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► **To cite this version:**

Manuel A. Gonzalez, María Tomé, Pedro López-Romero, Carmen Albo, Juan Carlos Sepúlveda, et al.. miR-335 orchestrates cell proliferation, migration and differentiation in human mesenchymal stem cells. *Cell Death and Differentiation*, 2010, 10.1038/cdd.2010.167 . hal-00601167

HAL Id: hal-00601167

<https://hal.science/hal-00601167>

Submitted on 17 Jun 2011

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miR-335 orchestrates cell proliferation, migration and differentiation in human mesenchymal stem cells

Running title: miR-335 controls hMSC activation

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ABSTRACT

In spite of the extensive potential of human mesenchymal stem cells (hMSCs) in cell therapy, little is known about the molecular mechanisms that regulate their therapeutic properties. We aimed to identify microRNAs (miRNAs) involved in controlling the transition between the resting and reparative phenotypes of hMSCs, hypothesizing that these miRNAs must be present in the undifferentiated cells and downregulated to allow initiation of distinct activation/differentiation programs. Differential miRNA expression analyses between undifferentiated and differentiated bone marrow-derived hMSCs revealed that miR-335 is significantly downregulated upon hMSC differentiation, being coexpressed in with its host coding-gene *MEST*. In addition, hMSCs derived from a variety of tissues express miR-335 at a higher level than human skin fibroblasts, and overexpression of miR-335 in hMSCs inhibited their proliferation and migration, as well as their osteogenic and adipogenic potential. Expression of miR-335 in hMSCs was upregulated by the canonical Wnt signaling pathway, which has been described as a positive regulator of MSC self-renewal, and downregulated by interferon gamma (IFN- γ), a pro-inflammatory cytokine that plays an important role in activating the immunomodulatory properties of hMSCs. Differential gene expression analyses, in combination with computational searches, defined a cluster of 62 putative target genes for miR-335 in hMSCs. Western blot and 3'UTR reporter assays confirmed *RUNX2* as a direct target of miR-335 in hMSCs. These results strongly suggest that miR-335 is important for the maintenance of the immature state and its downregulation is critical for the acquisition of reparative MSC phenotypes.

Keywords and abbreviations.

Keywords:

microRNA, mesenchymal stem cell, cell proliferation, cell migration, cell differentiation, osteogenesis.

Abbreviations:

hMSC: human mesenchymal stem cells.

fdr: false discovery rate.

INTRODUCTION

Mesenchymal stem cells (MSCs) are multipotent mesoderm-derived somatic stem cell precursors of non-hematopoietic connective tissues that are present in the stroma of virtually all mammalian organs, especially bone marrow and subcutaneous fat (reviewed in (1)). Upon activation by tissue damage, MSCs contribute to tissue repair processes through a multitude of activities, including cell proliferation, differentiation and migration, and the regulation of angiogenesis and immune responses. There is growing evidence, both in animal and clinical models, that administration of *ex vivo*-expanded human MSCs (hMSCs) has potential to ameliorate degenerative disorders, including cardiovascular, neurodegenerative, and autoimmune and inflammatory diseases; however, the specific molecular mechanisms underlying this therapeutic potential remain mostly unknown.

MicroRNAs (miRNAs) are a family of small (18-24 nucleotide), single-stranded non-coding RNAs which regulate gene expression in eukaryotic cells by controlling the translation (usually by repression), stability and localization of specific mRNA targets. Hundreds of conserved miRNA genes are known in many species, making miRNAs, after transcription factors, the second most diverse family of gene expression regulators. Computational predictions of miRNA targets indicate that each miRNA regulates hundreds of mRNAs, and that approximately one third of all mammalian protein-coding genes are regulated by miRNAs (2). Functional studies show that miRNAs participate in virtually each cellular process investigated, and that alterations in their expression levels might underlie human diseases, including cardiovascular disease and cancer. There are also data indicating that mammalian miRNAs can be imported into the nucleus (3) or even secreted from cells within small exocytic particles (4), suggesting the existence of currently unknown functions for this class of molecules.

Experimental evidence shows that miRNAs are critical regulators of stem cell biology. In mouse embryonic stem (ES) cells, knockout of Dicer or DGCR8, two key protein factors involved in miRNA biogenesis, impairs cell growth, with cells accumulating in G1, and severely reduces differentiation capacity (5, 6). Additionally, expression profiling demonstrates that ES cells express a unique repertoire of miRNAs compared with differentiated cell types (7). Molecular functions of ES cell-specific miRNAs have begun to be elucidated, and the results confirm their critical involvement in the regulation of ES cell pluripotency and self-renewal. MiRNAs also play an important role in controlling ES cell differentiation and commitment. As example, miR-134, miR-296 and miR-470 are upregulated by retinoic acid, and modulate ES cell differentiation through specific targeting of Nanog, Oct4 and Sox2 mRNAs (8).

To date, relatively few studies have examined miRNA function in MSCs. Gene expression microarray profiling has identified several miRNAs that are regulated during differentiation of MSCs into different cell lineages. Some of these miRNAs have been identified as regulators of osteogenic differentiation, including miR-125b (9), miR-26a (10), miR-196a (11), miR-204/211 (12), and miRs -148b, -27a, and -489 (13). Also, miR-335 has been recently identified as the most upregulated miRNA in bone marrow-derived human MSCs (hMSCs) in comparison with skin fibroblasts (14), which are otherwise phenotypically similar to hMSCs. However, the specific targets of most of these miRNAs remain to be determined, and the possible role of miRNAs in other therapeutically relevant MSC activities, such as in cell migration and proliferation, is unknown.

Here, we sought to identify miRNAs that may regulate the activation of the reparative phenotype of hMSCs, as well as some of their target genes.

Results

miR-335 is downregulated upon hMSC differentiation. We hypothesized that key miRNA regulators of MSC activation/differentiation in tissue repair should be expressed in the undifferentiated state, and downregulated early upon exposure to a differentiation signal. To identify these putative miRNAs, we used Agilent microarrays to obtain differential miRNA expression profiles of undifferentiated bone marrow-derived hMSCs *versus* the same cells cultured in the presence of adipogenic or osteogenic media. We also profiled human skin fibroblasts, since the target miRNAs should be expressed at comparatively low levels in more developmentally-restricted mesenchymal cell types. As we aimed to identify miRNAs potentially involved in the initial steps of hMSC activation/differentiation, cells were exposed to differentiation media for a relatively short period (nine days), instead of the 21-days commonly used for *in vitro* MSC differentiation assays.

Signal processing is a critical step in the analysis of the results of miRNA microarray experiments. We used a normalization algorithm that incorporates quantile normalization between arrays (15) to estimate a processed microRNA signal for the Agilent arrays. The quantile normalization, when applied to the background-corrected signal, showed significantly lower variability between replicates than the total gene signal normalized by the 75% percentile (Supplementary Figure S1).

The results showed no significant regulation (false discovery rate, $fdr < 15\%$) of miRNAs previously described as regulators of osteogenic (miR-26a, miR-27a, miR-125b, miR-148b, miR-196a, miR-489) or adipogenic differentiation (miR-103, miR-107, miR-143) under any of the conditions tested (Supplementary Figure S2A and Supplementary Table S1). Gene enrichment analysis of the predicted targets of miRNAs up- or downregulated in at least two conditions (see Methods) showed a significant ($p < 1E-06$) enrichment for genes involved in angiogenesis or signaling by Wnt, integrins, PDGF, cadherin, endothelin and TGF- β

(Supplementary Figure S2B and Supplementary Table S1). MiR-335 was the only miRNA significantly downregulated in all three “differentiated” cell populations (Figure 1a). Fold change (log₂) values were as follows: fibroblast vs. undifferentiated hMSCs, M=-5.684 (*fdr*=0.008); osteogenic differentiation vs. undifferentiated, M=-2.434 (*fdr*=0.029); adipogenic differentiation vs. undifferentiated, M=-1.141 (*fdr*=0.103). After validating the microarray results by real-time RT-PCR (Figure 1b), we selected miR-335 for further characterization.

