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Molecular footprints of local adaptation in two Mediterranean conifers

Research Article

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ABSTRACT

This study combines neutrality tests and environmental correlations to identify non-neutral patterns of evolution in candidate genes related to drought stress in two closely-related Mediterranean conifers, *Pinus pinaster* Ait. and *Pinus halepensis* Mill. Based on previous studies, we selected 12 amplicons covering six candidate genes that were sequenced in a large sample spanning the full range of these two species. Neutrality tests relatively robust to demography (*DHEW* compound test and *ML-HKA* test) were used to detect selection events at different temporal scales. Environmental associations between variation at candidate genes and climatic variables were also examined. These combined approaches detected distinct genes that may be targeted by selection, most of them specific to only one of the two conifers, despite their recent divergence (< 10 Ma). An exception was *4-coumarate: CoA ligase (4cl)*, a gene involved in the production of various important secondary products that appeared to play a role in local adaptation processes of both pines. Another remarkable result was that all significant environmental correlations involved temperature indices, highlighting the importance of this climatic factor as a selective driver on Mediterranean pines. The ability to detect natural selection at the DNA sequence level depends on the nature and the strength of the selection events, on the timescale at which they occurred and on the sensitivity of the methods to other evolutionary forces that can mimic selection (e.g. demography, population structure). Using complementary approaches can help to capture different aspects of the evolutionary processes that govern molecular variation at both intra- and interspecific levels.

INTRODUCTION

Identifying candidate genes underlying genetic differences for adaptive traits can help to understand how species have adapted to their environment and to predict how they will respond to future climatic changes, which is a special concern in regions such as the Mediterranean Basin, where a substantial decrease in precipitation and a pronounced warming is expected in the near future (Petit, Hampe, and Cheddadi 2005; Giorgi and Lionello 2008). The ability to detect the footprints of natural selection in population DNA sequence samples depends on the nature and the strength of the selection events (Nielsen 2005), on the evolutionary scale at which they occur (Zhai, Nielsen, and Slatkin 2009) and on the sensitivity of the methods to other evolutionary forces that can mimic selection (e.g. demography, population structure; Biswas and Akey 2006).

Positive selection, which drives the increase in frequency of advantageous mutations, is of particular interest because it underlies local adaptation. Recent or on-going positive selection events can be detected using methods based on polymorphism within species (Biswas and Akey 2006; Zhai, Nielsen, and Slatkin 2009). Recently, powerful methods based on site- and haplotype- frequency spectra that are relatively insensitive to demography or population structure have been developed (Zeng, Shi, and Wu 2007). Detecting selection events over a wider evolutionary scale (e.g. recurrent positive selection or ancient selective sweeps), can be efficiently done using methods contrasting within species polymorphism with among species divergence, as these methods are less sensitive to assumptions regarding recombination or demography. The *HKA* (Hudson-Kreitman-Aguadé) neutrality test (Hudson, Kreitman, and Aguade 1987) and its extensions (e.g. Wright and Charlesworth 2004) are among the most

powerful tests to detect positive selection (Zhai, Nielsen, and Slatkin 2009). In addition, the action of natural selection can be reflected in statistical associations between genetic and environmental data (Manel et al. 2003; Hancock et al. 2008; Coop et al. 2009). Several studies have shown that environmental heterogeneity influences the distribution of genetic diversity across plant populations (see Savolainen, Pyhäjärvi, and Knürr 2007, for forest trees; Nakazato, Bogonovich, and Moyle 2008; Montesinos et al. 2009). Detecting clinal genetic variation along environmental gradients can thus provide evidence for the action of selection (Gram and Sork 2001; Vasemagi and Primmer 2005; Parisod and Christin 2008). Approaches that detect adaptive clinal variation are very attractive because they can directly link environmental gradients with genotypes and phenotypes. However, detecting environmental associations depends on the spatial scale examined and the associations have to be controlled for selectively neutral processes that can also generate clines (e.g. Storz 2002; Hancock et al. 2008; Coop et al. 2009; Keller et al. 2009). Using environmental association approaches in conjunction with neutrality tests helps capturing different aspects of the evolutionary processes that govern molecular variation.

In the present study, we use the combined strategy outlined above to investigate patterns of polymorphism for a set of candidate genes related to drought stress in maritime pine (also known as cluster pine, *Pinus pinaster* Ait.), and in Aleppo pine (*Pinus halepensis* Mill.). *P. pinaster* is an economically important pine native to the occidental Mediterranean Basin and the European Atlantic front (Figure 1). This species is ecologically versatile, growing in a variety of substrates, in a wide range of elevations and under a range of Mediterranean and Atlantic climate regimes (semi-arid to very humid; Table 1). *P. halepensis*, a pine closely related to maritime pine, is one of the most abundant and widespread pine species of the Mediterranean Basin (Figure 1). This species, which also has a wide ecological breadth, is well adapted to dry

conditions as well as high-intensity fire regimes (Tapias et al. 2004), which makes it a species of choice for afforestation in xeric Mediterranean areas. These two conifers, because they have extensive and partially overlapping distributions across the Mediterranean Basin, may possibly display adaptation to similar sets of contrasted environmental conditions, *i.e.* regional climate types and spatial variability. Among the key environmental factors to *Pinus* distribution in Mediterranean climates, temperature and rainfall have been shown to constitute good biological descriptors (Richardson and Bond 1991; Soto et al. 2010). Additionally, several common garden studies have documented differences in drought tolerance and/or gene expression among Mediterranean pine provenances (for *P. pinaster* see Nguyen and Lamant 1989; Costa et al. 1998; Chambel, Climent, and Alía 2007; Aranda et al. 2009; for *P. halepensis* see Atzmon, Moshe, and Schiller 2004; Sathyan, Newton, and Loopstra 2005; Voltas et al. 2008). Overall, these studies point to Mediterranean pines as an interesting system to study local adaptation mediated by abiotic stress response. In that context, studying allelic (SNP-allele and haplotype) frequency in drought-related candidate genes along temperature or precipitation clines can help to improve our understanding of pine adaptation to environmental heterogeneity.

Although *P. pinaster* and *P. halepensis* share various ecological requirements and may have responded to their environment in a similar manner, they present quite contrasted biogeographic and demographic histories that can also have a strong effect on the pattern of variation of the candidate genes examined. *Pinus pinaster* has a long-presence in the western Mediterranean Basin surviving the last glaciations in multiple refugia located in southeastern Spain, northern Africa and the Atlantic coast of Portugal (Bucci et al. 2007). In contrast, *P. halepensis* would have undergone long-range colonization of the western Mediterranean from ancient populations located in Greece and Turkey (Bucci et al. 1998; Morgante, Felice, and

Vendramin 1998; Grivet et al. 2009). In addition, the dynamics of *P. halepensis* populations throughout the Mediterranean Basin highly depend on forest fires. This species-specific information on population dynamics has to be taken into account when looking at the molecular footprint of selection.

Based on previous population genomic studies identifying drought tolerance genes potentially under selection (see Eveno et al. 2008; Grivet et al. 2009) as well as on gene expression studies identifying genes affected by drought stress (Watkinson et al. 2003; Rani et al. 2009), we examine here patterns of nucleotide variation in an ortholog set of targeted candidate genes in *P. pinaster* and *P. halepensis*. We sample a broad geographic range of populations to: i) provide new evidence about positive selection acting on these genes, ii) identify the time scale at which selection events may have occurred, and iii) examine which environmental factors underlie molecular signatures of selection in both species. Our pluralistic approach provides insights on the adaptive strategy of two conifers that live under the same Mediterranean climate, but present distinct demographic, (re)colonization and life histories.

MATERIALS AND METHODS

Study species. *Pinus pinaster* Ait.: Maritime pine populations have been assessed using various molecular markers: chloroplast and mitochondrial (e.g. Burban and Petit 2003; Bucci et al. 2007), as well as nuclear (e.g. Salvador et al. 2000; Eveno et al. 2008). In particular, chloroplast markers were able to identify different gene clusters related to the history of the species (Bucci et al. 2007). Because of its economic importance, various programs of genetic improvement have been developed in *P. pinaster*, in particular for the Atlantic provenances. Within this framework,

several adaptive traits, such as growth, tolerance to drought and cold, and resistance to pests and diseases, have been the subject of genetic variability studies. Numerous approaches have been used to unravel the basis of quantitative traits in maritime pine: genetic (genetic cartography), physiologic (mechanisms implicated in traits), functional and structural genomic (candidate genes and proteins) and population genetic and genomic (genes under selection).

