heterogeneity is simulated such that there are actually two different two-locus combinations increasing/decreasing the phenotypic mean. Half of the individuals have one pair of functional SNPs (SNP1 and SNP2), and the other half have the other pair of functional SNPs (SNP3 and SNP4). Introducing the notations  $G_L$  ( $G_H$ ) as the multi-locus genotypes leading to a Low (High) phenotypic mean, traits are simulated according to the distributions specified below:

$$\begin{split} Y|_{g \in G_L, g' \in G_L} &\sim N(\mu_L, \sigma_{env}^2), \qquad Y|_{g \in G_L, g' \in G_H} \sim 0.5N(\mu_L, \sigma_{env}^2) + 0.5N(\mu_H, \sigma_{env}^2), \\ Y|_{g \in G_H, g' \in G_H} &\sim N(\mu_H, \sigma_{env}^2), \qquad Y|_{g \in G_H, g' \in G_L} \sim 0.5N(\mu_H, \sigma_{env}^2) + 0.5N(\mu_L, \sigma_{env}^2) \end{split}$$

Minor allele frequencies of all 4 functional SNPs are taken to be equal, i.e.,  $p \in \{0.1, 0.25, 0.5\}.$ 

Phenotypic mixing in genetics may occur when a percentage of individuals with high phenotypic mean have genotype combinations that are consistent with low phenotypic mean. In particular, a mixing proportion of  $w \in [0,1]$  of phenotypic mixture, trait values are simulated according to

 $Y|_{g\in G_L} \sim (1 - w\pi) N(\mu_L, \sigma_{env}^2) + w\pi N(\mu_H, \sigma_{env}^2), \quad Y|_{g\in G_H} \sim w(1 - \pi) N(\mu_L, \sigma_{env}^2) + [1 - w(1 - \pi)] N(\mu_H, \sigma_{env}^2)$ 

with mixing proportion either 25% (PM25) or 50% (PM50), and  $\pi$ , the probability of a multilocus genotype giving rise to a high phenotypic mean  $\mu_H$ .

#### Results

#### The impact of not correcting for lower-order effects

Table 2 gives an overview of MB-MDR empirical type I error rates in the presence and absence of noise (MG and GE). We observe that MB-MDR empirical type I error percentages are close to the nominal type I error percentage of 5%, when no correction for main effects is performed. When we adjust for main effects, type I error percentages are further reduced and seem to drop below the theoretical value. Similar trends are observed when genotyping errors and missing genotypes are introduced in the data.

#### TABLE 2: ABOUT HERE

Power estimates of MB-MDR to detect the correct interacting pair, SNP1 x SNP2 (in the absence of genetic heterogeneity) from error-free and noisy data are shown in Figure 2. The actual numerical results of the power profiles plotted in Figure 2 are presented in Table S1. This table also includes the corresponding empirical power estimates related to main effects adjusted analyses.

In the absence of any adjustment for lower order genetic effects (i.e., main effects), we notice that power profiles largely follow the same trajectory, except in the presence of 50% phenotypic mixture (PM50). For all scenarios of p, power increases with increasing  $g^2$  (Figure 2 and Table S1). Moreover, the power of MB-MDR (ranging from 54% to 100%, p=0.1, 38% to 100%, p=0.25, 33% to 100%, p=0.5 under M170 and from 44% to 100%, p=0.1, 43% to 100%, p=0.25, 39% to 100%, p=0.5 under M27 for error-free data; Table S1) is hardly affected by introducing small percentages of missing genotypes (MG5 in Figure 2), irrespective of the epistasis model under investigation. Power estimates for MG5 range from 42% to 100%, p=0.1, 33% to 100%, p=0.25, 28% to 100%, p=0.5 and from 33% to 100%, p=0.1, 34% to 100%, p=0.25, 31% to 100%, p=0.5 under M170 and M27, respectively (Table S1). For MG10, power obviously reduces further, but not in a dramatic way compared to MG5: power estimates reduce to a minimum of 31%, p=0.1, 25%, p=0.25, 25%, p=0.5 and to a minimum of 31%, p=0.1, 28%, p=0.25, 22%, p=0.5 for M170 and M27, respectively). When 5% genotyping errors are introduced in the population, systematically lower power curves are obtained than in the presence of randomly missing genotypes. However, high percentages of genotyping error (GE10) or high percentages of phenotypic mixture (PM50) generally lead to the lowest power performance of MB-MDR (Figure 2). For model M170 power estimates in the presence of 10% genotyping errors are in the range of 12% to 100%, p=0.1, 8% to 100%, p=0.25, 12% to 100%, p=0.5 and in the range of 9% to 100%, p=0.1, 20% to 100%, p=0.25, 26% to 100%, p=0.5 for model M27 (Table S1). High percentages of phenotypic mixture have a negative impact on MB-MDR power, which is also indicated by the minimally observed empirical power estimates for PM50. Power estimates for the latter are in the range of 3% to 98%, p=0.1, 3% to 97%, p=0.25, 2% to 95%, p=0.5 for M170 and in the range of 3% to 95%, *p*=0.1, 2% to 97%, *p*=0.25, 3% to 95%, *p*=0.5 for M27.

#### FIGURE 2: ABOUT HERE

Not surprisingly, there is a higher chance of identifying epistasis models for analyses without main effects correction as compared to analyses that do account for lower-order effects. The latter epistasis models usually involve other SNPs pairing with one of the functional SNPs (results not shown) and should therefore be considered as false positives. Empirically estimated false positive percentages, for a variety of scenarios, excluding GH settings, are reported in Table S2 ("No Correction" versus "Main Effects Correction" estimates). For error-free data, and no adjustments for main effects, the false positive percentage of MB-MDR of identifying a significant epistasis model not involving the actual functional pair of SNPs ranges from 28% to 100%, p=0.1, 6% to 53%, p=0.25, 6% to 7%, p=0.5 for M170 and from 15% to 99%, p=0.1, 26% to 100%, p=0.25, 38% to 100%, p=0.5 for M27. When main effects are accounted for in error-free data, the false positive percentage ranges from 3% to 39%, p=0.1, 3% to 12%, p=0.25, 3% to 6%, p=0.5 under M170 and from 3% to 7%, p=0.1, 3% to 21%, p=0.25, 2% to 98%, p=0.5 under M27 (Table S2). In general, Table S2 shows that irrespective of how the main effects adjustment is performed (using an additive or codominant model) and irrespective of the type of noisiness introduced, false positive percentages are typically lower than their "uncorrected" counterparts.

#### The impact of appropriately correcting an epistasis analysis for lower-order effects

Profiles for the empirical power estimates of MB-MDR to detect the correct two functional loci from error-free data with (additive and co-dominant) main effects correction and without main effects adjustment are plotted in Figure 3. Here, we observe that the power to identify the correct causal pair is reduced when a main effects correction is performed, with the lowest power levels obtained for codominant correction. The discrepancy between additive and codominant main effects adjustment is particularly pronounced for M27 and p=0.5. For M170 and p=0.5, the nature of the lower-order effects adjustment has virtually no influence on power. Power profiles for different sources of noise, according to main effects adjustment method, are given in Figure S1-i (missing genotypes), Figure S1-ii (genotyping errors) and Figure S1-iii (phenotypic mixture). The empirical power estimates used to generate Figures S1 are also presented in Table S1. Here, drawing conclusions is more subtle, although

generally speaking, empirical power estimates are smaller with co-dominant correction as opposed to additive correction.

#### FIGURE 3: ABOUT HERE

Numerical values on the effect of using different main effects adjustments on the false positive percentage to identify incorrect two-locus models can be derived from Table S2. For error-free data, the false positive percentages after additive main effects correction range from 5% to 39%, p=0.1, 5% to 12%, p=0.25, 3% to 6%, p=0.5 for M170 and from 4% to 7%, p=0.1, 4% to 21%, p=0.25, 9% to 98%, p=0.5 for M27. Using co-dominant coding to adjust for lower order effects, the false positive percentages range from 3% to 6%, p=0.1, 3% to 4%, p=0.25 or p=0.5 for M170 and from 3% to 6%, p=0.1, 3% to 3%, p=0.25 and from 2% to 4%, p=0.5 for M27. In fact, the practice of correcting an MB-MDR epistasis analysis using a codominant main effects model has the tendency to be over-conservative (Table S2).

#### Genetic heterogeneity

#### TABLE 4: ABOUT HERE

So far, we have not yet discussed the performance of MB-MDR for quantitative traits in the presence of genetic heterogeneity. Figure 4 shows empirical power curves to identify true genetic interactions in the presence of GH in a variety of simulation settings. Results are shown for MB-MDR analysis without main effects correction (Figure 4, row 1 for M170 and row 3 for M27) and with main effects correction (additive coding) adjustment (Figure 4, row 2 for M170 and row 4 for M27). As in non-GH settings, power estimates are larger when no correction for main effects is performed than when main effects are accounted for, with generally the most severe power loss observed for codominant main effects correction. However, when the contribution of main effects to the total genetic variance is ignored, false positive percentages rise as well, ranging from 7% to 100% for M27 and from 4% to 97% for

M170. When we adjust for main effects (additive coding), power estimates to identify the first pair (SNP1 x SNP2) drop to less than 50% for both M27 and M170, with the exception of M170. For the latter, and a genetic variance of 0.1, MB-MDR power is estimated to be 95% and 92% for p=0.25 and 0.5, respectively. Under a codominant correction, power estimates drop to less than 7% for both models with the exception of p=0.25 or 0.5 and  $g^2=0.05$  or 0.1. For the latter, power is estimated to be 15% and 26% for M170 and M27, respectively when p=0.1 and  $g^2=0.1$ . For M27, power=31%, p=0.25 and  $g^2=0.1$ . For M170, p=0.25 or 0.5, power estimates are around 30% and 88% for  $g^2=0.05$  and 0.1, respectively. Detailed information about empirical power estimates are given in Table S6.

