

# Epistasis between neurochemical gene polymorphisms and risk for ADHD

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Running Title: Exploration of epistasis in ADHD candidate genes

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# Abstract

A number of genes with function related to synaptic neurochemistry have been genetically associated with Attention-Deficit/Hyperactivity Disorder. However, susceptibility to the development of common psychiatric disorders by single variants acting alone, can so far only explain a small proportion of the heritability of the phenotype. It has been postulated that the unexplained "dark heritability" may at least in part be due to epistatic effects, which may account for the small observed marginal associations, and the difficulties with replication of positive findings. We undertook a comprehensive exploration of pair-wise interactions between genetic variants in 24 candidate genic regions involved in monoaminergic catabolism, anabolism, release, re-uptake and signal transmission, in a sample of 177 parent-affected child trios using a case-only design and a casepseudocontrol design using conditional logistic regression. Markerpairs thresholded on interaction odds ratio and p-value are presented. We detected a number of interaction odds ratios greater than 4.0, including an interesting correlation between markers in the ADRA1B and DBH genes in affected individuals, and several further interesting,

but smaller effects. These effects are no larger than you would expect by chance under the assumption of independence of all pair-wise relations, however independence is unlikely. Furthermore, the size of these effects is of interest and attempts to replicate these results in other samples are anticipated.

## Keywords

ADHD, epistasis, candidate gene

## Introduction

The heritability of Attention Deficit/Hyperactivity Disorder (ADHD) is well established, and genetic association data has been reviewed and meta-analysed recently by Gizer and colleagues<sup>1</sup>. The detection of DNA variants which increase risk for this disorder is important for biochemical and pharmacological research into this disorder, and may permit the facilitation of diagnosis, or refinement of the phenotype on the basis of a biological marker. Specifically, association between monoaminergic genes and ADHD is an active area of investigation, stimulated principally by the mode of action of current pharmacological treatments for the disorder, primary among which is methylphenidate, a known dopamine and noradrenaline re-uptake inhibitor. Associations between variants at the dopamine, serotonin and noradrenaline transporter and receptor genes, with ADHD, have been investigated, and meta-analyses support the presence of ADHD risk alleles with small to moderate effects at the DRD4, DRD5 and SLC6A3 genes<sup>1-3</sup> in candidate gene studies, and modest evidence at ADRA1A, TPH2 and DDC from a recent meta-analysis of genome-wide association studies<sup>4</sup>.

The influence of higher-order interactions between genes has been posited for common, complex genetic disorders on the basis of biological plausibility, and the presence of interactions influencing gene expression and protein function both in model organisms and in human disease susceptibility<sup>5-6</sup>. In the case of ADHD the most likely known risk genes inhabit the same biochemical neighbourhood, in that the functions of each gene product are related directly via proteinprotein interaction, or indirectly via regulatory pathways. Therefore there are grounds for prior supposition that common genetic variants in several risk genes may act synergistically to influence disease risk. Methods for detecting interaction, or epistasis, in case-control samples have received much attention recently<sup>9,10</sup>, particularly in the rapidly developing areas of machine learning. Methods for family-based samples are less well developed - usually adaptations of case-control methods (e.g. using matched case-pseudo-control samples). However, in addition to large family based samples being used in the latest generation of gene-mapping studies (e.g. the Autism Genome Project<sup>11</sup>; IMAGE:<sup>12</sup>), family-based samples were extensively used in candidate gene experiments prior to the emergence of genome-wide association studies. These are excellent genotype resources for exploring epistasis as these studies have genotyped a relatively small number of genetic polymorphisms within strong candidate genes.

The aim of this study was to use a subset of genetic markers which we expect to be enriched for true involvement in pathogenesis of ADHD, to: 1) attempt to detect any large epistatic effects which may exist; 2) explore epistatic effect sizes between candidate genes, for prioritisation of further investigation in this sample, and replication in other samples. To this end a limited number of variants in a range of strong candidate genes for ADHD were selected (see Methods). As this study was a focused exploration of interaction, we elected to use two principal techniques to analyse the data thoroughly for various forms of interaction effects. Thus, firstly, a case-only design was adopted in order to detect association between unlinked genes in affected individuals. This allows calculation of an "interaction odds ratio", equivalent to the interaction relative risk under the assumption of independence of the genes in the general population<sup>13</sup>. Secondly, a similar model was fitted to binary affection status by logistic regression<sup>14</sup>. This method examines patterns of transmission of alleles from each of the markers in a pair, thus is robust to bias arising from population stratification.