Pinus halepensis Mill.: Aleppo pine genetic variability has been studied with both chloroplast and nuclear markers (Bucci et al. 1998; Morgante, Felice, and Vendramin 1998; Grivet et al. 2009). A recent study revealed that the pattern of polymorphism observed in some candidate genes related to drought-tolerance in this species reflected long-range colonization and possibly natural selection during range expansion (Grivet et al. 2009). Various common garden studies have assessed Aleppo pine intraspecific variability in order to study the role of ecological factors in shaping adaptive strategies. Some experiments revealed adaptive variation to climate (total precipitation and dry season duration; Voltas et al. 2008) or reproductive features (Climent et al. 2008), highlighting thus the selective role of climate variables in determining population and family fitness in this species.

Sampling. Twelve populations of *P. pinaster* (77-122 individuals, depending on the gene; [Table S1](#)) and nine populations of *P. halepensis* (72-93 individuals; [Table S1](#)) were collected spanning the full range of each species ([Figure 1](#), [Table 1](#)). Populations were selected considering not only spatial distribution but also environmental heterogeneity in both species, prioritizing populations that represent contrasted environments ([Table 1](#)). Our sampling covers also different soil (siliceous, calcareous) and Mediterranean forest types. Finally, we included representations of the five traditional varieties or landraces described in maritime pine (Resch 1974): *mesogeensis*

(e.g. Olba), *atlantica* (e.g. Mimizan), *corteensis* (e.g. Pinia), *maghrebiana* (e.g. Tamrabta) and *renoui* (e.g. Tabarka), as well as all genecological groups normally considered in Aleppo pine: Western Europe, Eastern Europe and Northern Africa.

From each population, cones were collected from mother trees separated by at least 50 m without any phenotypic selection. Seeds from each mother tree were kept in individual paper bags and stored at 4 °C in a dry environment till DNA extraction (see below).

Spatial, environmental and genetic data.

Spatial and environmental data. Three spatial variables were recorded for each population: altitude, latitude and longitude (Table 1). Eight climatic variables were considered: annual mean temperature (AMT), temperature seasonality (expressed as the standard deviation across months multiplied by 100; TS), maximum temperature of the warmest month (MTWM), minimum temperature of the coldest month (MTCM), annual precipitation (AP), precipitation of wettest month (PWM), precipitation of driest month (PDM), and precipitation seasonality (coefficient of variation; PS). Climatic data for *P. pinaster*'s Iberian populations were obtained from a functional phytoclimatic model based on raw data from meteorological stations (Gonzalo 2007). Climatic data for *P. pinaster*'s non-Iberian populations as well as for all *P. halepensis*' populations were obtained from the WorldClim – Global Climate Data at 5 minute resolution (Hijmans et al. 2005) (Table 1). Graphical pairwise correlations between these 11 spatial and environmental variables are presented in Figure S1.

DNA extraction and candidate gene sequencing. Genomic DNA from *P. pinaster* (haploid) megagametophytes was extracted with a modified Dellaporta et al. (1983) protocol. DNA extraction for *P. halepensis* was carried out as reported in Grivet et al. (2009).

Candidate genes related to drought tolerance were originally identified on the basis of functional studies performed in *Pinus taeda* and other conifers, or derived from model species such as *Arabidopsis thaliana*, as described elsewhere (González-Martínez et al. 2006; Eveno et al. 2008; Grivet et al. 2009; Wachowiak, Balk, and Savolainen 2009). Altogether the candidate genes selected for this study belong to three well-known and relatively small multigene families: the ASR (*lp31-Pt* and *lp33-Pt*), dehydrin (*dhn2-Pp*, *dhn2-Ps* and *dhn5-Ps*) and 4-coumarate: CoA ligase (*4cl-Pt*) families. The ASR family is named after the Absciscic acid (ABA), stress and ripening response (Frankel et al. 2006). These proteins are also induced by water deficit stress (WDS). The dehydrin gene, *dhn2-Pp*, previously described in maritime pine (Eveno et al. 2008), is not orthologous to the one described in Scots pine, i.e. *dhn2-Ps* (Wachowiak et al. 2009).

Previously published primer pairs of these putative candidate genes were transferred from *P. taeda* (*Pt*), *P. sylvestris* (*Ps*) or *P. pinaster* (*Pp*) to either *P. pinaster* or *P. halepensis* or both, and three new primer pairs, which overlap with pre-described amplicons, were also designed in order to extend the coverage of these target genes (Table S2). PCR conditions for the two pines studied here are given in Table S3. Outgroup sequences for each amplicon were obtained from GenBank: *P. taeda* for amplicons *lp31-Pt*, *lp33-Pt*, *dhn2-Pp* and *4cl-Pt*, and *P. sylvestris* for *dhn5-Ps* and one *dhn2-Ps* amplicon; or produced with our newly designed primers (*P. nigra* for the second *dhn2-Ps* amplicon) (Table S2). Sequence data from this article have been deposited with the EMBL/GenBank Data Libraries under accession nos. HM481479-HM483364.

PCR products were purified and sequenced from both ends following standard protocols (for *P. pinaster*, see Eveno et al. 2008; for *P. halepensis*, see Grivet et al. 2009). Multiple sequence alignments and manual adjustments were done using SeqMan v7 (DNASTAR Lasergene software) and BIOEDIT (<http://www.mbio.ncsu.edu/BioEdit/page2.html>).

Chloroplast microsatellites. To control for associations between candidate genes and environmental data that could be due to neutral processes, we included variation of chloroplast microsatellites (the only neutral genetic markers available for the two species at the wide range scale) as a covariate in the multivariate logistic regressions (see *Environmental associations* section below). We performed population-based Principal Component Analysis (PCA) on chloroplast markers, and kept the Principal Components (PCs) that explained the majority of the overall inertia of the data. For *P. pinaster*, the three first PCs of the 16 most common haplotypes (determined on the basis of five chloroplast microsatellites) – accounting for 74% of the overall variation – were extracted from the lattice dataset (430 records \times 16 variables) of Bucci et al. (2007). For populations that did not match any of the 430 locations in this lattice, the mean of the closest 3-4 locations was computed (Table S4). For *P. halepensis*, PCs were computed in R (R Development Core Team) for each of the nine populations based on the 22 haplotypes determined from three chloroplast microsatellites (our unpublished data) and the three first PCs, which explain 82% of the overall variation, were used for subsequent analysis (Table S4).

Chloroplast microsatellites were also used to determine groups with similar evolutionary history to conduct neutrality tests (Figure 1). A previous study based on chloroplast microsatellites pointed at eight gene pools in *P. pinaster* (Bucci et al. 2007). Within the sampling used in the present study, Mimizan (Continental French lineage) and Pinia (Northern Italy and Corsican lineages) represent single lineages. Tabarka (Tunisia), Tamrabta (Moroccan Middle Atlas) and Sidi Meskour (Moroccan High Atlas) are all part of a North African lineage that combines western and eastern African origins. The rest of *P. pinaster* populations sampled are part of a wide Western Mediterranean lineage, except for Quatretonda and Oria that are considered marginal populations and may thus present a distinct evolutionary history (González-

Martínez et al. 2007a; Eveno et al. 2008). In *P. halepensis*, eastern Mediterranean populations (Elea in Greece and Shaharia in Israel) present a higher level of cpSSR diversity and substantial genetic differentiation from the rest and are considered single-population lineages (Grivet et al. 2009); Zaouia Ifrane (Morocco) and Aures Beni Melloul (Algeria) form part of the North African group, while all four Spanish populations and the Italian population of Imperia belong to the Western Mediterranean group. These regions (North Africa and Western Mediterranean) define relatively homogeneous zones of the Mediterranean Basin in terms of soil and climate (Barbéro et al. 1998).

Statistical analyses.

Gene diversity and divergence. Number of segregating sites (S), nucleotide diversity statistics (θ_π , Tajima 1989; θ_w , Watterson 1975), number of haplotypes (K), and haplotypic diversity (H_e) were computed for both North African and Western Mediterranean groups using scripts kindly provided by S.E. Ramos-Onsins (Department of Genetics, Faculty of Biology, University of Barcelona, Spain) and the program DnaSP v5 (Librado and Rozas 2009). Average divergence per site (K_{all}) for each geographical group was computed between each of the pines and the outgroup *Pinus taeda*, as well as between *P. pinaster* and *P. halepensis* themselves, using scripts also provided by S.E. Ramos-Onsins.