MiR-335 levels were also significantly reduced in human osteoblasts (Saos cell line) after differentiation (Supplementary Figure S4).

hMSCs from different tissues express higher levels than fibroblasts of miR-335 and its host gene *MEST*. To determine whether relatively high expression of miR-335 is specific to bone marrow-derived hMSCs or is a common characteristic of hMSCs, we analyzed miR-335 expression levels in hMSC populations obtained from bone marrow, subcutaneous adipose tissue and articular cartilage (Figure 2b). All hMSC isolates were tested at passage 5-7 at the same cell density (8000 cells/cm²), and were assayed for their differentiation potential to osteogenic, adipogenic, and chondrogenic lineages in the presence of specific differentiation factors (data not shown). All three populations express significantly higher levels of miR-335 than dermal fibroblasts, and miR-335 expression levels were significantly higher in hMSCs isolated from bone marrow or articular cartilage than in hMSCs derived from adipose tissue.

MiR-335 is encoded by the second intron of the *MEST* (mesoderm specific transcript homolog) gene (Figure 2a) (16). *MEST* expression, determined by real-time RT-PCR, correlated with the levels of mature miR-335 (Figure 2b; Spearman’s $r=0.5769$, $p=0.0008$), suggesting that the different hMSC isolates co-express mature miR-335 with its host gene and

not through a *MEST*-independent mechanism (such as a specific promoter or post-transcriptional regulation).

MEST expression levels also correlated with the levels of miR-335 under all other conditions tested in this study (Supplementary Figure S5).

miR-335 impairs hMSC proliferation, migration and differentiation. We next analyzed the effect of miR-335 overexpression in bone marrow-derived hMSCs. hMSCs were transduced with the lentiviral vector pLV-EmGFP-MIR335, which encodes the genomic sequence spanning miR-335, or with a control vector (pLV-EmGFP-Mock). Transduced cells were purified to >95% homogeneity (gfp-positive cells) by FACS. To avoid non-specific effects due to lentiviral gene silencing or to a high proviral copy number per cell, a multiplicity of infection of 5 was used, and only cells with medium-level gfp expression were selected (Supplementary Figure S3A). Real-time RT-PCR demonstrated a ~3-fold increase in miR-335 expression in pLV-EmGFP-MIR335-transduced cells compared with controls (Supplementary Figure S3B). Overexpression of miR-335 was not associated with any apparent changes in cell morphology or size (not shown).

When cultured over several passages, miR-335-overexpressing hMSCs showed a significant reduction in their proliferative activity compared with control cells (Figure 3a). However, miR-335 overexpression did not cause significant alterations to cell cycle kinetics (not shown) or the rate of apoptosis (Figure 3b).

hMSCs overexpressing miR-335 also showed an impaired migratory response to stimulation with fetal bovine serum (Figure 3c). Consistently, wild-type hMSCs transfected with a miR-335 inhibitor (Anti-miRTM-335, Ambion) showed increased migratory activity compared with cells transfected with a negative control Anti-miRTM oligonucleotide (Figure 3c, Supplementary Figure S3C). MiR-335-overexpressing hMSCs also showed impaired

migratory activity in an *in vitro* wound-healing assay compared with control cells (see Supplementary information videos). These results demonstrate that miR-335 is a negative regulator of hMSC migration.

The effect of miR-335 expression on hMSC differentiation capacity was monitored by comparing the *in vitro* differentiation responses of control and miR-335-transduced hMSCs to exposure to adipogenic or osteogenic stimuli for three weeks. Both adipogenic and osteogenic differentiation were significantly reduced by miR-335 overexpression. In particular, *in vitro* osteogenesis was almost completely abolished (Figure 3d), indicating a role for miR-335 in the regulation of hMSC differentiation programs. Analysis of expression of the differentiation markers osteopontin (osteogenic differentiation) and PPAR γ (adipogenic differentiation) confirmed the inhibitory effect of miR-335 in both processes (Figure 3e).

MiR-335 expression levels are regulated by signals that control MSC migration and differentiation. The apparent regulatory role of miR-335 in hMSC suggested that it might be regulated by signals known to control MSC biology. We therefore tested if canonical Wnt signaling, a negative regulator of MSC differentiation (17), affected miR-335 expression levels. Bone marrow-derived hMSCs treated with 10 mM LiCl or Wnt3a-containing conditioned medium upregulated miR-335 expression (Figure 4a, left panel). Moreover, addition of Dkk1 (100 ng/ml), a specific inhibitor of canonical Wnt signaling, significantly decreased miR-335 expression in the presence of exogenous Wnt3a (Figure 4a, right panel). Upregulation of miR-335 expression by Wnt3a was also observed in adipose-derived hMSCs and in skin fibroblasts (Figure 4b), although in a different extension, suggesting that this effect is common to all mesenchymal cell types.

We next evaluated the effect on miR-335 expression of IFN- γ signaling, another key regulator of MSC activity. IFN- γ has numerous actions on MSC, including the induction of

immunoregulatory activity, cell migration and osteogenic differentiation (18, 19). Treatment of bone marrow-derived hMSCs with human IFN- γ for 48 h significantly decreased expression of miR-335 (Figure 4c) in a dose-dependent manner, consistent with the lowered miR-335 expression during cell migration and osteogenic differentiation.

To identify possible binding sites in the *MEST* locus for transcription factors involved in β -catenin or IFN γ signaling pathways, we performed a multispecies (human, mouse and dog) sequence homology analysis for conserved binding sites for LEF/TCF (transcription factors involved in β -catenin signaling) and STAT1 (involved in IFN γ signaling). MULAN analysis with a matrix similarity of 0.95 showed one possible LEF1 and one possible TCF4 site in the 5 kb region upstream of the *MEST* transcriptional start site (corresponding to the longest human splicing variant), both mapping at -2.675 kb (Supplementary Figure S6). A similar analysis, with a matrix similarity of 0.90, identified three conserved potential STAT1 binding sites at -4.440, -4.384, and -4.303 kb (Supplementary Figure S6).

Potential miR-335 targets in hMSCs are enriched in transcription regulators of cell movement and differentiation. In order to identify potential miR-335 targets, we used a two-step approach (Figure 5a). First, we used three different miRNA target prediction programs, miRanda (20), TargetScan (21) and PicTar (22), to obtain a list of putative miR-335 targets. As these programs use different target scoring systems, we combined the three lists in a single non-redundant gene list (a total of 1838 genes, of which 1607 were mapped). Gene enrichment analysis using the PANTHER database revealed significant enrichment ($p < 0.001$) for molecular functions related to transcriptional and post-transcriptional gene regulation, and for biological processes ($p < 1E-06$) also related to regulatory activities, including mRNA transcription, protein modification, and signal transduction (Supplementary Figure S7).

Second, we performed a microarray gene expression analysis (Agilent Whole Human Genome Microarray Kit) on hMSCs transduced with pLV-EmGFP-MIR335 or pLV-EmGFP-mock. We focused on downregulated genes since mRNA expression levels tend to correlate negatively with the expression levels of their specific miRNAs (23). Using this approach, we identified 489 genes with decreased expression in miR-335-transduced cells that are thus candidate direct or indirect targets of this miRNA in hMSCs. Of these genes, 62 were also contained in the previously obtained list of predicted miR-335 targets (Supplementary Table S2), and were therefore classed as probable direct targets. This set of 62 genes was significantly enriched ($p < 0.05$) for transcription factors (21%) and membrane-bound signaling molecules (6.5%). A detailed analysis with the Ingenuity® Pathways Analysis software (IPA, Ingenuity® Systems, <http://www.ingenuity.com>) showed a predominant enrichment for functions involved in cell development, cell movement, gene expression, and the cell cycle (Benjamini-Hochberg multiple test-corrected p -value=1.37E-02-8.82E-02). Other significantly enriched biological functions included cell signaling, cell growth and proliferation, cell morphology, cell commitment, and cell death (Table 1). Using IPA, the two more representative gene regulatory networks were constructed, respectively containing 11 of the 20 potential miR-335 target genes involved in cellular movement (score 27) and 15 of the 17 genes involved in gene expression (score 42) (Figure 5b).

