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Abstract

Background: Gut hormones peptide YY (PYY) and glucagon-like peptide-1 (GLP-1) play an integral role in appetite control and energy homeostasis. Entero-endocrine L-cells can be stimulated by nutrients and or bile acids to co-secrete PYY and GLP-1. The aim of this study was to determine the response of bile acids, PYY,GLP-1 and ghrelin after a test meal.

Design: Twelve subjects with a BMI of 22.8 (0.52) kg/m² (mean (SEM)) received a 400 kcal test meal after which blood samples were taken every 30 minutes from 0 to 180 minutes. PYY, GLP-1 and ghrelin were measured by radioimmunoassays. Fractionated bile acids were measured by HPLC-MSMS.

Results: PYY positively correlated with glycochenodeoxycholic acid (GCDCA) (rs = 0.23, p = 0.03) and taurochenodeoxycholic acid (TCDCA) (rs = 0.26, p = 0.02). GLP-1 positively correlated with GCDCA (rs = 0.22, p = 0.047) and glycodeoxycholic acid (GDCA) (rs = 0.3, p = 0.005). Ghrelin negatively correlated with GDCA (rs = -0.45, p = < 0.0001), TCDCA (rs = -0.23, p = 0.034) and taurodeoxycholic acid (TDCA) (rs = -0.44, p = < 0.0001).

Conclusion: PYY and GLP-1 responses correlated with chenodeoxycholic acid (CDCA) counterparts, whereas ghrelin negatively correlated with deoxycholic acid (DCA) counterparts. Specific bile acids may thus differentially affect entero-endocrine cells.

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Introduction

Bile acids have several endocrine effects and are involved in three main signalling pathways; mitogenactivated protein kinase (MAPK) pathways (1,2); via the G-protein coupled receptor TGR5 (3,4), and via nuclear hormone receptors such as farnesoid X receptor a (FXRa) (5-7). Administration of bile acids to mice increases energy expenditure in brown adipose tissue via induction of the cyclic-AMP-dependent thyroid hormone activating enzyme type 2 iodothyronine deiodinase (D2) (8). In vitro treatment of brown adipocytes and human skeletal myocytes with bile acids increases D2 activity and oxygen consumption (8). In addition bile acid stimulation of FXRa activates a signalling pathway culminating in increased energy expenditure and decreased adiposity (9-11). In humans circulating bile acid concentrations correlate with measures of insulin sensitivity (12) and administration of bile acids has been found to prevent high fat diet-induced obesity and development of insulin resistance in mice (13). Modulation of bile acid homeostasis using bile acid sequestrants such as cholestyramine and colesevelam have been shown to improve glycaemic control in patients with type 2 diabetes mellitus (14-16). The mechanisms by which bile acids improve glycaemic control are largely thought to be FXRα-independent and are instead mediated by binding to TGR5, leading to cAMP generation and activation of the intracellular type 2 thyroid hormone deiodinase (17). Bile acids also act via the phosphoinositide 3-kinase (PI3 kinase)/AKT pathway, directly promoting insulin signalling and glycogen synthase activation, thus aiding insulin-dependent control of glucose metabolism in the liver (18). In addition bile acid activation of TGR5 has been found to stimulate GLP-1 production in vitro, which promotes insulin secretion and thus improves glucose metabolism.

Bile acids constitute a large family of molecules composed of a steroid structure with four rings and a five or eight carbon side-chain terminating in a carboxylic acid. Each bile acid has a specific number and orientation of hydroxyl groups (-OH). The principal primary bile acids in humans are cholic acid (CA) and chenodeoxycholic acid (CDCA). Ursodeoxycholic acid (UDCA) is the 7β epimer of CDCA. Bile acids can be conjugated to either glycine or taurine, forming bile salts. Glycochenodeoxycholic acid (GCDCA) and taurochenodeoxycholic acid (TCDCA) are the glycine and taurine conjugates of CDCA, respectively and glycocholic acid (GCA) and taurocholic acid (TCA) are the glycine and taurine conjugates of CA, respectively. Bile acids can also be conjugated in phase II metabolic reactions such as N-acetylglucosaminidation at the C7 position with a beta-hydroxyl group (19). Bacteria in the intestinal tract can convert primary bile acids into secondary bile acids by removing a hydroxyl group, for example CDCA becomes lithocholic acid (LA) and CA becomes deoxycholic acid (DCA) (Figure. 1.).