## <u>Methods</u>

#### Recruitment and Sample description

One hundred and seventy seven children and their families participated in the study including 5 families with two affected children. The families were recruited from child guidance clinics, ADHD support groups, and via direct clinician refferal. A stringent diagnosis of DSM-IV and/or ICD10 ADHD was made as described by Kirley et al<sup>15</sup>, using the Child and Adolescent Psychiatric Assessment (CAPA) and the Teacher Telephone Interview. Exclusion criteria included epilepsy, fragile X syndrome, foetal alcohol syndrome, pervasive developmental disorder, Tourette's syndrome, psychosis or IQ <70. Affected children met research criteria for ICD-10 hyperkinetic disorder (HKD: 100 children) and/or DSM-IV ADHD (combined: 112, inattentive: 21, or hyperactiveimpulsive: 11 subtypes), and/or a clinical diagnosis of ADHD not meeting full research criteria, but deemed to lie on the broad ADHD phenotypic spectrum. Therefore the present sample consisted of 172 parent - affected child trios.

#### Marker selection and Genotyping

Gene selection began with a set of candidate genes previously typed in this sample, and was expanded to include several genes found to be associated in other samples and to genes within neurochemical systems plausibly linked to ADHD (table 1; supplemental tables S1 and S2). This study was limited to 25 autosomal genes (CES1 and SLC6A2 were treated as one unit due to physical proximity). Markers within these genes were either (i) previously characterised in this sample; (ii) previously associated in other samples; (iii) selected due to known or suspected functional relevance (on gene or protein expression, protein sequence, and including presence in a known promoter region and lying within 500 base pairs of an exonic boundary); or (iv) in order to tag the remainder of the gene  $(r^2 \ge 0.8)$  insofar as gene size and marker density made possible. Markers and genes were placed on the human genome 18 physical map (hg18; NCBI build 36.3) using dbSNP (http://www.ncbi.nlm.nih.gov/SNP/), or by linear interpolation from flanking markers.

A total of 179 markers were genotyped. Single Nucleotide Polymorphisms (SNPs) were genotyped commercially at K-Bioscience using a competitive allele-specific polymerase chain reaction (PCR) - KASPar (K-Bioscience, Hoddesdon, UK; personal communication), or Taqman chemistries (Applied Biosystems, California, USA). Three markers, at the DBH (rs1611115 and rs6271) and COMT (rs4680) genes were genotyped in-house with Taqman assays on a 7900HT sequence analyzer (Applied Biosystems). Microsatellite and minisatellite markers were typed inhouse as described previously; these included the markers in SLC6A3 and DRD5<sup>16</sup>, DRD4<sup>17</sup>, TH<sup>18</sup> and SLC6A4<sup>19</sup>.

Multiallelic (> 2 alleles) markers were collapsed into 2 categories on the basis of 1) bimodality of the allele frequency histogram in founder individuals, 2) so as to approximate equifrequency of the down-coded "alleles". This might be expected to reflect the genetic history of the microsatellite. We did not use previously associated alleles at these markers as a basis for recoding<sup>20</sup>.

Only autosomal genetic markers with a minor allele frequency in founders of >0.05 were retained; only markers with over 80% genotype rate were included; Pedcheck<sup>21</sup> was used to detect Mendelisation errors, genotypes showing errors were removed for the entire trio; finally, markers which showed extreme deviation from Hardy-Weinberg equilibrium in the parents were excluded due to the possibility of miscalled genotypes. When markers within the same gene were in near perfect linkage disequilibrium in founders  $(r^2 \ge 0.98)$ , the one with least complete genotyping was discarded.