#### Discussion

Understanding the effects of genes on the development of complex diseases is a major aim of genetic epidemiology. Several studies have indicated that MDR has good power to identify gene-gene interactions in both simulated and real-life data.<sup>9</sup> Although MB-MDR has profiled itself as a promising extension of MDR accommodating study designs that are more complex than unrelated case-control settings<sup>2,3,5,6</sup>, a thorough investigation of its full potential, under a variety of real-life distorting factors, such as missing genotypes, genotyping errors, phenotypic mixtures and last but not least genetic heterogeneity, has never been carried out in the context of quantitative traits. This study has evaluated the power of MB-MDR for quantitative traits and unrelated individuals, in identifying gene-gene interactions for two different epistasis models. Scenarios with and without noisy data, as well as epistasis screening with and without lower-order effects adjustments have been considered. Although our simulations only involved 10 SNPS, conclusions about observed patterns largely remain the same when increasing the number of genetic markers (results not shown). Note that, an increasing number of SNPs, will lead to an increasing number of interacting pairs, resulting in an elevated multiple testing burden, and hence resulting in reduced power. A first important

finding is that MB-MDR adequately deals with one of the most major concerns in genetic association analysis studies (especially those targeting higher-order gene-gene interactions), namely avoiding that the overall type I rate is out of control (Table 2). The apparent slightly conservative results obtained when MB-MDR screening explicitly accounts for lower-order main effects, are not surprising. Indeed, under the general null hypothesis of no genetic association, adjusting for main effects involves over-fitting and hence unnecessary over-correction. However, all the empirical estimates of the MB-MDR type I error rate in Table 2 fall within the interval [0.025,0.075], satisfying Bradley's (1978)<sup>11</sup> liberal criterion of robustness. This criterion requires that the type I error rates are controlled for any level  $\alpha$  of significance, if the empirical type I error rate  $\hat{\alpha}$  is contained in the interval  $0.5\alpha \leq \hat{\alpha} \leq 1.5\alpha$ . Note that we remark that, since MB-MDR assesses global significance using resampling-based maxT adjusted p-values, the FWER will always be weakly controlled at 5%, provided the assumptions of the Westfall and Young approach<sup>7</sup> are attained.

A second important finding is that MB-MDR's power performance under different scenarios can be largely explained by the quantification of the actual genetic variance  $\sigma_{gen}^2$  and by the decomposition of the total genetic variance into contributions of main effects and epistasis, and/or by the decomposition of main effects into additive and dominance variance. Empirical decompositions based on classical variance components analysis of Sham (1998)<sup>12</sup>, are reported in Tables S3 for M170 and S4 for M27 in the absence of GH, and in Tables S5 in the presence of GH. Each of these estimates is based on simulation setting's sample size (750 000 individuals). These results support our theoretically derived variance components, which are summarized in Table 3 (details to be provided elsewhere).

In particular, the observed lowest power performances of non-GH settings for GE10 and PM50 can be explained by the fact that over-representation of the minor allele as well as introducing phenotypic mixture result in a loss of actual genetic variance (Table 1) and

therefore a loss of power. The theoretical results, indicating that a 50% reduction in total genetic variability is established when 50% phenotypic mixture is introduced in error-free data (Table 1) are supported by our empirical results (e.g., Tables S3 for M170 and S4 for M27, comparing  $\sigma_{gen}^2$  with  $g^2$ .

When 50% genetic heterogeneity is present, theory supports our empirical results in that the total actual genetic variance due to the two causal pairs of markers is twice the total actual genetic variance due to a single pair (Tables S5). Moreover, since we have introduced two possible genetic routes for an individual to be genetically predisposed for the trait of interest under GH (route 1 via SNP1xSNP2 or route 2 via SNP3xSNP4), the actual genetic variance in the pooled data will be half the genetic variance in the error-free data (see also Tables S5-ii for M170 and S5-iv for M27). The total genetic variance due to a single causal pair approximates  $g^2/4$  (Table S5-i and S5-iii), which is due to the fact that the 2 pairs have the same minor allele frequencies. Therefore, the theoretical genetic variance is split between the two pairs and thereafter between the 2 SNPs. MB-MDR was shown to be rather robust in the presence of missing genotypes and genotyping error. Note that MB-MDR that MB-MDR handles missing genotypes by using all available cases for the SNP pair under investigation. Hence, no individuals with missing data are a priori removed from the analysis, except when functional SNPs that are adjusted for in regression models have (partially) missing information.

A third finding is that accounting for important lower-order genetic effects in epistasis screening should be made standard. There is a debate about how to best model and test for both main effects and interactions or for interactions only when epistasis is present.<sup>13</sup> Although a fully non-parametric screening approach (e.g., such as MDR) is beautiful in that it does not require specifying particular genetic models, there is still a need to adjust for lower-order genetic effects via a parametric paradigm when targeting significant gene-gene

interaction models. The Model-Based MDR (MB-MDR) offers a flexible framework to make these adjustments. For MDR-like applications other than MB-MDR this is far from obvious. For instance, MDR for binary traits, Ritchie *et al.* (2003)<sup>9</sup> does not accommodate taking corrective measures for lower order effects. Although significant main effects can be filtered out prior to an MDR screening, this happens at the cost of missing out on genuinely true interactions.

Furthermore, examining the decomposition of the total genetic variance has shed more light on the scenarios in which an adjusted MB-MDR analysis is warranted. For instance, when the minor allele frequency of the causal loci is 0.5, model M170 is a pure epistatic model (Table S3: empirical estimates  $\sigma_{epi}^2/\sigma_{gen}^2$  approximate 1). Hence in this scenario the effects of correcting for main effects are taken to the extreme. Clearly, any correction for lower-order effects would be an over-correction. On the other hand, since there is no true evidence for main effects in this model, any adjustment for main effects will only remove a small portion of the variability (Table S3: M170, p=0.5; empirical estimates of  $\sigma_{main}^2/\sigma_{gen}^2$  are close to zero), resulting in false positives for the corrective analysis that are similar to those for the uncorrective analysis (Table S2: M170, p=0.5; empirical estimates close to 5% also when not adjusting for main effects). In effect, the contribution of main effects becomes increasingly important with increasing p for M27 ( $\approx$ 32%, p=0.1,  $\approx$ 61%, p=0.25 and  $\approx$ 85%, p=0.5) and the reverse holds for M170 ( $\approx$ 59%, p=0.1,  $\approx$ 11%, p=0.25 and  $\approx$ 0%, p=0.5) (Table 3, Table S3 and Table S4).

For model M170 and GH scenarios involving p either 0.25 or 0.5 for the causal pairs, the epistatic variance explains a relatively large proportion of the total genetic variance in the data  $(\sigma_{epi}^2/\sigma_{gen}^2 > 87\%;$  Table S5-ii), and correcting for main effects therefore has little effect on power. In contrast, for Model M170 and p=0.1 for the causal pairs, main effects do make an

important contribution to the total genetic variance ( $\sigma_{main}^2/\sigma_{gen}^2 > 57\%$ ; Table S5-i and Table S5-ii) compared to epistasis effects, which translates into a severe empirical power loss and power is dramatically reduced when proper accountancy for lower order effects is being made (Figure 4).

Summarizing, dealing with phenotypic mixtures and genetic heterogeneity will remain challenging for epistasis screening methods, for some time to come. Our empirical results suggest that more work is needed to better accommodate these particularities. Benefits may be gained from identifying the trait-specific factors (genetic or non-genetic) that best characterize mixed phenotypic populations. For genetic heterogeneity, the genes in which the loci are present can be part of different etiological pathways leading to the same disease or be part of the same pathway. According to Heidema et al. (2006)<sup>14</sup>, irrespective of the biological mechanism that gives rise to genetic heterogeneity, the association of the loci with the disease will be reduced if the total sample is used for measuring the association, as was done in this study. A method that is not robust in the presence of genetic heterogeneity will most likely suffer from a decrease in power to detect genetic effects. As our main effects corrective analyses have suggested, a way forward may be to use methods to identify the latent classes and to adapt the epistasis screening accordingly.

Finally, any epistasis screening should properly account for lower-order effects in order to be able to claim that an identified interaction involves a significant epistatic contribution to the total genetic variance.

#### Software

The implementation of MB-MDR used in this manuscript was coded in C++. It is available upon request from the first author (jmahachie@ulg.ac.be).

#### Acknowledgements

J.M. Mahachie John is a doctoral student funded by the Belgian Network BioMAGNet (Bioinformatics and Modeling: from Genomes to Networks), within the Interuniversity Attraction Poles Program (Phase VI/4), initiated by the Belgian State, Science Policy Office. All authors acknowledge research opportunities offered by the Belgian Network BioMAGNetand partial support by the IST Program of the European Community, under the PASCAL2 Network of Excellence (Pattern Analysis, Statistical Modeling and Computational Learning), IST-2007-216886. We also acknowledge the valuable discussions with Tom Cattaert (a Postdoctoral Researcher of the Fonds de la Recherche Scientifique – FNRS) on data generation and variance decomposition sections. In addition, F. Van Lishout acknowledges support by Alma in Silico, funded by the European Commission and Walloon Region through the Interreg IV Program.

#### **Conflicts of interest statement**

The authors declare no conflict of interest.

#### References

- 1. Ritchie MD, Hahn LW, Roodi N *et al*: Multifactor-dimensionality reduction reveals high-order interactions among estrogen-metabolism genes in sporadic breast cancer. *Am J Hum Genet* 2001; **69**: 138-147.
- Calle ML, Urrea V, vellalta G, Malats N, Van Steen K. Model-Based Multifactor Dimensionality Reduction for detecting interactions in high-dimensional genomic data; in: Department of Systems Biology UoV (ed), 2008a. <u>http://www.recercat.net/handle/2072/5001</u>
- 3. Calle ML, Urrea V, Vellalta G, Malats N, Van Steen K: Improving strategies for detecting genetic patterns of disease susceptibility in association studies. *Stat Med* 2008b; **27:** 6532-6546.
- 4. Lou XY, Chen GB, Yan L *et al*: A generalized combinatorial approach for detecting gene-by-gene and gene-by-environment interactions with application to nicotine dependence. *Am J Hum Genet* 2007; **80**: 1125-1137.
- 5. Cattaert T, Urrea V, Naj AC *et al*: FAM-MDR: a flexible family-base multifactor dimensionality reduction technique to detect epistasis using related individuals. *Public Library of Science ONE* 2010.