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Gut hormones peptide YY (PYY) and glucagon-like peptide-1 (GLP-1) play an integral role in appetite control and energy homeostasis. PYY and GLP-1 are secreted by L-cells of the distal small intestine in response to nutrients and bile acids (20-22). Exaggerated responses of PYY and GLP-1 may contribute to the weight loss and improvement in glycaemic control following gastric bypass surgery for the treatment of obesity. The mechanisms for increased concentrations of PYY and GLP-1 after gastric bypass are not entirely understood. The 'hindgut hypothesis', described by Mason (23), proposes that altered anatomy following bariatric surgery accelerates the delivery of nutrient-dense chyme to the distal intestine which stimulates increased gut hormones secretion. In addition plasma bile acid concentrations are doubled in individuals after gastric bypass surgery compared to obese individuals who have not had bariatric surgery (24) and it is possible that co-delivery of these bile acids within the chyme enhances the stimulation of gut hormone release post-gastric bypass.

A role of bile acids in appetite suppression was proposed as early as 1968 (25). Several studies indicate that bile acids deoxycholic acid and taurocholic acid combined with fatty acids are more significant than fatty acids alone in stimulating PYY release in the ileum (21,26-29). Bile acid infusions of taurochenodeoxycholic acid and deoxycholic acid have been shown to increase plasma PYY in rabbits (27,30) and deoxycholic acid infusion in the human colon stimulates PYY release in a dose-dependent manner (21). Importantly the concentrations of bile acids used in these studies are consistent with the physiological concentrations of bile acids in the stimulation of PYY release. However, not all bile acids have been found to stimulate PYY release with hyodeoxycholic acid and ursodeoxycholic acid having no effect on PYY release (27,32). The reason why PYY would be stimulated by specific bile acids is not clear. The post-prandial peak concentrations of GLP-1 and total bile acid concentrations inversely correlate with fasting and post-prandial glucose after gastric bypass (24).

Ghrelin is secreted from X/A-like cells of the fundus of the stomach (33,34) and may play a role in regulating pre-meal hunger, meal initiation and long-term energy balance (35,36). Circulating ghrelin concentrations increase during fasting and decrease after eating (37) while anticipation of meals may contribute to its secretion (38). The mechanisms by which ghrelin promotes hunger and increased energy intake are not entirely clear. There is some evidence to suggest that ghrelin may exert its effects by acting on the central nervous system (39). Obese subjects have lower fasting plasma ghrelin concentrations than lean controls (40), although appetite is commonly increased, suggesting that higher body mass is associated with

increased gastric responsiveness to ghrelin (41). A relationship between ghrelin and bile acids has not been described.

The aim of this study was to investigate the associations between the responses of bile acids, PYY, GLP-1 and ghrelin after a standard 400 kcal test meal.

Methods

All studies were performed according to the principles of the Declaration of Helsinki. The Local Research and Ethics Committee at King's College Hospital approved the study (05/Q0703/4). Written informed consent was obtained.

Subjects and Sample Collection

We studied 12 healthy male subjects (age 21.2 (0.46) years, BMI 22.8 (0.52) kg/m² (mean (SEM)). Subjects fasted overnight from 23:00 before ingesting a 400 kcal test meal (46.6g carbohydrate, 28.5 g fat and 10.4 g protein) at 09:00. EDTA blood samples were taken every 30 minutes from 0 to 180 minutes.