One of the predicted targets of miR-335 is RUNX2, a key transcription factor involved in osteogenesis. Although miRNAs can bind to coding regions and even to 5' regulatory regions, a single miR-335 binding site is predicted by the miRanda algorithm at nucleotides 3545-3567 in all the known *RUNX2* transcript variants (GenBank Accession Numbers NM_001024630, NM_001015051, NM_004348), corresponding to the 3'UTR region.

A western blot analysis of RUNX2 demonstrated that its protein levels are reduced in hMSCs by miR-335 exogenous overexpression (Figure 6a). To determine if miR-335 directly

modulates *RUNX2* expression levels in hMSCs, we cloned the full-length *RUNX2* 3'UTR downstream of the *Renilla* luciferase gene as a reporter, and assayed its expression in hMSCs transfected with a synthetic precursor or a specific inhibitor of miR-335. UTR reporters corresponding to *SOX4* and *UBE2F* genes were used as positive and negative controls, respectively, for the miR-335 inhibitory activity (24). Transfection of the miR-335 synthetic precursor significantly reduced the expression of UTR reporters for *SOX4* and *RUNX2*, but not the expression of the negative control gene *UBE2F* (Figure 6b, upper panel). Transfection with the specific inhibitor of miR-335 significantly increased the expression levels of UTR reporters for *SOX4* and *RUNX2*, but did not affect the levels of *UBE2F* (Figure 6b, bottom panel). These results indicate that miR-335 controls *RUNX2* expression in hMSCs by direct binding to its 3'UTR.

Finally, we have quantified the relative expression levels of *RUNX2* after osteogenic differentiation in hMSCs overexpressing miR-335. The results of these experiments (Figure 6c) showed a significant reduction of *RUNX2* expression levels in the cells transduced with the lentiviral vector encoding miR-335, compared to those transduced with a control lentiviral vector, which is in good agreement with the rest of the data indicating that *RUNX2* is a target of miR-335 in hMSCs.

Discussion

Our results support a central role for miR-335 in the gene regulatory network that controls the tissue-repair activities of hMSCs. Expression of this miRNA is high in undifferentiated multipotent hMSCs compared with their differentiated cell progeny, and is regulated by major signaling pathways that control MSC biology, such as those controlled by Wnt3a and IFN γ . The set of predicted miR-335 targets is enriched in genes whose products are regulators of

cellular movement and gene expression, including *RUNX2*, involved in the control of osteogenic differentiation.

There is growing evidence that miRNAs play a critical role in the biology of all stem cell types, including ES cells, germline stem cells, and somatic stem cells (SSCs). SSCs, which are present in adult somatic tissues, are usually less well-defined than embryonic or germline stem cells because a single tissue often contains several distinct SSC populations (or, as in the case with MSCs, similar SSCs can be isolated from different tissues) and they often cannot be purified to homogeneity. For these reasons, it has been difficult to identify the specific roles of a miRNA in a particular SSC population. Most miRNAs described as regulators of SSC biology are inducers of cell differentiation/commitment or inhibitors of self-renewal and proliferation. These include miR-203 in epidermal stem cells (25), and miR520h and others in hematopoietic stem cells (26). It also appears that miRNAs are involved in the self-renewal or maintenance of at least some types of SSCs, since targeted deletion of *Dicer* in mouse epidermis causes altered morphogenesis of hair follicles, with loss of stem cell markers (27).

Our results demonstrate that miR-335 is downregulated in hMSC cultures undergoing adipogenic or osteogenic differentiation, and is also expressed at comparatively lower levels in primary skin fibroblasts, which are mesenchymal cells with a limited differentiation potential. Our data are in complete accord with previous work identifying miR-335 as the most upregulated miRNA in bone marrow-derived hMSCs in comparison with fibroblasts (14).

Until recently, the only known specific biological role for miR-335 is in human breast cancer, where it has been proposed as a metastasis suppressor (24). Expression of miR-335 is lost in the majority of primary breast tumors from patients who relapse, and this is associated with poor distal metastasis-free survival (24). These results are in good agreement with our findings demonstrating that miR-335 is a negative regulator of hMSC migration. MiR-335

downregulation is also observed in drug-resistant ovarian cancer cells (28), cultured chondrocytes (versus cartilage tissue) (29), ethanol-treated neural progenitors (30), multiple myeloma cells and tumors (31), and muscle regeneration (32). In contrast, miR-335 is highly expressed in obesity models (33) and in fetal lung compared with adult tissue (34). During the preparation of this manuscript, a communication describing the direct targeting of *RBI* by miR-335 has been published (35). In this study, the authors establish the important role of this miRNA in the induction of p53-dependent cell cycle arrest after DNA damage. These results are in accordance with our findings demonstrating that miR-335 is a negative regulator of hMSC proliferation, and, taken together with our results, suggest an important role of *RBI* activity in regulation/activation of hMSC tissue repair activities.

miR-335 is encoded by the second intron of the maternally imprinted gene *MEST* (human chromosome 7q32) (16). Our results demonstrate that both miR-335 and *MEST* are coordinately upregulated in hMSCs from three different tissues (bone marrow, subcutaneous fat and articular cartilage) in comparison with skin fibroblasts. Moreover, adipose-derived hMSCs expressed significantly lower levels of miR-335 than the two other hMSC populations analyzed, indicating that the miRNA expression level in hMSC is dependant on the source tissue. The biological relevance of this finding remains to be determined, but our results suggest that miR-335 expression levels could be used to discriminate between hMSCs and fibroblasts, or even between different MSC populations with diverse differentiation potentials. Future studies are needed to confirm this finding.

We found no significant regulation of miRNAs previously described as regulators of osteogenic differentiation (miR-26a, miR-27a, miR-125b, miR-148b, miR-196a, miR-489, miR-204/211) or adipogenic differentiation (miR-103, miR-107, miR-143) under any of the conditions tested. This apparent discrepancy with published data might be related to the fact that our microarray analyses were performed on hMSCs at early differentiation stages, unlike

other studies that used terminally differentiated MSCs. Also, the method we used for signal processing (quantile normalization) (15) probably contributed to an overall reduction in the variation of gene expression between samples.

The finding that miR-335 expression impairs the proliferative and migratory capacities of primary bone marrow-derived hMSCs and inhibits both adipogenic and osteogenic differentiation suggests that miR-335 is part of a common regulatory pathway for cell proliferation, migration and differentiation in MSCs. The existence of a partially overlapping regulatory circuitry for these biological processes has already been demonstrated in MSCs (17), as well as in other SSC lineages (36). The role of canonical Wnt signaling pathway (Wnt3a) as a positive regulator of miR-335 expression in hMSCs agrees with evidence showing that Wnt3a inhibits osteogenic differentiation in MSCs (17). However, the observed reduction in proliferation capacity in hMSCs overexpressing miR-335 suggests that the reported proliferative effect of Wnt3a acts via miR-335-independent mechanisms. Also, it has been reported that Wnt3a induces ECM invasion in hMSCs (37), although it inhibits it in murine MSCs (38). Further studies are needed to determine the effects of Wnt3a in the migration capacity of hMSCs in different regulatory conditions, and if they are related to miR-335 up-regulation.

