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► **To cite this version:**

Nancy Dewi Yuliana, Alfi Khatib, Young Hae Choi, Robert Verpoorte. Metabolomics for Bioactivity Assessment of Natural Products. *Phytotherapy Research*, 2010, 10.1002/ptr.3258 . hal-00599835

HAL Id: hal-00599835

<https://hal.science/hal-00599835v1>

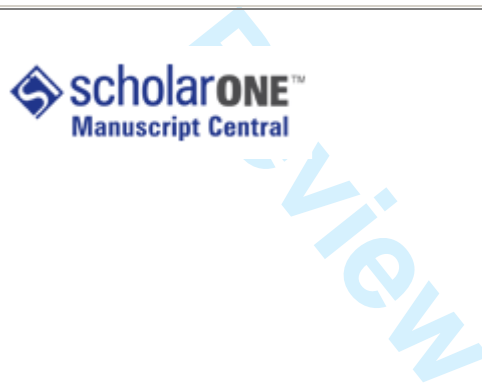
Submitted on 11 Jun 2011

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Journal:	<i>Phytotherapy Research</i>
Manuscript ID:	PTR-10-0117.R2
Wiley - Manuscript type:	Review
Date Submitted by the Author:	30-May-2010
Complete List of Authors:	Dewi Yuliana, Nancy; Institue of Biology, Leiden University Khatib, Alfi; Bogor Agricultural University Choi, Young Hae; Institute of Biology, Leiden University Verpoorte, Robert; Institue of Biology, Leiden University
Keyword:	natural product, drug discovery, metabolomics, bioactivity



Metabolomics for Bioactivity Assessment of Natural Products

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Natural products have historically been a rich source of lead molecules in drug discovery based on their capability to create unique and diverse chemical structures. However, it is also true that the vast number of metabolites typically present in natural products and their huge dynamic range results in the loss of many possibly bioactive natural compounds, becoming an inextricable obstacle for drug development. Recently, new strategies which favor a holistic approach as opposed to the traditional reductionist methods previously used, have been introduced with the purpose of overcoming the bottlenecks in natural product research. This approach is based on the application of new technologies, including metabolomics, for example. Metabolomics allows a systematic study of a complex mixture such as phytochemical preparation, which can be linked to observations obtained through biological testing systems without the need of isolating active principles. This may put drug discovery from natural products back in the limelight again. In this review paper, the description of some examples of successful metabolomics applications in several important fields related to drug discovery from natural sources aims at raising the potential of metabolomics in reducing the gap between natural products (NP) and modern drug discovery demand.

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7 **Key words:** natural product, drug discovery, bioactivity, metabolomics.
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For Peer Review

Introduction

In recent years the development of novel drugs has become increasingly difficult, expensive and time consuming. It is estimated that more than 800 million USD and an average of 14.2 years are needed for a novel drug application (NDA) approval. Both factors are rapidly increasing with time while the output of newly launched drugs has dropped (Butler, 2004; Dickson and Gagnon, 2004). The introduction of high throughput screening (HTS), a process which allows the testing of a large number of samples on certain molecular targets, was expected to shorten the discovery route. To fully use its high potential, HTS requires a large number of compounds to screen, in the order of thousands of samples per day (Mishra *et al.*, 2008). In order to meet this demand, high throughput synthesis and combinatorial chemistry were developed. Concomitantly, further development in the field of structural biology and genomics, e.g. cloning and expression technology resulting in the production of various purified human receptors and enzymes which could be studied at a molecular level, produced a sharp increase in the number of possible molecular targets, demanding more drug-based libraries of a wide chemical diversity to screen (Koehn and Carter, 2005). The combination of these new strategies seemed very powerful and the delivery of a large number of novel leads was expected (Ortholand and Ganesan, 2004). However, these expectations were shown to be greatly overrated as the number of new active compounds that made it to the market in the past two decades reached its lowest level ever in 2002 with an insignificant improvement in 2003 (Newman and Cragg, 2007; Newman *et al.*, 2003; Ortholand and Ganesan, 2004). Sorafenib, a multikinase inhibitor indicated for advanced renal cancer, is the single synthetic drug derived from combinatorial chemistry approved by Food and Drug Administration (FDA) for clinical