- 6. Mahachie John JM, Baurecht H, Rodriguez E *et al*: Analysis of the high affinity IgE receptor genes reveals epistatic effects of FCER1A variants on eczema risk. *Allergy* 2010; **65**: 875-882.
- 7. Westfall PH, Young SS: *Resampling-based multiple testing*, New York: Wiley, 1993.
- 8. Evans DM, Marchini J, Morris AP, Cardon LR: Two-Stage Two-Locus Models in Genome-Wide Association. *PLoS Genet* 2006; **2:** e157.
- 9. Ritchie MD, Hahn LW, Moore JH: Power of multifactor dimensionality reduction for detecting gene-gene interactions in the presence of genotyping error, missing data, phenocopy, and genetic heterogeneity. *Genet Epidemiol* 2003; **24**: 150-157.
- 10. Akey JM, Zhang K, Xiong M, Doris P, Jin L: The effect that genotyping errors have on the robustness of common linkage-disequilibrium measures. *Am J Hum Genet* 2001; **68**: 1447-1456.
- 11. Bradley JV: Robustness? *British Journal of Mathematical and Statistical Psychology* 1978; **31:** 144-152.
- 12. Sham P: *Statistics in Human Genetics (Arnold Applications of Statistics Series)* New York Toronto Johnson Wiley & Sons Inc., 1998.
- 13. Verhoeven KJF, Cassela G, McIntyre LM: Epistasis:Obstacle or advantage for mapping complex traits? *PLoS One* in revision.
- 14. Heidema AG, Boer J, Nagelkerke N, Mariman E, van der A D, Feskens E: The challenge for genetic epidemiologists: how to analyze large numbers of SNPs in relation to complex diseases. *BMC Genetics* 2006; **7:** 23.

#### Titles and legends to figures

Figure 1 Graphical presentation of the key steps involved in MB-MDR analysis

**Figure 2** Empirical power estimates of MB-MDR as the percentage of analyses where the correct interaction (SNP1 x SNP2) is significant at the 5% level, for error-free and noise-induced simulation settings. Results are shown for MB-MDR analysis without main effects adjustment and simulated scenarios other than GH

**Figure 3** Empirical power estimates of MB-MDR as the percentage of analyses where the correct interaction (SNP1 x SNP2) is significant at the 5% level, for error-free simulation settings. Legend: no main effects adjustment (—), main effects adjustment via additive coding (...), and co-dominant coding (---)

**Figure 4** Empirical power estimates of MB-MDR as the percentage of analyses where the correct interactions (SNP1 x SNP2) and/or (SNP3 xSNP4) are significant at the 5% level, in the presence GH. First 2 rows: MB-MDR without and with main effects correction for model M170 respectively. Last 2 rows: MB-MDR without and with main effects correction for model M27 respectively. Main effects are corrected for via additive coding. Different definitions for power are adopted: power to identify both interacting pairs SNP1 x SNP2 and SNP3 x SNP4 (cyan); power to identify SNP1 x SNP2 (black); power to identify SNP3 x SNP4 (magenta), power to identify at least one of the interactive pairs (coral)

#### Supplementary Material

**Figure S1-i** Empirical power estimates for MB-MDR as the percentage of analyses where the correct interaction (SNP1 x SNP2) is significant at the 5% level, in the presence of 5% or 10% missing genotypes. Legend: no main effects adjustment (—), main effects adjustment via additive coding (...), and co-dominant coding (---)

**Figure S1-ii** Empirical power estimates for MB-MDR as the percentage of analyses where the correct interaction (SNP1 x SNP2) is significant at the 5% level, in the presence of 5% or 10% genotyping errors. Legend: no main effects adjustment (—), main effects adjustment via additive coding (...),and co-dominant coding (---)

Figure S1-iii Empirical power estimates for MB-MDR as the percentage of analyses where the

correct interaction (SNP1 x SNP2) is significant at the 5% level, in the presence of 25% or 50%

phenotypic mixture. Legend: no main effects adjustment (---), main effects adjustment via

additive coding (...), and co-dominant coding (---)

Table 1 Proportion  $\sigma_{gen}^2 / g^2$  of the total genetic variance in error-free data that is due to genetics in the error-prone data, exhibiting either 5% (GE5) or 10% (GE10) genotyping errors, or 25% (PM25) or 50% (PM50) phenotypic mixture.

Model	р	(	ЭE	PM		
		5%	10%	25%	50%	
	0.1	0.673	0.494	0.563	0.250	
M27	0.25	0.857	0.742	0.563	0.250	
	0.5	0.926	0.858	0.563	0.250	
	0.1	0.667	0.489	0.563	0.250	
M170	0.25	0.701	0.507	0.563	0.250	
	0.5	0.740	0.546	0.563	0.250	

 Table 2 Type I error percentages for data generated under the general null hypothesis of no genetic association in the absence and presence of noise

MG5(MG10) = 5% (10%) missing genotypes and GE5(GE10)=5%(10%) genotyping errors

			Main Eff	ects Correction
р	Noisiness	No Correction	Additive	Codominant
0.1	None	0.055	0.055	0.049
0.25	None	0.051	0.038	0.036
0.5	None	0.054	0.039	0.030
0.1	MG5	0.046	0.039	0.038
0.25	MG5	0.051	0.034	0.036
0.5	MG5	0.052	0.044	0.047
0.1	MG10	0.046	0.041	0.041
0.25	MG10	0.054	0.043	0.043
0.5	MG10	0.048	0.045	0.038
0.1	GE5	0.051	0.037	0.037
0.25	GE5	0.048	0.038	0.036
0.5	GE5	0.038	0.035	0.031
0.1	GE10	0.049	0.037	0.032
0.25	GE10	0.049	0.033	0.031
0.5	GE10	0.054	0.039	0.039

Table 3 Theoretically derived proportions of the genetic variance in error-prone or error-free data due to main effects (additive and dominance) or epistasis. Results are presented for 5% and 10% GE scenarios. "Other" scenarios refer to error-free settings, MG5, MG10, PC25, PC50 and GH50.

			$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$		$\sigma^2_{\it epi}/\sigma^2_{\it gen}$			
Model	р	GE5	GE10	Other	GE5	GE10	Other	
	0.1	0.373	0.420	0.319	0.627	0.580	0.681	
M27	0.25	0.635	0.659	0.609	0.365	0.341	0.391	
	0.5	0.865	0.873	0.857	0.135	0.127	0.143	
	0.1	0.650	0.701	0.581	0.350	0.299	0.419	
M170	0.25	0.139	0.161	0.118	0.861	0.839	0.882	
	0.5	0.000	0.000	0.000	1.000	1.000	1.000	
			$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$			$\sigma^2_{\scriptscriptstyle dom}/\sigma^2_{\scriptscriptstyle main}$		
Model	р	GE5	GE10	Other	GE5	GE10	Other	
	0.1	0.957	0.979	0.947	0.043	0.021	0.053	
M27	0.25	0.865	0.884	0.857	0.135	0.116	0.143	
	0.5	0.680	0.698	0.667	0.320	0.302	0.333	
	0.1	0.837	0.898	0.780	0.163	0.102	0.220	
M170	0.25	0.447	0.502	0.400	0.553	0.498	0.600	
	0.5	0.957	0.979	0.947	0.043	0.021	0.053	

# **APPENDIX:** Supplementary Material

Table S1 Empirical power of MB-MDR to detect the correct two functional loci, excluding
scenarios of simulated genetic heterogeneity

				Model 27			Model 170	
			No	Main Effe	ects Correction	No	Main Effe	ects Correction
р	$g^2$	Noisiness	Correction	Additive	Codominant	Correction	Additive	Codominant
		None	0.446	0.056	0.018	0.542	0.006	0.006
		MG5	0.338	0.018	0.004	0.414	0.014	0.004
		MG10	0.308	0.012	0.004	0.312	0.008	0.000
	0.01	GE5	0.180	0.012	0.002	0.194	0.000	0.000
	-	GE10	0.094	0.000	0.000	0.116	0.000	0.000
	-	PM25	0.152	0.016	0.010	0.180	0.004	0.002
		PM50	0.034	0.000	0.000	0.028	0.000	0.000
		None	0.904	0.264	0.146	0.930	0.144	0.072
		MG5	0.862	0.240	0.108	0.908	0.096	0.046
		MG10	0.784	0.190	0.068	0.800	0.070	0.040
	0.02	GE5	0.620	0.092	0.030	0.666	0.050	0.004
		GE10	0.366	0.018	0.010	0.444	0.006	0.002
		PM25	0.506	0.084	0.018	0.554	0.010	0.010
		PM50	0.134	0.012	0.006	0.126	0.002	0.000
		None	0.996	0.592	0.384	0.996	0.392	0.182
	-	MG5	0.990	0.526	0.272	0.992	0.348	0.182
	-	MG10	0.960	0.412	0.240	0.976	0.244	0.102
0.1	0.03	GE5	0.912	0.232	0.070	0.922	0.136	0.008
		GE10	0.648	0.084	0.060	0.748	0.054	0.002
	-	PM25	0.834	0.210	0.096	0.856	0.094	0.046
	-	PM50	0.286	0.038	0.016	0.284	0.006	0.004
		None	1.000	0.884	0.842	1.000	0.904	0.554
	-	MG5	1.000	0.850	0.774	1.000	0.790	0.442
	-	MG10	1.000	0.772	0.684	1.000	0.680	0.370
	0.05	GE5	0.994	0.598	0.488	1.000	0.488	0.098
	-	GE10	0.940	0.272	0.238	0.970	0.198	0.026
	-	PM25	0.990	0.562	0.346	1.000	0.314	0.168
	-	PM50	0.598	0.150	0.050	0.664	0.034	0.030
		None	1.000	1.000	1.000	1.000	1.000	0.958
	-	MG5	1.000	0.992	0.998	1.000	1.000	0.928
	F	MG10	1.000	0.990	0.994	1.000	0.996	0.928
	0.1	GE5	1.000	0.978	0.978	1.000	0.982	0.638
		GE10	1.000	0.978	0.978	1.000	0.750	0.030
		PM25	1.000	0.904	0.874	1.000	0.942	0.662
	ŀ	PM50	0.950	0.904	0.266	0.986	0.942	0.002
		1 101.50	0.950	0.4/8	0.200	0.980	0.220	0.114