Bile acid and hormone measurement

Fractionated bile acids were measured using HPLC-MSMS as previously described by Tagliacozzi *et al* (42). Bile acids analysed were chenodeoxycholic acid (CDCA), deoxycholic acid (DCA), cholic acid (CA), ursodeoxycholic acid (UDCA), lithocholic acid (LA) and their respective glycine (G-) and taurine (T-) conjugates. The method was linear between 0.1-10 μ mol/L for all bile acids and their conjugates with intraassay coefficient of variations (CV) ranging from 3.6 - 8.0 % and inter-assay CVs ranging from 1.5 - 6.8 %. The lower limit of quantitation was 0.1 μ mol/L. LCA and UDCA and their conjugates and CDCA were below the detection limit. Glucose was measured using the glucose oxidase method on the Advia 2400 analyser (Siemens Healthcare Diagnostics, Frimley, UK). Insulin was measured using a two site sandwich chemiluminescent immunoassay on the Advia Centaur analyser (Siemens Healthcare). The intra-assay CV was 3.3% and inter-assay CV was 4.8%. PYY was measured by an established in-house radioimmunassay as previously described by Adrian *et al* (43). The assay measured the biologically active components, both the full length (PYY₁₋₃₆) and the fragment (PYY₃₋₃₆). The detection limit was 5 pmol/L and the intra- and inter-assay CVs were 5.8% and 9.8%, respectively. GLP-1 was measured by an established in-house radioimmunoassay as previously described by Kreymann *et al* (44). The detection limit was 7.5 pmol/L and the intra-assay CV was 6.1%. Ghrelin was measured by the commercially available Phoenix Pharmaceutical assay kit as previously described by Patterson *et al* (45). The assay measured total human ghrelin (acylated and des-acylated) and the intra-assay CV was 8.8%

Statistical Analysis

Results are reported as mean (SEM) and correlation co-efficient (rs). Statistical analysis was performed using Analyse-It for Excel (Analyse-It, Leeds, UK). Data were normally distributed and a Spearman correlation was used. Significance is shown by a P < 0.05.

Results

None of the unconjugated bile acids changed significantly throughout the study period but all glycine and taurine conjugated bile acids and total bile acids increased significantly (Table. 1.). The predominant bile acid measured was GCDCA, followed by GCA and GDCA. All other bile acids had peak concentrations of < 1.00 µmol/L. Table. 2. and Figure. 2. show the changes in plasma hormones and glucose before and after the standardised 400 kcal meal.

Correlations were sought between individual types of bile acids, PYY, GLP-1 and ghrelin. Bile acids which significantly correlated with PYY were GCDCA (rs = 0.23, p = 0.03), total glycine conjugated bile acids (rs = 0.23, p = 0.03), TCDCA (rs = 0.26, p = 0.02) (Figure. 3.), total taurine conjugated bile acids (rs = 0.24, p = 0.03) and total bile acids (rs = 0.26, p = 0.02). Bile acids which significantly correlated with GLP-1 were GCDCA (rs = 0.22, p = 0.047), GDCA (rs = 0.3, p = 0.005), total glycine conjugated bile acids (rs = 0.26, p = 0.02) and total bile acids (rs = 0.26, p = 0.02) (Figure. 3.). Bile acids which significantly correlated with GLP-1 were GCDCA (rs = 0.22, p = 0.047), GDCA (rs = 0.3, p = 0.005), total glycine conjugated bile acids (rs = 0.26, p = 0.02) and total bile acids (rs = 0.26, p = 0.02) (Figure. 3.). Bile acids which significantly correlated with ghrelin were GDCA (rs = - 0.45, p = < 0.0001) (Figure. 3.), TCDCA (rs = - 0.23, p = 0.03), TDCA (rs = - 0.44, p = < 0.0001), total taurine conjugated bile acids (rs = - 0.22, p = 0.044).

Discussion

The CDCA group of bile acids GCDCA and TCDCA correlated with PYY. GCDCA also correlated with GLP-1. GCDCA and TCDCA share a common chemical structure with -OH hydroxylation at C7 and a single hydrogen ion at C12 (Figure. 1.). The positive correlation found between GCDCA and PYY and GLP-1 and TCDCA and PYY suggests that the specific chemical structure of bile acids may play a role in stimulating Lcells. UDCA and its conjugates and CDCA also share this chemical structure but concentrations were below

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the detection limit of our assay and therefore we are unable to comment whether a similar correlation is possible with PYY and GLP-1.

The DCA group of bile acids GDCA, TDCA and TCDCA correlated negatively with ghrelin. DCA counterparts are hydroxylated at both C7 and C12. Bile acids may contribute through an endocrine mechanism to the inhibition of ghrelin secretion from P/D1 cells of the fundus of the stomach post-prandially. Ghrelin and cholecystokinin (CCK) are negatively correlated (46). CCK is a hormone released from the I-cells of the small intestine in response to lipid ingestion which stimulates the release of bile from gallbladder, and also acts as a satiety signal. It is possible that increased CCK levels post-prandially stimulate increased bile acid secretion which may in turn inhibit ghrelin synthesis through neural or endocrine mechanisms.

