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Toxicity to Bivalve Hemocytes of Pathogenic *Vibrio* Cytoplasmic Extract

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Abstract: Using a chemiluminescence (CL) test, it had been previously demonstrated that *Vibrio pectenicida*, which is pathogenic to *Pecten maximus* larvae, was able to inhibit completely the CL activity of *P. maximus* hemocytes and partially inhibit those of *Crassostrea gigas*. Conversely, a *Vibrio* sp. strain, S322, pathogenic to *C. gigas* larvae was more active in reducing the CL activity of oyster hemocytes than of scallop hemocytes. Using this same CL biotest, *V. pectenicida* and S322 cytoplasmic extracts were shown to reproduce CL inhibition while the cytoplasmic extract of a nonpathogenic strain (U1, *Pseudoalteromonas*) was without effect. Moreover, cytoplasmic extract as well as live *V. pectenicida* cells provoked, within a few hours, death of *P. maximus* hemocytes adhering to a glass slide. After partial purification, it was shown that toxic activities of *V. pectenicida* cytoplasmic extract was due to a toxin, named VHKT (for vibrio hemocyte-killer toxin), which is heat stable, acid and protease resistant, and less than 3 kDa in molecular weight. Attempts to purify VHKT by reverse-phase (C18) HPLC separated activity into the fraction eluted by water at a retention time of 4.02 min.

Key Words: *Vibrio pectenicida*; *Pecten maximus*; *Crassostrea gigas*; chemiluminescence; toxin; hemocyte; bivalves.

INTRODUCTION

Previously, Lambert and Nicolas (1998) demonstrated specific inhibition of the chemiluminescence (CL) activity of bivalve hemocytes by pathogenic vibrios. *Vibrio pectenicida* (strain A496) virulent to *Pecten maximus* larvae (Nicolas *et al.*, 1996; Lambert *et al.*, 1998) completely suppressed the CL activity of scallop hemocytes but only partially suppressed the activity of *Crassostrea gigas* hemocytes. With another *Vibrio* sp. (S322), pathogenic to oysters, the opposite results were obtained. The hypothesis proposed to explain this specificity was that it was due to specific attachment by pili or flagella to the hemocyte membrane or possibly due to different toxins.

Toxins of vibrios involved in human disease, including *V. cholerae*, *V. vulnificus*, and *V. parahaemolyticus*, have been studied extensively. Some toxins from vibrios which are virulent to fish, such as *V. anguillarum*, have also been characterized. For bivalve larvae and other molluscs, few results have been reported. The activity of toxins was demonstrated by exposure of larvae to diluted bacterial culture supernatants (Umbreit and Tripp, 1975; Brown and Roland, 1984). Nottage and Birkbeck (1986) purified a ciliostatic toxin from *V. alginolyticus* with a low molecular weight. In the case of *V. pectenicida* (Lambert, 1998), inoculation of cytoplasmic extract to axenic culture of *P. maximus* larvae caused as high mortality as live bacteria.

In the present work, using the CL test described by Lambert and Nicolas (1998), we attempted to demonstrate the presence of a toxic molecule in the bacterial cytoplasm of *V. pectenicida* and *Vibrio* sp. strain S322 and to further purify and characterize the *V. pectenicida* toxin.

MATERIALS AND METHODS

Bivalves

Thirty adult scallops and oysters, 3 or 4 years old, were collected in the Bay of Brest (Brittany). Scallops were maintained at 15°C in a 450-L seawater tank equipped with a sand filter and air-lift system. They were fed just close to satiation on a four-microalgae mixture which included *Isochrysis affinis galbana* (1 to 2.6 × 10⁹ cells/day/animal), *Pavlova lutheri* (1 to 2.6 × 10⁹ cells/day/animal), *Chaetoceros calcitrans* (0.5 to 1.3 × 10⁹ cells/day/animal), and *Skeletonema costatum* (0.3 to 0.5 × 10⁹ cells/day/animal). For *C. gigas*, the same

conditions were applied, except for the temperature (18°C) and the diet (1.1 to 2.9 × 10⁹ cells/day/animal). They were maintained, for at least 2 weeks, under constant conditions of temperature and feeding before sampling.

Inoculated Bacteria

Three strains were used: strain A496 of *V. pectenicida* (CIP 105229), strain S322, pathogenic to oyster larvae (*C. gigas*), and strain U1, collected from the dominant flora of *P. maximus* larval cultures. The affiliations of both U1 and S322 were obtained by sequencing the 16S rDNA and by phylogenetic analysis. Strain S322 was close to *V. orientalis*, whereas U1 was located inside the *Pseudoalteromonas* genus. Before inoculation, the three strains were cultured in Marine broth (Difco) at 18°C, without shaking, for 48 h, which corresponded to the end of the exponential phase. The bacterial cells were collected by centrifugation (5000g, 10 min), washed twice in sterile seawater (SSW), and resuspended in SSW. The concentration was checked by measurement of optical density (OD) at 540 nm, for which the correlation between direct counts and OD had previously been established (Lambert, 1998).