Neutrality tests. All neutrality tests were performed at the regional level (see group definition above) considering all sequenced gametes in each amplicon, using one sequence from another pine species as outgroup when appropriate, and considering all substitutions as well as only silent sites (except for the *MK* test where all sites must be used, see below). First, we conducted neutrality tests based on within-species population genetic data with (to determine ancestral

states) or without outgroup, including Tajima's D test (Tajima 1989), Fay and Wu's H normalized test (Zeng et al. 2006), both based on the site-frequency spectrum (SFS), Ewens-Watterson EW homozygosity test (Watterson 1978), and the $DHEW$ neutrality test (Zeng, Shi, and Wu 2007) – a compound test that combines the properties of D , H normalized and EW tests – using scripts provided by K. Zeng (State Key Laboratory of Biocontrol and Key Laboratory of Gene Engineering of the Ministry of Education, Sun Yat-sen University, Guangzhou, China). Second, we used methods that compare patterns of polymorphism within species and divergence between species. The McDonald-Kreitman (MK) test, which compares the ratio of nonsynonymous and synonymous mutations between and within species (McDonald and Kreitman 1991), was computed using DnaSP v5 (Librado and Rozas 2009). $ML-HKA$ (Maximum-Likelihood multilocus Hudson-Kreitman-Aguadé) neutrality tests were computed using the *mlhka* program (Wright and Charlesworth 2004). This test is an extension of the HKA test that allows to test specific locus or groups of loci against a group of neutrally evolving genes for their levels of polymorphism and divergence. First, each amplicon was compared to all the rest and the selection parameter K was computed. The K parameter measures the degree to which diversity is increased or decreased with respect to divergence. Second, for significant tests, the analysis was repeated considering only amplicons from different genes in order to avoid potential linkage. All runs consisted in 100,000 cycles of the Markov chain; long runs of 1,000,000 cycles were also computed for significant tests as well as different starting values for the divergence time parameter (T). However, no differences were found in the results and only basic runs are presented here. Standard likelihood ratio tests (LRTs) were used to compare among models considering one amplicon under selection at a time and the null model of no selection (degrees of freedom equal one). The $ML-HKA$ test was performed using different outgroups to gain insights

on evolutionary times at which positive selective wave(s) may have taken place: i) with *P. taeda* as outgroup for both *P. pinaster* and *P. halepensis* amplicons. In this case, the time frame would correspond to the split between the *Trifoliae* section (to which *P. taeda* belongs) and the *Pinus* section (to which the Mediterranean pines belong) in the Oligocene (~25 Ma; Gernandt et al. 2005; Gernandt et al. 2008). Three amplicons were discarded from these analyses because they had *P. sylvestris* or *P. nigra* as outgroups (two from *dhn2-Ps*, and one from *dhn5-Ps*); ii) with *P. halepensis* as outgroup for *P. pinaster* amplicons and *P. pinaster* as outgroup for *P. halepensis* amplicons. The time frame, in this case, would correspond to the diversification of Mediterranean piness (in the Miocene, ~10 Ma; Gernandt et al. 2008). One amplicon was discarded from this series of tests (*dhn2-Ps_b*; see Table S2 for amplicon nomenclature) due to bad alignment between species and, thus, the possible amplification of a paralogous fragment in *P. halepensis* (*dhn2-Ps_b* from *P. halepensis* had more similarity with the outgroup, *P. nigra*, than with its close relative *P. pinaster*).

Environmental associations. We tested if SNP-allele or haplotype frequencies at candidate loci correlated with climatic variables using logistic regression. We first carried out series of univariate logistic regressions to test for association between SNP/haplotype frequencies and environmental variables using the program SAM (Joost, Kalbermatten, and Bonin 2008). We considered a correlation as significant only when two likelihood ratio tests (G and Wald tests) rejected the null hypothesis of no association between the genetic and the environmental variables (at the 5% level). A strict Bonferroni correction was applied to correct for multiple testing of univariate models.

In a second step, whenever a SNP/haplotype was found significantly correlated with an environmental variable, we performed a multivariate logistic regression using neutral marker

PCs (see above) as covariates (together with the SNP and the environmental variables) to control for neutral processes (e.g. postglacial migrations, geographical isolation) that may have also generated clines in the absence of any local adaptation. Multiple logistic regressions were performed in *R* (R Development Core Team) using the *glm* function (assuming that allelic counts at SNPs were binomially distributed) for each SNP/haplotype separately. Model selection was carried out using the *drop1* function and likelihood ratio tests (LRTs). Whenever the models compared were not nested (and LRTs were thus not appropriate), we used Akaike's Information Criterion (AIC). Tests for environmental associations were performed at the amplicon level (counting the SNPs located within overlapping regions only once) and including only populations with more than three gametes sampled (average number of gametes sampled per population: 7.98 in *P. pinaster* and 9.27 in *P. halepensis*, see Table S1).

RESULTS

Gene diversity and divergence

Overall primer transfer rates across the two pine species were high (91.6% for *P. pinaster* and 83.3% for *P. halepensis*) and we obtained a total of 533 kbp and 380 kbp of sequence in *P. pinaster* and *P. halepensis*, respectively. All sequences represented putative orthologs (determined on the basis of similarities with available sequences in GenBank and construction of phylogenies including multiple gene members of the target families in different conifers) except probably for *dhn2-Ps_b* in *P. halepensis* (see Methods). Maritime pine showed a higher level of gene diversity than Aleppo pine (Table 2), and this result held when considering only the nine orthologous amplicons in the two species (data not shown). In total, 131 SNPs (24 non-

synonymous) were detected in *P. pinaster* (4 to 27 per amplicon) and 65 SNPs (12 non-synonymous) in *P. halepensis* (1 to 21 per amplicon) (Table 2). There were only four shared polymorphisms (i.e. 2 % of polymorphic sites were shared) across species, with two of them (those within *4cl-Pt*) appearing only in one *P. pinaster* individual. Number of insertions/deletions was similar between the two species for the same locus, but these indels were not shared across species. None of them caused frame shifts. At the haplotype level, not only the number of haplotypes substantially differed between the two species (86 for *P. pinaster* vs. only 59 for *P. halepensis*) but also their frequency spectrum: *P. pinaster* had multiple common haplotypes along with some less frequent ones, while *P. halepensis* generally displayed one major haplotype together with some low-frequency ones (Figure S2). Finally, there was also a contrasted pattern of gene diversity between the regional groups defined by chloroplast microsatellites in *P. pinaster*: the Western Mediterranean group displayed higher nucleotide diversity than the North African group (average θ_w per population of 0.0039 vs. 0.0022). The SNP and haplotypes detected were used to conduct neutrality tests, as well as to detect associations with environmental variables.

Divergence at the DNA sequence level was examined using two outgroups: *P. taeda*, a New World pine, and each of the Mediterranean pines as reference for the other (i.e. *P. halepensis* was used as outgroup for *P. pinaster* and *vice versa*). For both species, nucleotide divergence per site with *P. taeda* ($K_{all} = 0.0397$ for *P. pinaster*; $K_{all} = 0.0498$ for *P. halepensis*) was higher than with the other Mediterranean species ($K_{all} = 0.0235$ for *P. pinaster* vs. *P. halepensis*) except for *dhn2-Pp_b* in *P. pinaster* that displayed a slightly higher divergence between the Mediterranean pines than between each of them with *P. taeda*.

Neutrality tests

Neutrality tests rejected the null neutral model for distinct genes depending on the species and the geographic group (Table 3). The D and H_{norm} tests identified some genes departing significantly from expectation under the neutral model that are not detected by the more robust $DHEW$ test: $lp31-Pt_a$ and $lp31-Pt_b$ in the North African group for *P. pinaster*; $lp33-Pp$ in the Western Mediterranean group for *P. halepensis*. As shown below, these two tests are probably reflecting demographical events and not selective processes. More relevantly, two fragments of the $4cl-Pt$ gene were identified as potential targets of selection by $DHEW$ tests: $4cl-Pt_b$ in the Western Mediterranean group for *P. pinaster* and $4cl-Pt_d$ in the North African group for *P. halepensis*. Considering all polymorphic sites or only silent sites did not change the qualitative results of the test (Table 3 and Table S5).

None of the MK tests showed any departure from the null hypothesis of an equal ratio of nonsynonymous to synonymous variation within and between species (data not shown). The $ML-HKA$ tests based on silent sites (with *P. taeda* as outgroup) revealed uncoupled patterns of polymorphism and divergence in only one amplicon in *P. pinaster* in the Western Mediterranean group ($dhn2-Pp_b$), while in *P. halepensis* one amplicon in this same group ($lp33-Pp$) and one in the North African group ($4cl-Pt_b$) were significant (Table 3). These results still stood when analyses were restricted to only one amplicon per gene (in order to avoid potential linkage between fragments from the same gene) and when fragments potentially under selection were not used as control loci (data not shown). When all substitution sites were included in the analyses, three more genes were significant for *P. halepensis* in the Western Mediterranean group ($dhn2-Ps_a$, $4cl-Pt_a$, $4cl-Pt_b$) (Table S5). In contrast, when Mediterranean pines were used as outgroup for each other, patterns of polymorphism and divergence either considering all sites

(Table S5) or silent sites (Table 3) seemed to evolve neutrally and a similar trend of the maximum likelihood estimate of the selection parameter (*ML-HKA*'s *K*) was observed in the two series of analyses.