In contrast with the effect of Wnt3a, IFN γ decreased miR-335 expression in hMSCs. Considered together with the anti-migratory and differentiation effects of miR-335, this action might account, at least partially, for the previously described pro-migratory and pro-osteogenic activities of IFN γ (19). This cytokine plays a key role in the therapeutic activity of MSCs, as it is involved in the activation of their immunomodulatory phenotype (18). IFN γ signaling, both intercellular and autocrine, might thus regulate an unsuspected large number of genes in MSCs through the inhibition of miR-335 expression. At this time, we do not know the precise molecular mechanism by which IFN γ downregulates miR-335 expression in MSC,

but it might involve an indirect effect through its previously described inhibitory action on the canonical Wnt signaling pathway (39).

Comparative genomics is a powerful new tool for identifying and characterizing functional sequences. The presence in the 5' upstream region of the *MEST* locus of highly conserved potential LEF1 and TCF4-binding sites (95% similarity between human, mouse and dog) and potential STAT1-binding sites is compatible with the regulation of the *MEST*/miR-335 locus by both Wnt3a and IFN γ ; however, further studies are needed to confirm these putative transcription factor binding sites.

Consistent with our experimental results, the set of 62 potential direct gene targets of miR-335 is significantly enriched in genes related to cell movement and gene expression, pointing to a key regulatory role of miR-335 in MSC biology. In particular, we have experimentally validated RUNX2, a transcription factor essential for osteogenic differentiation, as a direct miR-335 target. Recently, miR-204 and miR-211 have been described as negative regulators of RUNX2 (12). Taken together, these results demonstrate that osteogenic differentiation of mesenchymal precursors is regulated by multiple miRNAs through the control of RUNX2 expression levels. Since miR-335 is differentially expressed in MSCs derived from different tissues and donors, it is tempting to speculate that such a difference in miR-335 expression might correlate with their action on bone homeostasis in physiological or in pathological conditions. Further studies are needed to evaluate this possibility.

To our knowledge, this is the first demonstration of the involvement of a specific miRNA in the coordinated regulation of MSC proliferation, migration and differentiation. Our findings are summarized in the model presented in Figure 7. miR-335 is down-regulated in MSCs in response to tissue damage signals, such as IFN γ , which leads to de-repression of miR-335 target genes involved in MSC proliferation, migration, and osteogenic

differentiation. These results have implications for the understanding of the major molecular mechanisms controlling the therapeutic activity of hMSCs *versus* their maintenance in an undifferentiated state, and strongly suggest an important role of miR-335 in tissue homeostasis.

Materials and Methods

Cell Culture. This study was carried out according to guidelines of the Instituto de Salud Carlos III (Spain). hMSCs from bone marrow, adipose tissue, and articular cartilage were isolated and characterized as previously described (40). Human skin fibroblasts were obtained from Inbiobank (San Sebastián, Spain). The 293T and the Saos-2 cell lines were purchased from The American Type Culture Collection (CRL-11268, Manassas, VA). All cell lines were maintained in expansion medium, consisting of Dulbecco's modified Eagle's medium (DMEM) supplemented with 1 g/l D-glucose, supplemented with 10% heat-inactivated fetal calf serum (FCS), 2 mM L-glutamine, and 50 µg/ml gentamicin; all culture reagents were from Invitrogen (Carlsbad, CA). Cells were cultured in a humidified 37°C incubator at 5% CO₂.

For Wnt stimulation, cells were exposed to different concentrations of conditioned medium from Wnt3a-expressing QT6 cells. Control and Wnt3a-expressing QT6 cells were kindly provided by Dr. D. Büscher (Cellerix SL; Madrid, Spain).

When indicated, several factors (DKK1, R&D Systems, Minneapolis, MN; IFN γ , PeproTech, Rocky Hill, NJ) or chemicals (KCl/LiCl, Sigma-Aldrich, St. Louis, MO) were added to the hMSCs cells to evaluate its effect both on the modulation of miR-335 expression or on the several properties of the cells. In each case, the range of concentrations used is indicated.

Lentiviral Vectors. The lentiviral vectors pLV-EmGFP-MIR335 (encoding the human miR-335 gene) and pLV-EmGFP-Mock (encoding a non-specific shRNA sequence) were based on pRRLsin18.PPT.CMV.GFP.Wpre, kindly provided by Dr. Luigi Naldini (41). Detailed construction procedures for both plasmids are provided in Supplementary Information.

Lentiviral particles were produced by transient transfection of 293T cells (41). Lentiviral transduction was carried out by overnight incubation of 3×10^5 hMSCs in 10 cm plates with 3 ml culture medium containing lentiviral particles at a multiplicity of infection (MOI) = 5. Transduced (gfp⁺) cells were purified 48 h after transduction by fluorescence-activated cell sorting (FACS) using a FACSAriaII SORP cell sorter (BD Biosciences).

Transfection of miRNA precursors and inhibitors. Anti-miRTM miRNA inhibitors (Ambion, Austin, TX) are chemically modified, single stranded nucleic acids designed to specifically bind and inhibit endogenous miRNAs. Pre-miRTM miRNA precursors (Ambion) are synthetic double-stranded oligoribonucleotides designed to match the sequence of specific miRNA:miRNA* duplexes. Depending on the experiment, Anti-miRTM or Pre-miRTM oligos at a final concentration of 50 nM were transfected (Lipofectamine 2000, Invitrogen) into 2×10^4 hMSCs cultured in 24-well plates (BD Biosciences, Franklin Lakes, NJ) according to the oligonucleotide manufacturer's protocol.

Real-Time Reverse Transcriptase Polymerase Chain Reaction. Total RNA was isolated from cultured cells with a miRNeasy Mini Kit (Qiagen, Valencia, CA). Human miR-335, *MEST*, *SPP1* (osteopontin), *PPARG*, and *RUNX2* transcripts were quantified by real-time reverse transcriptase polymerase chain reaction (RT-PCR) using the corresponding TaqMan® Gene Expression Assays (Applied Biosystems, Foster City, CA). RNU48 and GAPDH were

used as endogenous normalization controls for miRNAs and protein-coding genes, respectively.

Analysis of cell proliferation, cell cycle, and apoptosis. Cell proliferation was assessed by seeding 6×10^4 cells/2 ml/well in 6-well plates (Corning, Lowell, MA, USA) and culturing for 30 days. The medium was replaced every 3-4 days, and the cells were harvested and counted once a week. Cumulative population doubling (CPD) was calculated as $\log_{10}(\text{number of cells harvested}) - \log_{10}(\text{number of cells seeded})/\log_{10}(2)$.

For cell cycle profiling, approximately 1×10^6 methanol:acetic acid-fixed cells were incubated with 10 μM propidium iodide and 10 μM RNase A for 1 h at 37°C and analyzed by flow cytometry using a Becton-Dickinson LSR cytometer. Data were analyzed with the Summit v4.3 software (Dako Inc, Carpinteria, CA, USA).

For apoptosis analysis, cells were harvested and centrifuged at 2000 x g for 5 min at room temperature, washed twice with PBS and resuspended in 1X binding buffer (10 mM HEPES/NaOH, pH7.4, 140 mM NaCl, 2.5 mM CaCl_2) at a concentration of 1×10^6 cells/ml. 5 μl of Annexin V-DY634 (Immunostep, Salamanca, Spain) were added to 4×10^5 cells suspended in 400 μl of binding buffer. As a negative control, 2×10^5 cells were used without addition of Annexin V antibody. Cells were vortexed gently and incubated in the dark for 15 min at room temperature. Cells were centrifuged at 1000 x g for 5 min, and 1X binding buffer (500 μl) and PI (5 μl) were added to each tube. The samples were analyzed by FACScan (BD Biosciences) using the Cell Quest software (BD Biosciences).