Bacterial Extracts

Extracts were obtained from the three bacterial cultures. Strain A496 was cultured first in Marine broth 2216E (Difco) in a 250-mL conical flask. After 48 h growth, the culture was transferred to a 15-L fermentor using growth conditions of 25°C, pH 7.8, and 100% oxygen saturation. Bacterial concentration was checked by regular measurement of OD at 540 nm. The stationary phase was reached after 26 h, when 41 g of bacterial cells, about 1.5 × 10¹³ cells, was collected using continuous centrifugation. Cells were suspended in 100 mL of sterile 1% (w/v) NaCl and then disrupted by a double high-pressure treatment (18,000 psi) in a French Press system. The bacterial suspension was then centrifuged (100,000g, 40 min) to separate membrane and cytoplasmic fractions. The supernatant (cytoplasmic extract) was carefully removed, 0.2 µm filtered (Minisart SRP 25 filter, Sartorius), divided into 1.6-mL volumes in Eppendorf tubes, and frozen at -80°C.

The same protocol was used for strains U1 and S322, but they were cultured only in 200-mL flasks. About 2.3 × 10¹¹ cells for U1 and 7.2 × 10¹⁰ cells for S322 were collected from the cultures.

The yield of the French Press system was evaluated by protein measurement (Lowry method: Waterborg and Matthews, 1984) before and after treatment. The result was in agreement (72%) with those (75 to 100%) given by Hancock and Poxton (1988). Taking account of these assays, 1 mL of cytoplasmic extract corresponded to 1.1 × 10⁸ cells for A496, 0.25 × 10⁸ cells for S322, and 0.32 × 10⁸ for U1.

CL Assays

Hemolymph collection in scallop and oyster was as described previously by Lambert and Nicolas (1998). The protocol for CL assays was first described by Bache`re *et al.* (1991) and adapted by Lambert and Nicolas (1998). Briefly, the hemolymph, diluted with modified anti-aggregant Alsever solution (MAS, Bache`re *et al.*, 1988), was distributed into plastic scintillation vials at 1 × 10⁵ hemocytes per vial. This solution was made up to 2 mL with luminol solution (200 mL) and SSW.

Generation of chemiluminescence was measured with a liquid scintillation counter set to measure repeated sequential counts. Results were expressed in counts per minute (cpm). At the end of the experiment, the CL activity levels in control hemocyte samples after stimulation with zymosan particles, 80 particles per hemocyte (the optimal concentration previously determined by Bachère *et al.*, 1991), were compared to the CL activity levels of hemocytes previously in contact for 80 min with bacteria or bacterial extract. Each set of hemocytes was made up of four replicates, i.e., four vials. In each experiment, one set of four vials without hemocytes was also used to determine the “zero level” and one set, with hemocytes but not stimulated by zymosan and not exposed to bacteria or bacterial extracts, was used to determine the “base level.” Significant differences between the CL responses of treated groups were determined at the peak of CL activity due to zymosan stimulation using variance analysis ($n = 4$, $\alpha = 0.05$).

Five CL assays were carried out to determine the effect of A496, S322, and U1 cytoplasmic extracts and then six CL assays to identify the features of the toxic molecule.

CL Assays Using Cytoplasmic Extracts of A496, U1 and S322

A first CL experiment was performed using live cells of A496 and A496 cytoplasmic extracts. In addition to control, zero, and base sets of hemocytes, one set of four vials of *P. maximus* hemocytes was inoculated with 100 cells of *V. pectenicida* (strain A496) per hemocyte, the optimal ratio previously determined by Lambert and Nicolas (1998), and four sets, respectively, were inoculated with 0.8, 4, 8, and 16 μL of A496 bacterial extract.

A second experiment was performed to compare the activity of 25 μL of A496 cytoplasmic extract to that of 83 μL of U1 cytoplasmic extract, which corresponded to the same quantity of bacterial cells (about 2.4×10^9) as 25 μL of A496 extract. A third CL experiment was done to study the toxic effect of S322 bacterial extract on oyster hemocytes. The same protocol as for A496 strain was used except that the four other sets were exposed to 2, 20, 66, and 200 μL of S322 cytoplasmic extract, respectively. To compare the levels of activity of A496 and S322 extracts, two CL assays were performed with both types of hemocytes (oyster and scallop). In each vial, 100,000 hemocytes were distributed, and then 16 μL of A496 extract was inoculated, i.e., 2.0×10^9 equivalent bacteria. To obtain the same level of "equivalent bacteria," 800 μL of S322 extract was used. This extract was concentrated to 50 μL by vacuum evaporation before inoculation. A CL assay using evaporated extract (vacuum, 80°C) had previously shown no loss of activity.