Environmental associations

Environmental associations were examined at both the SNP and the haplotype level, and, overall, similar associations were detected. Nonetheless, a few correlations differed, with some found only at the haplotype level, which highlights how these two levels complement each other as haplotypes may reflect interactions among linked mutations. Within the two species, all significant associations detected between SNPs or haplotypes and climatic variables involved temperature indices as environmental variables.

As many as 23 significant associations were initially found in *P. pinaster* (non-corrected model as provided by SAM, see Methods). Only three associations remained after integrating neutral marker PCs as covariates, two of them with spatial variables (one with altitude, one with latitude) and one (*4cl-Pt_c_229*) with the minimum temperature of the coldest month (MTCM) (Figure 2; Table S6). At the haplotype level, initially 32 associations were found in maritime pine but only six remained in the corrected model: two (*4cl-Pt_c_A* and *4cl-Pt_c_B*) with altitude and one (*dhn5-Ps_C*) with annual mean temperature (AMT), one (*dhn5-Ps_E*) with maximum temperature of the warmest month (MTWM), and two (*4cl-Pt_c_A* and *4cl-Pt_c_B*) with minimum temperature of the coldest month (MTCM) (Figure 2; Table S7). In the case of *P. halepensis*, only one association was found by the corrected models, both at the SNP (between *4cl-Pt_c_131* and temperature seasonality, TS) and at the haplotype (*4cl-Pt_c_B* and TS) level (Figure 2; Tables S6 and S7).

In summary, association analysis identified two loci exhibiting significant correlations with temperature indices: *4cl-Pt_c* that is associated with MTCM in *P. pinaster* and with TS in *P. halepensis* at both the SNP and haplotype levels, and *dhn5-Ps* that is associated with AMT and MTWM at the haplotype level in *P. pinaster* only.

DISCUSSION

In this study, we assessed the impact of natural selection on the same set of candidate genes related to drought tolerance in two widespread Mediterranean pine species. Our results revealed distinct selection patterns according to species, geographic regions and loci. Below, we discuss these findings in the light of the history of each species and the specificities of each of the methods used to reveal footprints of selection.

Neutrality tests

Neutrality tests examining selection events based on site- and haplotype-frequency spectrum identified distinct genes potentially targeted by selection. The *D* and/or *H_{norm}* tests detected three loci departing from neutrality that were not detected by the *DHEW* compound tests. These results may be explained by the sensitivity of the *D* and *H_{norm}* tests to demographic factors and different degrees of background selection (Zeng et al. 2006). Especially, Tajima's *D* is sensitive to background selection and population growth while the *H_{norm}* is more sensitive to population shrinkage and subdivision. In a previous study in *P. halepensis* based on ten candidate genes, we have shown that this species has undergone historical bottlenecks and that the western populations of the species harbored some signatures of this demographic event (Grivet et al.

2009). In addition, based on the present data set and extensive coalescent simulation, we have found that observed values of D and H_{norm} for *P. pinaster* both in the Western Mediterranean and North African groups reject the standard neutral model and suggest past bottlenecks in this species too (see [Figure S3](#)). To test the robustness of the *DHEW* compound test, neutral coalescent simulations under realistic bottlenecks scenarios were further simulated in *P. pinaster* (1,000 coalescent simulations per scenario) and the significant values for this test were recorded. In all cases, the number of significant tests obtained was equal or lower than the number expected by chance (see [Figure S3](#)), highlighting the robustness of the *DHEW* test in the presence of bottleneck events and geographic population structure of magnitudes similar to the ones found in maritime and Aleppo pines. In addition to our simulations, the *DHEW* test has been shown to be robust to recombination and to have high sensitivity to detect positive selection (Zeng et al. 2007), as it combines powerful (and insensitive to recombination) haplotype-frequency spectrum tests (i.e the *EW* test) with SFS statistics, such as the D test that maintains power to detect positive selection across a wide period of time (Zeng et al. 2006; Zhai, Nielsen, and Slatkin 2009). Only two on-going events of selection were detected by the *DHEW* test, one in *P. pinaster* and one in *P. halepensis*. Both cases involved the *4cl-Pt* gene (albeit in different geographical regions and distinct amplicons), suggesting a potential role of *4cl* in local adaption in both pine species (see below).

Among the tests examining polymorphism within species and divergence among species, the *MK* test did not detect any loci under selection. While this test is somewhat robust to demography, it seems less powerful in detecting positive selection than *HKA* type tests (Zhai, Nielsen, and Slatkin 2009). The *ML-HKA* tests for both silent and all sites, using *P. taeda* as outgroup, identified three amplicons with uncoupled levels of polymorphism and divergence in

the two pines. Two genes (*dhn2-Pp_b* in *P. pinaster* in the Western Mediterranean group and *4cl-Pt_b* in *P. halepensis* in the North African group) showed high levels of diversity compared to divergence (selection parameter higher than one), a pattern compatible with balancing selection. One other gene (*lp33-Pp* in the Western Mediterranean group in *P. halepensis*) showed low diversity compared to divergence (selection parameter lower than one), which could reflect the transient reduction in variability occurring during a selective sweep. Three other genes showed uncoupled levels of polymorphism and divergence in *P. halepensis* when examining all sites, but not when examining silent sites alone. Two of these genes (*4cl-Pt_a* and *4cl-Pt_b* in the Western Mediterranean group) had an excess of nucleotide diversity, which could reflect recent balancing selection acting on non-synonymous sites, while for one gene (*dhn2-Pp_a* in the Western Mediterranean group) purifying selection reducing diversity in non-synonymous sites could explain the observed pattern. Evidence for extensive purifying selection in conifers comes from an approximately four-fold nucleotide diversity at synonymous compared to non-synonymous sites in most species (see Table 1 in González-Martínez et al. 2010), including maritime ($dN/dS = 0.169$) and Aleppo ($dN/dS = 0.344$) pines (this study). Although demographical events can result in a significant *ML-HKA* test, its multilocus nature combined with the *HKA* framework should produce a more robust test than those based on comparing different aspects of polymorphism at a single locus (Wright and Charlesworth 2004).

In order to gain power by increasing sample size while taking the specific history of each species into account, we performed the neutrality tests in groups of populations that present similar evolutionary history (i.e. North African and Western Mediterranean groups). However, grouping populations may bias the output of the SFS-based neutrality tests as it can increase the proportion of singletons, leading to an excess of low frequency variants and thus to more

negative D (Städler et al. 2009). In our study, populations were grouped according to homogeneous gene pools (see Bucci et al. 2007 for *P. pinaster* and Grivet et al. 2009 for *P. halepensis*), the consequence of which should be that no substantial differences in the number of rare variants at the population and group levels are found. The extensive gene flow normally found in conifers with large and continuous distribution would also support this approach. Nevertheless, to check that our results were not biased because of population grouping, we performed neutrality tests at the population level too and found that values of D test at the group level were not more negative than at the single population level (data not shown). Moreover, results of the H_{norm} and $DHEW$ tests should not be affected by the accumulation of singletons as the first is based on intermediary and high frequency variants while the second integrates all variants of the site frequency spectrum. Thus, our analyses appear reasonably robust to population grouping.

Selective events appear to have affected distinct genes in *P. pinaster* and in *P. halepensis* despite their overlapping environment and close phylogenetic relationship, a result that may be connected to the different histories of the two pines that have likely resulted in different selective pressures. In *P. pinaster*, selective events were detected only within the Western Mediterranean group, while in *P. halepensis* footprints of selection were identified both in the Western Mediterranean and North African groups. These results point to different geographical selection pressures that may have led to the process of regional adaptation of the pines (see Barbéro et al. 1998 for a description of the different Mediterranean environmental zones; and Gómez and Zamora 2000; Nakazato, Bogonovich, and Moyle 2008; Montesinos et al. 2009 for some examples on adaptive variation across heterogeneous environments). It is also noticeable that *P. pinaster* had lower levels of nucleotide variation within the North African

range. This fact can be attributed to population history and/or interpreted in the light of more extreme environmental conditions constraining population sizes and/or resulting in stronger selection in this range.

