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Differential tissue distribution and specificity of phenoloxidasés from the Pacific oyster *Crassostrea gigas*

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1 **Differential tissue distribution and specificity of phenoloxidasés from the Pacific oyster**
2 *Crassostrea gigas*

3

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19 Abreviations :

20 PO: phenoloxidase; HLS: haemocyte lysate supernatant; PPD: p-phenylenediamine; PTU : 1-
21 phenyl-2-thiourea; CTAB: cethyltrimethylammonium bromide; MBTH: 3-methyl-2-
22 benzothiazolinone hydrazone; Tris HCl: trizma hydrochloride ; AS: ammonium sulfate; SDS:
23 sodium dodecyl sulfate; TEMED: N,N,N',N'-tetramethylethylenediamine; BSA: bovine
24 serum albumin.

25

26 **Abstract:** Phenoloxidases (POs) play a key role in melanin production, are involved in
27 invertebrate immune mechanisms, and have been detected in different bivalves. Recently, we
28 identified catecholase- and laccase-like PO activities in plasma and haemocyte lysate
29 supernatant (HLS) of the Pacific oyster *Crassostrea gigas*. To go further in our investigations,
30 the aims of this study were (i) to determine the tissue distribution of PO activities in *C. gigas*,
31 and (ii) to identify and characterise the different sub-classes of POs (i.e. tyrosinase,
32 catecholase and/or laccase) involved in these oxido-reductase activities. With dopamine and
33 p-phenylenediamine (PPD) but not with L-tyrosine used as substrates, PO-activities were
34 detected by spectrophotometry in the gills, digestive gland, mantle, and muscle. These results
35 suggest the presence of catecholase and laccase but not of tyrosinase activities in oyster
36 tissues. The highest activity was recovered in the digestive gland. PO-like activities were all
37 inhibited by 1-phenyl-2-thiourea (PTU) and by the specific laccase inhibitor,
38 cethyltrimethylammonium bromide (CTAB). With dopamine as substrate, the catecholase
39 inhibitor 4-hexylresorcinol (4-HR) only inhibited PO in the muscle. SDS-PAGE zymographic
40 assays with dopamine and PPD elicited a unique ~40 kDa protein band in the muscle. In the
41 other tissues, laccase-like activities could be related to ~10 kDa and/or ~200 kDa protein
42 bands. The ~10 kDa protein band was also detected in plasma and HLS, confirming the
43 presence of a laccase in the later compartments, and probably in most of the tissues of
44 *C.gigas*. This is the first time to our knowledge that a ~10 kDa protein band is associated to a
45 laccase-like activity in a mollusc species, contributing to the characterisation of
46 phenoloxidase activities in marine bivalves.

47

48 **Key Words:** bivalve; phenoloxidase; laccase; catecholase; zymography

49

50 **1. Introduction**

51 Phenoloxidases (POs, EC 1.14.18.1) are a class of copper proteins widely distributed in
52 bacteria, fungi, plants and animals (Cerenius et al. 2008). They play a key role in melanin
53 production and are implicated in immune defence mechanisms in invertebrates. This class of
54 enzymes include tyrosinases (EC 1.14.18.1), catecholases (EC 1.10.3.1) and laccases (EC
55 1.10.3.2), all capable of o-diphenol oxidation. However, among these three enzymes, only
56 tyrosinases can hydroxylate monophenols (e.g. L-tyrosine), and only laccases can oxidise p-
57 diphenols and aromatic compounds containing amine groups (e.g. p-phenylenediamine, PPD)
58 (Thurston 1994, Solomon et al. 1996). In addition to that, a panel of inhibitors exert different
59 actions on these three types of enzymes: while 1-phenyl-2-thiourea (PTU) inhibits the three
60 types of PO activities (Williamson 1997, Jordan and Deaton 2005), 4-hexylresorcinol (4-HR)
61 inhibits tyrosinase and catecholase but not laccase activities (Dawley and Flurkey 1993,
62 Zavarzina and Zavarzin 2006) and cethyltrimethylammonium bromide (CTAB) specifically
63 inhibits laccase activity (Walker and McCallion 1980). Recently, we conducted a study to
64 identify PO activities present in the haemolymph of the Pacific oyster *Crassostrea gigas*
65 (Luna-Acosta et al. 2010a). By using different PO substrates, such as L-tyrosine, L-3,4-
66 dihydroxyphenylalanine (L-DOPA), dopamine or PPD, and different PO inhibitors, such as
67 PTU, 4-HR and CTAB, results suggested the presence of both catecholase- and laccase-like
68 activities in the plasma, and the presence of a laccase-like activity in the haemocyte lysate
69 supernatant (HLS, Luna-Acosta et al. 2010a). Our interest in *C. gigas* comes from the fact
70 that this organism dominates over all other molluscs with respect to global distribution and
71 aquaculture production, but suffers from massive summer mortality each year (Cheney et al.
72 2000). Summer mortality of *C. gigas* has been suggested to be the result of a complex
73 interaction between the host, pathogens and environmental factors (Cheney et al. 2000).
74 Importantly, studies in *C. gigas* have shown that PO activities, usually detected by using the

75 o-diphenol substrates L-DOPA or dopamine, can be modulated by environmental factors,
76 such as the presence of heavy metals or hydrocarbons (Gagnaire et al. 2004a, Bado-Nilles et
77 al. 2008, Luna-Acosta et al. 2010b). In addition to that, a gene coding for a laccase in the
78 haemocytes from *C. gigas* was modulated in the presence of hydrocarbons (Bado-Nilles et al.
79 2010). To the best of our knowledge, studies on POs in *C. gigas* have only been carried out in
80 the haemolymphatic compartment. However, POs may be present in other body tissues in
81 bivalves, e.g. in the prismatic shell layer (Nagai et al. 2007) or in the byssus gland (Hellio et
82 al. 2000). A better characterisation and localisation of POs in *C. gigas* is needed to expand
83 our knowledge on the immune defence mechanisms in this organism and therefore to a better
84 understanding of the potential causes of summer mortality events.