Features of the Toxic Molecule

For the next eight CL assays, in addition to control, zero, and base sets, one set of hemocytes was inoculated with 25 μL of A496 cytoplasmic extract and one set with the same inoculum after different chemical or physical treatments (see below). To evaluate the effect of the treatment, a control set of hemocytes was inoculated with sterile 1% (w/v) NaCl after the same treatment (control treatment). The treatments were

(1) cytoplasmic extract heated for 10 min at 70°C or 10 min at 100°C. Precipitated proteins were discarded after centrifugation (10,000g, 10 min);

(2) cytoplasmic extract incubated for 1 h at 35°C, with a final concentration of 0.4 mg \times mL⁻¹ of proteinase K (Sigma). Proteinase K was then inhibited by the addition of 1 mM (final concentration) PMSF (phenylmethanesulfonyl fluoride, Sigma);

(3) protein precipitation with perchloric acid; 1.6 mL of A496 cytoplasmic extract was diluted by the addition of 5.07 mL of 1% (w/v) NaCl. After pH measurement, 0.51 mL of perchloric acid (HClO₄, 70%, 0.9 M final concentration) was added. The protein deposit was discarded after centrifugation (10,000g, 10 min), and the supernatant neutralized by adding about 1.3 mL of 2.5 M, K₂CO₃;

(4) protein precipitation with trichloroacetic acid (TCA); 1 mL of 15% (w/v) trichloroacetic acid solution was added to 1 mL of A496 cytoplasmic extract. After centrifugation (10,000g, 10 min), the pH of the supernatant was neutralized by adding about 3 mL of a mixture of 2.5 mL freon (1.1.2 trichlorotrifluoroethane 99%) and 0.5 mL amine (tri-*n*-octylamine). The amine-Freon method allowed neutralization of the pH of the supernatant in a constant volume.

Lipid extraction was performed in two ways. The first used butanol; 1 mL of extract was mixed with 1 mL of butanol. After vigorous shaking and centrifugation for 5 min at 9000g, the upper phase containing butanol and lipids was collected. A second extraction was performed under the same conditions and the two phases were again separated. Butanol from the lipid phase was evaporated and lipids were dissolved in 1 mL of 1% NaCl, 0.1% DMSO (dimethyl sulfoxide) solution. Two "control treatments" corresponding to each phase were made using the same conditions. The second method used was that of Bligh and Dyer (1959); 1 mL of cytoplasmic extract was mixed with 2 mL of ethanol and 1 mL of chloroform. After shaking, the mixture was homogenous. On adding 1 mL of ice-cold water and 1 mL of chloroform, and shaking, chloroform containing lipids was separated. Both phases were collected, evaporated, and redissolved in 10% NaCl (with 0.1% DMSO for the lipid phase). For each phase, one "control treatment" was performed.

Separation by Ultrafiltration

The A496 cytoplasmic extract, previously treated with trichloroacetic acid to eliminate large macromolecules, was filtered by centrifugation (3000 to 5000g) using Microsep Filtron tubes calibrated at molecular weight cutoffs of 30, 10, 3, and 1 kDa.

HPLC Chromatography

Two assays were performed. In the first, 200 μL of A496 cytoplasmic extract, previously treated with TCA and 1 kDa filtered, was separated by HPLC on a reverse-phase C-18 column (Hypersil, 250 mm in length), which was eluted with a methanol gradient from 5 to 50%, 1 mL \times min $^{-1}$ (Pump: L-6200A Merck-Hitachi), and with a 206-nm detector (L-4250C UV-Vis Merck detector); a total of 14 fractions were collected at 2-min intervals using a Gilson FL203B fraction collector. From the elution profile, fractions 1–3 were combined. Activities of the 12 fractions, previously vacuum evaporated (80°C), were tested using the CL assays. In the second assay, 100 μL of A496 cytoplasmic extract, previously treated with TCA and 1 kDa filtered, was separated by HPLC on a reverse-phase C-18 column (Chromasil, 250 mm in length), eluted with water, 2 mL \times min $^{-1}$ (Pump: L-6200A Merck-Hitachi), and absorbent material detected at 230 nm (L-4250C UV-Vis Merck detector). Nine fractions were collected and the activity of the fractions, previously vacuum evaporated (80°C), was then tested using CL assays.

Cytotoxic Effect

To demonstrate the toxic activity of A496 strain and A496 cytoplasmic extract on the viability of the hemocytes of *P. maximus*, 100,000 hemocytes were withdrawn from two scallops and dispersed onto a glass slide in 80 mL of SSW. The experiment was done in triplicate. A first set of slides was used as a control. In the second set, 100 bacterial cells per hemocyte (A496 strain) were added. In the third set, 25 mL of the A496 cytoplasmic extract was used. After 6 h incubation in a wet chamber at 20°C, the viability of hemocytes was checked using the acridine orange/ethidium bromide method (Mishell and Shiigi, 1980). Counting of viable cells was made on 200 cells per slide and the comparison of means was made using variance analysis ($n = 3$, $\alpha = 0.05$).