#### **Table S1 Continued**

				Model 27			Model 170	)
			No		ects Correction	No	Main Effe	ects Correction
р	$g^2$	Noisiness	Correction	Additive	Codominant	Correction	Additive	Codominant
		None	0.424	0.038	0.024	0.376	0.300	0.180
		MG5	0.340	0.042	0.030	0.332	0.266	0.154
		MG10	0.284	0.018	0.010	0.248	0.184	0.082
	0.01	GE5	0.298	0.012	0.010	0.232	0.184	0.084
		GE10	0.198	0.018	0.006	0.084	0.070	0.032
		PM25	0.146	0.010	0.000	0.144	0.112	0.048
		PM50	0.024	0.002	0.000	0.026	0.014	0.006
		None	0.912	0.178	0.138	0.908	0.864	0.710
		MG5	0.836	0.150	0.108	0.846	0.780	0.616
		MG10	0.780	0.130	0.104	0.766	0.714	0.532
	0.02	GE5	0.840	0.136	0.088	0.674	0.626	0.440
		GE10	0.696	0.078	0.050	0.428	0.342	0.210
		PM25	0.480	0.046	0.040	0.440	0.384	0.250
		PM50	0.116	0.012	0.006	0.110	0.080	0.040
		None	0.998	0.500	0.444	0.992	0.990	0.952
		MG5	0.984	0.426	0.356	0.992	0.984	0.936
		MG10	0.970	0.306	0.252	0.972	0.960	0.856
0.25	0.03	GE5	0.980	0.310	0.242	0.922	0.886	0.752
		GE10	0.930	0.212	0.130	0.728	0.668	0.484
		PM25	0.796	0.122	0.096	0.788	0.722	0.546
		PM50	0.224	0.016	0.006	0.234	0.182	0.090
		None	1.000	0.878	0.838	1.000	1.000	1.000
		MG5	1.000	0.822	0.784	1.000	1.000	1.000
		MG10	1.000	0.740	0.678	1.000	1.000	0.998
	0.05	GE5	1.000	0.730	0.630	0.998	1.000	0.994
		GE10	1.000	0.546	0.384	0.978	0.964	0.906
		PM25	0.992	0.434	0.362	0.990	0.978	0.934
		PM50	0.580	0.078	0.054	0.588	0.528	0.328
		None	1.000	1.000	1.000	1.000	1.000	1.000
		MG5	1.000	1.000	1.000	1.000	1.000	1.000
		MG10	1.000	0.996	0.996	1.000	1.000	1.000
	0.1	GE5	1.000	1.000	0.992	1.000	1.000	1.000
		GE10	1.000	0.986	0.982	1.000	1.000	1.000
		PM25	1.000	0.960	0.932	1.000	1.000	1.000
		PM50	0.970	0.342	0.288	0.970	0.954	0.878

#### **Table S1 Continued**

				Model 27			Model 170	)
			No	Main Effe	ects Correction	No	Main Effe	ects Correction
р	$g^2$	Noisiness	Correction	Additive	Codominant	Correction	Additive	Codominant
		None	0.386	0.050	0.000	0.332	0.256	0.206
		MG5	0.306	0.034	0.002	0.284	0.206	0.152
		MG10	0.222	0.024	0.000	0.252	0.168	0.124
	0.01	GE5	0.272	0.022	0.000	0.188	0.128	0.096
		GE10	0.260	0.015	0.000	0.116	0.068	0.028
		PM25	0.140	0.020	0.002	0.100	0.058	0.032
		PM50	0.034	0.006	0.000	0.024	0.012	0.008
		None	0.848	0.200	0.000	0.850	0.798	0.770
		MG5	0.780	0.152	0.000	0.792	0.728	0.668
		MG10	0.732	0.110	0.000	0.700	0.632	0.606
	0.02	GE5	0.782	0.132	0.000	0.612	0.540	0.454
		GE10	0.740	0.092	0.000	0.374	0.284	0.212
		PM25	0.400	0.044	0.002	0.426	0.346	0.288
		PM50	0.098	0.012	0.000	0.088	0.052	0.036
		None	0.978	0.460	0.016	0.984	0.988	0.978
		MG5	0.978	0.354	0.004	0.974	0.960	0.948
		MG10	0.926	0.266	0.000	0.942	0.910	0.898
0.5	0.03	GE5	0.966	0.344	0.004	0.898	0.860	0.832
	F	GE10	0.956	0.246	0.002	0.680	0.616	0.516
		PM25	0.704	0.116	0.000	0.780	0.688	0.618
		PM50	0.224	0.020	0.000	0.200	0.138	0.098
		None	1.000	0.832	0.034	1.000	1.000	1.000
		MG5	1.000	0.768	0.026	0.998	0.998	0.998
		MG10	1.000	0.714	0.024	0.998	1.000	1.000
	0.05	GE5	1.000	0.738	0.012	0.998	1.000	0.998
		GE10	0.998	0.658	0.006	0.958	0.948	0.916
		PM25	0.982	0.366	0.008	0.968	0.952	0.950
		PM50	0.514	0.078	0.000	0.466	0.358	0.294
		None	1.000	0.998	0.316	1.000	1.000	1.000
		MG5	1.000	0.998	0.286	1.000	1.000	1.000
		MG10	1.000	0.994	0.168	1.000	1.000	1.000
	0.1	GE5	1.000	0.998	0.266	1.000	1.000	1.000
		GE10	1.000	0.988	0.142	1.000	1.000	1.000
		PM25	1.000	0.902	0.052	1.000	1.000	1.000
		PM50	0.946	0.338	0.004	0.950	0.928	0.912

# Table S2 False positive percentage of analyses with identified significant epistasis models other than the correct two functional interacting loci, in the absence of GH. Estimates exceeding 0.05 are highlighted in bold.

				Model 27			Model 170	)
			No	Main Effe	ects Correction	No	Main Effe	ects Correction
р	g <sup>2</sup>	Noisiness	Correction	Additive	Codominant	Correction	Additive	Codominant
		None	0.148	0.038	0.034	0.278	0.062	0.056
		MG5	0.124	0.046	0.044	0.216	0.046	0.040
		MG10	0.126	0.050	0.044	0.200	0.034	0.032
	0.01	GE5	0.114	0.046	0.044	0.146	0.046	0.044
	-	GE10	0.102	0.048	0.038	0.154	0.058	0.048
	-	PM25	0.082	0.050	0.054	0.142	0.038	0.040
		PM50	0.064	0.046	0.046	0.086	0.044	0.038
		None	0.282	0.048	0.044	0.556	0.054	0.032
		MG5	0.290	0.048	0.038	0.534	0.034	0.020
		MG10	0.242	0.040	0.030	0.470	0.054	0.040
	0.02	GE5	0.224	0.034	0.038	0.374	0.050	0.034
		GE10	0.182	0.038	0.032	0.286	0.028	0.030
		PM25	0.156	0.042	0.038	0.286	0.044	0.044
	-	PM50	0.084	0.044	0.034	0.120	0.052	0.046
		None	0.534	0.042	0.032	0.792	0.092	0.052
	-	MG5	0.436	0.026	0.024	0.766	0.038	0.024
		MG10	0.432	0.046	0.042	0.742	0.088	0.066
0.1	0.03	GE5	0.338	0.058	0.050	0.608	0.052	0.036
		GE10	0.232	0.034	0.030	0.436	0.042	0.032
		PM25	0.240	0.042	0.034	0.434	0.054	0.038
		PM50	0.122	0.044	0.044	0.182	0.050	0.042
		None	0.754	0.036	0.034	0.986	0.160	0.038
	ļ	MG5	0.750	0.022	0.018	0.988	0.100	0.030
	ľ	MG10	0.694	0.048	0.034	0.970	0.096	0.038
	0.05	GE5	0.682	0.054	0.056	0.920	0.062	0.032
	ļ	GE10	0.514	0.034	0.022	0.754	0.076	0.062
	ļ	PM25	0.480	0.050	0.040	0.776	0.080	0.038
	ļ	PM50	0.184	0.048	0.038	0.288	0.060	0.042
		None	0.990	0.072	0.056	1.000	0.394	0.050
		MG5	0.988	0.050	0.030	1.000	0.340	0.030
	ŀ	MG10	0.970	0.066	0.048	1.000	0.302	0.042
	0.1	GE5	0.944	0.068	0.050	1.000	0.172	0.044
	ŀ	GE10	0.884	0.050	0.036	0.996	0.078	0.030
	ŀ	PM25	0.822	0.050	0.036	0.992	0.158	0.038
	ŀ	PM50	0.404	0.034	0.036	0.690	0.060	0.036