Environmental associations

Environmental associations identified two loci that were correlated with temperature, suggesting the importance of this climatic variable as selective agent (Saxe et al. 2001; Jump et al. 2006). One of these genes was common for both pines (*4cl-Pt* correlated with MTCM in *P. pinaster* and with TS in *P. halepensis*), while the other gene association was only significant in *P. pinaster* (*dhn5-Ps* correlated with AMT and MTWM). Some of the correlations between genetic and climatic data in *P. pinaster* were due basically to the extreme values of a few North African populations (Sidi Meskour and Tamrabta for *4cl-Pt_c*; Tabarka for *dhn5-Ps_C*; see Figures 2 and 3) and may not represent true adaptive responses to environmental gradients but local adaptation to particular environments or genetic drift due to population isolation (see references in Alleaume-Benharira, Pen, and Ronce 2006; Rosenblum, Hickerson, and Moritz 2007). In contrast, the other associations (*4cl-Pt_c* in *P. halepensis* and *dhn5-Ps_E* in *P. pinaster*) showed more robust patterns. For instance, *dhn5-Ps_E* tended to be absent from populations characterized by the highest MTWM (Arenas de San Pedro in central Spain, Tamrabta in Moroccan Middle Atlas and Tabarka in coastal Tunisia) regardless of their geographical location (Table 1 and Figure 3).

Environmental correlations with allelic variation can be spuriously inflated by neutral processes that may also generate genetic clines, such as population history or population genetic structure. This is particularly true for forest trees that may have followed a postglacial

colonization pathway overlapping temperature and rainfall clines. Here, to control for confounding associations between candidate genes and environmental data, we included variation of chloroplast microsatellites as a covariate in the multivariate logistic regressions. In *P. pinaster*, the use of cpSSRs has allowed a more accurate description of genetic structure (Bucci et al. 2007) than previous studies based on biochemical markers, such as terpenes or allozymes (e.g. Baradat and Marpeau-Bezard 1988; Salvador et al. 2000). Recently, nuclear molecular data for a geographically limited set of populations (nuSSRs and SNPs) (Eveno et al. 2008; our unpublished results) have identified similar gene pools as Bucci et al. (2007). More extensive sampling with highly-polymorphic biparentally-inherited nuclear markers could, in principle, reveal a more complex spatial genetic pattern affecting the outcome of some of the multivariate logistic regressions reported in this study. However, because the available chloroplast data set we used strongly reflects the species' history, they are expected to 'correct' for the presence of overall neutral genetic gradients. Accordingly, including these markers as covariates identified a substantial fraction of the correlations initially retained as significant as false positives (83.6 % in *P. pinaster* and 85.7% in *P. halepensis*) (Tables S6 and S7).

The dehydrin gene family

Neutrality tests and environmental associations both point to dehydrins as potential targets of natural selection in *P. pinaster*. Some dehydrins were also suggested to be under selection in a study based on detection of outlier loci in this species (Eveno et al. 2008). Dehydrins displayed also non-neutral patterns of nucleotide diversity in *Pinus sylvestris* populations showing divergence for cold tolerance in Europe (Wachowiak et al. 2009) and were associated with carbon isotope discrimination (and, thus, potentially with drought tolerance) in *P. taeda*

(González-Martínez et al. 2008). Altogether these studies suggest the involvement of dehydrins in the adaptive response of pines to abiotic stress.

Dehydrins are part of a relatively small multigene family of intracellular stabilizers that plays a major role in cell protection against desiccation. These proteins are produced in response to any type of stress that causes dehydration at the cellular level, such as cold, drought or salinity (Close 1997). Changes in dehydrin gene expression have been reported in response to drought and/or cold stress in many plants such as cowpea (Ismail, Hall, and Close 1999), barley (Suprunova et al. 2004), wheat (Lopez et al. 2001), apple (Wisniewski et al. 2008), tomato (Weiss and Egea-Cortines 2009) and blueberry (Panta, Rieger, and Rowland 2001); while transgenic experiments confirmed the role of dehydrin genes in enhancing tolerance to drought or freezing stress in plants (e.g. in *Arabidopsis*, Puhakainen et al. 2004). In conifers, at least eight dehydrin genes have been identified in *Pinus sylvestris* (Joosen et al. 2006) and *Picea abies* (Yakovlev et al. 2008), and dehydrin expression has been shown to increase under wounding, cold and drought stress (Richard et al. 2000; Watkinson et al. 2003). Thus, it is not surprising to find in our study significant associations between dehydrins and temperatures, as critical low temperatures can cause tissue injury while high temperatures accompany dehydration, both stimulating the accumulation of dehydrins (Lewitt 1980; Ingram and Bartels 1996; Rizhsky, Liang, and Mittler 2002). Although our results, and others in *P. pinaster*, suggest that selection may have acted on some of the dehydrins tested, further genetic association and functional studies are necessary to confirm the role of dehydrins in local adaptation of this Mediterranean pine.

The 4-coumarate: CoA ligase (4cl) gene

There was converging evidence both from neutrality tests (*DHEW* in *P. pinaster* and both *DHEW* and *ML-HKA* in *P. halepensis*) and environmental associations that the *4-coumarate: CoA ligase (4cl)* gene may be under selection in the two Mediterranean pines studied. Preliminary results on phenotypical association corrected by neutral gradients in *P. pinaster* also point at *4cl-Pt*. Indeed, multiple regressions reveal significant associations between polymorphism in *4cl-Pt_c* at the SNP and haplotype levels and total height in populations growing in sites characterized by dry and intermediate humidity (Table S8). The *4cl* family has been extensively studied in plants, where it is encoded by four to five genes in the fully sequenced genomes of *Arabidopsis*, rice and poplar (reviewed in Souza et al. 2008). The *4cl* gene is involved, among other processes, in the production of basic enzymes of the phenylpropanoid metabolism that are important metabolites acting as protectants against biotic and abiotic stresses (Rani et al. 2009). The *4cl* gene also encodes key enzymes in the biosynthesis of lignin and several studies have demonstrated its involvement in plant growth (e.g. Yun et al. 2005; Wagner et al. 2009; Yun et al. 2009). Thus, we expect that changes in *4cl* function would have significant repercussions on tree physiology and morphology. Implication of *4cl* in pine morphology and physiology has been shown in gene association studies (for *P. taeda* see González-Martínez et al. 2007b) as well as in gene suppression studies (for *P. radiata* see Wagner et al. 2009).

Neutrality tests versus Environmental correlations

The different approaches used in this study suggested different loci under selection within each of the two Mediterranean pine species. This fact has to be connected to the specificities of the statistical methods used, whose performance has been recently studied under various

demographic and recombination scenarios (Zeng et al. 2007; Zeng, Shi, and Wu 2007; Ramírez-Soriano et al. 2008; Zhai, Nielsen, and Slatkin 2009). Within-species site and haplotype frequency spectrum-based methods are suitable to detect ongoing or recently fixed selective sweeps - *EW* tending to be more powerful around the time when a selected mutation reaches fixation (Zeng, Shi, and Wu 2007). In contrast, tests comparing levels of (within-species) polymorphism and (among-species) divergence are able to detect the cumulative effects of positive selection events over a wider evolutionary scale. Within this category, the *ML-HKA* test is suitable to quantify the amount of selection in the genome and, due to its multilocus nature, it is expected to be more robust to changes in population size such as population bottlenecks and expansion than traditional tests (Wright and Charlesworth 2004). Simulations have shown that the *HKA* test is relatively insensitive to change of divergence time as most of its power comes from the transient reduction in variability occurring during selective sweeps (Zhai, Nielsen, and Slatkin 2009). However, we only obtained significant tests when using the less closely-related outgroup (i.e. a New World pine, *P. taeda*). This points to either lower power when divergence time from the outgroup is low or, alternatively, to selection events that took place before the split of the two Mediterranean pines considered. In our study, none of the *EW* neutrality tests were significant (data not shown), while both the *DHEW* and the *ML-HKA* tests revealed distinct genes potentially under selection. As a consequence, potential selective events detected by the neutrality tests assessed in this study would correspond to either on-going or relatively old selective events (i.e. before the diversification of the Mediterranean pines in the Miocene, ~10 Ma; Gernandt et al. 2008).

Last generation tests, such as *DHEW* or *ML-HKA*, are powerful for detecting positive selection and are relatively insensitive to other evolutionary forces, but still do not integrate

recombination rate, a factor that can produce substantial biases (Nielsen et al. 2007; Ramírez-Soriano et al. 2008), nor provide insights on selection drivers. Methods based on correlation between genetic and environmental data are appealing because they aim at understanding species-specific adaptations and processes that connect an organism to its environment by looking at allelic frequencies or the genetic structure of populations (Foll and Gaggiotti 2006; Joost, Kalbermatten, and Bonin 2008). These correlation methods are also more appealing than methods based on detection of outliers (e.g. Eveno et al. 2008) as they target particular selection drivers and provide a hypothesis testing framework. In addition, they are well suited for high-throughput genotyping even for non-model species (Namroud et al. 2008; Eckert et al. forthcoming). However, correlation methods are not without limitations, as it may be challenging to find the environmental factors that are relevant for each species adaptation and associations have to be controlled for historical and demographical processes, in particular for allelic clines produced by postglacial migrations in Europe and the Americas.