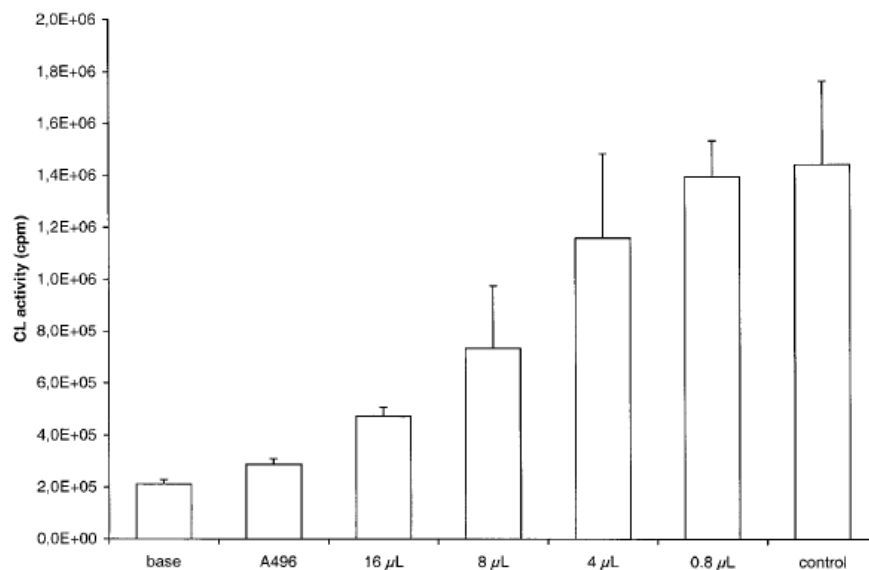


FIG. 1. CL activity (\pm SD, $n = 4$) of *P. maximus* hemocytes reached after stimulation by zymosan particles (base, nonstimulated hemocytes; control, hemocytes stimulated without prior contact with bacteria or bacterial extract; A496, hemocytes stimulated after 80 min of contact with 0.8, 4, 8, or 16 μL of A496 cytoplasmic extract).

RESULTS

Activity of *V. pectenicida* (A496 Strain) Cytoplasmic Extract

Results presented in Fig. 1 show the capacity of A496 bacterial extract to inhibit the CL activity of *P. maximus* hemocytes stimulated by zymosan particles. The inhibition was directly linked to the dose of bacterial extract. For 0.8 and 4 μL there was no significant inhibition compared to the control. For 8 μL , the inhibition was significant (ANOVA $n = 4$, $\alpha = 0.05$), and a 16- μL dose provoked an effect equivalent to that induced by live A496 bacteria. These results showed clearly that the cytoplasmic extract of the strain A496 had the same dose-dependent capacity as live bacteria to inhibit the CL activity of *P. maximus* hemocytes.

Activity of U1 Cytoplasmic Extract

Results presented in Fig. 2 show a significant difference (ANOVA $n = 4$, $\alpha = 0.05$) after stimulation by zymosan between CL activity levels of hemocytes previously in contact with either U1 extract or A496 extract. Previous contact with U1 cytoplasmic extract did not affect the CL activity of hemocytes and the CL level reached was not significantly different from that of the control. At the same time, there was a strong inhibition of the hemocyte CL activity after contact with A496 extract (>50%). This result confirmed the toxic capacity of *V. pectenica* (strain A496) cytoplasmic extract against the CL activity of *P. maximus* hemocytes.