Cell Differentiation Assays. hMSCs were seeded at 2×10^4 cells/cm² in expansion medium, and after 24 h medium was replaced by the corresponding induction medium. Osteogenic medium contained expansion medium supplemented with 10 mM β -glycerophosphate, 0.1

μ M dexamethasone, and 0.2 mM ascorbic acid. Adipogenic medium contained expansion medium supplemented with 0.01 μ M dexamethasone, 0.5 mM IBMX (3-isobutyl-1-methyl xanthine), and 60 μ M indomethacin. In all cases, induction medium was replaced every 3–4 days and, on day 9 or 21 (depending on the experiment) cells were processed for histochemical analysis. Cells were fixed with 70% ethanol (1 h, 4°C) and stained (5 min). For osteogenic assays, cells were stained with 40 mM Alizarin Red, pH 4.1. Cells cultured in adipogenic medium were stained with 2% Oil Red O. Analysis of the expression of marker genes for osteogenic (osteopontin) and adipogenic (PPAR γ) differentiation was performed by real-time RT-PCR as described above.

Cell Migration Assays. For transwell migration assays, hMSCs (1×10^4) were cultured in medium containing 0.2% FCS in a 24-well tissue culture insert with an 8 μ m pore size membrane (BD Biosciences). Fresh culture medium containing 10% FCS was added to the bottom well. After 6 h, the filter membranes were fixed with 70% ethanol and mounted in mounting medium containing DAPI. Migration of hMSCs was determined by counting the number of DAPI-stained nuclei on the underside of the membrane under x 100 magnification, using CellProfilerTM image analysis software (<http://www.cellprofiler.org>).

For wound-healing assays, a "wound" was created in a cell monolayer by scraping with a sterile plastic tip, and time-lapse video recordings were made by sequentially capturing microscopic images at 30 min intervals over a total period of 46 h.

Western blot analysis. Whole cell lysates for western blotting were extracted with lysis buffer containing 50 mM Tris (pH 8), 150 mM NaCl, 1 mM EDTA, 0.1% Triton X-100, and 0.25% sodium deoxycholate and protease inhibitor cocktail (Roche, Rotkreuz, Switzerland). Protein samples were resolved by 10% SDS-PAGE and gels transferred to nitrocellulose

membranes (GE Healthcare, Little Chalfont, UK). Membranes were blocked with 5% non-fat milk for 1 h at room temperature and subsequently incubated overnight at 4°C with primary antibodies at the following dilutions: RUNX2, 1:500 (Sigma-Aldrich) and β -actin, 1:5000 (AbCam, Cambridge, MA). Signals were detected using the appropriate horseradish peroxidase-conjugated secondary antibody (Dako, Glostrup, Denmark). The blots were visualized by an enhanced chemiluminescence system (ECL; GE Healthcare) according to the manufacturer's instructions, and relative intensity of the specific bands were quantified by densitometry using the ImageJ software (<http://rsb.info.nih.gov/ij/>).

UTR reporter assays. The full-length 3'UTRs of the putative miR-335 target gene *RUNX2*, as well as the negative control *UBE2F* and the positive control *SOX4* genes were amplified from human genomic DNA (Novagen, Madison, WI, USA) and individually cloned into the PsichcekTM-2 dual luciferase reporter vector (Promega, Madison, WI, USA). hMSCs were then co-transfected with each reporter construct and with a miRNA synthetic precursor (Pre-miRTM-335 or Pre-miRTM Negative Control #2, Ambion) or antagonist (Anti-miRTM-335 or Anti-miRTM Negative Control #1, Ambion) at a final concentration of 50 nM as described above. Cells were lysed 30 h after transfection and the ratio of *Renilla* to firefly luciferase was measured with the dual luciferase assay (Promega). Normalized *Renilla* to firefly ratios were determined in the presence or absence of miR-335 inhibition.

Microarray Analyses. The Agilent Human microRNA Microarray v2.0 (G4470B, Agilent Technologies) was used to identify miRNAs highly expressed in undifferentiated hMSCs. miRNA expression profiles of undifferentiated hMSCs were compared with profiles of human skin fibroblasts and of the same hMSC lines after 9 days of adipogenic or osteogenic induction.

The Agilent Whole Human Genome Microarray Kit (G4112F, Agilent Technologies) was used to identify genes downregulated in hMSCs exogenously overexpressing miR-335. The mRNA expression profile of hMSCs transduced with lentiviral vector pLV-EmGFP-MIR335 was compared with that of hMSCs transduced with the control vector pLV-EmGFP-Mock.

A full description of the experimental procedures, data processing and statistical analysis used for both types of microarrays is included in the Supplementary information. All microarrays have been submitted to the Gene Expression Omnibus database at <http://www.ncbi.nlm.nih.gov/geo>; accession number GSE19232.

Bioinformatic analysis. Computational miRNA target prediction analyses were carried out using the algorithms miRanda (<http://www.microrna.org/microrna/home.do>), TargetScan (<http://www.targetscan.org>) and PicTar (<http://pictar.mdc-berlin.de>).

Human, mouse and dog *MEST* loci were aligned, and the extent of DNA sequence homology was computed with the web-based program MULAN (<http://mulan.dcode.org>) (42). Using MULAN and multiTF (<http://www.multitf.dcode.org>) with the TRANSFAC professional V10.2 library database (<http://www.biobase.de>), binding sites for LEF1, TCF4 (0.95 matrix similarity) and STAT1 (0.9 matrix similarity) were predicted in the 5 kb upstream region of the human *MEST* locus.

Target gene functions were profiled with PANTHER software (<http://www.pantherdb.org/tools>) and Ingenuity® Pathways Analysis software (<http://www.ingenuity.com>).

Statistical Analysis. Statistical analyses of experimental data were conducted with Prism 3.0 (Graphpad Software Inc., San Diego, CA). Unless otherwise stated, data were compared by

the Mann-Whitney U-test for statistical significance. Results were considered statistically significant at $P \leq 0.05$.

Conflict of interest

The authors declare no conflict of interest.

Acknowledgements. We thank S. Calleja and R. Alvarez (CNIC) for technical help with microarrays and real-time RT-PCR; P. Fernández and J.C. Ramírez (CNIC) for lentivirus production; F. Cabo (CNIC) for bioinformatics and statistical support; C. Carreiro and A. Esteban (CNIC) for preparation and maintenance of plasmid stocks; M. García-Arranz (Hospital Universitario La Paz, Madrid, Spain), C. Trigueros, A. Izeta and A. García (F. Inbiomed, San Sebastián, Spain) for biological samples; S. Bartlett (CNIC) for English revision; and M. Ramón for secretarial support. This work was supported by grants to AB from the Spanish Plan Nacional de Salud y Farmacia/CICYT (SAF 2008-02099), Comunidad Autónoma de Madrid (P-BIO-0306-2006) and Red de Terapia Celular del Instituto de Salud Carlos III (TerCel); MAG is supported by the “Miguel Servet” Program (CP07/00306) of the Instituto de Salud Carlos III (Ministerio de Ciencia e Innovación, Spain); JCS is supported by the ACI-PLAN-E Program (Ministerio de Ciencia e Innovación, Spain). The CNIC is supported by the Spanish Ministry of Science and Innovation and the Pro-CNIC Foundation.