85 In this general context, our goal was to determine the distribution and the nature of PO
86 activities (tyrosinase, catecholase, and laccase) in different oyster body compartments, namely
87 gills, digestive gland, mantle, muscle, plasma and HLS. PO activities were determined by
88 spectrophotometry using different PO substrates (L-tyrosine, dopamine and PPD) and PO
89 inhibitors (PTU, 4-HR, CTAB). Electrophoretic techniques using polyacrylamide gels are
90 useful to detect PO enzymes and their associated molecular weights in crude extracts without
91 the necessity of enzyme purification (Cardenas and Dankert 2000, Decker et al. 2001, Dicko
92 et al. 2002, Perdomo-Morales et al. 2007). Hence, SDS-PAGE zymographic assays were
93 carried out on crude and partially purified samples from the different oyster compartments.
94 Differences between tissues, in terms of PO-like activity and molecular weight characteristics,
95 are discussed.

96

97 **2. Materials and methods**

98 *2.1. Chemicals and materials*

99 L-tyrosine, dopamine, p-phenylenediamine (PPD), 1-phenyl-2-thiourea (PTU), 4-
100 hexylresorcinol (4-HR), cethyltrimethylammonium bromide (CTAB), 3-methyl-2-
101 benzothiazolinone hydrazone (MBTH), trizma hydrochloride (Tris HCl), sodium chloride
102 (NaCl), ammonium sulfate (AS), sodium dodecyl sulfate (SDS), trizma base, glycine,
103 N,N,N',N'-tetramethylethylenediamine (TEMED), ammonium persulfate, glacial acetic acid,
104 Coomassie brilliant blue, bovine serum albumin (BSA), copper sulfate and bicinchoninic acid
105 were obtained from Sigma-Aldrich (France). Magnesium chloride ($MgCl_2$) and calcium
106 chloride ($CaCl_2$) were obtained from Acros organics (France). Acrylamide/Bis acrylamide
107 30% was obtained from Bio-Rad.

108

109 2.2. *Oysters*

110 Three years old Pacific oysters, *Crassostrea gigas* (n= 30; mean \pm SD; weight: 75.5 ± 8.7 g;
111 length: 9 ± 3 cm) were purchased during October-November 2008 from shellfish farms in
112 Aytré Bay (Charente Maritime, France), on the French Atlantic coast, and were processed
113 immediately after their arrival in the laboratory.

114

115 2.3. *Collection of oyster tissues*

116 After opening the oyster shells by cutting off the adductor muscle, a quantity (0.5-1 ml) of
117 haemolymph was withdrawn directly from the pericardial cavity with a 1-ml syringe equipped
118 with a needle (0.9 x 25 mm), and the haemolymph from 10 oysters was pooled to reduce
119 inter-individual variation (Gagnaire et al. 2004b). Haemolymph samples were centrifuged
120 (260 g, 10 min, 4°C) to separate the cellular fraction (i.e. haemocytes) from plasma, as
121 described previously (Hellio et al. 2007). Gills, digestive gland, mantle and muscle were
122 removed from oysters and pooled. Three replicates from 10 oysters were prepared per tissue.

123 Haemocytes, gills, digestive gland, mantle and muscle were homogenized at 4°C in Tris
124 buffer (0.1 M Tris HCl, 0.45 M NaCl, 26 mM MgCl₂ and 10 mM CaCl₂) adjusted to pH 7.
125 Haemocytes were lysed using a Thomas-Potter homogenizer (IKA-Labortechnik, clearance
126 0.13-0.18mm) at 200 rpm for 1 min. Gills, digestive gland, mantle and muscle were
127 homogenized, as described previously (Luna-Acosta et al. 2010b), using an Ultra Turrax (T25
128 basic, IKA-WERKE) at 19 000 rpm for 1 min followed by twelve up and down strokes of
129 Thomas-Potter homogenizer at 200 rpm for 1 min (IKA-Labortechnik RW 20.n, size 0.13-
130 0.18mm, BB). All homogenized samples were centrifuged at 10 000 g for 10 min at 4°C. The
131 resulting haemocyte lysate supernatant (HLS) and tissue supernatants were collected for
132 enzymatic studies.

133 Aliquots (100 µl) of plasma, HLS and tissue samples were stored at -80°C. Each aliquot was
134 used only once per microplate for spectrophotometric analysis, or per gel running for
135 zymographic studies.

136

137 *2.4. Partial purification*

138 A previous analysis, by using different concentrations of saturated ammonium sulfate (0, 30,
139 40, 60, 70, 80, 100%), revealed that precipitation with 60% saturated ammonium sulfate (60P-
140 SAS) was the best condition for protein concentration to detect PO-like activity for oyster
141 tissues, i.e. the gills, digestive gland, mantle and muscle (data not shown), and was in
142 agreement with other studies (Cong et al. 2005, Liu et al. 2006). Therefore, proteins of
143 collected supernatants from oyster tissues were brought to 60% saturation concentration by
144 addition of solid ammonium sulfate at 4°C, and allowed to stand overnight. The resulting
145 precipitate was collected by centrifugation (15 500 g for 10 min), dissolved in a small volume
146 of Tris buffer , and dialysed at 4°C against distilled water for 12h and twice against Tris

147 buffer for 8h. Crude plasma samples were concentrated with Centricon-5 centrifugal
148 concentration units (Amicon™).

149

150 *2.5. Phenoloxidase assays*

151 Phenoloxidase-like (PO-like) activity was measured spectrophotometrically by recording the
152 formation of o-quinones, as described previously (Luna-Acosta et al. 2010a). PO assays were
153 conducted in 96-well microplates (Nunc, France). Dopamine or p-phenylenediamine (PPD)
154 were used as substrates, at final concentrations of 100 mM and 50 mM, respectively.
155 Dopamine (100 mM) was prepared just before being used in Tris buffer. At 25°C, 10 µl of
156 sample was incubated with 80 µl of dopamine and 50 µl of Tris buffer. Several control wells
157 were systematically used: ‘buffer control’ containing only buffer, ‘sample control’ containing
158 only sample and buffer, and ‘non-enzymatic control’ containing only substrate and buffer,
159 always in a final volume reaction of 140µl. Immediately after dopamine addition, PO-like
160 activity was monitored during 4h by using a VersaMax™ microplate reader (Molecular
161 Devices) and by following the increase of absorbance at 490 nm. Because of solubility
162 constraints, the protocol was slightly modified in the case of PPD: the sample was incubated
163 with 7 µl of PPD (50 mM diluted in methanol) and 123 µl of buffer (no effect of methanol
164 was observed on the enzymatic reactions). PO-like activity was monitored during 2h at 420
165 nm. For all conditions, the experiments were performed with three pooled oyster samples.
166 Each pool was tested in triplicate wells and average rates were calculated by dividing the sum
167 of replicate measurements from the three oyster pools, by the number of measurements, i.e. 9
168 (3 replicate measurements x 3 oyster pools).