Together, neutrality tests and environmental association approaches complement each other by looking at different evolutionary scales and types of selection. In our study, they detected a relatively-high number of genes showing non-neutral patterns of evolution, a result that can be attributed to a selection of candidate genes based on earlier studies (González-Martínez et al. 2006; Eveno et al. 2008; Grivet et al. 2009). This fact supports a candidate gene approach with targeted genes, at least for organisms that have large genomes (e.g. conifers), which, so far, preclude dense genome-wide sampling.

Conclusion

629 Our pluralistic approach revealed a dynamic action of natural selection in space and time within
630 maritime and Aleppo pines (Felsenstein 1976; see Vasemagi and Primmer 2005 for other
631 examples). Selection events along with environmental associations were detected; some of these
632 events differ between the two species reflecting individual histories (recolonization,
633 demography, adaptation) while others are shared, which translates partly as a common history of
634 these closely-related and partially sympatric Mediterranean pines. The simultaneous search for
635 patterns of selection in two closely-related species can help understanding the evolutionary
636 forces responsible for adaptive responses, and thus provides an effective way of assessing the
637 degree of local adaptation, a key factor to integrate in future management and conservation
638 strategies (Wright and Gaut 2005).

639

SUPPLEMENTARY MATERIAL

Table S1. Sampling size per amplicon (six putative candidate genes) and per population for *P. pinaster* and *P. halepensis*.

Table S2. Characteristics of the primers used to amplify the candidate genes in *P. pinaster* and *P. halepensis*.

Table S3. PCR conditions for amplicon amplification (six putative candidate genes) for *P. pinaster* and *P. halepensis*.

Table S4. First three Principal Components (PCs) of the chloroplast haplotype frequency for *P. pinaster* and *P. halepensis*.

Table S5. Polymorphism, divergence and neutrality tests (all sites) in *P. pinaster* and *P. halepensis* for the phylogeographical groups defined with chloroplast microsatellites.

Table S6. SAM output (uncorrected models) showing the significant associations (at 5%) between a given allele (SNP) at a given locus and a given environmental variable for *P. pinaster* and *P. halepensis*.

Table S7. SAM output (uncorrected models) showing the significant associations (at 5%) between a given haplotype and a given environmental variable for *P. pinaster* and *P. halepensis*.

Table S8. Phenotypic data for *P. pinaster* populations (unpublished data kindly provided by R. Alía and P.H. Garnier-Géré).

Figure S1. Graphical pairwise relationships between the 11 spatial and environmental variables recorded.

Figure S2. Haplotype distribution in *P. pinaster* and *P. halepensis* for the nine common amplicons.

663 **Figure S3.** Demographic inferences in *P. pinaster* using multilocus coalescent simulation of
664 genetic bottleneck scenarios.
665

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Table 1. Spatial coordinates and climatic variables for *P. pinaster* and *P. halepensis* populations.

Population	Country	Spatial variables			Climatic variables							
		Altitude (m)	Latitude (degrees)	Longitude (degrees)	AMT (°C)	TS	MTWM (°C)	MTCM (°C)	AP (mm)	PWM (mm)	PDM (mm)	PS
<i>P. pinaster</i>												
Arenas de San Pedro	Spain	733	40.194822	-5.116213	14.2	668.9	33.4	1.4	1318	199	12	60.8
Cazorla	Spain	1100	37.919675	-2.925765	11.5	650.2	30.6	-1.4	1257	179	11	58.3
Coca	Spain	800	41.254705	-4.497827	12.3	655.9	31.2	-0.6	454	55	15	30.0
Cómpeta	Spain	903	36.834265	-3.953989	14.0	512.1	27.5	4.7	899	132	5	64.1
Olba	Spain	1002	40.173309	-0.622966	12.4	600.7	28.3	0.6	509	63	24	31.5
Oria	Spain	1223	37.531165	-2.351138	13.1	633.1	30.6	0.4	357	46	5	46.2
Quatretonda	Spain	435	38.971645	-0.358844	15.3	553.7	30.3	3.8	777	120	9	52.1
Mimizan	France	19	44.134167	-1.303056	13.2	490.9	24.9	3.1	1235	149	62	24.3
Pinia	France	15	42.021083	9.464861	15.6	515.5	27.0	6.3	585	80	8	49.6
Tabarka	Tunisia	121	36.958397	8.703792	17.7	573.4	31.8	6.7	916	162	4	72.3
Tamrabta	Morocco	1758	33.600000	-5.016667	11.8	665.3	31.3	-3.2	721	96	8	54.4
Sidi Meskour	Morocco	1931	31.439375	-6.903864	11.4	711.3	32.5	-5.2	514	71	5	56.4
<i>P. halepensis</i>												
Cabanellas	Spain	210	42.235556	2.790000	14.8	575.5	27.4	3.9	713	97	35	26.0
Carratraca	Spain	650	36.841111	-4.834444	15.7	568.6	30.6	4.4	693	106	3	69.4
S'avall	Spain	10	39.287222	3.047778	16.8	525.9	28.7	6.8	566	92	5	54.7
Tarrasa	Spain	117	41.466667	2.100000	15.9	551.1	27.7	5.8	619	83	29	29.5
Imperia	Italy	109	43.900000	8.050000	15.1	550.5	27.5	4.6	804	107	18	38.5
Zaouia Ifrane	Morocco	1512	33.570000	-5.140000	11.5	668.7	31.2	-3.8	849	123	8	57.4

Aures Beni Melloul	Algeria	936	35.166667	6.833333	13.4	657.2	31.2	-0.1	375	53	10	36.1
Elea	Greece	155	37.766667	21.533333	16.7	592.0	30.8	5.4	808	150	6	77.1
Shaharia	Israel	236	31.600000	34.833333	20.0	496.4	31.8	8.0	397	100	0	116

AMT: annual mean temperature; TS: temperature seasonality (STD *100); MTWM: maximum temperature of the warmest month; MTCM: minimum temperature of the coldest month; AP: annual precipitation; PWM: precipitation of the wettest month; PDM: precipitation of the driest month; PS: precipitation seasonality (CV *100).

Table 2. Gene diversity (all sites) for 12 amplicons from six putative candidate genes across the twelve *P. pinaster* populations and the nine *P. halepensis* populations.

Amplicon	<i>P. pinaster</i>							<i>P. halepensis</i>						
	<i>N</i>	<i>L</i>	<i>S</i>	θ_{π}	θ_w	<i>K</i>	<i>He</i>	<i>N</i>	<i>L</i>	<i>S</i>	θ_{π}	θ_w	<i>K</i>	<i>He</i>
<i>lp31-Pt</i>														
<i>a</i>	122	456	12	6.26	4.89	6	0.717	79	353	3	2.48	1.72	4	0.460
<i>b</i>	111	560	7	3.91	2.37	3	0.446	74	488	7	3.93	2.94	4	0.464
<i>lp33-Pp</i>	97	449	9	4.29	3.89	9	0.798	90	375	1	0.17	0.53	2	0.065
<i>dhn2-Pp</i>														
<i>a</i>	120	472	12	6.06	4.74	11	0.807	89	448	2	0.20	0.88	3	0.088
<i>b</i>	85	596	21	8.42	7.03	11	0.811	93	346	4	0.55	2.26	5	0.105
<i>dhn2-Ps</i>														
<i>a</i>	92	743	27	8.10	7.13	14	0.899	na	na	na	na	na	na	na
<i>b</i>	80	513	15	5.92	5.90	14	0.855	92	457	11	1.93	4.73	6	0.204
<i>dhn5-Ps</i>	77	449	4	4.00	1.81	5	0.735	na	na	na	na	na	na	na
<i>4cl-Pt</i>														
<i>a</i>	91	551	13	5.67	4.64	4	0.678	88	461	10	5.88	4.30	13	0.805
<i>b</i>	92	256	7	3.56	5.37	5	0.291	79	688	21	10.67	6.18	13	0.793
<i>c</i>	87	543	4	0.69	1.46	4	0.341	78	528	3	2.47	1.15	5	0.697
<i>d</i>	na	na	na	na	na	na	na	72	452	3	2.80	1.37	4	0.484
<i>Total</i>	1054	5588	131	-	-	86	-	834	4596	65	-	-	59	
<i>Mean</i>	-	-	-	5.17	4.48	-	0.671	-	-	-	3.11	2.61	-	0.416

N : number of sequences; L : total analyzed length in base pair; S : polymorphic sites; θ_{π} : Tajima's nucleotide diversity $\times 10^{-3}$ (Tajima 1989) and θ_w : Watterson's nucleotide diversity (Watterson 1975) per site ($\times 10^{-3}$); K : number of haplotypes; He : haplotype diversity; na: amplicons that failed to transfer across species.