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4 use (Newman and Cragg, 2007). The fact, this increased HTS screening capacity and
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6 number of novel molecular targets has failed to produce an increase of new lead
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8 compounds poses the question on whether this is due to limitations in the innovative
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10 aspects of the discovery process itself, e.g. generation of compounds limited chemical
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12 diversity, or because of the potential of natural products (NP) has been de-emphasized
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14 in the past few years (Butler, 2004).
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19 The possibility of combining NP extract libraries with HTS also seems interesting,
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21 though not always an easy task. Several reasons for the incompatibility of NP and HTS
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23 have been described (Butler, 2004; Ortholand and Ganesan, 2004). The complexity of
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25 NP extract which is a complex mixture of mostly uncharacterized compounds which
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27 may mask the bioactivity of target compounds or cause false positive results is the most
28
29 common hurdle. The large dynamic range (e.g. the difference in the levels of major
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31 compounds as compared to that of minor compounds, differences in polarity, in boiling
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33 or evaporating points, etc) which may affect compounds solubility makes the integration
34
35 of NP and HTS more complicated. The differences between areas where plants are
36
37 grown, the methods of production and processing of plants are also important since
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39 these might affect the chemical profile of the NP (Jahangir *et al.*, 2008; Wang *et al.*,
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41 2005). The intellectual property issue in natural product outsourcing is another problem.
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45 Thus, the drug discovery process using plant sources was almost abandoned (Dickson
46
47 and Gagnon, 2004; McChesney *et al.*, 2007). Companies such as Glaxo Wellcome,
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49 Smith Kline & French, and Pfizer, gradually eliminated their NP screening programmes
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51 while others such as Merck and Novartis continued their exploration but focussed on
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53 microbial sources (Ortholand and Ganesan, 2004).
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5 The failure to increase the number of new drugs delivered to the market has placed
6
7 the drugs discovery programs of pharmaceutical companies under scrutiny. Looking
8
9 back to the past two decades, of the 877 novel medicines developed between 1981 and
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11 2002, 6% were NP, 27% were NP derivatives and 16% were synthetic products
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13 developed on the basis of a NP model (Newman *et al.*, 2003), indicating that NP is still
14
15 an important source of novel leads for therapeutical drugs. This situation has led to a
16
17 renewed interest in NP in the drug discovery programs. Therefore, it is a challenging
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19 task to make NP available for the drug discovery arena.
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24 Most NP drug discovery work follows ‘the silver bullet’ approach as is usual in
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26 present Western medicine, in which the search is focused on single compounds with a
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28 particular biological activity (single target-single compound paradigm) tested at
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30 molecular level (e.g. receptor binding inhibition or enzyme screening) (Verpoorte *et al.*,
31
32 2005). This approach which has been responsible for the successful delivery of many
33
34 potent drugs in the past, such as the statins to lower cholesterol level and huperzine A
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36 for Alzheimer’s disease (Li *et al.*, 2008b), is not actually the best approach for NP
37
38 studies since the presence of synergism and pro-drugs in an extract can be misleading
39
40 (Verpoorte *et al.*, 2005). It is also one of the reasons for the long time line and high
41
42 costs which the pharmaceutical companies have to deal with when working with NP.
43
44 Shifting the silver bullet paradigm to a more holistic approach, which is actually the
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46 philosophy of some traditional medicine such as Traditional Chinese Medicine (TCM)
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48 and Ayurveda, is now evolving into a new trend (Ulrich-Merzenich *et al.*, 2007;
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50 Verpoorte *et al.*, 2005; Wang *et al.*, 2005). In a holistic approach, a whole herbal
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52 preparation, which consists of numerous multi components, is viewed as one active
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54 entity with effects on multiple targets in living organisms (Li *et al.*, 2008b; Ulrich-
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4 Merzenich *et al.*, 2007). For such an approach, the response of the whole system needs
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6 to be observed with all possible tools such as clinical trials or experimental animal
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8 models, where both classic physiological observations (e.g. blood pressure, analgesic
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10 activity, sedative) and modern molecular observation (gene expression, proteome,
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12 metabolome) can be used to obtain better insight into different changes in organism.
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14 This is known as a systems biology approach (Verpoorte *et al.*, 2005).
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19 Among these ‘omics’ methods, metabolomics, a relatively new member, is thought
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21 to be the most informative as regards understanding systems biology since it reflects
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23 the genotype (Sumner *et al.*, 2003). Therefore it is a valuable tool to use in the provision
24
25 of the information required in this new approach which is particularly interesting for the
26
27 study of medicinal plants. It thus offers new opportunities for NP in the drug discovery
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29 arena.
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34 This review will discuss the chemical diversity of compounds present in NP as
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36 compared to synthetic compounds, the obstacles which might limit NP integration with
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38 the new emerging drug discovery technologies, the shift in tendency from a reductionist
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40 to a holistic approach in NP research and how metabolomics can help to narrow the gap
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42 between NP and the demand of modern drug discovery, and helping to turn NP and
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44 medicinal plants into a unique drug source.
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49 **1. Natural products as a drug source; their advantages and limitations**