Supplementary information accompanies the paper on the Cell Death and Differentiation website (<http://nature.com/cdd>)

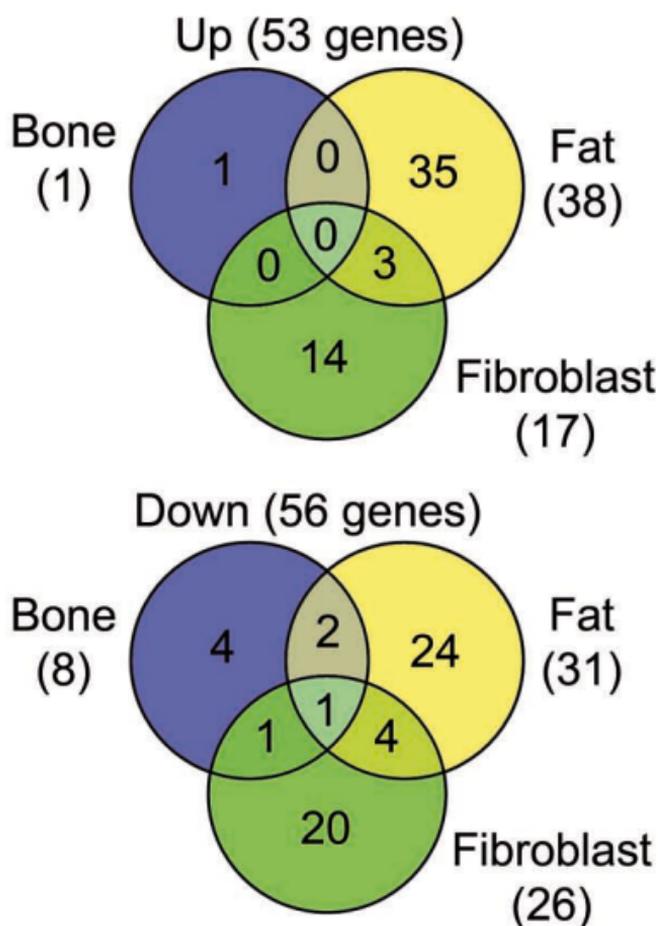
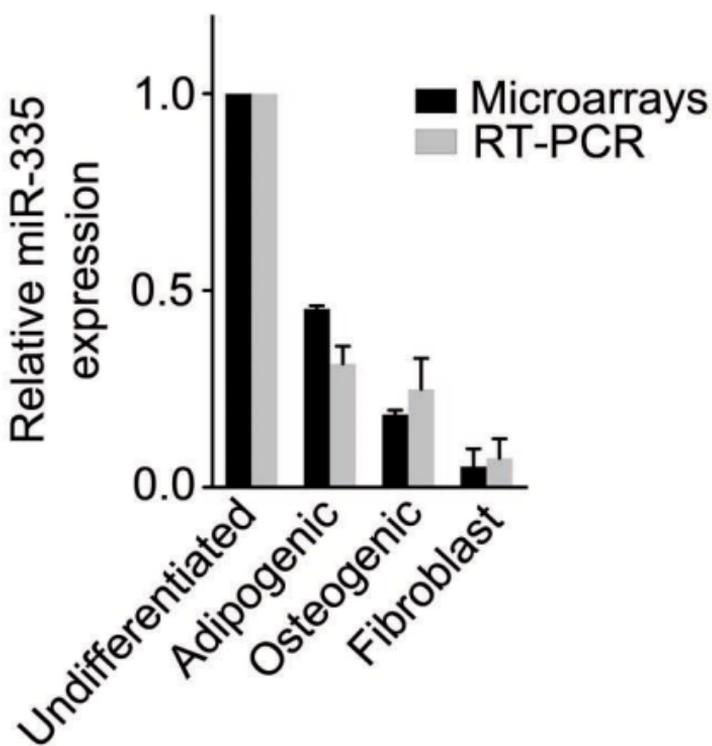
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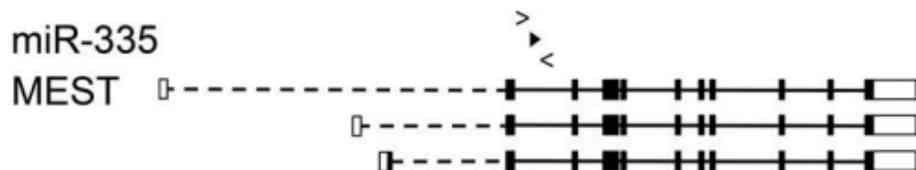
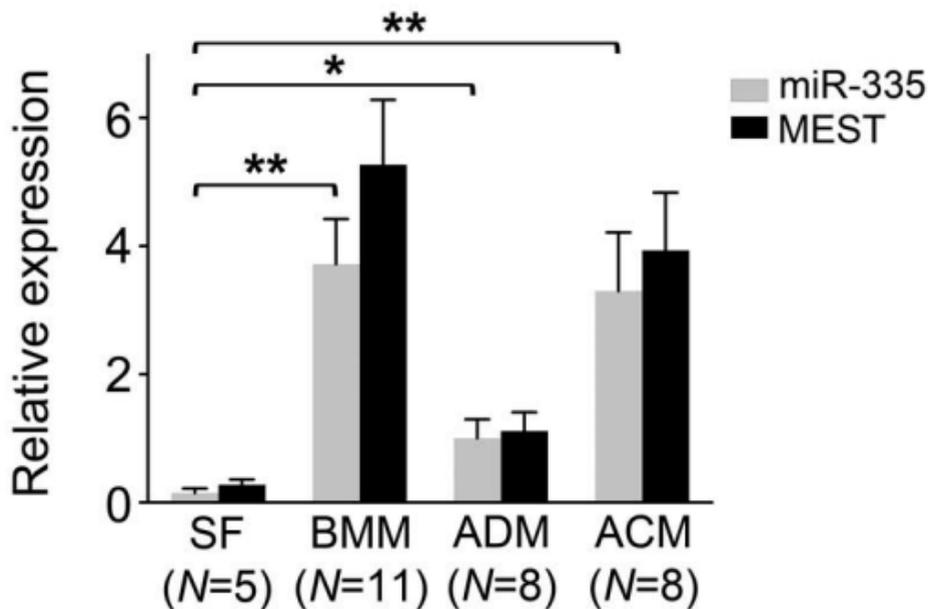
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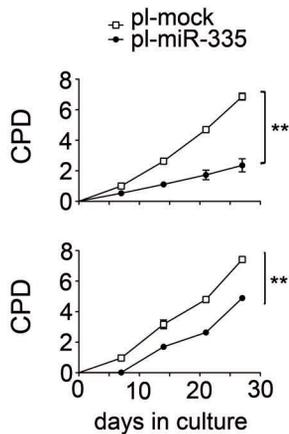
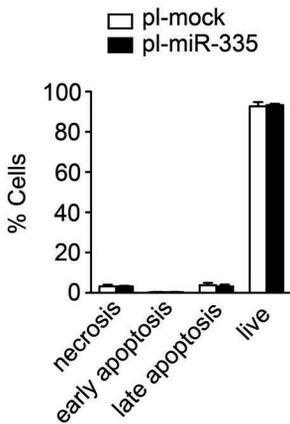
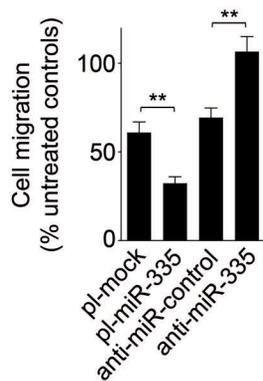
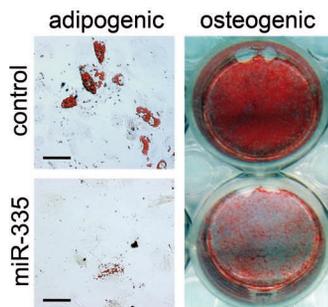
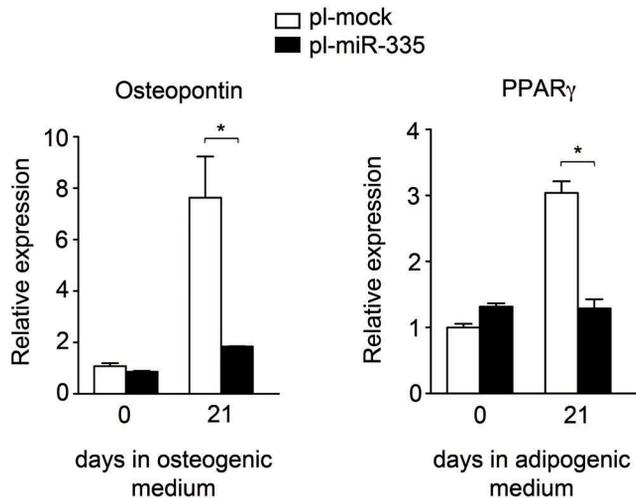
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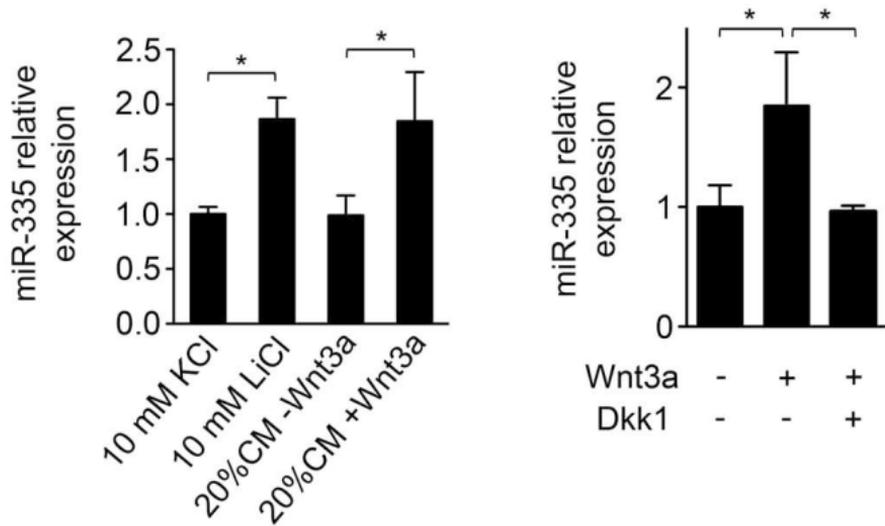
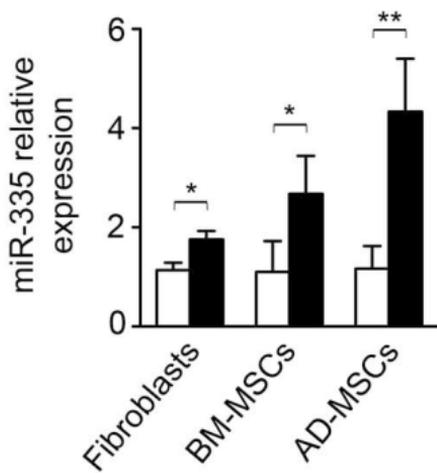
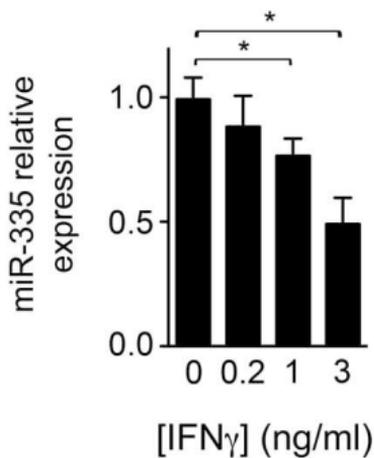
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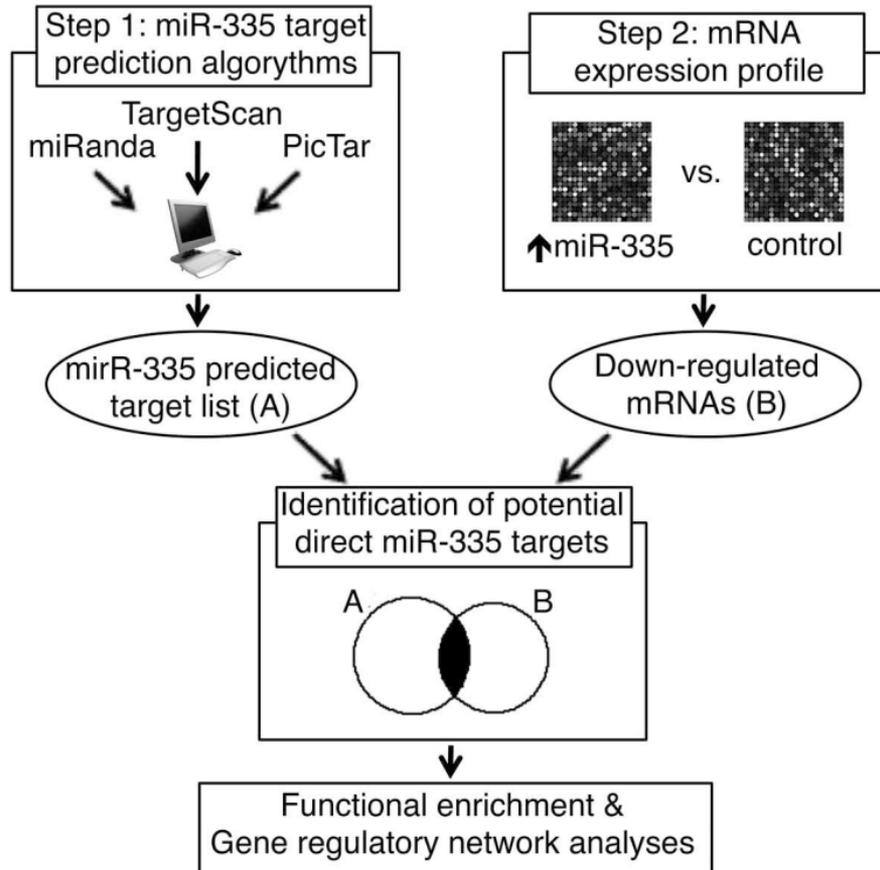
a**DIFFERENTIALLY EXPRESSED miRNAs**
(vs. undifferentiated MSCs)**b**