Table 3a. Polymorphism, divergence and neutrality tests (silent sites) in *P. pinaster* for the phylogeographical groups defined with chloroplast microsatellites (see text for details).

Amplicon	<i>S</i>	<i>D</i>	<i>H_{norm}</i>	<i>DHEW</i> <i>P</i> -value	<i>ML-HKA's K</i> (<i>P. taeda</i>)	<i>K_s</i> (<i>P. taeda</i>)	<i>ML-HKA's K</i> (<i>P. halepensis</i>)	<i>K_s</i> (<i>P. halepensis</i>)
Western Mediterranean ^a								
<i>lp31-Pt</i>								
<i>a</i>	8	2.172	-0.635	0.958	0.961	38.350	1.292	20.310
<i>b</i>	6	1.834	-1.380	0.913	0.698	44.250	0.720	24.360
<i>lp33-Pp</i>	6	1.133	0.911	0.984	0.696	60.060	1.839	15.990
<i>dhn2-Pp</i>								
<i>a</i>	7	0.269	-0.853	0.406	1.605	38.160	1.307	31.580
<i>b</i>	19	0.186	0.825	0.727	3.256 *	31.940	1.975	39.720
<i>dhn2-Ps</i>								
<i>a</i>	15	0.631	0.703	0.562	ptna	ptna	phna	phna
<i>b</i>	8	-0.126	0.232	0.209	ptna	ptna	d	d
<i>dhn5-Ps</i>	2	1.960	0.136	0.907	ptna	ptna	phna	phna
<i>4cl-Pt</i>								
<i>a</i>	2	0.422	0.067	0.610	0.307	31.480	0.250	26.870
<i>b</i>	2	-1.041	-3.173 *	0.014 *	0.741	43.930	0.446	41.220
<i>c</i>	1	0.214	0.565	0.663	0.542	23.080	0.000	1.190
<i>d</i>	na	na	na	na	na	na	na	na
North Africa ^b								
<i>lp31-Pt</i>								
<i>a</i>	7	-0.276	-3.065 *	0.336	0.878	39.300	0.952	22.280

<i>b</i>	6	-2.046	-3.813	0.073	0.820	44.250	0.880	26.150
<i>lp33-Pp</i>	5	0.014	0.926	0.899	0.614	57.860	1.946	13.040
<i>dhn2-Pp</i>								
<i>a</i>	4	0.799	-0.554	0.615	0.868	36.610	0.792	28.620
<i>b</i>	13	1.095	0.361	0.718	2.036	32.780	1.362	33.600
<i>dhn2-Ps</i>								
<i>a</i>	6	0.190	-0.232	0.322	ptna	ptna	phna	phna
<i>b</i>	6	-1.116	-1.223	0.085	ptna	ptna	d	d
<i>dhn5-Ps</i>	2	0.688	-0.809	0.540	ptna	ptna	phna	phna
<i>4cl-Pt</i>								
<i>a</i>	8	2.797	0.411	0.995	1.279	39.160	0.442	28.880
<i>b</i>	3	2.266	0.090	0.982	1.303	48.190	1.017	34.730
<i>c</i>	2	0.201	0.387	0.494	1.039	25.240	0.000	3.410
<i>d</i>	na	na	na	na	na	na	na	na

D: Tajima's *D* (Tajima 1989); *H_{norm}*: Fay and Wu's normalized *H* (Zeng et al. 2006); *DHEW* test: compound test (Zeng et al. 2007); *ML-HKA*'s *K*: Selection parameter of the Maximum-Likelihood multilocus Hudson-Kreitman-Aguadé neutrality tests (Wright and Charlesworth 2004); the species used as outgroup is given in parenthesis; *S*: number of segregating sites; *K_s*: average proportion of nucleotide differences between species per silent site $\times 10^{-3}$.

na: amplicons that failed to transfer across species; ptna: *P. taeda* outgroup sequence not available; phna: *P. halepensis* outgroup sequence not available; d: discarded because possible paralog.

P-values [P neutrality test (neutral) \leq P neutrality test (observed)]: 0 < *** < 0.001 < ** < 0.01 < * < 0.05

^a Western Mediterranean: Arenas de San Pedro, Cazorla, Coca, C mpeta and Olba.

^b North Africa: Tabarka, Tamrabta and Sidi Meskour.

Table 3b. Polymorphism, divergence and neutrality tests (silent sites) in *P. halepensis* for the phylogeographical groups defined with chloroplast microsatellites (see text for details).

Amplicon	<i>S</i>	<i>D</i>	<i>H_{norm}</i>	<i>DHEW</i> <i>P</i> -value	<i>ML-HKA's K</i> (<i>P. taeda</i>)	<i>K_s</i> (<i>P. taeda</i>)	<i>ML-HKA's K</i> (<i>P. pinaster</i>)	<i>K_s</i> (<i>P. pinaster</i>)
Western Mediterranean ^a								
<i>lp31-Pt</i>								
<i>a</i>	2	1.689	-1.714	0.860	0.597	34.590	0.452	20.960
<i>b</i>	4	0.535	-0.948	0.501	0.889	48.280	1.380	23.090
<i>lp33-Pp</i>	1	-0.629	-4.343 *	0.270	0.145 *	69.100	0.361	14.020
<i>dhn2-Pp</i>								
<i>a</i>	0	nps	nps	nps	nps	55.770	nps	44.020
<i>b</i>	0	nps	nps	nps	nps	57.180	nps	38.220
<i>dhn2-Ps</i>								
<i>a</i>	na	na	na	na	na	na	na	na
<i>b</i>	2	-0.595	0.317	0.382	ptna	ptna	d	d
<i>dhn5-Ps</i>	na	na	na	na	na	na	na	na
<i>4cl-Pt</i>								
<i>a</i>	6	0.752	-1.222	0.599	2.236	43.260	1.618	22.500
<i>b</i>	16	2.386	-0.320	0.985	2.279	52.000	1.242	30.660
<i>c</i>	2	1.701	-1.305	0.867	1.350	31.970	2.976	9.440
<i>d</i>	3	2.324	-0.981	0.965	0.674	54.780	ppna	ppna
North Africa ^b								
<i>lp31-Pt</i>								
<i>a</i>	2	1.818	-1.053	0.898	0.819	34.140	1.123	12.710

<i>b</i>	2	1.882	0.602	0.909	0.530	49.000	0.924	22.700
<i>lp33-Pp</i>	0	nps	nps	nps	nps	69.410	nps	9.550
<i>dhn2-Pp</i>								
<i>a</i>	1	-0.774	0.289	0.313	0.329	56.280	0.525	19.310
<i>b</i>	1	-1.147	0.160	0.189	0.215	57.320	0.205	33.600
<i>dhn2-Ps</i>								
<i>a</i>	na	na	na	na	na	na	na	na
<i>b</i>	0	nps	nps	nps	ptna	ptna	d	d
<i>dhn5-Ps</i>	na	na	na	na	na	na	na	na
<i>4cl-Pt</i>								
<i>a</i>	4	1.306	-0.201	0.791	2.072	44.030	1.815	22.450
<i>b</i>	14	2.316	0.044	0.981	3.251 *	50.760	1.951	31.290
<i>c</i>	2	1.077	0.221	0.834	1.665	27.720	3.309	5.200
<i>d</i>	3	-1.494	-6.584	0.009 **	0.860	60.020	ppna	ppna

D: Tajima's *D* (Tajima 1989); *H_{norm}*: Fay and Wu's normalized *H* (Zeng et al. 2006); *DHEW* test: compound test (Zeng et al. 2007); *ML-HKA*'s *K*: Selection parameter of the Maximum-Likelihood multilocus Hudson-Kreitman-Aguadé neutrality tests (Wright and Charlesworth 2004); the species used as outgroup is given in parenthesis; *S*: number of segregating sites; *K_s*: average proportion of nucleotide differences between species per silent site $\times 10^{-3}$.

na: amplicons that failed to transfer across species; ptna: *P. taeda* outgroup sequence not available; ppna: *P. pinaster* outgroup sequence not available; d: discarded because possible paralog; nps: monomorphic fragment.

P-values [P neutrality test (neutral) \leq P neutrality test (observed)]: 0 < *** < 0.001 < ** < 0.01 < * < 0.05

^a Western Mediterranean: Cabanellas, Carratraca, Imperia, S'avall and Tarrasa.

^b North Africa: Aures Beni Melloul and Zaouia Ifrane.

Figure 1. Location of the 12 *P. pinaster* and the 9 *P. halepensis* populations and the distribution of the two species. Grey areas represent the two groups (Western Mediterranean and North African) defined by chloroplast SSRs (see text for details).