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51 Factors which distinguish NP from synthetic chemicals include their incomparable
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53 molecular diversity and biological functionality (Nisbet and Moore, 1997). Starting with
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55 only a few building blocks of NP provided by the biosynthetic pathways, the action of
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57 various “decorating” enzymes provide a large diversity of end products with different
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4 functionalities (Nisbet and Moore, 1997; Pieters and Vlietinck, 2005; Verpoorte, 1998).
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6 Compared to products resulting from combinatorial chemistry libraries which usually
7
8 lack chirality and structure rigidity, two important requirements for pharmaceutical
9
10 drug-like properties (Nisbet and Moore, 1997), NP have privileged structures which are
11
12 able to interact with proteins and other biological targets for specific purposes (Koehn
13
14 and Carter, 2005).
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19 To address the question of how the unique structural properties of NP are able to
20
21 complement the chemical space of synthetic compounds, the comparison of the major
22
23 structural differences between combinatorial compounds, drugs, and NP molecules has
24
25 been made. It is obvious that current commercial drugs and NP share more similarities
26
27 in several topological pharmacophore patterns. The important differences between NP
28
29 and synthetic compounds include the number of chiral centers, the prevalence of
30
31 aromatic rings, the presence of complex ring systems, and the degree of the saturation of
32
33 the molecule as well as the number and ratios of different heteroatoms (Feher and
34
35 Schmidt, 2003). Natural products have a higher number of chiral centers, a higher
36
37 number of oxygen atoms instead of nitrogen, sulfur and halogen containing groups
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39 present than in synthetic compounds. Natural products have a lower ratio of aromatic
40
41 ring atoms to total heavy atoms, and a higher number of solvated hydrogen bond donors
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43 and acceptor functional groups. These characteristics are favorable attributes for a drug
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45 candidate profile (Feher and Schmidt, 2003; Henkel *et al.*, 1999; Newman and Cragg,
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47 2007; Newman *et al.*, 2003). With these unique properties, in general, NP actually have
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49 more drug-like properties than synthetic compounds (Feher and Schmidt, 2003; Koehn,
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51 2008).
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However, a NP-based drug discovery project also poses some challenges, mostly connected with their presence within a complex matrix with all kind of active compounds (Bindseil *et al.*, 2001; Koehn, 2008; Koehn and Carter, 2005; McChesney *et al.*, 2007; Ortholand and Ganesan, 2004; Pieters and Vlietinck, 2005; Schmid *et al.*, 1999):

(1) The first challenge is dereplication which is the rapid identification of known and unknown compounds from partially purified mixtures. In a number of test systems, common plant products which have been found to be active e.g. unsaturated fatty acids on adenosine and GABA receptors or tannins on enzymes based bioassays, have to be identified at the very beginning of the study of an active extract.

(2) Interaction between metabolites present in extracts. Extracts consist of mixtures of many compounds requiring an elaborate purification to isolate new pure compounds. Another problem of complex extracts is the presence of antagonism or synergisms which might occur between components in a mixture, posing a major intellectual challenge which can be painstakingly time consuming.

(3) Structure determination by spectroscopic methods.

(4) Large scale production of the active component is sometimes an obstacle as the identified NP lead might be a minor compound of the extract, or the supply of the material for extraction might be limited.

(5) Sourcing of authenticated plants, a very important issue for quality control. For example, some plants have congener substitutes which have an active principal content which may be different to the original one.