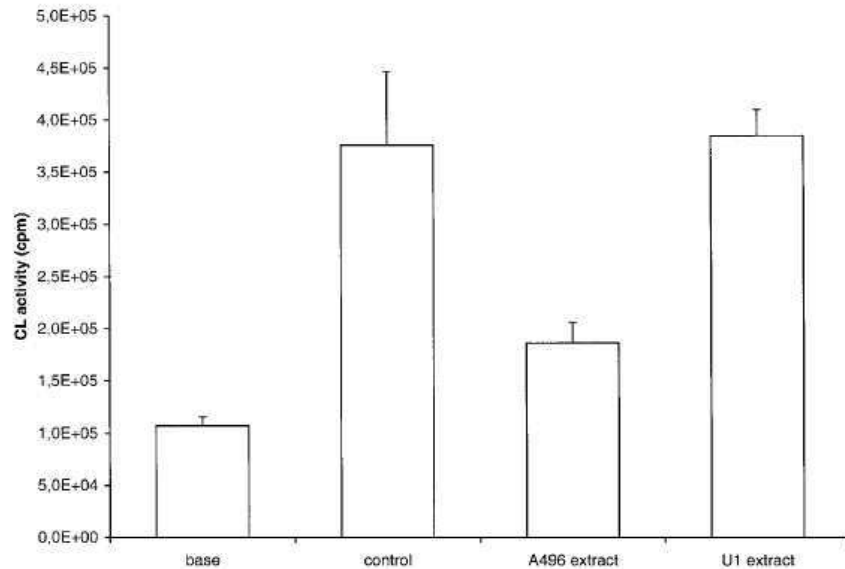


FIG. 2. CL activity (\pm SD, $n = 4$) of *P. maximus* hemocytes reached after stimulation by zymosan particles (base, nonstimulated hemocytes; control, hemocytes stimulated without prior contact with bacteria or bacterial extract; A496 extract, hemocytes stimulated after 80 min of contact with 25 μ L of A496 cytoplasmic extract, about 2.4×10^9 bacteria-equivalent. U1 extract, hemocytes stimulated after 80 min of contact with 83 μ L of U1 cytoplasmic extract, about 2.4×10^9 bacteria-equivalent).

Activity of S322 Cytoplasmic Extract on Oyster Hemocytes

Results presented in Fig. 3 show the capacity of S322 bacterial extract to inhibit the CL activity of *C. gigas* hemocytes stimulated by zymosan particles. The inhibition was directly linked to the dose of bacterial extract. For 2 μ L there was no significant inhibition compared to the control. For 20, 66, and 200 μ L, the inhibition was significant (ANOVA, $n = 4$, $\alpha = 0.05$). These results showed that the cytoplasmic extract of the strain S322 had the same dose-dependent capacity as live bacteria to inhibit the CL activity of *C. gigas* hemocytes. As shown with *V. pectenica*, these results confirmed the presence of a toxic molecule in the cytoplasm of this pathogenic vibrio.

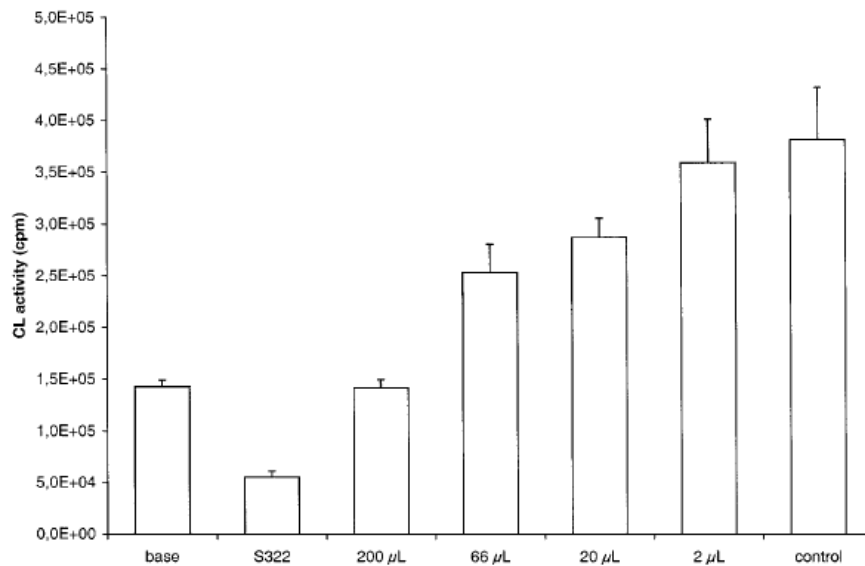


FIG. 3. CL activity (\pm SD, $n = 4$) of *C. gigas* hemocytes reached after stimulation by zymosan particles (base, nonstimulated hemocytes; control, hemocytes stimulated without prior contact with bacteria or bacterial extract; S322, hemocytes stimulated after 80 min of contact with 100 bacterial cells of *Vibrio* sp. strain S322; 200, 66, 20, 2 μ L, hemocytes stimulated after 80 min of contact with 200, 66, 20, or 2 μ L of S322 cytoplasmic extract).

A496 and S322 Cytoplasmic Extract Activity: Loss of Specificity

As shown in Table 1, there was no significant difference between CL activity reached by scallop and oyster hemocytes after zymosan stimulation, in the case of prior contact with S322 or A496 cytoplasmic extracts. Therefore, it seemed that live A496 and S322 cells were able to inhibit specifically the CL activity of the hemocytes of their usual host (*P. maximus* for A496 and *C. gigas* for S322), but this specificity of action disappeared when using cytoplasmic extracts. The fact that there was strong inhibition in all cases strongly confirmed the presence of a toxic molecule.

TABLE 1

Comparison of the CL Activity Level (cpm: mean, $n = 4$) of Hemocytes (*Pecten maximus* and *Crassostrea gigas*) Stimulated with Zymosan Particles, According to the Type of Cytoplasmic Extract (*Vibrio* sp. Strain S322 or *Vibrio pectenica* Strain A496) Previously in Contact with Them

	<i>Pecten maximus</i> hemocytes (CL activity in cpm)	<i>Crassostrea gigas</i> hemocytes (CL activity in cpm)
A496 extract	4.16×10^4 (ns)	0.50×10^6 (ns)
S322 extract	5.12×10^4 (ns)	1.31×10^6 (ns)
Control	9.93×10^5 (*)	1.20×10^7 (*)

Note. ns, no significant difference between A496 and S322 extract.