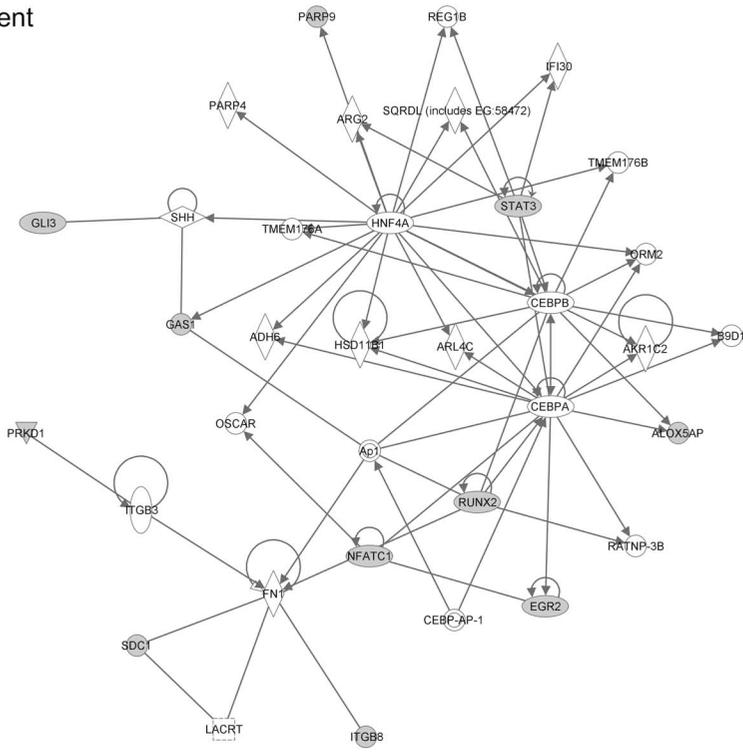
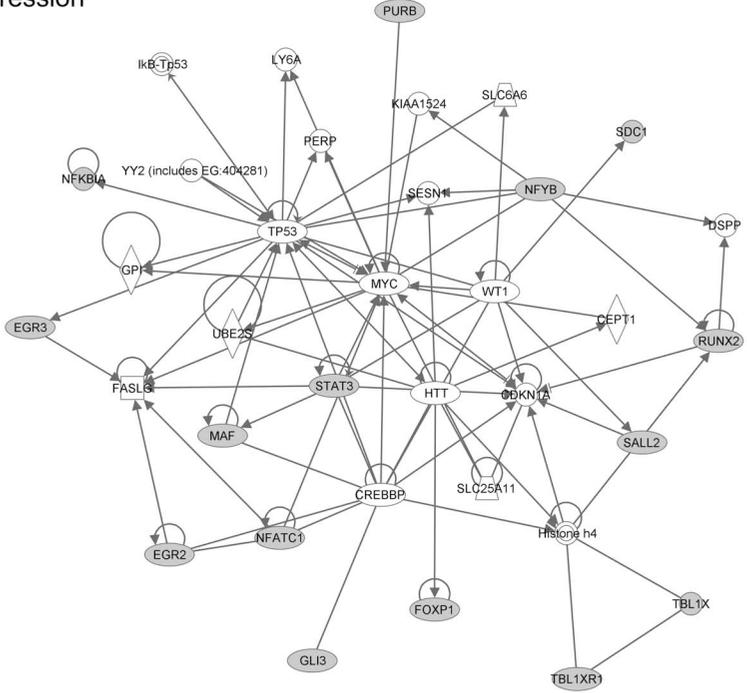
a**b**