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4 Last but not least, biodiversity and intellectual property issues can be another
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6 complicating factor related to this context.
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10 11 **2. Previous efforts to increase the number of compounds for drug candidates**

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13 Because of the demand to speed up the drug discovery timeline and to reduce costs, the
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15 *in vitro* screening system has become the preferred method. The use of animal tests or
16
17 isolated organs was abandoned for more specific testing targets at a molecular level
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19 (reductionist mode) (Verpoorte *et al.*, 2006). Following this trend, there are two crucial
20
21 prerequisites to start the drug discovery process: a large high quality library of
22
23 candidates to be screened, and a reliable and sensitive assay to screen the activity.
24
25 Combinatorial chemistry was developed to address the first need (Williard *et al.*, 1996).
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27 Based on the total chemical space, this technology is theoretically able to provide 10^{30} –
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29 10^{100} compounds (Macarron, 2006; Myers, 1997).
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35 The second need was met by high throughput screening (HTS) as the response to the
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37 availability of hundreds to thousands of compounds resulting from combinatorial
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39 chemistry. HTS is a fast and cost-effective technology designed to select drug
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41 candidates (Bleicher *et al.*, 2003; Gómez-Hens and Aguilar-Caballo, 2007). It is
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43 defined as an automatic approach in drug discovery which is characterized by
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45 miniaturization to allow the use of minute quantities of samples and reagent.
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47 Continuous innovations keep taking place to maximize the quantity of screened
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49 compounds and to minimize the cost. The miniaturization of screening assay aimed at
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51 increasing HTS capacity is achieved by increasing the screening plate density and
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53 minimizing screening volume (Landro *et al.*, 2000). The system which enables the
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55 screening of 120000 -240000 compounds per day has been created by the use of high
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4 density 1536 wells plates. Higher capacity plates of 3456 wells are also available but
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6 used rarely due to the limitation determined by the low liquid volume delivery
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8 (Bajorath, 2002; Smith, 2002; Williams, 2004). Compounds found to be active in HTS
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10 are called “hits”, but the distance from a hit to a lead is very great. This requires more
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12 extensive screening for other activities, including toxicological evaluations. In this
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14 process similar compounds will be produced to optimize the activity profile.
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19 However, compounds obtained from the screening are not always (physico)
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21 chemically and pharmacologically suitable for further development as a lead. While the
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23 *in vitro* profile is maybe very promising, ADMET (absorption, distribution, metabolism,
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25 elimination, toxicity) properties are often poor (Bleicher *et al.*, 2003). In most cases the
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27 compound will require modifications to become a lead and drug candidate. Thus, the
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29 crucial bottleneck of lead development is not only hit identification, but the potential of
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31 the candidate to become a new drug (Bleicher *et al.*, 2003).
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36 Therefore it is necessary not to focus only on quantity but also consider quality of
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38 both library and assays. One suggestion made on this matter, was to introduce simple
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40 ADMET parameters at the early stage of screening and the more sophisticated ones at a
41
42 later stage (Walters and Namchuk, 2003). The appropriate application of virtual
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44 screening as a complement to a screening campaign to enrich the hit rate and reduce the
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46 number of false positive has been suggested as well (Bajorath, 2002; Bleicher *et al.*,
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48 2003; Entzeroth, 2003; Smith, 2002). Most virtual screening is guided by the structure
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50 of the target protein and 3-dimensional descriptors of small molecular structures that fit
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52 into the protein active site. Further approaches such as the identification of active
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54 compounds by clustering and partitioning techniques, compound filtering to eliminate
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56 undesired compounds (e.g. toxicant and groups with poor aqueous solubility and
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4 passive absorption) can also be applied (Bajorath, 2002). Understanding the mechanism
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6 of action of a biological target is another approach. Several techniques have been
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8 reported, such as immobilization and stabilization of the target protein upon its ligand
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10 binding, as well as the use of nuclear magnetic resonance (NMR) spectroscopy coupled
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12 to other analytical methods to learn about the drug-protein interaction (Entzeroth, 2003).
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14 For the targets without sufficient bio-structural information, such as in the area of G-
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16 protein coupled receptors (GPCRs), a ligand motif-based library design approach can be
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18 applied (Bleicher *et al.*, 2003)
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24 The aforementioned efforts, however, did not successfully fulfill the high expectation
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26 of increasing productivity. The tremendous amount of money spent was not
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28 counterbalanced with the number of new approved drugs (Smith, 2002). However, all
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30 these methods will result in all companies having the same basic information on targets
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32 and probably ending up with the same compound, reducing the chance for real
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34 innovation with this approach will be limited. This has encouraged reconsidering NP as
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36 a drug source.
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42 **3. Efforts to combine natural products with new technologies**