## Table S2 Continued

				Model 27			Model 170	
	2		No	Main Effe	ects Correction	No	Main Effe	ects Correction
р	$g^2$	Noisiness	Correction	Additive	Codominant	Correction	Additive	Codominant
		None	0.260	0.044	0.034	0.060	0.050	0.026
		MG5	0.212	0.044	0.036	0.074	0.056	0.048
		MG10	0.218	0.060	0.048	0.076	0.070	0.048
	0.01	GE5	0.250	0.060	0.052	0.088	0.066	0.060
	ſ	GE10	0.210	0.032	0.026	0.052	0.038	0.042
	ſ	PM25	0.158	0.042	0.044	0.060	0.046	0.038
	ſ	PM50	0.082	0.048	0.048	0.072	0.044	0.038
		None	0.556	0.048	0.032	0.108	0.048	0.034
	Ē	MG5	0.522	0.050	0.038	0.092	0.058	0.042
		MG10	0.534	0.050	0.036	0.092	0.058	0.044
	0.02	GE5	0.518	0.066	0.046	0.094	0.052	0.040
		GE10	0.452	0.066	0.056	0.080	0.044	0.032
		PM25	0.268	0.056	0.040	0.076	0.052	0.040
		PM50	0.128	0.040	0.030	0.054	0.048	0.032
		None	0.850	0.064	0.028	0.122	0.056	0.034
		MG5	0.786	0.054	0.028	0.134	0.056	0.042
		MG10	0.728	0.052	0.040	0.110	0.054	0.044
0.25	0.03	GE5	0.738	0.072	0.036	0.106	0.048	0.030
		GE10	0.694	0.050	0.026	0.120	0.056	0.030
		PM25	0.492	0.018	0.018	0.074	0.038	0.028
		PM50	0.198	0.058	0.042	0.064	0.040	0.032
		None	0.992	0.098	0.034	0.242	0.082	0.044
		MG5	0.976	0.078	0.032	0.208	0.062	0.028
		MG10	0.964	0.078	0.038	0.190	0.064	0.034
	0.05	GE5	0.984	0.076	0.024	0.206	0.070	0.034
		GE10	0.934	0.066	0.020	0.176	0.056	0.038
	ſ	PM25	0.786	0.054	0.026	0.132	0.058	0.034
	ľ	PM50	0.354	0.062	0.046	0.086	0.046	0.040
Ī		None	1.000	0.212	0.028	0.528	0.120	0.032
	ſ	MG5	1.000	0.196	0.040	0.502	0.110	0.032
	Ē	MG10	1.000	0.178	0.036	0.462	0.078	0.022
	0.1	GE5	1.000	0.216	0.040	0.412	0.112	0.036
	ſ	GE10	0.998	0.154	0.040	0.358	0.088	0.040
	ľ	PM25	0.992	0.108	0.044	0.286	0.090	0.032
	ľ	PM50	0.748	0.052	0.028	0.122	0.058	0.042

## Table S2 Continued

		ŀ	I	Model 27			Model 170	
	2	Maisian	No	Main Effe	ects Correction	No	Main Effe	ects Correction
р	g <sup>2</sup>	Noisiness	Correction	Additive	Codominant	Correction	Additive	Codominant
		None	0.378	0.088	0.022	0.056	0.046	0.036
		MG5	0.324	0.056	0.022	0.044	0.034	0.024
		MG10	0.308	0.066	0.026	0.058	0.068	0.048
	0.01	GE5	0.362	0.066	0.030	0.058	0.048	0.042
		GE10	0.322	0.077	0.026	0.038	0.022	0.014
		PM25	0.210	0.084	0.036	0.044	0.032	0.034
		PM50	0.112	0.052	0.036	0.046	0.038	0.028
		None	0.772	0.216	0.036	0.074	0.054	0.044
	-	MG5	0.750	0.168	0.026	0.052	0.032	0.030
		MG10	0.692	0.130	0.028	0.044	0.050	0.038
	0.02	GE5	0.744	0.140	0.036	0.032	0.026	0.022
	-	GE10	0.684	0.130	0.038	0.054	0.042	0.032
	-	PM25	0.420	0.100	0.036	0.050	0.044	0.034
	-	PM50	0.160	0.062	0.034	0.064	0.052	0.054
		None	0.954	0.376	0.042	0.044	0.034	0.028
		MG5	0.948	0.284	0.046	0.060	0.042	0.036
		MG10	0.910	0.242	0.054	0.064	0.030	0.028
0.5	0.03	GE5	0.924	0.270	0.038	0.052	0.034	0.030
		GE10	0.912	0.224	0.032	0.052	0.038	0.034
		PM25	0.648	0.154	0.026	0.046	0.038	0.032
		PM50	0.270	0.088	0.044	0.044	0.040	0.030
		None	0.998	0.608	0.028	0.058	0.056	0.040
		MG5	1.000	0.576	0.036	0.042	0.032	0.024
	-	MG10	0.998	0.522	0.042	0.038	0.028	0.036
	0.05	GE5	1.000	0.594	0.042	0.060	0.042	0.030
		GE10	1.000	0.500	0.038	0.034	0.038	0.036
		PM25	0.968	0.314	0.042	0.048	0.042	0.036
	Ī	PM50	0.458	0.124	0.040	0.036	0.032	0.026
ľ		None	1.000	0.978	0.028	0.056	0.038	0.030
	ľ	MG5	1.000	0.956	0.036	0.054	0.046	0.034
	ľ	MG10	1.000	0.928	0.040	0.048	0.044	0.040
	0.1	GE5	1.000	0.982	0.024	0.052	0.040	0.028
	Ī	GE10	1.000	0.912	0.032	0.054	0.038	0.028
	ľ	PM25	1.000	0.732	0.026	0.050	0.036	0.030
	ľ	PM50	0.884	0.288	0.034	0.036	0.028	0.028

р	g <sup>2</sup>	Noisiness	$\sigma^2_{add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{_{epi}}$	$\sigma^2_{\scriptscriptstyle epi}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle gen}$
		None	0.005	0.803	0.006	0.592	0.004	0.408	0.011
		MG5	0.005	0.755	0.006	0.576	0.004	0.424	0.010
		MG10	0.005	0.820	0.006	0.575	0.004	0.425	0.010
	0.01	GE5	0.003	0.828	0.004	0.633	0.002	0.367	0.006
	-	GE10	0.003	0.910	0.003	0.702	0.001	0.298	0.005
	_	PM25	0.003	0.803	0.004	0.601	0.002	0.399	0.006
		PM50	0.001	0.790	0.001	0.566	0.001	0.434	0.002
		None	0.009	0.772	0.012	0.581	0.008	0.419	0.020
	_	MG5	0.009	0.785	0.012	0.593	0.008	0.407	0.020
		MG10	0.009	0.789	0.012	0.581	0.008	0.419	0.020
	0.02	GE5	0.007	0.834	0.008	0.648	0.005	0.352	0.013
		GE10	0.006	0.912	0.007	0.709	0.003	0.291	0.010
		PM25	0.005	0.787	0.006	0.577	0.005	0.423	0.011
		PM50	0.002	0.794	0.003	0.578	0.002	0.422	0.005
		None	0.013	0.772	0.017	0.590	0.012	0.410	0.030
		MG5	0.013	0.772	0.017	0.561	0.013	0.439	0.030
		MG10	0.014	0.787	0.018	0.588	0.013	0.412	0.031
0.1	0.03	GE5	0.011	0.837	0.013	0.651	0.007	0.349	0.020
		GE10	0.009	0.894	0.010	0.682	0.005	0.318	0.015
		PM25	0.007	0.779	0.010	0.572	0.007	0.428	0.017
		PM50	0.003	0.799	0.004	0.576	0.003	0.424	0.008
		None	0.023	0.770	0.030	0.580	0.021	0.420	0.051
		MG5	0.024	0.800	0.030	0.589	0.021	0.411	0.050
		MG10	0.022	0.767	0.029	0.576	0.021	0.424	0.050
	0.05	GE5	0.018	0.841	0.021	0.649	0.011	0.351	0.032
	-	GE10	0.015	0.899	0.016	0.685	0.007	0.315	0.024
	-	PM25	0.013	0.777	0.017	0.592	0.011	0.408	0.028
	-	PM50	0.006	0.786	0.007	0.567	0.005	0.433	0.012
		None	0.045	0.783	0.057	0.583	0.041	0.417	0.098
		MG5	0.046	0.792	0.058	0.582	0.041	0.418	0.099
		MG10	0.047	0.793	0.059	0.593	0.041	0.407	0.100
	0.1	GE5	0.037	0.835	0.044	0.654	0.023	0.346	0.068
		GE10	0.031	0.896	0.034	0.705	0.023	0.295	0.000
		PM25	0.031	0.782	0.034	0.581	0.014	0.419	0.049
		PM50	0.011	0.783	0.014	0.577	0.021	0.423	0.025

Table S3 Empirical variance decomposition for model M170, in the absence of GH. Whereas  $g^2$  represents the total genetic variance corresponding to error-free data,  $\sigma_{gen}^2$  now refers to the empirical total genetic variance.

# Table S3Continued

р	$g^2$	Noisiness	$\sigma^2_{\scriptscriptstyle add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle epi}$	$\sigma^2_{\scriptscriptstyle epi}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle gen}$
		None	0.000	0.429	0.001	0.103	0.009	0.897	0.010
		MG5	0.000	0.460	0.001	0.100	0.009	0.900	0.010
		MG10	0.000	0.329	0.001	0.142	0.009	0.858	0.010
	0.01	GE5	0.000	0.436	0.001	0.135	0.006	0.865	0.007
		GE10	0.000	0.454	0.001	0.166	0.004	0.834	0.005
		PM25	0.000	0.436	0.001	0.113	0.005	0.887	0.006
		PM50	0.000	0.441	0.000	0.126	0.002	0.874	0.002
		None	0.001	0.408	0.002	0.117	0.018	0.883	0.020
		MG5	0.001	0.414	0.002	0.124	0.017	0.876	0.020
		MG10	0.001	0.391	0.003	0.148	0.017	0.852	0.019
	0.02	GE5	0.001	0.473	0.002	0.142	0.012	0.858	0.014
		GE10	0.001	0.556	0.002	0.158	0.009	0.842	0.010
		PM25	0.000	0.395	0.001	0.107	0.010	0.893	0.011
		PM50	0.000	0.361	0.001	0.130	0.005	0.870	0.005
		None	0.001	0.410	0.004	0.121	0.026	0.879	0.030
		MG5	0.001	0.355	0.003	0.113	0.027	0.887	0.030
		MG10	0.001	0.379	0.003	0.117	0.026	0.883	0.030
0.25	0.03	GE5	0.001	0.455	0.003	0.132	0.018	0.868	0.021
		GE10	0.001	0.485	0.003	0.172	0.013	0.828	0.015
		PM25	0.001	0.450	0.002	0.110	0.015	0.890	0.016
		PM50	0.000	0.377	0.001	0.124	0.007	0.876	0.007
		None	0.002	0.414	0.006	0.118	0.044	0.882	0.050
		MG5	0.002	0.389	0.006	0.120	0.044	0.880	0.050
		MG10	0.003	0.452	0.006	0.115	0.045	0.885	0.051
	0.05	GE5	0.002	0.474	0.005	0.147	0.030	0.853	0.035
		GE10	0.002	0.547	0.004	0.160	0.021	0.840	0.025
		PM25	0.001	0.407	0.003	0.124	0.024	0.876	0.028
		PM50	0.001	0.389	0.001	0.118	0.011	0.882	0.013
		None	0.005	0.407	0.011	0.114	0.088	0.886	0.100
		MG5	0.005	0.403	0.011	0.113	0.089	0.887	0.100
		MG10	0.004	0.367	0.011	0.115	0.088	0.885	0.100
	0.1	GE5	0.004	0.454	0.010	0.139	0.060	0.861	0.070
		GE10	0.004	0.508	0.008	0.155	0.044	0.845	0.052
		PM25	0.003	0.400	0.007	0.121	0.049	0.879	0.056
		PM50	0.001	0.366	0.003	0.120	0.022	0.880	0.025