# Data Analysis

An allelic case-only test for pair-wise interaction of SNPs<sup>22</sup> was performed, using the proband from each pedigree. In order to exclude the effect of linkage disequilibrium between SNPs, all intragenic SNP pairs, and all intergenic SNP pairs separated by less than 500 kilobases were excluded. Tables with any zero cell count were discarded. A chi-square test (1 d.f.) was performed on the 2x2 tables, and an odds ratio estimate was calculated.

The null distribution of the odds ratio for each pair of SNPs examined was simulated by fixing genotypes at the first marker, and permuting genotypes at the second. Two-sided p-values for the odds ratios were estimated from 50 000 permutations, or from 1 000 000 permutations for marker pairs which showed an initial empirical p-value less than 0.001. 95% confidence intervals for the interaction Odds Ratios were generated from 1 000 bootstrap samples, for each marker pair. A global, family-wise p-value was also calculated from the proportion of the 50 000 permutations in which an odds ratio was exceeded in any of the pair-wise tests.

Interactions were also tested in a logistic regression model, implemented on a matched case-pseudocontrol sample<sup>14</sup>. Fifteen pseudocontrols were generated for each trio, for each marker pair, from all combinations of parental two-marker genotypes. Two models were fitted to the data in R (http://cran.r-project.org) with the *coxph* function from the *survival* package, using robust variance estimation, stratified on a family ID variable (equivalent to a conditional logistic regression). An interaction parameter was tested by contrasting the model with a nested model containing main effects alone using a likelihood ratio chi-square test with one degree of freedom. Only additive allelic effects were examined. For each marker pair, perfectly collinear variables were handled by removing one from the model. A pseudo- $\mathbb{R}^2$  for each model representing how much of the variance in the dependent variable (affection status) is explained by the genetic terms was also calculated.

#### <u>Results</u>

Twenty-seven markers were excluded (see Methods) due to low genotyping rate, 4 due to low minor allele frequency in parents, and 3 due to extreme deviation from Hardy-Weinberg equilibrium in parents (goodness-of-fit Chi-square, p < 0.00033 = 0.05/177). Interaction between one pair of genes (CES1-SLC6A2) was not examined due to physical proximity <500 kbp. Four markers were removed due to near perfect linkage disequilibrium in parents with neighbouring markers ( $r^2$ = 0.98 - 1.00). Table 1 shows genic positions, marker number and spacing. The Chi-square test statistic had very low correlation with the intermarker distance, for syntenic markers (supplemental figure S3).

The remaining 141 markers had a mean missing genotype rate of 0.075 in affected individuals. We estimated from single-marker missingness that 82.4% (142) of cases would be genotyped at both markers of a markerpair, on average across all between-gene pairs. The observed average was 85.3%. Mean minor allele frequency in founders was 0.304 (range: 0.051 - 0.498). The mean expected frequency of minor allele - minor allele conjunctions over all marker pairs in cases was 0.0874 (range: 0.0023 - 0.2468), and approximately 64% of expected minor allele minor allele pair frequencies were greater than 0.05. The observed mean minor allele - minor allele frequency in cases was 0.0871, with 71.0% being greater than 0.05. A total of 9 269 SNP-SNP case-only interaction tests were performed within the 276 gene pairs. After exclusion of sparse tables 9 262 tests (99.9%) remained. A QQ plot of the chi-square statistic showed some deflation of the statistic relative to the expected 1 degree of freedom chi-squared distribution, with a "stepped" appearance, most likely due to non-independence of tests (supplemental figure S2). Results are presented sorted by the odds ratio estimate, or by p-value (table 2 and supplemental table S3).

The largest odds ratio observed was 7.27 between rs129882 in Dopamine Beta-Hydroxylase (DBH), and rs2283135, a relatively rare variant in the SLC18A2 gene. Scaled to lie above 1, nine estimated interaction odds ratios exceeded 4.0, of which five included the rarest variant in the study: rs916455 in DRD4, which had a minor allele frequency of 3% in cases. For most of these the bootstrap confidence interval calculation failed at either the upper or lower bound due to a sparse table. A total of 20 marker pairs showed an Odds Ratio > 3.0, including 12 pairs to which rs916455 contributed.