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44 Many studies reported possible approaches which could potentially help to overcome
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46 the above mentioned problems in the NP drug discovery process. Solutions to address
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48 the incompatibility of NP extracts with HTS technology were proposed, including the
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50 generation of a high quality NP compound library, the improvement of fractionation
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52 methods, and of dereplication and identification steps.
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59 **3.1. Constructing a high quality natural product library**

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Several strategies to construct better NP libraries and to improve methods to screen these libraries have been reviewed. The advantages and limitations of a crude extract library, pre-fractionated library, or pure compound library have been described (Koehn, 2008). Crude extract libraries are simple to make, the obstacles appearing in the identification and dereplication steps. A pre-fractionated library can be effective to overcome these hurdles since the less complex fractions require fewer purification steps and the interference of unwanted compounds is reduced, while more investment is required both for fraction preparation and for the HTS assay since several samples are obtained from a single extract. The pure compound library is preferred since it is able to move forward directly after finding a hit.

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The strategy employed to build a fraction library as a compound source for HTS from Chinese herbal formulations reported to treat atherosclerosis has been described. Medium pressure liquid chromatography with a gradient solvent system followed by reverse phase semi-preparative HPLC was used as a filtering method to remove unfavorable compounds lacking desirable ADME properties and to prepare semi-crude extracts with different polarity. The fractions were characterized and identified by LC-DAD-ESI-MS. A component detection algorithm (CODA) was used to reduce noise and background of the chromatograms which then were presented as a histogram describing compound molecular distribution. Small molecules (less than 500 Da) were characterized by retention time, molecular weight, UV absorbance, and crude drug origin. In this way, 36 major compounds of each fraction were identified and after cell based multiples bioassay screening, validation and dereplication steps, 9 hits were found (Liu *et al.*, 2008).

3.2. Improving fractionation methods

Another study focused on sample fractionation using an automatic multiple solid phase extraction (SPE) method (Schmid *et al.*, 1999). Several different solid phases were used thus reducing the complexity of an extract from a microbial fermentation broth, leading to a more reliable and reproducible screening result.

3.3. Improving dereplication and identification steps

A fast dereplication and identification of novel active compounds can be achieved by direct coupling of the bioassay to liquid chromatography (Bajorath, 2002; van Elswijk and Irth, 2002). An example is the post-column acetylcholine esterase (AChE) inhibitor assay. After centrifugal partition chromatography (CPC) pre-fractionation and bioactivity testing, the most active *Narcissus* 'Sir Winston Churchill' fraction was injected into an online biochemical detection system coupling HPLC--UV-MS for AChE inhibitor activity. This system enabled a fast dereplication, allowing the identification of known compounds. It proved very useful for the detection of novel active compounds in the presence of known active compounds. For example, beside the well known galantamine, another active compound, unguiminorine, was identified (Ingkaninan *et al.*, 2000). A similar technique was applied to screen for natural phosphodiesterase and estrogen receptor ligands (Schenk *et al.*, 2003; Schobel *et al.*, 2001).

Sometimes, the information of molecular weight and UV spectra provided by combination of LC-DAD-MS is not sufficient for the identification of active compounds. The use of electrospray ionization Fourier Transform Ion Cyclotron Resonance MS (ESI-FTICR-MS) as a rapid and informative approach for direct affinity