# Table S3 Continued

р	g <sup>2</sup>	Noisiness	$\sigma^2_{\scriptscriptstyle add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle epi}$	$\sigma^2_{\scriptscriptstyle epi}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle gen}$
		None	0.000	0.374	0.000	0.000	0.010	1.000	0.010
		MG5	0.000	0.778	0.000	0.001	0.010	0.999	0.010
		MG10	0.000	0.758	0.000	0.001	0.011	0.999	0.011
	0.01	GE5	0.000	0.795	0.000	0.000	0.007	1.000	0.007
		GE10	0.000	0.591	0.000	0.002	0.005	0.998	0.005
		PM25	0.000	0.363	0.000	0.002	0.006	0.998	0.006
		PM50	0.000	0.898	0.000	0.002	0.002	0.998	0.002
		None	0.000	0.318	0.000	0.001	0.020	0.999	0.020
		MG5	0.000	0.516	0.000	0.000	0.020	1.000	0.020
		MG10	0.000	0.461	0.000	0.001	0.020	0.999	0.020
	0.02	GE5	0.000	0.885	0.000	0.001	0.015	0.999	0.015
		GE10	0.000	0.768	0.000	0.001	0.011	0.999	0.011
		PM25	0.000	0.424	0.000	0.002	0.011	0.998	0.011
		PM50	0.000	0.795	0.000	0.001	0.005	0.999	0.005
	0.03	None	0.000	0.197	0.000	0.000	0.030	1.000	0.030
		MG5	0.000	0.649	0.000	0.000	0.031	1.000	0.031
		MG10	0.000	0.850	0.000	0.000	0.031	1.000	0.031
0.5		GE5	0.000	0.248	0.000	0.000	0.022	1.000	0.022
		GE10	0.000	0.893	0.000	0.001	0.016	0.999	0.016
		PM25	0.000	0.906	0.000	0.000	0.017	1.000	0.017
		PM50	0.000	0.228	0.000	0.001	0.008	0.999	0.008
		None	0.000	0.561	0.000	0.000	0.050	1.000	0.050
		MG5	0.000	0.552	0.000	0.000	0.050	1.000	0.050
		MG10	0.000	0.974	0.000	0.000	0.050	1.000	0.050
	0.05	GE5	0.000	0.851	0.000	0.000	0.037	1.000	0.037
		GE10	0.000	0.852	0.000	0.000	0.027	1.000	0.027
		PM25	0.000	0.899	0.000	0.000	0.027	1.000	0.027
		PM50	0.000	0.675	0.000	0.001	0.012	0.999	0.012
		None	0.000	0.979	0.000	0.000	0.100	1.000	0.100
		MG5	0.000	0.881	0.000	0.000	0.100	1.000	0.100
		MG10	0.000	0.863	0.000	0.000	0.100	1.000	0.100
	0.1	GE5	0.000	0.924	0.000	0.000	0.075	1.000	0.075
		GE10	0.000	0.756	0.000	0.000	0.055	1.000	0.055
		PM25	0.000	0.918	0.000	0.000	0.056	1.000	0.056
		PM50	0.000	0.850	0.000	0.000	0.026	1.000	0.026

Table S4 Empirical variance decomposition for model M27, in the absence of GH. Whereas  $g^2$  represents the total genetic variance corresponding to error-free data,  $\sigma_{gen}^2$  now refers to the empirical total genetic variance.

р	$g^2$	Noisiness	$\sigma^2_{add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle epi}$	$\sigma^2_{\scriptscriptstyle epi}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle gen}$
		None	0.003	0.948	0.003	0.324	0.007	0.676	0.010
		MG5	0.003	0.954	0.003	0.298	0.007	0.702	0.010
		MG10	0.003	0.941	0.003	0.314	0.007	0.686	0.010
	0.01	GE5	0.002	0.964	0.002	0.363	0.004	0.637	0.006
		GE10	0.002	0.984	0.002	0.407	0.003	0.593	0.005
		PM25	0.002	0.947	0.002	0.318	0.004	0.682	0.006
		PM50	0.001	0.937	0.001	0.345	0.002	0.655	0.003
		None	0.006	0.953	0.006	0.309	0.013	0.691	0.020
		MG5	0.006	0.954	0.006	0.322	0.014	0.678	0.020
		MG10	0.006	0.939	0.006	0.308	0.014	0.692	0.021
	0.02	GE5	0.005	0.969	0.005	0.372	0.009	0.628	0.014
		GE10	0.004	0.981	0.004	0.408	0.006	0.592	0.010
		PM25	0.003	0.963	0.004	0.330	0.007	0.670	0.011
		PM50	0.002	0.971	0.002	0.332	0.003	0.668	0.005
	0.03	None	0.009	0.951	0.009	0.320	0.020	0.680	0.030
		MG5	0.009	0.942	0.010	0.313	0.021	0.687	0.031
		MG10	0.009	0.952	0.010	0.321	0.020	0.679	0.030
0.1		GE5	0.007	0.943	0.007	0.368	0.012	0.632	0.020
		GE10	0.006	0.979	0.006	0.408	0.009	0.592	0.015
		PM25	0.005	0.960	0.005	0.322	0.011	0.678	0.017
		PM50	0.002	0.926	0.002	0.308	0.005	0.692	0.007
		None	0.015	0.953	0.016	0.312	0.034	0.688	0.050
		MG5	0.016	0.945	0.017	0.324	0.035	0.676	0.051
		MG10	0.015	0.945	0.017	0.311	0.035	0.689	0.051
	0.05	GE5	0.013	0.953	0.013	0.372	0.033	0.628	0.034
		GE10	0.012	0.985	0.010	0.419	0.021	0.581	0.025
		PM25	0.010	0.938	0.009	0.313	0.014	0.687	0.023
		PM50	0.008	0.938	0.009	0.313	0.009	0.681	0.023
		None	0.004	0.932					
		MG5	0.030	0.945	0.031 0.033	0.314 0.326	0.068	0.686 0.674	0.100 0.101
		MG10	0.031	0.940	0.033	0.320	0.000	0.684	0.101
	0.1	GE5	0.030	0.942	0.032	0.310	0.070	0.634	0.102
	0.1	GE10	0.024	0.933	0.023	0.300	0.045	0.634	0.008
		PM25							
		PM50	0.016	0.946	0.017	0.314	0.038	0.686	0.055
		FINIOU	0.007	0.941	0.008	0.313	0.017	0.687	0.025

# Table S4 Continued

р	g <sup>2</sup>	Noisiness	$\sigma^2_{add}$	$\sigma_{add}^2/\sigma_{main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{epi}$	$\sigma_{epi}^2/\sigma_{gen}^2$	$\sigma^2_{gen}$
	U		<i>U<sub>add</sub></i>	$O_{add} / O_{main}$	0 <sub>main</sub>	O <sub>main</sub> /O <sub>gen</sub>	0 <sub>epi</sub>	O <sub>epi</sub> / O <sub>gen</sub>	0 gen
		None	0.005	0.856	0.006	0.605	0.004	0.395	0.010
		MG5	0.005	0.855	0.006	0.630	0.004	0.370	0.010
		MG10	0.005	0.845	0.006	0.624	0.004	0.376	0.010
	0.01	GE5	0.005	0.865	0.006	0.649	0.003	0.351	0.009
		GE10	0.004	0.900	0.005	0.667	0.002	0.333	0.007
		PM25	0.003	0.867	0.004	0.623	0.002	0.377	0.006
		PM50	0.001	0.861	0.001	0.571	0.001	0.429	0.003
		None	0.010	0.865	0.012	0.617	0.008	0.383	0.020
		MG5	0.011	0.869	0.012	0.625	0.007	0.375	0.020
		MG10	0.010	0.851	0.012	0.613	0.007	0.387	0.019
	0.02	GE5	0.010	0.856	0.011	0.637	0.006	0.363	0.018
		GE10	0.009	0.892	0.010	0.658	0.005	0.342	0.015
		PM25	0.006	0.859	0.007	0.624	0.004	0.376	0.011
		PM50	0.003	0.871	0.003	0.628	0.002	0.372	0.005
	0.03	None	0.016	0.852	0.019	0.609	0.012	0.391	0.031
		MG5	0.016	0.864	0.019	0.610	0.012	0.390	0.030
		MG10	0.015	0.861	0.018	0.606	0.012	0.394	0.030
0.25		GE5	0.014	0.870	0.017	0.642	0.009	0.358	0.026
		GE10	0.013	0.885	0.015	0.652	0.008	0.348	0.022
		PM25	0.009	0.864	0.010	0.610	0.007	0.390	0.017
		PM50	0.004	0.853	0.005	0.623	0.003	0.377	0.007
		None	0.026	0.855	0.031	0.609	0.020	0.391	0.050
		MG5	0.026	0.855	0.030	0.605	0.020	0.395	0.050
		MG10	0.026	0.851	0.030	0.611	0.019	0.389	0.050
	0.05	GE5	0.024	0.864	0.028	0.641	0.015	0.359	0.043
		GE10	0.022	0.873	0.025	0.666	0.012	0.334	0.037
		PM25	0.015	0.864	0.017	0.605	0.011	0.395	0.028
		PM50	0.007	0.856	0.008	0.618	0.005	0.382	0.012
		None	0.052	0.861	0.060	0.609	0.039	0.391	0.099
		MG5	0.053	0.862	0.062	0.609	0.040	0.391	0.102
		MG10	0.052	0.853	0.061	0.611	0.039	0.389	0.100
	0.1	GE5	0.046	0.855	0.054	0.632	0.031	0.368	0.085
		GE10	0.044	0.885	0.049	0.656	0.026	0.344	0.075
		PM25	0.029	0.856	0.033	0.599	0.022	0.401	0.056
		PM50	0.013	0.858	0.016	0.612	0.010	0.388	0.025