Limitations of our study included the homogenous nature of our volunteers and therefore the data can not be extrapolated to females or older people. The detection limit of the assay was insufficient for determination of a correlation with UDCA, its conjugates and CDCA. The number of subjects were too small to draw firm conclusions, but the data suggested that the total bile acid and GCDCA response may have been biphasic in 9 of 12 subjects (Figure. 4.). It is possible that with 30 minute sampling the peaks and troughs may have been missed in all subjects. The biphasic response of bile acids in some of our subjects may also have accounted for the low rs values of our correlations.

Conclusion

This study suggests that post-prandial conjugated bile acids correlate with appetite regulating gut peptides PYY, GLP-1 and ghrelin. The post-prandial responses of the CDCA counterparts primarily correlated with PYY and GLP-1 responses, whereas the post-prandial DCA counterparts correlated with changes in ghrelin. Specific bile acids may play a part in entero-endocrine cell stimulation or inhibition. Targeted experiments, specifically with bile acids hydroxylated at C7 may now be considered to determine the therapeutic value of these bile acids in reducing weight through stimulation of PYY and GLP-1 and inhibition of ghrelin.

Table. 1. Baseline and peak values (mean (SEM)) of glycine and taurine conjugated bile acids in plasma

following a 400 kcal test meal.

Bile Acid	Baseline (µmol/L)	Peak (µmol/L)	P value
GCDCA	0.38 (0.05)	3.82 (0.39)	< 0.0001
GDCA	0.11 (0.02)	1.13 (0.22)	0.0006
GCA	0.15 (0.02)	1.45 (0.19)	< 0.0001
Total glycine	0.77 (0.08)	6.97 (0.77)	< 0.0001
conjugated bile acids			
TCDCA	0.03 (0.01)	0.34 (0.14)	0.048
TDCA	0.02 (0.01)	0.16 (0.04)	0.007
ТСА	0.01 (0.01)	0.14 (0.05)	0.022
Total taurine	0.16 (0.09)	0.66 (0.21)	0.012
conjugated bile acids			
Total bile acids	1.41 (0.14)	8.33 (0.85)	< 0.0001

Table. 2. Baseline and peak values (mean (SEM)) of plasma hormones and glucose following a 400 kcal test meal.

PYY (pmol/L) 21.3 (1.98) 34.9 (2.54) < 0.001
GLP-1 (pmol/L) 52.1 (4.63) 72.5 (4.54) < 0.001 Ghrelin (pmol/L) 1026 (133.5) 550 (75.3) < 0.001
Ghrelin (pmol/L) 1026 (133.5) 550 (75.3) < 0.001 Glucose (mmol/L) 4.6 (0.08) 5.6 (0.10) < 0.001
Glucose (mmol/L) 4.6 (0.08) 5.6 (0.10) < 0.001 Insulin (mU/L) 5.7 (1.57) 48.8 (6.42) < 0.001
Insulin (mU/L) 5.7 (1.57) 48.8 (6.42) < 0.001

Figure. 1. The structure of bile acids.



Bile acid	R1 (C3)	R2 (C7)	R3 (C12)	
CA	ОН	ОН	ОН	
CDCA	ОН	ΟΗ (α)	Н	
DCA	ОН	Н	ОН	
LCA	ОН	Н	Н	
UDCA	ОН	ΟΗ (β)	Н	
Legend:				

Legend:

X unconjugated - OH, X tauro- conjugated - NHCH₂CH₂CH₂SO₃H, X glyco- conjugated - NHCH₂COOH, CA cholic acid, CDCA - chenodeoxycholic acid, DCA - deoxycholic acid, LCA - lithocholic acid, UDCA ursodeoxycholic acid



Figure. 2. Changes in plasma levels of gut hormones following a 400 kcal test meal in 12 subjects.

Expressed as mean (SEM).

a) PYY, b) GLP-1 and c) ghrelin





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Figure. 3. Scatter graph demonstrating the correlation between a) taurochenodeoxycholic acid (TCDCA) and PYY; b) total bile acids and GLP-1 and c) glycochenodeoxycholic acid (GDCA) and ghrelin.





Figure. 4. Line graphs demonstrating the response of a) glycochenodeoxycholic acid (GCDCA) and b) total bile acids following a 400 kcal test meal in 9 subjects. Expressed as mean (SEM).



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