The smallest p-value observed was  $5.4 \ge 10^{-5}$  between rs11953285 and rs1611115, in the ADRA1B and DBH genes, respectively (Table 2A). We note that no global p-value dipped below 0.97, between SLC18A2 and

DBH, and all marker-pairs excluded from Table 2 showed a global pvalue = 1.

The conditional logistic regression approach yielded maximum pseudo- $R^2$  values of 0.033 (partial model) to 0.045 (full model), indicating a small fraction of the variance in affection status could be accounted for by the interaction terms. Correlation with the top case-only test results was strong (Spearman rho for p-values = 0.776); over 80% of interaction Odds Ratios were in the same direction, i.e. with the same allele (supplemental figure 5). Table 2 and supplmentary table S3 show the results from the logistic regression.

#### Discussion

From Pierce and Ahsan<sup>13</sup>, we estimate that a single pair-wise case-only analysis with our sample size may have had good power to detect relatively large interaction relative risks (>2), with interacting minor allele frequencies of approximately 0.3. Considering a range of minor allele frequencies from 0.05 to 0.5, the least power would be obtained in the case where interacting loci both have risk alleles with frequency of 0.05. In this case 80% power would be obtained for an odds ratio of 4.86 (p < 0.05). If one locus had risk allele frequency of 0.05, and the other locus 0.5, 80% power is obtained at OR = 2.24.If both risk alleles had frequency 0.5, an odds ratio of 2.5 could be detected with 80% power at a significance level of 5.1 x  $10^{-6}$ . Effects of these magnitudes are large by the standards seen to date in genetic association studies of ADHD, however with the large number of tests between all markers in the candidate genes we examined, we did not expect to be able to achieve an acceptable gobal false positive level, nor in fact was this acheived. Therefore, we leave these findings as exploratory, to be used for prioritisation of future studies in independent samples. In a more detailed exploration of the interaction model (using, for example a four-parameter model of additive x additive, additive x dominance, dominance x additive, dominance x dominance) it was universally the case that unreliable parameter estimates were obtained for the top marker-pair interactions, and the model likelihoods did not converge. This may simply reflect our modest sample size.

The presence of markers with a low MAF in the top interactions ranked by the odds ratios is expected, due to unreliability of odds ratios derived from tables with small cell sizes as indicated by wide confidence intervals, and as reflected in the p-values for these tests (see supplemental figure 4). When the Odds Ratio is sensitive to small observed cell counts, the empirical p-value is of use to judge whether a particularly large estimate would be likely to occur by chance alone.

It is probable that several factors exist which preclude the independence of the individual pair-wise interaction tests performed with either the case-only design, or the regression. These include linkage disequilibrium within genes to due ancestral co-segregation. We hope to have robustly excluded any effect due to LD between syntenic markers by using a severe minimum distance between genes of 500 kilobasepairs. However, adjacent genes frequently share features at a functional level and in expression profile<sup>23</sup>, and a large minimum distance may exclude some true functional interactions. Other potential confounders may also cause correlation between genes. These include, but are probably not limited to, higher-order interaction effects on disease, on survival to age of sampling, and membership of cryptic subpopulations. We have not tested the first of these due to sample size limitations, and to avoid a large increase in the number of tests and increased computational burden. Epistatic effects on survival to sampling cannot be excluded, however no reports of marginal effects at these genes exist in the literature. Unfortunately this sample was not genotyped at sufficient loci to allow an estimate of population stratification, or to allow a genomic correction to be

applied, however, the sample is ethnically homogeneous therefore we do not expect any severe bias. This is supported by the strong correlation between the case-only and the family trio-based logistic regression results, which is not susceptible to the cryptic population structure in the sample.

As this study was intended to be hypothesis-generating, no threshold defining statistical significance was enforced. Formal statistical evaluation of the evidence for interaction at the gene pair level is not trivial, as it would require consideration of both the number of markers within each gene and the extent of linkage disequilibrium, as well as higher order effects. Methods such as that of Chatterjee and colleagues<sup>10</sup> are of interest, but are not yet applicable to familybased or case-only designs. The LD patterns within each gene are relatively low (supplemental figure 1), and use of a gene-based permutation procedure which controls for LD will be important when a threshold for statistical significance is applied.