169 For enzymatic oxidation, the results were systematically corrected for non-enzymatic
170 autoxidation of the substrate and were expressed in specific activity (SA), i.e. in international
171 units (IU) per mg of protein. One IU is defined as the amount of enzyme that catalyzes the

172 appearance of 1 μmole of product per min (Fenoll et al. 2002) under the above conditions
173 using molar extinction coefficient of dopamine and PPD reactions products of 3 300 $\text{M}^{-1} \text{cm}^{-1}$
174 (Waite 1976) and 43 160 $\text{M}^{-1} \text{cm}^{-1}$ (Eggert et al. 1996, Paranjpe et al. 2003), respectively.

175

176 *2.6. Phenoloxidase inhibition assays*

177 Working solutions of inhibitors were prepared just before being used in Tris buffer. PO
178 inhibition assays were performed by preincubating 10 μl of the specific PO inhibitor PTU
179 (5 mM, final concentration), the specific tyrosinase and catecholase inhibitor 4-HR (1 mM,
180 final concentration), or the specific laccase inhibitor CTAB (1 mM, final concentration), with
181 10 μl of sample for 20 minutes. Then, PO assay was carried out with dopamine (100 mM,
182 prepared in Tris buffer) or PPD (50 mM, prepared in methanol). Appropriate controls were
183 used as described before. Experiments were performed with three pooled oyster samples.
184 Each pool was tested in triplicate wells and average rates were calculated.

185

186 *2.7. Protein assays*

187 Protein concentrations were determined by the slightly modified Lowry method, as described
188 previously (Smith et al. 1985), using bovin serum albumin as standard.

189

190 *2.8. Gel electrophoresis and zymography*

191 To associate PO enzyme activities with individual proteins, and estimate the molecular
192 weights of the enzymes, SDS-PAGE and 1 D-zymography were used. Aliquots of the
193 different oyster tissues (equivalent to 76 μg of proteins for gills, 76 μg for digestive gland, 57
194 μg for mantle, 40 μg for muscle, 47 μg for plasma and 1.55 μg for HLS) were mixed with
195 sample buffer (65 mM Tris HCl pH 6.8, 25% glycerol, 2% SDS, 0.01% Bromophenol blue).
196 Samples were then applied to 7% SDS-PAGE gels or 15% SDS-PAGE gels in non reducing

197 conditions (i.e. without boiling samples after the addition of sample buffer) and with an upper
198 gel of 4% using a Mini-PROTEAN III Cell (Bio-Rad). Electrophoresis was carried out
199 according to the method of Laemmli (1970) at 110V for 2h45. Two gels containing the same
200 samples were run and processed in parallel. For each tissue, samples previously brought to 0,
201 30, 40 or 60% saturation concentration by addition of solid ammonium sulfate at 4°C were
202 runned per gel. After electrophoresis, SDS-PAGE gels were washed 2 x 10 min in distilled
203 water and 2 x 10 min in Tris buffer.

204 The first SDS-PAGE gel was stained with a solution containing 100 mM L-tyrosine and 5
205 mM MBTH (to detect tyrosinase activity), 100 mM dopamine and 5 mM MBTH (to detect
206 catecholase activity), or 100 mM PPD (to detect laccase activity). MBTH was used, according
207 to the method of Dicko et al. (2002), to trap o-quinone products originating from the
208 oxidation of phenolic compounds by phenoloxidases. All substrates were dissolved in Tris
209 buffer. The gels were developed for 1 h, at 25°C and then rinsed with distilled water several
210 times, dried at room temperature and photographed.

211 The second SDS-PAGE gels were immediately washed with distilled water and stained with
212 Coomassie brilliant blue R-250 for visualizing total proteins. The molecular weight of PO
213 activity bands were estimated with pre-stained molecular weight markers (Broad Range
214 Markers, Tebu Bio, France) that were run together with samples (data not shown).

215 In order to test the specificity of the zymographic assay, a purified laccase from *Trametes*
216 *versicolor* (20 µg) and a purified superoxide dismutase (SOD) from bovine erythrocytes (20
217 µg) were included in the activity gels.

218

219 2.9. Statistical analysis

220 All values are reported as mean \pm standard deviation (SD). Statistical analysis was carried out
221 with SYSTAT 11.0. Values were tested for normality (Shapiro test) and homogeneity of

222 variances (Bartlett test). In some cases, logarithmic transformations (Log_{10}) were used to meet
223 the underlying assumptions of normality and homogeneity of variances. For normal values,
224 one-way nested ANOVA tests were used followed by a Tukey post-hoc test. For non normal
225 values, Kruskal-Wallis tests were applied, followed by Dunn's multiple comparisons test (Zar
226 1984). The statistical significance was designed as being at the level of $p < 0.05$.

227

228 **3. Results**

229 *3.1. Spectrophotometric studies*

230 Different PO substrates (L-tyrosine, dopamine and PPD), the common PO inhibitor, PTU, the
231 tyrosinase and catecholase inhibitor, 4-HR, and the laccase inhibitor, CTAB, were used.
232 When L-tyrosine was used as substrate, no PO-like activity was detected in any of the tissues
233 that were tested, i.e. gills, digestive gland, mantle, and muscle (data not shown). When
234 dopamine and PPD were used as substrates, PO-like activity was detected in all oyster tissues
235 (Fig. 1). PO-like activity was inhibited by PTU. The inhibition was total in muscle with
236 dopamine as substrate (Fig. 1g), and in digestive gland (Fig. 1d), mantle (Fig. 1f), and muscle
237 (Fig. 1h) with PPD as substrate. PO-like activity was insensitive to 4-HR except in the muscle
238 with dopamine as substrate (Fig. 1g). By contrast, PO-like activity was fully (or almost fully)
239 inhibited by the laccase inhibitor CTAB (1 mM) in all the oyster tissues with both dopamine
240 and PPD as substrates (Fig. 1).

241 Since fresh weight differs between the different analyzed tissues (i.e. the gills, digestive
242 gland, mantle and muscle), tissue distribution of PO-like activity was also examined in terms
243 of recovery of enzymatic activity (Table 1). With dopamine as substrate, the highest total PO-
244 like activity was recovered in the digestive gland, followed by the gills, mantle and muscle
245 (Table 1). With PPD as substrate, the total PO-like activity was considerably higher in the
246 digestive gland compared to the other compartments.