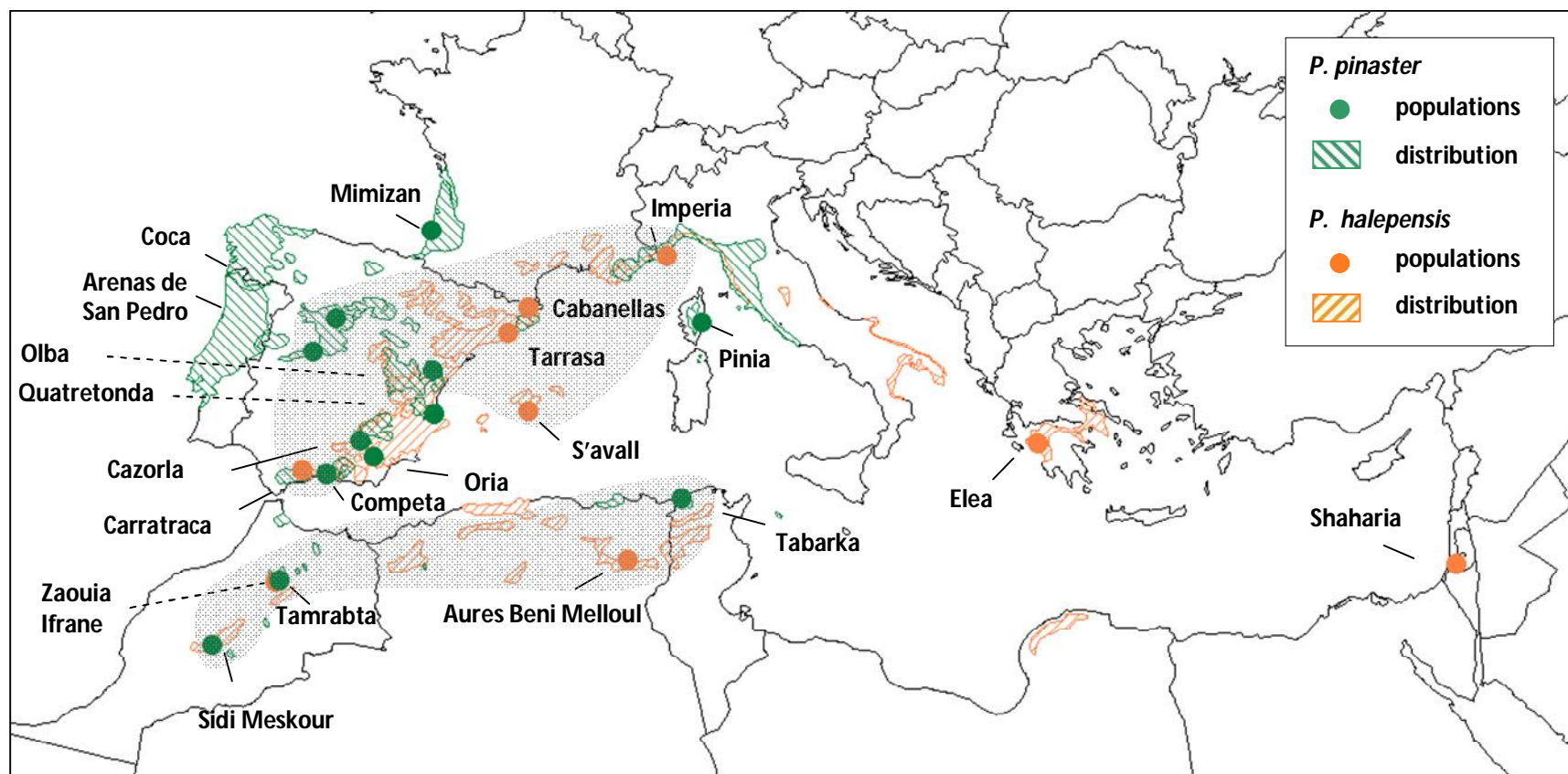


Figure 2. Predicted and observed patterns of clinal variation at *4cl-Pt_c*. Sigmoid curves denote the clines in either allelic (SNP position given) or haplotypic (haplotypes represented by capital letters) frequencies under a logistic regression model. The logistic regression model uses a climatic variable (MTCM or TS) after controlling for neutral processes (i.e. using the PCs of neutral molecular markers as covariates) for *P. pinaster* and *P. halepensis*. MTCM: minimum temperature of the coldest month; TS: temperature seasonality.

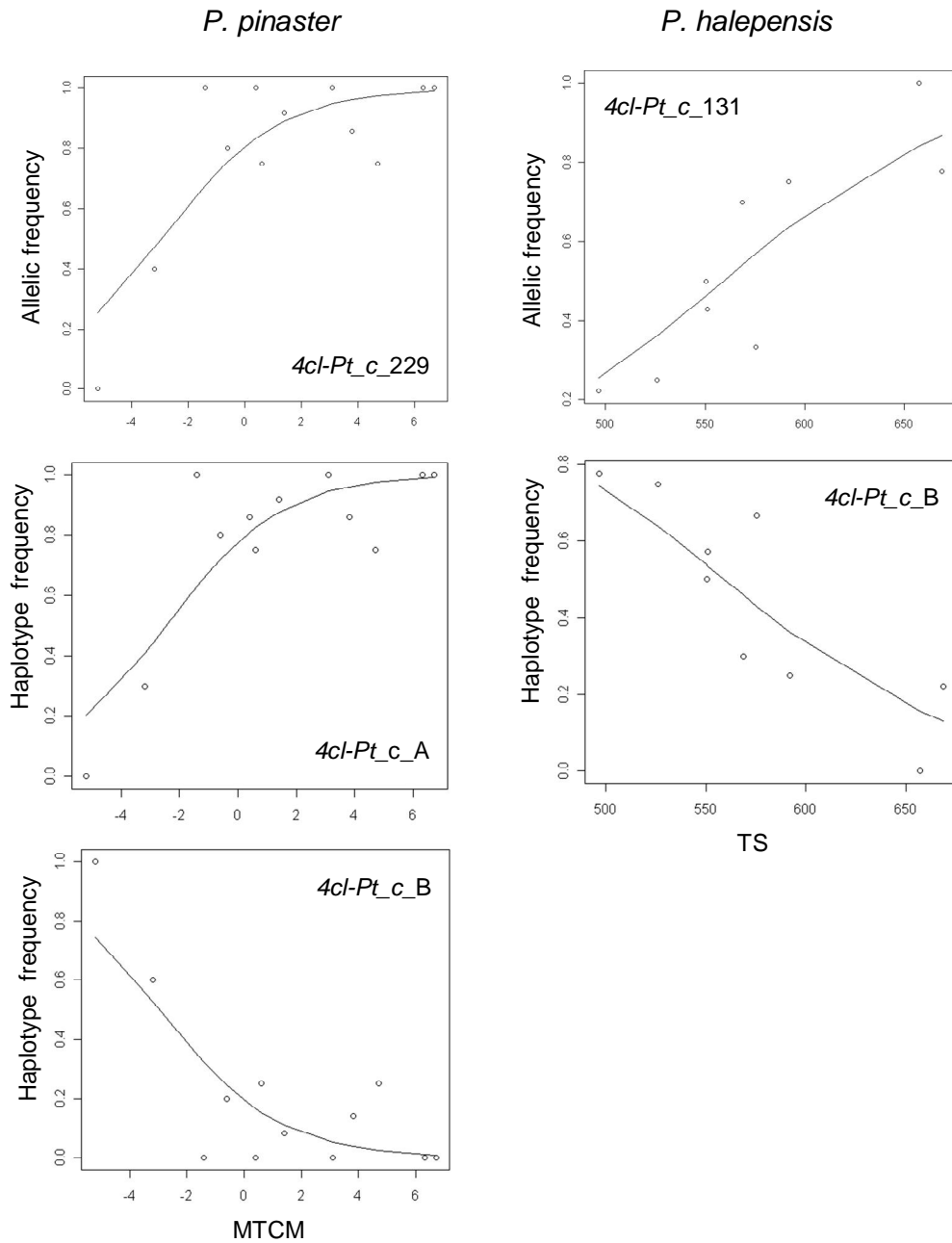


Figure 3. Upper section: *dhn5-Ps* haplotype distribution showing the proportion of haplotypes C (in grey), E (in black) and all others (in white) in *P. pinaster*. The size of the pie is proportional to the sample size at each location. **Lower section:** expected (sigmoid curves) and observed covariation in haplotype frequencies of *dhn5-Ps_C* and *dhn5-Ps_E* and climatic variables. Expected curves were obtained under a logistic regression model including a single environmental variable (AMT or MTWM) after controlling for neutral processes (i.e. using the PCs of neutral molecular markers as covariates) for *P. pinaster*. AMT: annual mean temperature; MTWM: maximum temperature of the warmest month.