### **Table S4 Continued**

р	g <sup>2</sup>	Noisiness	$\sigma^2_{\scriptscriptstyle add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{\scriptscriptstyle epi}$	$\sigma^2_{\scriptscriptstyle epi}/\sigma^2_{\scriptscriptstyle gen}$	$\sigma^2_{_{gen}}$
		None	0.006	0.667	0.009	0.844	0.002	0.156	0.010
		MG5	0.006	0.698	0.009	0.845	0.002	0.155	0.010
		MG10	0.006	0.662	0.009	0.854	0.001	0.146	0.010
	0.01	GE5	0.005	0.693	0.008	0.874	0.001	0.126	0.009
		GE10	0.005	0.705	0.008	0.875	0.001	0.125	0.009
		PM25	0.003	0.653	0.005	0.851	0.001	0.149	0.006
		PM50	0.001	0.708	0.002	0.857	0.000	0.143	0.002
		None	0.012	0.675	0.017	0.852	0.003	0.148	0.020
		MG5	0.011	0.655	0.017	0.863	0.003	0.137	0.020
		MG10	0.011	0.660	0.017	0.846	0.003	0.154	0.020
	0.02	GE5	0.011	0.697	0.016	0.859	0.003	0.141	0.018
		GE10	0.010	0.702	0.015	0.877	0.002	0.123	0.017
		PM25	0.006	0.667	0.010	0.850	0.002	0.150	0.011
		PM50	0.003	0.677	0.004	0.859	0.001	0.141	0.005
	0.03	None	0.017	0.664	0.026	0.855	0.004	0.145	0.030
		MG5	0.017	0.680	0.026	0.865	0.004	0.135	0.030
		MG10	0.017	0.679	0.025	0.856	0.004	0.144	0.030
0.5		GE5	0.016	0.683	0.024	0.870	0.004	0.130	0.027
		GE10	0.016	0.717	0.022	0.870	0.003	0.130	0.026
		PM25	0.009	0.661	0.014	0.864	0.002	0.136	0.016
		PM50	0.004	0.646	0.007	0.866	0.001	0.134	0.008
		None	0.028	0.669	0.042	0.854	0.007	0.146	0.049
		MG5	0.029	0.669	0.043	0.858	0.007	0.142	0.050
		MG10	0.028	0.659	0.042	0.851	0.007	0.149	0.050
	0.05	GE5	0.027	0.669	0.040	0.871	0.006	0.129	0.046
		GE10	0.026	0.693	0.038	0.876	0.005	0.124	0.043
		PM25	0.016	0.678	0.024	0.858	0.004	0.142	0.028
		PM50	0.007	0.659	0.011	0.856	0.002	0.144	0.013
		None	0.058	0.672	0.086	0.862	0.014	0.138	0.100
		MG5	0.057	0.669	0.085	0.855	0.014	0.145	0.099
		MG10	0.057	0.667	0.085	0.856	0.014	0.144	0.100
	0.1	GE5	0.055	0.682	0.080	0.864	0.013	0.136	0.093
		GE10	0.052	0.701	0.074	0.875	0.011	0.125	0.085
		PM25	0.032	0.663	0.049	0.861	0.008	0.139	0.056
		PM50	0.014	0.665	0.021	0.855	0.004	0.145	0.025

Table S5-i Empirical variance decomposition of the total genetic variance due to (SNP1,SNP2) in model M170, for several GH settings.

Whereas  $g^2$  represents the total genetic variance corresponding to error-free data,  $\sigma_{gen,pair}^2$  now refers to the empirical total genetic variance due to a single interactive pair.

$p_1$	<b>p</b> <sub>2</sub>	g <sup>2</sup>	$\sigma^2_{\scriptscriptstyle add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen,pair}$	$\sigma^2_{\scriptscriptstyle epi}$	$\sigma^2_{\scriptscriptstyle epi}/\sigma^2_{\scriptscriptstyle gen,pair}$	$\sigma^2_{\scriptscriptstyle gen, pair}$
		0.01	0.001	0.789	0.001	0.552	0.001	0.448	0.003
		0.02	0.002	0.789	0.003	0.562	0.002	0.438	0.005
0.1	0.1	0.03	0.004	0.793	0.005	0.596	0.003	0.404	0.008
		0.05	0.006	0.770	0.008	0.604	0.005	0.397	0.013
		0.1	0.012	0.782	0.015	0.582	0.012	0.418	0.026
		0.01	0.000	0.436	0.000	0.133	0.002	0.867	0.003
		0.02	0.000	0.329	0.001	0.112	0.005	0.888	0.005
0.25	0.25	0.03	0.000	0.382	0.001	0.117	0.007	0.883	0.008
		0.05	0.000	0.398	0.001	0.119	0.011	0.881	0.012
		0.1	0.001	0.393	0.003	0.109	0.022	0.891	0.025
		0.01	0.000	0.527	0.000	0.001	0.003	0.999	0.003
		0.02	0.000	0.490	0.000	0.001	0.005	0.999	0.005
0.5	0.5	0.03	0.000	0.764	0.000	0.000	0.007	1.000	0.007
		0.05	0.000	0.770	0.000	0.002	0.013	1.000	0.013
		0.1	0.000	0.991	0.000	0.000	0.025	1.000	0.025

Table S5-ii Empirical variance decomposition of the total genetic variance due to (SNP1,SNP2) and (SNP3,SNP4) in model M170, for several GH settings.

Whereas  $g^2$  represents the total genetic variance corresponding to error-free data,  $\sigma_{gen,2pairs}^2$  now refers to the empirical total genetic variance due the two causal marker pairs.

$p_1$	<b>p</b> <sub>2</sub>	g <sup>2</sup>	$\sigma^2_{add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{main}/\sigma^2_{gen,2pairs}$	$\sigma^2_{\scriptscriptstyle epi}$	$\sigma_{\scriptscriptstyle epi}^{\scriptscriptstyle 2}/\sigma_{\scriptscriptstyle gen,2pairs}^{\scriptscriptstyle 2}$	$\sigma^2_{{\it gen},2{\it pairs}}$
		0.01	0.002	0.770	0.003	0.568	0.002	0.432	0.005
		0.02	0.004	0.789	0.006	0.569	0.004	0.431	0.010
0.1	0.1	0.03	0.007	0.795	0.009	0.594	0.006	0.407	0.015
		0.05	0.012	0.771	0.015	0.593	0.011	0.408	0.026
		0.1	0.023	0.781	0.029	0.583	0.021	0.417	0.050
		0.01	0.000	0.405	0.001	0.132	0.004	0.868	0.005
		0.02	0.000	0.386	0.001	0.117	0.009	0.883	0.010
0.25	0.25	0.03	0.001	0.360	0.002	0.119	0.013	0.881	0.015
		0.05	0.001	0.416	0.003	0.117	0.022	0.883	0.025
		0.1	0.002	0.396	0.006	0.112	0.044	0.888	0.050
		0.01	0.000	0.794	0.000	0.002	0.005	0.998	0.005
		0.02	0.000	0.719	0.000	0.001	0.010	0.999	0.010
0.5	0.5	0.03	0.000	0.603	0.000	0.000	0.014	1.000	0.014
		0.05	0.000	0.694	0.000	0.001	0.026	0.999	0.026
		0.1	0.000	0.777	0.000	0.000	0.050	1.000	0.050

Table S5-iii Empirical variance decomposition of the total genetic variance due to (SNP1,SNP2) in model M27, for several GH settings.

Whereas g <sup>2</sup> represents the total genetic variance corresponding to error-free data, $\sigma_{gen, pair}^2$ now
refers to the empirical total genetic variance due to a single interactive pair.

<b>p</b> 1	<b>p</b> <sub>2</sub>	$g^2$	$\sigma^2_{\scriptscriptstyle add}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen,pair}$	$\sigma^2_{_{epi}}$	$\sigma_{\scriptscriptstyle epi}^2/\sigma_{\scriptscriptstyle gen,pair}^2$	$\sigma^2_{_{gen,pair}}$
		0.01	0.001	0.956	0.001	0.320	0.002	0.680	0.003
		0.02	0.001	0.947	0.002	0.287	0.004	0.713	0.005
0.1	0.1	0.03	0.002	0.934	0.002	0.293	0.006	0.707	0.008
		0.05	0.004	0.955	0.004	0.338	0.008	0.662	0.013
		0.1	0.008	0.940	0.008	0.324	0.017	0.676	0.025
		0.01	0.001	0.853	0.002	0.557	0.001	0.443	0.003
	0.25	0.02	0.003	0.859	0.003	0.607	0.002	0.393	0.005
0.25		0.03	0.004	0.856	0.005	0.615	0.003	0.386	0.007
		0.05	0.006	0.857	0.007	0.605	0.005	0.395	0.012
		0.1	0.013	0.845	0.016	0.604	0.010	0.396	0.026
		0.01	0.001	0.669	0.002	0.849	0.000	0.151	0.003
		0.02	0.003	0.671	0.004	0.868	0.001	0.132	0.005
0.5	0.5	0.03	0.004	0.645	0.007	0.869	0.001	0.131	0.008
		0.05	0.007	0.662	0.011	0.863	0.002	0.137	0.013
		0.1	0.014	0.667	0.021	0.855	0.004	0.145	0.025

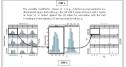
Table S5-iv Empirical variance decomposition of the total genetic variance due to (SNP1,SNP2) and (SNP3,SNP4) in model M27, for several GH settings.