247 3.2. SDS-PAGE zymographic assays

248 When gels were stained with L-tyrosine, no bands were observed, and this, for all the oyster
249 tissues tested, i.e. the gills, digestive gland, mantle, muscle, plasma and HLS (data not
250 shown). However, PO-like activity was detected in all oyster tissues that were analyzed by
251 SDS-PAGE zymographs, with both dopamine and PPD as substrates.

252 PO substrates such as dopamine or PPD can oxidise non-enzymatically. This leads to different
253 intermediary products of the melanization cascade such as quinone radicals. Unspecific
254 reactions between quinone radicals issued from non-enzymatically oxidation reactions and
255 radical species that could be produced by SOD-like proteins are thus likely to take place in
256 zymographic studies (Eibl et al. 2010). However, PO-like activity was detected in the
257 presence of laccase from *T. versicolor* but not in the presence of SOD from bovine
258 erythrocytes (data not shown), confirming that our zymographic conditions were well adapted
259 to discriminate between true PO-like activities and other enzymatic activities involving
260 radical species.

261 With dopamine as substrate, the activity detected for the gills and the mantle corresponded to
262 one upper band with an estimated molecular weight of ~200 kDa and to a lower band with an
263 estimated molecular weight of ~10 kDa (Fig. 2a). In both tissues, most of the PO activity
264 appeared in the higher molecular mass band. For the digestive gland the activity corresponded
265 to an upper band of ~200 kDa and for the plasma and the HLS to one lower band with an
266 estimated molecular weight of ~10 kDa (Fig. 2a). Similarly, an upper band with a molecular
267 weight of ~200 kDa was observed in the presence of PPD for the gills and the mantle, but not
268 for the digestive gland (Fig. 2b), and a lower band with a molecular weight of ~10 kDa was
269 detected in the presence of PPD for the gills, digestive gland, mantle, crude plasma and crude
270 HLS (Fig. 2b). Again, most activity in samples from the gills and the mantle was evident in
271 higher molecular mass bands. The bands of ~10 kDa are not likely to be an artefact since they

272 stained differentially with dopamine and PPD, depending on the tissue that was analyzed, e.g.
273 stained with PPD but not with dopamine in the digestive gland. Contrary to the other tissues,
274 the activity detected for the muscle, with dopamine and PPD as substrates, corresponded to a
275 band with an estimated molecular weight of ~40 kDa.

276

277 **4. Discussion**

278 POs are of widespread occurrence in bacteria, fungi, plants, invertebrates and vertebrates
279 (Sanchez-Ferrer et al., 1995; Cerenius et al., 2008). Despite the importance of the reactions
280 and the functional roles associated to POs, the enzymes belonging to the PO class have not
281 been thoroughly characterised in molluscs and especially in bivalve species. In the present
282 study, spectrophotometric analyses were conducted to identify PO-like activity in different
283 tissues from *C. gigas*. L-tyrosine, dopamine and PPD were used as substrates and different
284 PO inhibitors were tested. When L-tyrosine was used as substrate, no PO-like activity was
285 detected in any of the tissues that were tested, i.e. the gills, digestive gland, mantle, and
286 muscle. Similar results were recently obtained with plasma and HLS (Luna-Acosta et al.
287 2010a). These data suggest the total absence of tyrosinase-like activity in *C. gigas*. Inhibition
288 assays in the present study were thus only conducted with dopamine and PPD as substrates.

289 The choice on PO specific inhibitors was based on a previous study carried out on the PO
290 inhibitors described in the literature (Luna-Acosta et al. 2010a). Indeed, PO substrates can
291 chemically oxidise in the absence of PO (autoxidation) and many PO inhibitors described in
292 the literature such as reducing agents (e.g. 2-mercaptoethanol or sodium azide) can inhibit
293 autoxidation reactions (Luna-Acosta et al. 2010a). Such inhibitors are likely to react with the
294 substrate and/or the quinone intermediates derived from the autoxidation reaction. Therefore,
295 this type of inhibitors should be avoided for identifying PO activity. Among PO inhibitors
296 described in different species, PTU was chosen because it has been described as a common

297 inhibitor of all POs (Arias et al. 2003, Zufelato et al. 2004), 4-HR as a tyrosinase and a
298 catecholase but not a laccase specific inhibitor (Dawley and Flurkey 1993, Zavarzina and
299 Zavarzin 2006) and CTAB as a laccase but not a tyrosinase or a catecholase specific inhibitor
300 (Walker and McCallion 1980, Martinez-Alvarez et al. 2008). Moreover, no inhibitory effect
301 was observed with these chemicals in dopamine or PPD autoxidation reactions (Luna-Acosta
302 et al. 2010a).

303 In the present study, PO-like activity in the muscle was completely inhibited in the presence
304 of PTU (5 mM) and 4-HR (1 mM) with dopamine as substrate, and in the presence of PTU (5
305 mM) and CTAB (1 mM) with PPD as substrate (Fig. 1), suggesting the presence of
306 catecholase and laccase in this tissue. These results are in agreement with those obtained
307 previously with plasma (Luna-Acosta et al. 2010a). However, in the previous study, no
308 inhibition was exerted by CTAB with plasma and with dopamine as substrate (Luna-Acosta et
309 al. 2010a), while, in the present study, an inhibitory effect by CTAB was observed in the
310 muscle and with dopamine as substrate. Since 4-HR and CTAB are specific catecholase and
311 laccase inhibitors (van Doorn and Vaslier 2002), respectively, results in the muscle suggest
312 the presence of a laccase sensitive to inhibition by 4-HR, or a catecholase sensitive to
313 inhibition by CTAB.

314 Interestingly, PO-like activities in all other tissues were partially inhibited in the presence of
315 PTU and completely inhibited in the presence of CTAB with dopamine and PPD as
316 substrates, suggesting the presence of laccase activity in different tissues from *C. gigas* (Fig.
317 1). These results are in agreement with those obtained previously with HLS (Luna-Acosta et
318 al. 2010a).

319 In zymographic studies, catecholase- and/or laccase- but not tyrosinase-like activities were
320 detected in the gills, digestive gland, mantle, muscle, plasma and HLS (Fig. 2). This coincides
321 with properties of the Asian swimming crab *Charybdis japonica* (Liu et al. 2006) and the

322 eastern oyster *C. virginica* (Jordan and Deaton 2005), and differs from tyrosinase-type POs
323 from other invertebrates, such as the vinegar fly *Drosophila melanogaster* (Asada et al. 1993),
324 the bloodfluke planorb *Biomphalaria glabrata* (Bai et al. 1997), the Manila clam *Ruditapes*
325 *philippinarum* (Cong et al. 2005) and the Sydney rock oyster *Saccostrea glomerata*
326 (Aladaileh et al. 2007).