The most studied gene - gene interaction in ADHD is for a joint effect of the dopamine receptor 4 gene (DRD4) and the dopamine transporter gene (SLC6A3) VNTR polymorphisms, with some suggestive results<sup>24-26</sup>. We were unable to demonstrate strong effects between DRD4 and SLC6A3. There is also a report in the literature of an interaction between the Noradrenaline transporter (SLC6A2 / NET1) and the catechol-Omethyltransferase (COMT) genes affecting quantitative traits derived from an ADHD self-report scale<sup>27</sup>. Pairs of markers from these genes are amongst our top results, ranked by p-value: COMT marker rs737866, when paired with three SLC6A2 markers had Odds Ratios of 1.89 (95% CI: 1.27-2.83; p = 0.000866) with rs36017, 1.66 (1.14-2.41; p = 0.0085) with rs1345429, and 1.76 (1.23 - 2.68; p = 0.00128) with rs5569. These three SLC6A2 variants are relatively highly correlated with each other (r<sup>2</sup> from 0.38 to 0.90; all D' > 0.97), which may explain the correlation of odds ratios (see Supplmentary materials, figure S1(0)).

It has been hypothesised that functional disruption of serotoninrelated genes (SLC6A4, HTR1A, HTR1B) may act synergistically at the synapse to increase extracellular serotonin levels<sup>28</sup>. We did observe modest interaction effects (table 2) between two of these genes -SLC6A4 and HTR1B (we did not examine HTR1A). While this did not stand out as a finding in the case-only analysis, the logistic regression showed a large interaction odds ratio estimate of 3.24 (suppl. table S3).

Two pairs of interactions with the smallest p-values stand out. ADRA1B

showed a minimum p-value of 0.00005 with DBH (odds ratio = 4.56). DBH converts dopamine (DA) to noradrenaline (NA), and in particular the SNP rs1611115 (-1021C $\rightarrow$ T) in the DBH gene promoter has been shown to have a strong effect on plasma enzyme activity<sup>29</sup>. Here the "low-activity" C allele (implying lack of NA and accumulation of DA) shows a synergistic effect with an ADRA1B allele to increase risk. An interaction at or near this SNP could potentially explain some of the heterogeneous marginal association findings at this gene<sup>1</sup>. There is a dearth of well-targetted studies on the ADRA1B gene product, and its central neural functions are unknown.

The second pair involves HTR2A with ADRA1A, showing a minimum p-value of 0.0008, with an odds ratio = 1.96 (see table 2, supplemental table S3). Linking these results, one publication has suggested that a HTR2A-mediated pathway may compensate for ADRA1B inactivity in a mouse hyperactivity model<sup>30</sup>, raising the possibility that the products of these two genes may have some complementary function in a dopaminergic regulation that is also targetted by amphetamines (including methylphenidate).

Of course, a reasonable disease mechanism may be deduced from any subset of interactions that one selects, especially since these genes and polymorphisms were preselected for this very reason - this discussion on the potential functional relevance of these results is purely speculative, especially in the absence of known functional effects of the markers studied. However, these results have implicated a number of gene-pairs with interesting effect sizes, and suggestive of a functionally consistent set of interactions which may converge on a common neurochemical phenotype, and which should be prioritised in future studies.

While the detection of a statistical interaction may imply a biological (physico-chemical) interaction between the genes and/or gene products or via regulatory mechanisms, proof and further elucidation require extensive cell and molecular experiments. We also note that the statistical tests performed were exploratory, and we place greater emphasis on estimates of the effect sizes demonstrated here; which must, however, must await replication in a larger, independent sample. Here we were able to primarily investigate interaction in the form of a deviation from multiplicative effect on the risk ratio at pairs of markers in or near strong candidate genes. Interactions may lead to a marginal effect on disease risk at either or both of the loci, however such marginal effects may appear considerably smaller than the true, unmeasured interaction, and therefore on a genome-wide scale power to detect such effects will be low, while controlling the family-wise error rate. For this reason we feel that directed studies of interaction such as the present one may be fruitful, when small but suggestive marginal effect sizes at the genes of interest have been demonstrated. This exploration of interactions between a select set of candidate genes in our sample of ADHD cases has found some interesting results, justifying such further study.