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4 screening of bovine carbonic anhydrase II (bCAII) ligands from extracts was reported
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6 (Poulsen *et al.*, 2006). The bCAII-ligand affinity is based on the non-covalent binding
7
8 between the enzyme and the ligand. The electrospray ionization technique is known as a
9
10 gentle ionization process that can transfer intact non-covalent complexes from the
11
12 solution phase into the gas phase while the high resolution of the FTICR-MS allows the
13
14 very accurate deduction of mass-to-charge ratio (m/z) of the protein-ligand complex
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16 from which even the molecular weight of this ligand can be calculated. This method was
17
18 applied by separating small molecules from macromolecules by size exclusion
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20 chromatography (SEC), after the protein was added to the extract. This protein-ligand
21
22 complex was then subjected to online SEC-ESI-FTICR-MS or directly infused into ESI-
23
24 FTICR-MS, allowing the rapid determination of molecular weight of the active
25
26 compound with an error below 0.08 Da. Bioactive compounds which were present in
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28 the mixture could be detected down to the level of 0.02% dry weight. HPLC coupled to
29
30 an on-line postcolumn antioxidant detection system in combination with absorbance
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32 spectrometry and quadrupole time-of-flight high-resolution MS (QTOF-MS) allowed
33
34 the identification of compounds responsible for the antioxidant capacity of raspberry
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36 (*Rubus idaeus*) and their level during raspberry ripening stages and in different cultivars
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38 and different genetic background (Beekwilder *et al.*, 2005).
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47 In a recent report, NMR based screening of bioactive NP and structure elucidation of
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49 the active compound in a single step was described (Politi *et al.*, 2005). This study
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51 aimed at detecting ligands from plants that bind to a specific target protein, lectin. The
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53 interaction between a ligand present in the plant extract was detected by comparing the
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55 saturation transfer difference (STD) and NOESY spectra of target protein with and
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57 without extract. The most important structural information was provided by two
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4 dimensional NMR data such as TOCSY, HSQC, HMBC and selective 1D-COSY.
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6 Several LC-NMR hyphenation techniques and their application to advance plant extract
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8 profiling have been described (Exarchou *et al.*, 2005). The advantage and limitation of
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10 continuous flow LC-NMR, stop flow LC-NMR, LC-NMR/MS, LC-SPE-NMR, and
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12 application of cryogenic and microflow technology have also been reviewed. The
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14 performance of these LC-NMR coupling technologies depends on many factors such as
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16 the physical connection of LC-NMR, flow-through probe design, and factors affecting
17
18 NMR sensitivity (solvent compatibility, solvent suppression, the adequate sample
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20 volume). Practical examples of the application of various hyphenated LC-NMR
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22 techniques for rapid screening, dereplication, and structure determination of active
23
24 compounds from natural extract has been published (Wolfender *et al.*, 2006).
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33 **3.4. Virtual screening**

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35 The application of virtual screening of known NP has been proposed (Rollinger *et al.*,
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37 2008). NP are still an incredible rich source of new drugs and because of the better
38
39 understanding of protein-ligand interaction properties, NP virtual screening can
40
41 constitute an alternative way to accelerate the drug discovery process. Many advantages
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43 have been described including higher capacity, reduction in the number of experiments
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45 for testing and isolating compounds, the ability to calculate the possible interaction of
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47 compounds and targets, and the possibility of applying additional filters such as ADME
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49 and drug-like properties to improve the hit quality. Strategies to combine virtual
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51 screening and classical methods in NP exploration were proposed; (1) Direct screening
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53 of a 3D database of NP using available target information and pharmacophore model of
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55 specific activity, followed by the isolation of hits once the results are confirmed by
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4 experimental data. (2) Extracts were roughly screened experimentally to select the
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6 active ones, or the specific activity was explored through an ethnopharmacological
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8 approach, the result of which was then virtually screened using a pharmacophore model
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10 of the target, followed by isolation.
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13 14 15 16 **4. Metabolomics** 17

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19 Metabolomics is one of the new ‘omics’ fields, joining genomics, transcriptomics, and
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21 proteomics (Rochfort, 2005). The general aim of metabolomics is the qualitative and
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23 quantitative analysis of all metabolites (the metabolome), present in an organism, at a
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25 specific time and under specific influence (Colquhoun, 2007; Hall, 2006; Mendes *et al.*,
26
27 2005; Ulrich-Merzenich *et al.*, 2007; Verpoorte *et al.*, 2007). A metabolome is the entire
28
29 set of small molecules present in a cell, tissue, or organism under certain conditions and
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31 at certain point in time (Ulrich-Merzenich *et al.*, 2007).
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35 Metabolome components can be viewed as gene expression end products, thus its
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37 quantitative and qualitative measurement may provide a broad insight of the
38
39 biochemical status of an organism and can be used to monitor gene function (Sumner *et*
40
41 *al.*, 2003). The use of transcriptomics and proteomics profiles to predict gene function
42
43 has been criticized because of limitations such as the fact that the changes in the
44
45 transcriptome or proteome do not always correlate to biochemical phenotypes or that the
46
47 translated protein may or may not be enzymatically active (Nobeli and Thornton, 2006;
48
49 Rochfort, 2005; Sumner *et al.*, 2003). The lack of a data base source may result in
50
51 limited information since the identification of mRNA and proteins is based on sequence
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53 similarity or database matching. Therefore, among the ‘omics’ technologies,
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55 metabolomics can be viewed as the most functional approach (Sumner *et al.*, 2003). The
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4 combined information obtained from genomics, proteomics and metabolomics should
5
6 be able to provide an integrated understanding of a cell or organism (Sumner *et al.*,
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8 2003; Ulrich-Merzenich *et al.*, 2007).
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11 12 13 14 **4.1. Detection method**