327 In the present study, when dopamine or PPD were used as substrates, PO-positive bands of
328 ~10, 40 or 200 kDa were detected in *C. gigas* depending on the tissue. These tissue-dependent
329 differences in molecular weights may be due to (i) the activation state of POs or (ii) the
330 existence of polymeric forms of the enzyme. Indeed, the molecular weights of POs vary
331 depending on the activation state, animal tissue and animal species that are studied (Table 2).
332 In molluscs, the molecular weights of POs estimated by exclusion chromatography or SDS-
333 PAGE electrophoresis are in the range of 35 to 381 kDa, and in invertebrates, POs occur as
334 monomers, dimers, tetramers or pentamers (Jaenicke and Decker 2003). Thus, differences in
335 molecular weights in the present study may be explained by the existence of polymeric forms
336 of the enzyme. Generally, the molecular weight of monomeric forms is about 40 to 45 kDa,
337 and generally each subunit possesses two copper atoms (Prota et al. 1981). In the present
338 study, a PO-positive band of ~40 kDa was detected in the muscle of *C. gigas*, suggesting that
339 a monomeric form of a laccase sensitive to inhibition by 4-HR, or a catecholase sensitive to
340 inhibition by CTAB could be present in the muscle of this species. In the other analyzed
341 tissues, PO-positive bands of ~10 or ~200 kDa were detected, and in the haemolymphatic
342 compartments, PO-positive bands of ~10 kDa were detected. Results of the upper band of
343 ~200 kDa are in agreement with the large range of molecular weights of PO reported for
344 molluscs, i.e. from 35 to 381 kDa (Table 2). However, to our knowledge, this is the first time
345 that a PO-positive band of ~10 kDa is reported in a mollusc species. A PO-positive band of
346 ~10 kDa was detected in a non-mollusc aquatic invertebrate, the red swamp crayfish

347 *Procambarus clarkii* (Cardenas and Dankert 2000), suggesting that *C. gigas* possesses PO
348 with characteristics (i.e. PO-like activity and molecular weight) comparable to that of
349 arthropods.

350 Overall, in the present study, differences between tissues were observed in terms of (i)
351 substrate affinity (ii) effect of PO inhibitors, and (iii) number and molecular weight of the
352 bands detected by zymography. The data presented here suggest that zymography can be a
353 useful way of characterising PO-like activities present in *C. gigas*. Interestingly, results of the
354 present study revealed that numerous differences exist between the tissues and the
355 haemolymphatic compartment in *C. gigas*, both in terms of PO-like activities and in terms of
356 proteins that may be responsible for these activities. Our results indicate that at least three
357 oligomeric forms of POs coexist in the Pacific oyster. This comparative study gives first
358 evidences of structure-function relationships of tissue POs in *C. gigas*, contributing to the
359 understanding of tissue-specific heterogeneity of PO activities in this marine organism. As
360 POs are involved in immune response, tests based on modifications in oligomeric forms and
361 functions of this class of enzymes, and more particularly of laccase, could be used as a probe
362 to measure health conditions in this economically important species.

363

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372 **References**

- 373 Aladaileh, S., Rodney, P., Nair, S.V., Raftos, D.A., 2007. Characterization of phenoloxidase
374 activity in Sidney rock oysters (*Saccostrea glomerata*). *Comp. Biochem. Physiol. B* 148, 470-
375 480.
- 376 Arias, M.E., Arenas, M., Rodriguez, J., Soliveri, J., Ball, A.S., Hernandez, M., 2003. Kraft
377 pulp biobleaching and mediated oxidation of a nonphenolic substrate by laccase from
378 *Streptomyces cyaneus* CECT 3335. *Appl. Env. Microbiol.* 69, 1953-1958.
- 379 Asada, N., Fukumitsu, T., Fujimoto, K., Masuda, K., 1993. Activation of prophenoloxidase
380 with 2-propanol and other organic compounds in *Drosophila melanogaster*. *Insect Biochem.*
381 *Mol. Biol.* 23, 515-520.
- 382 Bado-Nilles, A., Gagnaire, B., Thomas-Guyon, H., Le Floch, S., Renault, T., 2008. Effects of
383 16 pure hydrocarbons and two oils on haemocyte and haemolymphatic parameters in the
384 Pacific oyster, *Crassostrea gigas* (Thunberg). *Toxicol. In Vitro* 22, 1610-1617.
- 385 Bado-Nilles, A., Renault, T., Faury, N., Le Floch, S., Quentel, C., Auffret, M., Thomas-
386 Guyon, H. 2010. *In vivo* effects of LCO soluble fraction on immune-related functions and
387 gene transcription in the Pacific oyster, *Crassostrea gigas* (Thunberg). *Aquat. Toxicol.* 97,
388 196-203.
- 389 Bai, G., Brown, J., Watson, C., Yoshine, T., 1997. Isolation and characterization of
390 phenoloxidase from egg masses of the gastropod mollusc, *Biomphalaria glabrata*. *Comp.*
391 *Biochem. Physiol. B* 118, 463-469.
- 392 Cardenas, W., Dankert, J., 2000. Cresolase, catecholase and laccase activities in haemocytes
393 of the red swamp crayfish. *Fish Shellfish Immunol.* 10, 33-46.
- 394 Cerenius, L., Lee, B., Söderhäll, K., 2008. The proPO-system: pros and cons for its role in
395 invertebrate immunity. *Trends Immunol.* 29, 263-271.