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<u>Conflict of Interest</u> The authors declare no conflict of interest.

Supplementary information is available at the European Journal of Human Genetic's website.

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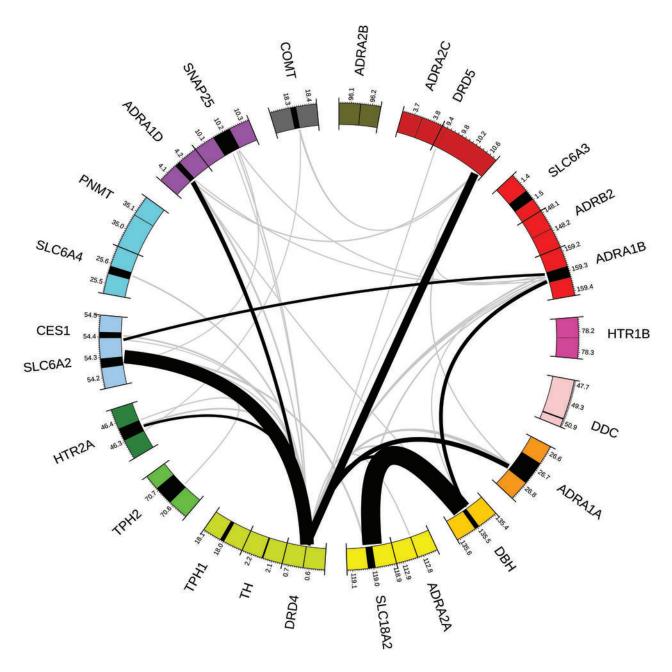
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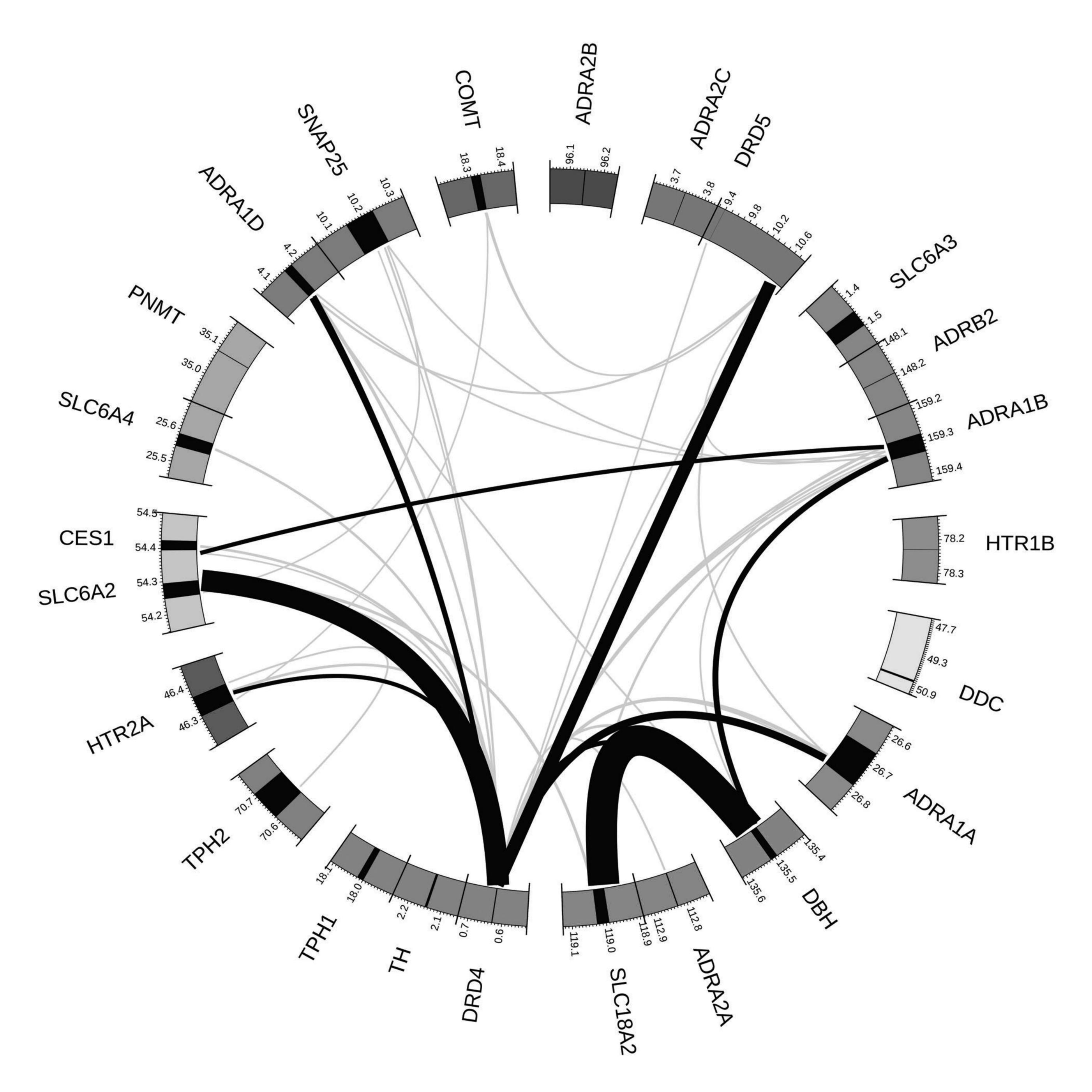
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## Titles and legends to figures