* Significant difference between control and extracts (ANOVA, $n = 4$, $\alpha = 0.05$).

Characterization of the Toxic Molecule

The results of the eight CL assays performed to define some basic features of the toxic molecule from the A496 cytoplasmic extract showed that, in all cases, the CL activity of hemocytes in the control treatment did not show any significant difference compared with the nontreated hemocytes, i.e., there was no interference

between the different treatments on toxic activity. For fractions which led to a significant decrease of the CL activity of *P. maximus* hemocytes (ANOVA, $n = 4$, $\alpha = 0.05$), there was no decline in toxic activity if the A496 cytoplasmic extract had previously been heated to 70° or 100°C or treated with proteinase K or perchloric or trichloroacetic acid. The main toxic activity was found in the nonlipid fraction of the cytoplasmic extract using butanol extraction, but the activity was present both in lipid and nonlipid fractions using the Bligh and Dyer method. After ultrafiltration, it was not possible to differentiate statistically (ANOVA, $n = 4$, $\alpha = 0.05$) the inhibition of hemocyte CL activity depending on whether they were previously in contact with A496 cytoplasmic extracts filtered at 30, 10, 3, or 1 kDa and the unfiltered control. These results showed that the toxic activity was probably due to a small molecule (<3 kDa), which was heat resistant, acid and protease resistant, and water soluble. Results after lipid extraction could suggest that there was more than one molecule, or a complex one, which was partly lipid. Nevertheless, there was strong activity in the water fraction.

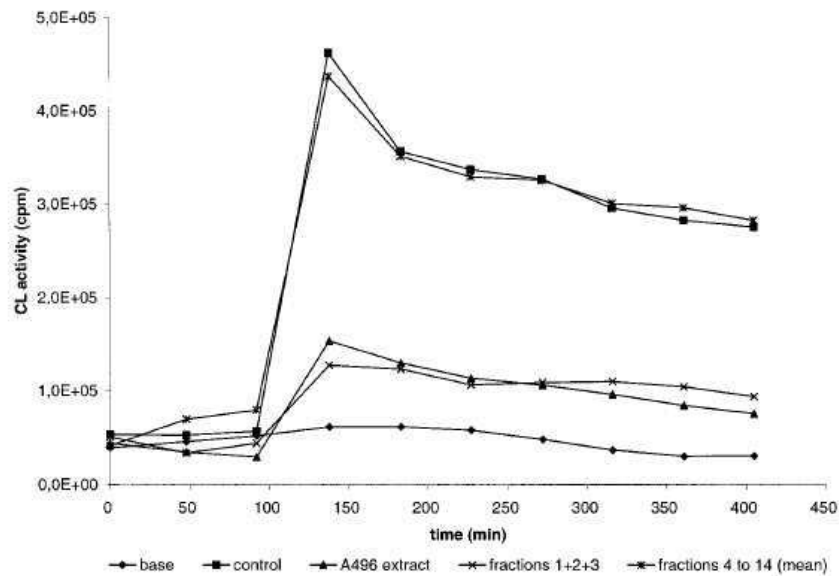


FIG. 4. Variation in CL activity (cpm) of *P. maximus* hemocytes, nonstimulated (base), stimulated by zymosan particles (at $t = 92$ min) without prior contact with bacterial extract (control), or stimulated by zymosan particles after prior contact with fractions of A496 cytoplasmic extract, TCA treated, 1 kDa filtered, eluted on HPLC with a C18 (Hypersil) reverse-phase column with a 5/50% methanol gradient; fraction 1 + 2 + 3, combined fractions corresponding to the first elution peak; fraction No. 4 to 14, mean of the results obtained using fractions 4 to 14 tested one by one. Statistical analysis (ANOVA, $n = 4$, $\alpha = 0.05$) on means at the peak level ($t = 138$ min) showed no significant differences between "fractions 4 to 14" (calculated individually) and "control" as well as no significant differences between "A496 extract" and "fraction 1 + 2 + 3."

Purification Assays

Using HPLC with a C18 reverse-phase column to separate the A496 cytoplasmic extract which had been previously TCA treated and 1 kDa filtered, it appeared (Fig. 4) that in the case of elution with a methanol gradient from 5 to 50%, the toxic activity was eluted in the first peak, corresponding to fractions 1 + 2 + 3. Using water as the elution solution (Fig. 5), the main toxic activity appeared in fraction 5, with a retention time 4.02 min. These results confirmed that the toxic molecule was in the water fraction of the extract.