- 396 Cheney, D., Macdonald, E., Elston, R., 2000. Summer mortality of Pacific oysters
397 *Crassostrea gigas* (Thunberg): Initial findings on multiple environmental stressors in Puget
398 Sound, Washington. *J. Shellfish Res.* 19, 353-359.
- 399 Cong, R., Sun, W., Liu, G., Fan, T., Meng, X., Yang, L., Zhu, L., 2005. Purification and
400 characterization of phenoloxidase from clam *Ruditapes philippinarum*. *Fish Shellfish*
401 *Immunol.* 18, 61-70.
- 402 Dawley, R.M., Flurkey, W.H., 1993. Differentiation of tyrosinase and laccase using 4-
403 hexylresorcinol, a tyrosinase inhibitor. *Phytochemistry.* 33, 281-284.
- 404 Decker, H., Ryan, M., Jaenicke, E., Terwilliger, N., 2001. SDS-induced phenoloxidase
405 activity of hemocyanins from *Limulus polyphemus*, *Eurypelma californicum*, and *Cancer*
406 *magister*. *J. Biol. Chem.* 276, 17796-17799.
- 407 Dicko, M., Hilhorst, R., Gruppen, H., Laane, C., van Berkel, W., Voragen, A., 2002.
408 Zymography of monophenolase and o-diphenolase activities of polyphenol oxidase. *Anal.*
409 *Biochem.* 306, 336-339.
- 410 Eggert, C., Temp, U., Eriksson, K., 1996. The lignolytic system of the white rot fungus
411 *Pycnoporus cinnabarinus*: Purification and characterization of the laccase. *App. Env.*
412 *Microbiol.* 62, 1151-1158.
- 413 Eibl, J., Abdallah, Z., Ross, G., 2010. Zinc–metallothionein: a potential mediator of
414 antioxidant defence mechanisms in response to dopamine-induced stress. *Can. J. Physiol.*
415 *Pharmacol.* 88, 305–312.
- 416 Fan, T., Li, M., Wang, J., Yang, L., Cong, R., 2009. Purification and characterization of
417 phenoloxidase from *Octopus ocellatus*. *Acta Biochim. Biophys. Sin.* 41, 865-872.
- 418 Fenoll, L., Rodriguez-Lopez, J., Garcia-Molina, F., Garcia-Canovas, F., Tudela, J., 2002.
419 Unification for the expression of the monophenolase and diphenolase activities of tyrosinase.
420 *Int. U. Biochem. Mol. Biol. Life* 54, 137-141.

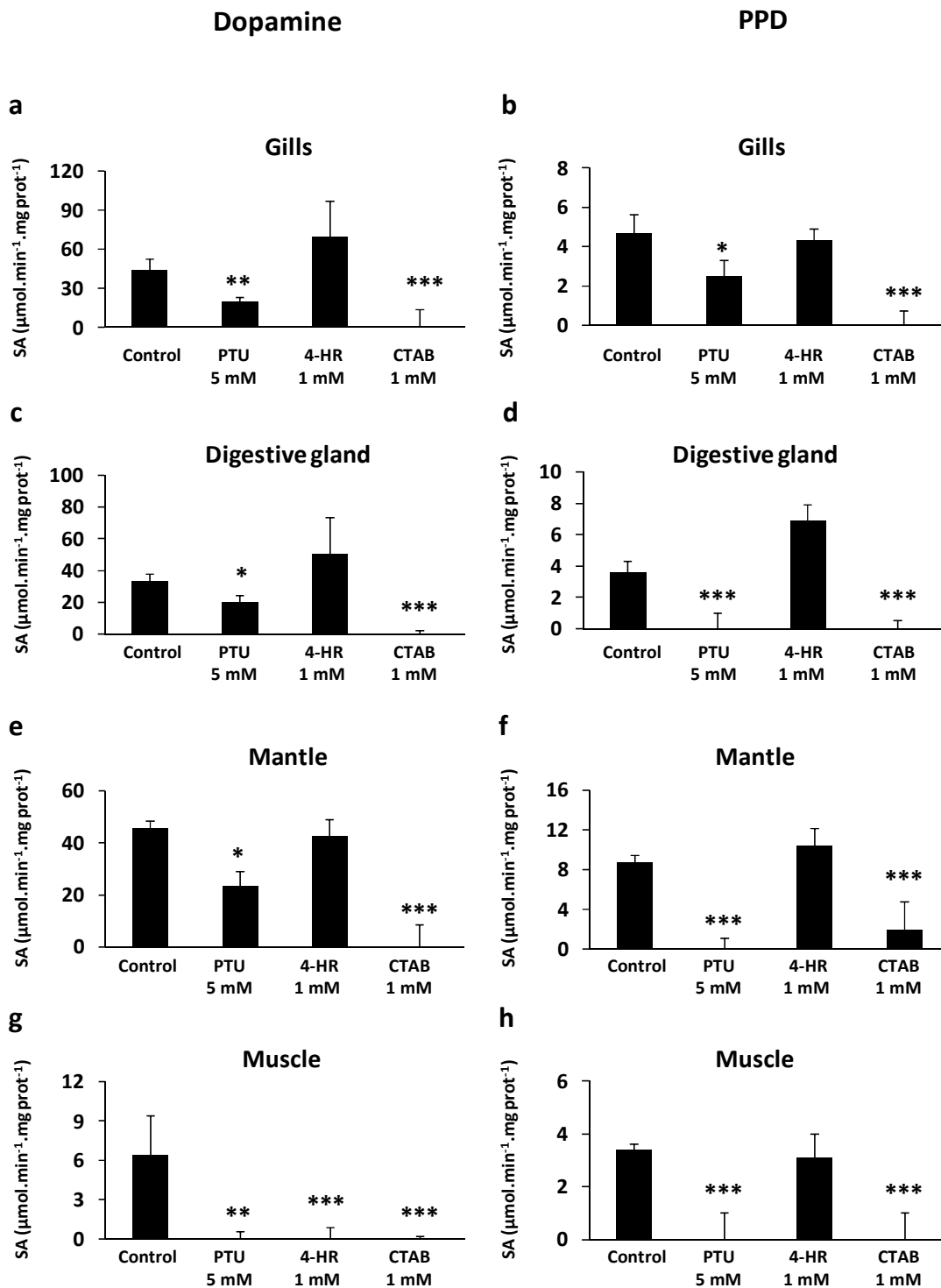
- 421 Gagnaire, B., Thomas-Guyon, H., Renault, T., 2004a. *In vitro* effects of cadmium and
422 mercury on Pacific oyster, *Crassostrea gigas* (Thunberg), haemocytes. Fish Shellfish
423 Immunol. 16, 501-512.
- 424 Gagnaire, B., Renault, T., Thomas-Guyon, H., 2004b. *In vitro* and *in vivo* effects of mercury
425 on haemocytes of Pacific oyster, *Crassostrea gigas* (Thunberg): development of techniques
426 evaluating estuarine pollution. In: Vème Congrès International de Limnologie-
427 Océanographie, Paris, France.
- 428 Hellio, C., Bourgougnon, N., Le Gal, Y., 2000. Phenoloxidase (E.C. 1.14.18.1) from *Mytilus*
429 *edulis* byssus gland: purification, partial characterization and application for screening
430 products with potential antifouling activities. Biofouling 16, 235-244.
- 431 Hellio, C., Bado-Nilles, A., Gagnaire, B., Renault, T., Thomas-Guyon, H., 2007.
432 Demonstration of a true phenoloxidase activity and activation of a ProPO cascade in Pacific
433 oyster, *Crassostrea gigas* (Thunberg) *in vitro*. Fish Shellfish Immunol. 22, 433-440.
- 434 Jaenicke, E., Decker, H., 2003. Tyrosinases from crustaceans form hexamers. Biochem. J.
435 371, 515-523.
- 436 Jordan, P., Deaton, L., 2005. Characterization of phenoloxidase from *Crassostrea virginica*
437 hemocytes and the effect of *Perkinsus marinus* on phenoloxidase activity in the hemolymph
438 of *Crassostrea virginica* and *Geukensia demissa*. J. Shellfish Res. 24, 477-482.
- 439 Laemmli, U. K., 1970. Cleavage of structural proteins during the assembly of the head of
440 bacteriophage T4. Nature 227, 680-685.
- 441 Liu, G., Yang, L., Fan, T., Cong, R., Tang, Z., Sun, W., Meng, X., Zhu, L., 2006. Purification
442 and characterization of phenoloxidase from crab *Charybdis japonica*. Fish Shellfish Immunol.
443 20, 47-57.