Figure 1. Visual representation of the interaction relative risk estimates for the top 0.5% of variant-variant interactions. <u>Legend:</u> chromosomes are colour-coded; gene spans in black. Note that the regions around the DDC and DRD5 genes are rescaled reltive to other genes (1:17 and 1:5 respectively) due to distance to the furthest marker; interaction links are shaded by quantile: grey = 99.5<sup>th</sup> -99.9<sup>th</sup>, black = >99.9<sup>th</sup>; arc widths are proportional to the interaction Odds Ratio.





		Position	1 (b.p.)		Mean marker spacing (bp)	
Gene	Chr	start	stop	#markers		
ADRA2B	2	96, 142, 350	96, 145, 615	1	-	
ADRA2C	4	3, 738, 094	3, 740, 051	1	-	
DRD5	4	9, 392, 356	9, 394, 731	3	663, 473	
ADRA1B	5	159, 276, 318	159, 332, 595	11	4, 433	
ADRB2	5	148, 186, 349	148, 188, 381	2	33	
SLC6A3	5	1, 445, 910	1, 498, 538	2	17, 933	
HTR1B	6	78, 228, 667	78, 229, 839	5	470	
DDC	7	50, 493, 628	50, 600, 648	2	3, 744, 601	
ADRA1A	8	26, 661, 584	26, 778, 839	14	8, 550	
DBH	9	135, 491, 306	135, 514, 287	11	2, 315	
ADRA2A	10	112, 826, 780	112, 830, 652	3	1,859	
SLC18A2	10	118, 990, 706	119, 027, 085	5	9, 173	
DRD4	11	627, 305	630, 703	5	1,011	
TH	11	2, 141, 735	2, 149, 611	1	-	
TPH1	11	17, 998, 660	18, 018, 911	3	3, 526	
TPH2	12	70, 618, 893	70, 712, 488	12	6, 266	
HTR2A	13	46, 305, 514	46, 369, 170	8	8, 701	
SLC6A2	16	54, 248, 057	54, 295, 201	9	6, 526	
CES1	16	54, 394, 265	54, 424, 576	3	22, 743	
PNMT	17	35, 078, 033	35, 080, 254	3	3, 093	
SLC6A4	17	25, 547, 506	25, 587, 080	5	9, 879	
ADRA1D	20	4, 149, 278	4, 177, 659	11	2, 359	
SNAP25	20	10, 147, 477	10, 236, 065	16	6,019	
COMT	22	18, 309, 263	18, 337, 496	5	6, 396	

based on human genome build hg18.