a**b****c****d****e**

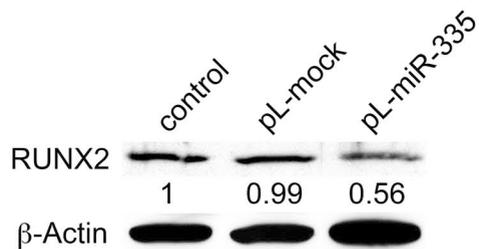
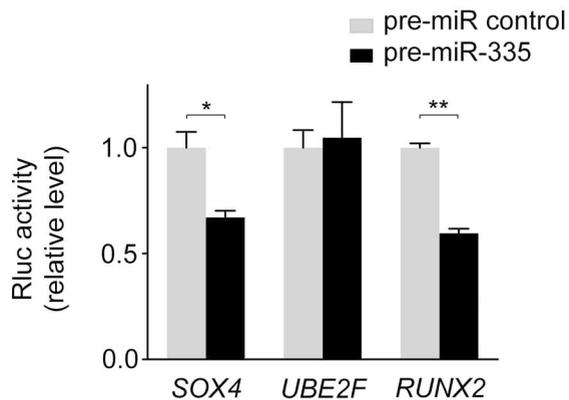
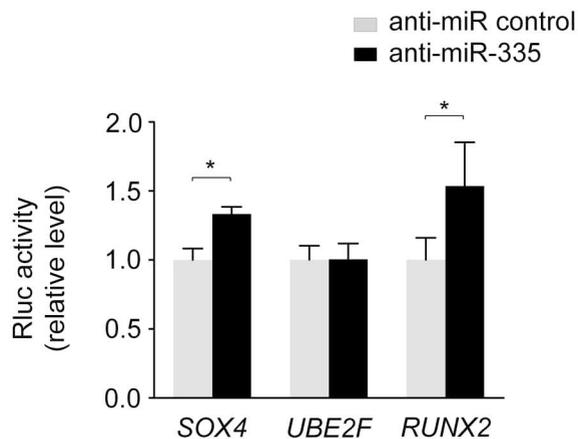
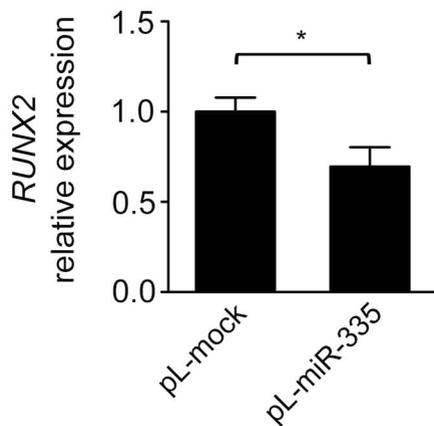
a**b****c**

a



b**Cell movement****Gene expression**

◇ Enzyme	▽ Kinase	○ Transmembrane Receptor
○ Transcription Regulator	△ Phosphatase	◻ Transporter
□ Growth Factor	⊙ Group or Complex	○ Other
◊ Peptidase	◻ G-protein Coupled Receptor	

a**b****c**

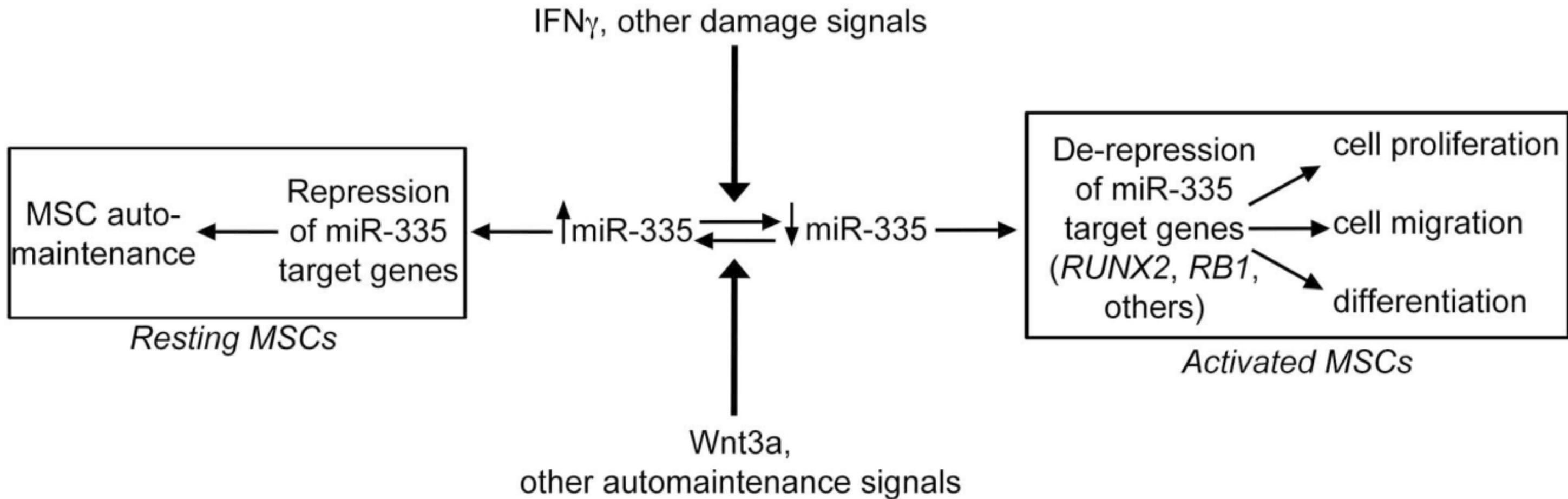


Table 1. Most significantly enriched molecular and cellular functions found using the IPA software in the list of 62 genes identified as potential miR-335 targets in hMSCs (B-H p-value: Benjamini-Hochberg multiple test-corrected p-value).

Relevant functions	B-H p-value	# molecules
Cellular Development	1.37E-02 – 8.82E-02	21
Cellular Movement	1.37E-02 – 8.82E-02	20
Gene Expression	1.37E-02 – 8.82E-02	17
Cell Cycle	1.37E-02 – 8.82E-02	3
Cell Signaling	1.76E-02 – 6.70E-02	4
Cellular Growth and Proliferation	2.47E-02 – 8.82E-02	19
Cell Morphology	2.58E-02 – 8.52E-02	11
Cellular Compromise	2.58E-02 – 5.84E-02	6
Cell Death	2.58E-02 – 8.82E-02	18