- 444 Luna-Acosta, A., Rosenfeld, E., Amari, M., Fruitier-Arnaudin, I., Bustamante, P., Thomas-
445 Guyon, H., 2010a. First evidence of laccase activity in the Pacific oyster *Crassostrea gigas*.
446 Fish Shellfish Immunol. 28, 719-726.
- 447 Luna-Acosta, A., Bustamante, P., Godefroy, J., Fruitier-Arnaudin, I., Thomas-Guyon, H.,
448 2010b. Seasonal variation of pollution biomarkers to assess the impact on health status of
449 juvenile Pacific oysters *Crassostrea gigas* exposed *in situ* Env. Sci. Pol. Res. 17, 999-1008.
- 450 Martinez-Alvarez, O., Montero, P., Gomez-Guillen, C., 2008. Evidence of an active laccase-
451 like enzyme in deepwater pink shrimp (*Parapenaeus longirostris*). Food Chem. 108, 624-632.
- 452 Maruyama, N., Etoh, H., Sakata, K., Ina, K., 1991. Studies on phenoloxidase from *Mytilus*
453 *edulis* associated with adhesion. Agric. Biol. Chem. 55, 2887-2889,
- 454 Nagai, K., Yano, M., Morimoto, K., Miyamoto, H., 2007. Tyrosinase localization in mollusc
455 shells. Comp. Biochem. Physiol. B 146, 207-214.
- 456 Naraoka, T., Uchisawa, H., Mori, H., Matsue, H., Chiba, S., Kimura, A., 2003. Purification,
457 characterization and molecular cloning of tyrosinase from the cephalopod mollusk,
458 *Illex argentinus*. Eur. J. Biochem. 270, 4026-4038. Paranjpe, P., Karve, M., Padhye, S.,
459 2003. Characterization of tyrosinase and accompanying laccase from *Amorphophallus*
460 *campanulatus*. Indian J. Biochem. Biophys. 40, 40-45.
- 461 Perdomo-Morales, R., Montero-Alejo, V., Perera, E., Pardo-Ruiz, Z., Alonso-Jiménez, A.,
462 2007. Phenoloxidase activity in the hemolymph of the spiny lobster *Panulirus argus*. Fish
463 Shellfish Immunol. 23, 1187-1195.
- 464 Prota, G., Ortonne, J., Voulot, C., Khatchadourian, C., Nardi, G., Palumbo, A., 1981.
465 Occurrence and properties of tyrosinase in ejected ink of cephalopods. Comp. Biochem.
466 Physiol. B 118, 463-469.
- 467 Renwrantz, L., Schmalmack, W., Redel, R., Friebel, B., Schneewei, H., 1996. Conversion of
468 phenoloxidase and peroxidase indicators in individual haemocytes of *Mytilus edulis*

- 469 specimens and isolation of phenoloxidase from haemocyte extract. *Comp. Biochem. Physiol.*
470 B 165, 647-658.
- 471 Sanchez-Ferrer, A., Rodriguez-Lopez, J.N., Garcia-Canovas, F., Garcia-Carmona, F. 1995.
472 Tyrosinase: a comprehensive review of its mechanism. *Biochim. Biophys. Acta* 1247, 1-11.
- 473 Smith, P., Khron, R., Hermanson, G., Mallia, A., Gartner, F., Provanzano, M., Fujimoto, E.,
474 Goeke, N., Olson, B., Goeke, N., Olson, B., Klenk, D., 1985. Measurement of a protein using
475 bicinchoninic acid. *Anal. Biochem.* 150, 76-85.
- 476 Solomon, E., Sunduran, U., Machonkin, T., 1996. Multicopper oxidases and oxygenases.
477 *Chem. Rev.* 96, 2563-2606.
- 478 Thurston, C., 1994. The structure and function of fungal laccases. *Soc. Gen. Microbiol.* 140,
479 19.
- 480 van Doorn, W., Vaslier, N., 2002. Wounding-induced xylem occlusion in stems of cut
481 chrysanthemum flowers: roles of peroxidase and catechol oxidase. *Postharvest Biol.*
482 *Technol.* 26, 275-284.
- 483 Waite, J., 1976. Calculating extinction coefficients for enzymatically produced o-quinones.
484 *Anal. Biochem.* 75, 211-218.
- 485 Waite, J., Wilbur, K., 1976. Phenoloxidase in the periostracum of the marine bivalve
486 *Modiolus demissus* Dillwyn. *J. Exp. Zool.* 195, 359-367.
- 487 Walker, J., McCallion, R., 1980. The selective inhibition of *ortho*- and *para*-diphenol
488 oxidases. *Phytochemistry* 19, 373-377.
- 489 Williamson, P., 1997. Laccase and melanin in the pathogenesis of *Cryptococcus neoformans*.
490 *Front. Biosc.* 2, 99-107.
- 491 Zar, J., 1984. *Biostatistical analysis*. Prentice-Hall, New Jersey.
- 492 Zavarzina, A.G., Zavarzin, A.A., 2006. Laccase and tyrosinase activities in lichens.
493 *Microbiology.* 75, 546-556.