Locus 1			Locus 2			Case-only test		Regression test		
gene	marker	chr:pos	gene	marker	chr:pos	Odds Ratio (95% C.I.)	p-value (1)	Odds Ratio	p-value	full model pseudo-R <sup>2</sup> (3)
ADRA1B	rs11953285	5:159, 324, 38 9	DBH	rs1611115	9:135,490,336	4.56 (2.10- 20.02)	0.000054	4.83	0.0004	0.018
SLC6A3	Intron8 VNTR	5:1,464,855	SNAP25	rs362562	20:10, 191, 186	2.41 (1.46- 4.23)	0.000558	1.60	0.0871	0.008
ADRA1B	rs7737796	5:159, 302, 00 7	SNAP25	rs363006	20:10,228,083	2.43 (1.50- 4.37)	0.000711	2. 27	0.0102	0.016
ADRA1A	rs472865	8:26, 754, 388	DRD4	exon 3 VNTR	11:629,989	3.64 (1.61- 14.41	0.00100	3.80	0.0048	0.015
ADRA1B	rs6884105	5:159,281,02 1	SNAP25	rs363006	20:10,228,083	2.18 (1.38- 3.85)	0.00104	2.13	0.0215	0.008
ADRA1A	rs486354	8:26, 766, 207	DRD4	120 bp duplication	11:625,946	2.46 (1.42- 5.21)	0.00134	2.66	0.0111	0.012
DBH	rs129882	9:135, 513, 49 0	SLC18A2	rs2283135 (2)	10:118,989,92 2	7.27 (N/A)	0.00178	9.45	0.0037	0.016
DRD4	rs12720373	11:626,688	CES1	rs3815589	16:54, 384, 420	2.47 (1.38- 5.36)	0.00204	2. 21	0.0222	0.015
DRD5	D4S2928 (2)	4:10, 612, 311	DRD4	exon 3 VNTR	11:629,989	5.84 (N/A)	0.00250	3.66	0.0664	0.007
CES1	rs3815589	16:54, 384, 42 0	ADRA1D	rs4815670	20:4, 164, 864	2.03 (1.23- 3.47)	0.00314	2.03	0.0252	0.010
ADRA1A	rs472865	8:26, 754, 388	DBH	rs2797849	9:135,491,762	2.33 (1.36- 4.71)	0.00328	2.28	0.0220	0.011
ADRA1B	rs13162302	5:159,299,85 3	ADRA1D	rs6052456	20:4, 173, 573	2.67 (1.41- 6.64)	0.00410	3. 82	0.0010	0.021
DRD5	D4S2928 (2)	4:10, 612, 311	COMT	rs4680	22:18,331,271	2.60 (1.38- 5.27)	0.00416	2.04	0.0904	0.004
HTR1B	rs1228814	6:78, 230, 539	SLC6A4	rs1872924	17:25, 570, 472	2.06 (1.23- 4.09)	0.00456	3. 24	0.0068	0.016
SLC18A2	rs363279	10:119,026,6 15	SLC6A2	rs9930182	16:54, 298, 798	3.38 (1.59- 13.27)	0.00474	4. 15	0.0033	0.017

Legend:

VNTR = Variable Number of Tandem Repeats polymorphism; chr:pos = chromosome: position (base pairs from pter; hg18); odds ratio were rescaled to lie above 1, confidence intervals were derived from 1,000 bootstrap samples, "N/A" indicates that a ceiling or floor effect was observed in the bootstrap estimates, due to sparse contingency table; see supplementary materials for the full results. (1) - empirical p-value from 50,000 or 1,000,000 (in bold) Monte Carlo permutations.

- (2) Minor allele frequency < 0.1 in parents.
- (3) McFadden pseudo- $\mathbb{R}^2$  for the full model including main effects and interaction.

NB: The p-values presented are unadjusted for multiple testing; none of them are significant at p < 0.05 after correction for the number of tests.