Whereas g <sup>2</sup> represents the total genetic variance corresponding to error-free data, $\sigma_{gen,2pairs}^2$ now	
refers to the empirical total genetic variance due the two causal marker pairs.	

<b>p</b> 1	<b>p</b> <sub>2</sub>	$g^2$	$\sigma^2_{_{add}}$	$\sigma_{\scriptscriptstyle add}^2/\sigma_{\scriptscriptstyle main}^2$	$\sigma^2_{\scriptscriptstyle main}$	$\sigma^2_{\scriptscriptstyle main}/\sigma^2_{\scriptscriptstyle gen,2pairs}$	$\sigma^2_{_{epi}}$	$\sigma_{\scriptscriptstyle epi}^{\scriptscriptstyle 2}/\sigma_{\scriptscriptstyle gen,2pairs}^{\scriptscriptstyle 2}$	$\sigma^2_{{\scriptscriptstyle gen},2{\scriptscriptstyle pairs}}$
		0.01	0.002	0.953	0.002	0.314	0.004	0.686	0.005
		0.02	0.003	0.953	0.003	0.307	0.007	0.693	0.010
0.1	0.1	0.03	0.004	0.939	0.005	0.307	0.011	0.693	0.015
		0.05	0.008	0.948	0.008	0.322	0.017	0.678	0.025
		0.1	0.015	0.935	0.016	0.323	0.034	0.677	0.050
		0.01	0.003	0.868	0.003	0.603	0.002	0.398	0.005
		0.02	0.005	0.861	0.006	0.625	0.004	0.375	0.010
0.25	0.25	0.03	0.008	0.856	0.009	0.609	0.006	0.391	0.015
		0.05	0.013	0.858	0.015	0.610	0.010	0.391	0.025
		0.1	0.026	0.847	0.031	0.607	0.020	0.393	0.050
		0.01	0.003	0.675	0.004	0.851	0.001	0.149	0.005
		0.02	0.006	0.666	0.008	0.845	0.002	0.155	0.010
0.5	0.5	0.03	0.009	0.656	0.013	0.867	0.002	0.133	0.015
		0.05	0.014	0.659	0.022	0.858	0.004	0.142	0.025
		0.1	0.028	0.665	0.042	0.852	0.007	0.149	0.050

Table S6 Power of MB-MDR to detect different power definitions from genetic heterogeneity settings (Pair1=SNP1 x SNP2, Pair2=SNP3 x SNP4), and false positives, FP (any pair that is not strictly a functional pair). Considered analyses methods are MB-MDR without main effects correction, with additive or codominant lower order effects adjustment. False positive percentages higher than 0.05 are highlighted in bold.

	l		er unu			Model M2				I	Model M17	0		
						Pair1	Pair 1				Pair1	Pair1		
	$p_1$	<b>p</b> <sub>2</sub>	$g^2$	Pair1	Pair2	or	and	FP	Pair1	Pair2	or	and	FP	
						Pair2	Pair2				Pair2	Pair2		
			0.01	0.014	0.022	0.036	0.000	0.074	0.018	0.022	0.040	0.000	0.110	
			0.02	0.102	0.144	0.234	0.012	0.130	0.138	0.108	0.224	0.022	0.172	
Ę	0.1	0.1	0.03	0.296	0.306	0.508	0.094	0.212	0.294	0.298	0.512	0.080	0.380	
ctio			0.05	0.650	0.572	0.852	0.370	0.336	0.690	0.688	0.906	0.472	0.688	
No Main Effects Correction			0.1	0.936	0.958	0.994	0.900	0.742	0.976	0.980	1.000	0.956	0.968	
			0.01	0.020	0.016	0.036	0.000	0.116	0.016	0.020	0.036	0.000	0.072	
cts	0.05	0.05	0.02	0.100	0.094	0.186	0.008	0.200	0.096	0.078	0.168	0.006	0.076	
ffe	0.25	0.25	0.03	0.226	0.266	0.442	0.050	0.368	0.224	0.196	0.378	0.042	0.066	
пE			0.05	0.564	0.596	0.830	0.330	0.628	0.510	0.546	0.792	0.264	0.112	
<b>Jai</b>			0.1	0.980	0.964	1.000	0.944	0.962	0.960	0.972	1.000	0.932	0.202	
0 N			0.01	0.020	0.044	0.064	0.000	0.148	0.016	0.022	0.038	0.000	0.052	
Z			0.02	0.070	0.072	0.134	0.008	0.278	0.074	0.072	0.138	0.008	0.062	
	0.5	0.5	0.03	0.206	0.174	0.332	0.048	0.520	0.162	0.172	0.306	0.028	0.054	
			0.05	0.490	0.488	0.750	0.228	0.810	0.494	0.430	0.710	0.214	0.036	
			0.1	0.938	0.958	0.996	0.900	1.000	0.954	0.918	1.000	0.872	0.080	
			0.01	0.002	0.002	0.004	0.000	0.024	0.002	0.000	0.002	0.000	0.030	
Ę	0.1	0.1	0.1	0.02	0.006	0.006	0.012	0.000	0.036	0.002	0.002	0.004	0.000	0.038
ctio	0.1		0.03	0.040	0.024	0.064	0.000	0.036	0.002	0.012	0.014	0.000	0.036	
ITE			0.05	0.096	0.106	0.192	0.010	0.014	0.030	0.034	0.062	0.002	0.060	
C			0.1	0.476	0.482	0.718	0.240	0.058	0.280	0.286	0.494	0.072	0.128	
cts		0.25	0.01	0.000	0.000	0.000	0.000	0.030	0.012	0.006	0.018	0.000	0.042	
ffe	0.25		0.02	0.008	0.004	0.012	0.000	0.028	0.078	0.040	0.116	0.002	0.048	
п Н	0.25		0.03	0.012	0.016	0.028	0.000	0.044	0.152	0.168	0.284	0.036	0.036	
Aai			0.05	0.056	0.066	0.118	0.004	0.042	0.438	0.496	0.740	0.194	0.028	
Additive Main Effects Correction			0.1	0.348	0.346	0.586	0.108	0.074	0.954	0.962	0.998	0.918	0.056	
litiv			0.01	0.000	0.004	0.004	0.000	0.054	0.006	0.006	0.012	0.000	0.030	
Add	0.5	0.5	0.02	0.008	0.008	0.016	0.000	0.064	0.044	0.038	0.076	0.006	0.042	
4	0.5	0.5	0.03	0.016	0.014	0.030	0.000	0.118	0.108	0.112	0.204	0.016	0.036	
			0.05	0.066	0.048	0.110	0.004	0.180	0.430	0.336	0.606	0.160	0.026	
			0.1	0.312	0.356	0.572	0.096	0.500	0.920	0.898	0.994	0.824	0.054	
			0.01	0.000	0.000	0.000	0.000	0.016	0.000	0.000	0.000	0.000	0.024	
tior	0.1	0.1	0.02			0.004	0.000			0.000	0.002	0.000	0.026	
reci	0.1	0.1	0.03	0.016	0.008	0.024	0.000	0.024	0.004	0.008	0.012	0.000	0.032	
Cor			0.05	0.036	0.042	0.078	0.000	0.010	0.024	0.030	0.054	0.000	0.028	
ts (			0.1	0.260	0.264	0.454	0.070	0.028	0.154	0.162	0.294	0.022	0.028	
fec			0.01	0.000	0.000	0.000	0.000	0.014	0.000	0.000	0.000	0.000	0.034	
Ef	0.25	0.25	0.02	0.006	0.006	0.012	0.000	0.018	0.036	0.030	0.066	0.000	0.038	
lain	0.23	0.23	0.03	0.008	0.012	0.020	0.000	0.030	0.072	0.070	0.136	0.006	0.022	
Codominant Main Effects Correction			0.05	0.044	0.032	0.072	0.004	0.026	0.296	0.316	0.540	0.072	0.018	
lan			0.1	0.310	0.280	0.514	0.076	0.028	0.874	0.854	0.976	0.752	0.014	
mir			0.01	0.000	0.000	0.000	0.000	0.024	0.002	0.006	0.008	0.000	0.022	
iope	0.5	0.5	0.02	0.000	0.000	0.000	0.000	0.018	0.024	0.016	0.038	0.002	0.028	
Cc	0.5	0.5	0.03	0.000	0.000	0.000	0.000	0.014	0.078	0.064	0.132	0.010	0.032	
			0.05	0.002	0.000	0.002	0.000	0.018	0.324	0.296	0.526	0.094	0.014	
			0.1	0.010	0.014	0.024	0.000	0.018	0.894	0.876	0.992	0.778	0.038	



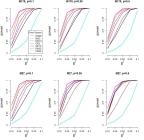
Highlies, Lew (@) and Ho Evidence (Mark) labeling to determined by importing the direction of the cellstate assistant for sensition and its significance, using a liberal thresholding, 0.13; High init cellstars inform, and no evidence rath are proted and Statel. For sensitivity mith X numbing this single test statistic Wire for backset/facture.



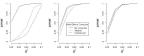
Another set of Africtura's chosen and steps 1-2 are repeated until no more factors need to be investigated.



Accessing joint significance of k-factor models, hardso correcting for multiple testing



M170, p=0.1



1227 am2 1

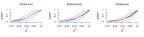
M27. p=0.28

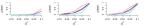
1427, pm0.5



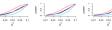




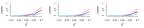






















1627\_1005.p=0.1







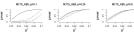
M27\_M010, p=0.1

0.05 8.1

M27\_M310.p=0.25













M27\_085.p=0.1







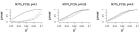
M27.0E10, p=0.1

0.05 8.1

M27\_GE10, p=0.25



















M27\_PC25, p=0.1