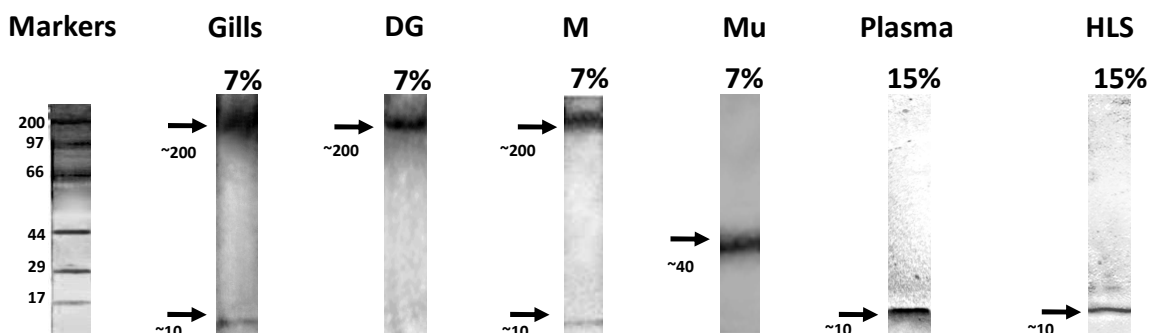
494 Zufelato, M.S., Lourenço, A.P., Simões, L.P., Jorge, J.A., Bitondi, M.M., 2004.
495 Phenoloxidase activity in *Apis mellifera* honey bee pupae, and ecdysteroid-dependent
496 expression of the prophenoloxidase mRNA. *Insect Biochem. Mol. Biol.* 34, 1257-1268.
497

498 **Figure 1. Inhibition of phenoloxidase-like activity in precipitated protein fractions from**
 499 **the gills, digestive gland, mantle and muscle.** Both dopamine (a, c, e, g) and PPD (b, d, f, h)
 500 were used as substrates. ‘Control’ corresponds to the condition without inhibitor. PO inhibitor
 501 concentrations correspond to final concentrations in the assay. Mean \pm SD $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{mg}$
 502 prot^{-1} , n = 9; *statistical difference of $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$, respectively.

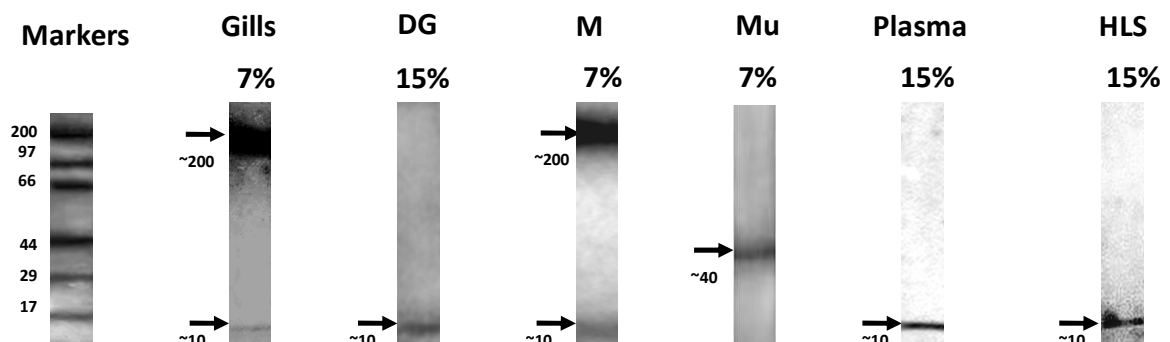


504 **Figure 2. Zymograms for the determination of phenoloxidase activities from different *C.***
 505 ***gigas* tissues after partial purification by precipitation with 60% of sulfate ammonium**
 506 **saturation for the gills, digestive gland (DG), mantle (M), muscle (Mu) and plasma or**
 507 **with crude sample for the haemocyte lysate supernatant (HLS). Samples were run on 7%**
 508 **or 15% analytical SDS-PAGE gels and stained with (a) 100 mM dopamine and 5 mM MBTH**
 509 **and with (b) 100 mM PPD. Gills, DG, M, Mu, plasma and HLS were loaded at a protein**
 510 **concentration of 76, 76, 57, 40, 57 and 1.55 μ g, respectively. The arrows indicate the bands**
 511 **showing PO-like activity. Their estimated molecular weights (in kDa) are indicated below the**
 512 **arrows.**
 513

a. Stained with 100 mM dopamine/5 mM MBTH



b. Stained with 100 mM PPD



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517 **Table 1. Total phenoloxidase-like activity (Mean \pm standard deviation, $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{mg}$**
518 **$\text{prot}^{-1}\cdot\text{g}$ fresh weight, n=3) from different oyster body tissues with dopamine and PPD as**
519 **substrates.**

520

Body tissue	Total phenoloxidase-like activity	
	Dopamine	PPD
Gills	1.8 \pm 0.3	0.2 \pm 0.0
Digestive gland	3.0 \pm 0.4	50.4 \pm 4.3
Mantle	0.9 \pm 0.1	0.2 \pm 0.0
Muscle	0.5 \pm 0.2	0.3 \pm 0.1

521

522 **Table 2 Phenoloxidases from molluscs: characteristics reported in the literature**

Vernacular name	Scientific name	Molecular weight (kDa)	Localisation	Reference
Freshwater snail	<i>Biomphalaria glabrata</i>	35	Egg mass	Bai et al. 1997
Common octopus	<i>Octopus vulgaris</i>	205	Ink	Prota et al. 1981
Ocellated octopus	<i>Octopus ocellatus</i>	153.8	Ink	Fan et al. 2009
Argentine shortfin squid	<i>Illex argentinus</i>	127.6	Ink	Naraoka et al. 2003
Common cuttlefish	<i>Sepia officinalis</i>	125	Ink	Prota et al. 1981
European squid	<i>Loligo vulgaris</i>	135	Ink	Prota et al. 1981
Blue mussel	<i>Mytilus edulis</i>	381, 316 49, 135, 260	Haemocyte lysate supernatant (HLS) Foot gland	Renwranz et al. 1996 Maruyama et al. 1991
Ribbed mussel	<i>Modiolus demissus</i>	70	Periostracum	Waite and Wilbur (1976)
Manila clam	<i>Ruditapes philippinarum</i>	76.9	Haemolymph	Cong et al. 2005
Japanese pearl oyster	<i>Pinctada fucata</i>	43, 49	Prismatic shell layer	Nagai et al. 2007
Eastern oyster	<i>Crassostrea virginica</i>	133	Haemocyte membranes supernatant (HMS)	Jordan and Deaton 2005

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