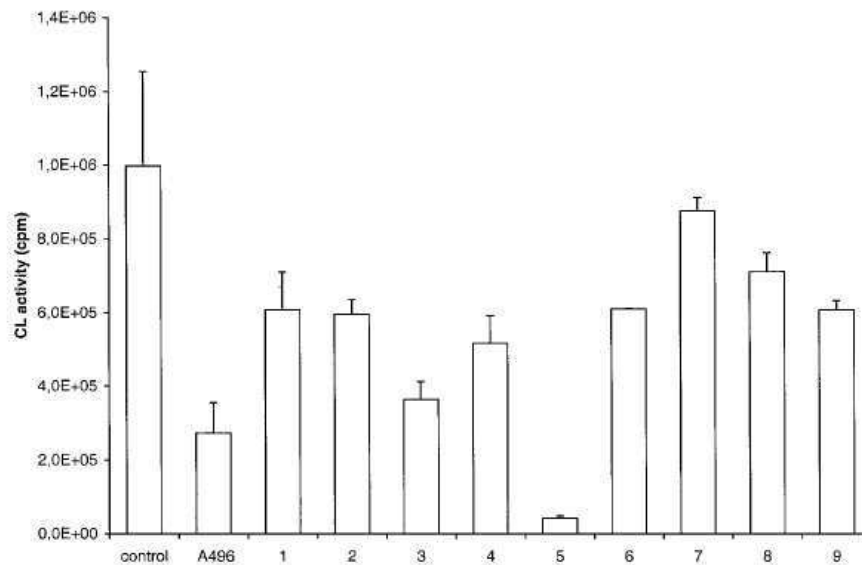


FIG. 5. CL activity (\pm SD, $n = 4$) of *P. maximus* hemocytes reached after stimulation by zymosan particles, control, hemocytes stimulated without prior contact with bacterial extract; A496, hemocytes stimulated after 80 min of contact with 25 μ L of A496 cytoplasmic extract; 1 to 9, hemocytes stimulated after 80 min of contact with one of the nine fractions of A496 cytoplasmic extract TCA treated, 1 kDa filtered, and eluted using water on HPLC with a C18 (Chromasil) reverse-phase column.

Cytotoxic Effect on Hemocytes: Glass Slide Experiment

After 6 h, the mean percentage of viable hemocytes in control slides was 76.1%, but only 30.5% in the case of hemocytes in contact with A496 bacterial cells and 7.2% in the case of hemocytes in contact with A496 cytoplasmic extract (significant difference versus control, $n = 3$, $\alpha = 0.05$). In the last two cases (bacterial cells and cytoplasmic extract) hemocytes were rounded without typical pseudopodia and filopodia.

DISCUSSION

In the present work, contact of hemocytes with the cytoplasmic extract of *V. pectenicida* (strain A496) and *Vibrio* sp. (strain S322) provoked the same CL inhibition as bacterial cells, whereas the cytoplasmic extract of U1 strain (nonpathogenic), tested under the same conditions, was unable to inhibit the CL activity. A dose response effect was also observed in both vibrio extracts.

It is not easy to correlate bacterial pathogenic activity with toxic activity. For instance, Egidius (1987) emphasized the difficulties of some authors in relating the hemolytic capacity of *V. anguillarum* to its pathogenic activity. However, in the case of *V. pectenicida*, the ability to induce mortality in *P. maximus* larvae seems partly due to its capacity to kill hemocytes. Toxic molecules in the bacterial cytoplasm reproduced the inhibition of CL activity and accelerated hemocyte death. *V. pectenicida* was also probably able to interfere with other phagocytic cells of the digestive gland. In this way, through its internal toxins, *V. pectenicida* may disrupt the defence system of the larvae and the digestive system too.

However, to produce similar CL inhibition activity in 100 live bacterial cells per hemocyte (1×10^7 bacterial cells per vial) using cytoplasmic extract required about 2.4×10^9 bacteria equivalents per vial, e.g., about 276 times more. This phenomenon can be explained by the fact that only some of the toxic factors were present in the cytoplasmic extract; either active molecules were in the form of a less active or inactive precursor or they were bound to the membrane and only partially released by the French Press procedure. The chemical and physical treatments applied to obtain the extract could also explain part of the loss of activity. Moreover, if whole bacteria were actively phagocytosed, leading to higher cytoplasmic concentrations of toxicant, there was no similar active phagocytosis using free toxins.

Another question is the mode of action of this toxin. In a first attempt, two main hypotheses can be suggested: either the toxin acts specifically on one or another step of the metabolic respiratory burst (RB) pathway or its cytotoxic activity is sufficiently strong to lead quickly to hemocyte death. This last hypothesis seems the more probable. In fact, the experiments on glass slides showed the capacity of vibrio extracts to lead to

hemocyte death within 6 h, although it is likely that it occurred earlier. Be that as it may, the present work allowed some features of the toxic molecule of *V. pectenica* to be determined. One of the active factors, which we named VHKT, for vibrio hemocyte-killer toxin, was heat stable, protease and acid resistant, and water soluble. It was also a small molecule, less than 3 kDa, and all these features make it unlikely that VHKT is a protein. Using HPLC, the active fraction of the *V. pectenica* extract was present in the fraction eluted with water. Unfortunately, RMN analysis on the active fraction (No. 5, reverse-phase C18 chromasyl, retention time 4.02 min) showed a complex composition, necessitating further purification steps.

However, from available data, it was interesting to compare VHKT to some other toxins from vibrios. Sears and Kapers (1996) reviewed the toxins from *V. cholerae*, *V. fluvialis*, *V. hollisae*, *V. parahaemolyticus*, *V. mimicus*, and *V. metschnikovii* but the activities or properties of these toxins were not comparable to those of VHKT. By focusing on vibrio species pathogenic for marine animals, the ciliostatic toxin described by Nottage *et al.* (1989) was close to VHKT in terms of thermoresistance and low molecular weight (5 kDa). But tests made using the method described by Nottage *et al.* (1989) to demonstrate ciliostatic activity showed that *V. pectenica*, either live bacteria or cytoplasmic extracts, were not effective in reducing the ciliary activity of *Mytilus edulis* gills (data not shown). Thus, it is likely that VHKT is a new toxin but further characterization of the molecule is required.

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REFERENCES

- Bachère, E., Chagot, D., and Grizel, H. 1988. Separation of *Crassostrea gigas* hemocytes by density gradient centrifugation and counterflow centrifugal elutriation. *Dev. Comp. Immunol.* **12**, 549–559.
- Bachère, E., Hervio, D., and Mialhe, E. 1991. Luminol-dependent chemiluminescence by hemocytes of two marine bivalves, *Ostrea edulis* and *Crassostrea gigas*. *Dis. Aquat. Org.* **11**, 173–180.
- Bligh, E. G., and Dyer, J. 1959. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.* **37**, 911–917.
- Brown, C., and Roland, G. 1984. Characterization of exotoxin produced by a shellfish-pathogenic *Vibrio* sp. *J. Fish Dis.* **7**, 117–126.
- Egidius, E. 1987. Vibriosis: Pathogenicity and pathology. A review. *Aquaculture* **6**, 15–28.
- Hancock, I. C., and Poxton, I. R. 1988. Bacterial cell surface technique. In “Modern Microbiological Methods,” p. 56. Wiley–Interscience, New York.
- Lambert, C. 1998. “Etude des Infections à Vibrionaceae chez les Larves de Mollusques Bivalves à Partir d’un Modèle Larve de *Pecten maximus*.” Thesis doctoral, Université de Bretagne Occidentale, Brest, France.
- Lambert, C., and Nicolas, J. L. 1998. Specific inhibition of chemiluminescent activity by pathogenic vibrios in hemocytes of two marine bivalves: *Pecten maximus* and *Crassostrea gigas*. *J. Invertebr. Pathol.* **71**, 53–63.
- Lambert, C., Nicolas, J. L., Cilia, V., and Corre, S. 1998. *Vibrio pectenica* sp. nov. a pathogen of scallop (*Pecten maximus*) larvae. *Int. J. Syst. Bacteriol.* **48**, 481–487.
- Mishell, B. B., and Shiigi, S. M. 1980. In “Selected Methods in Cellular Immunology” (B. B. Mishell and S. M. Shiigi, Eds.), pp. 21–22. Freeman, San Francisco.
- Nicolas, J. L., Corre, S., Gauthier, G., Robert, R., and Ansquer, D. 1996. Bacterial problems associated with scallop (*Pecten maximus*) larval culture. *Dis. Aquat. Org.* **27**, 67–76.
- Nottage, A. S., and Birkbeck, T. H. 1986. Toxicity to marine bivalves of culture supernatant fluids of the bivalve-pathogenic *Vibrio* strain NCMB 1338 and other marine vibrios. *J. Fish Dis.* **9**, 249–256.
- Nottage, A. S., Sinclair, P. D., and Birkbeck, T. H. 1989. Role of low-molecular-weight ciliostatic toxins in vibriosis of bivalve mollusks. *J. Aquat. Anim. Health* **1**, 180–186.
- Sears, C. L., and Kaper, J. B. 1996. Enteric bacterial toxins: Mechanisms of action and linkage to intestinal secretion. *Microbiol. Rev.* **60**, 167–215.
- Umbreit, T. H., and Tripp, M. R. 1975. Characterization of the factors responsible for death of fish injected with *Vibrio anguillarum*. *Can. J. Microbiol.* **21**, 1271–1274.
- Waterborg, J. H., and Matthews, H. R. 1984. The Lowry method for protein quantification. In “Methods in Molecular Biology, Vol. 1, Proteins” (John M. Walker, Ed.), pp. 1–3. Humana Press, Clifton, New Jersey.