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16 Compared to proteins and RNA, secondary metabolites have a much more diverse
17
18 atomic arrangement which results in broad variations in chemical and physical
19
20 properties, such as molecular weight, polarity, solubility, and volatility (Dunn and Ellis,
21
22 2005), and can be present in a large range of concentrations (Moco *et al.*, 2006). This
23
24 generates technological difficulties in the simultaneous profiling of all metabolites
25
26 (Sumner *et al.*, 2003). There are thus considerable challenges in metabolomics
27
28 experiments such as sampling, sample preparation, instrumental analysis, and then in
29
30 the interpretation of the tremendous amount and diversity of data arising from the
31
32 metabolomics analysis (Bhalla *et al.*, 2005; Rochfort, 2005), the construction of
33
34 consolidated metabolite libraries and the development of metabolite-specific data
35
36 management systems (Moco *et al.*, 2006). Therefore, metabolomics based work still
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38 depends on further improvement of sampling, analyzing and computational technologies
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40 (Bhalla *et al.*, 2005; Rochfort, 2005).
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47 The identification and quantification techniques applied to metabolomic based work
48
49 have been comprehensively reviewed (Bhalla *et al.*, 2005; Fiehn, 2002; Kell, 2004;
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51 Sumner *et al.*, 2003; van der Greef and Smilde, 2005; Verpoorte *et al.*, 2008). Many
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53 analytical methods with better reliability, robustness, selectivity, and resolution, have
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55 been developed in which instead of targeting on a few specific compounds, the aim is to
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4 identify and quantitate multiple targets in order to obtain an overview of compounds
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6 classes (Fiehn, 2002).
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9 The progressive development of ionization and mass detection methods, and the
10 ability to detect a wide range of molecular weight and polarities have become a strong
11 point for the use of MS and tandem MS techniques in metabolomics (Verpoorte *et al.*,
12 2008). GC coupled to MS (GC-MS) has been used to identify and elucidate the
13 structures of 326 different compounds from *Arabidopsis thaliana* leaf (Fiehn *et al.*,
14 2000) and to identify and classify 322 tomato volatiles from 94 genotypes according to
15 their biochemical precursor similarity (Tikunov *et al.*, 2005)
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26 Liquid chromatography (LC) is an alternative tool particularly for non-volatile plant
27 metabolites though it requires more technical support than GC (Dunn and Ellis, 2005;
28 Nobeli and Thornton, 2006). An open access database dedicated to LC-MS based
29 metabolomics of tomato fruit (*Solanum lycopersicum*) has been presented. A C18 base
30 RP LC-photodiode array detection (PDA)-electrospray ionization (ESI)-quadrupole
31 time-of-flight (QTOF)-MS method was developed for the detection and putative
32 identification of predominantly semipolar secondary metabolites. Some previously
33 unreported novel compounds from tomato fruit were identified by this means (Moco *et*
34 *al.*, 2006).
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47 The use of laser desorption/ionization mass spectrometry on porous silica (DIOS)
48 and cluster analysis to discriminate the metabolic footprints of mutant yeast and wild
49 type yeast has been reported (Vaidyanathan *et al.*, 2005). Compared to matrix- assisted
50 laser desorption/ionization (MALDI) mass spectrometry, DIOS-MS is much simpler,
51 allowing the analysis of small molecules in absence of matrix ions with little or no
52 fragmentation and better tolerability to moderate amounts of contaminants in the
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4 biological sample. Both methods are also especially effective for analysis of proteins
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6 and their derivatives (Thomas *et al.*, 2001).
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10 Although MS sensitivity is probably the best among analytical methods, there are
11 some limitations for its application in metabolomics. For example, the different
12 sensitivity of the detection of molecules and its reproducibility are important issues.
13
14 Because of different detector response, calibration curves are needed for each individual
15
16 compound for absolute quantification, this being totally unrealistic considering the
17
18 hundreds of signals that appear in a metabolomic profile. In this case, NMR
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20 spectrometry is the method of choice, in absence of which only a relative
21
22 quantification of the individual compounds in different experiments can be made
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28 (Verpoorte *et al.*, 2008)
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31 The application of NMR for metabolic profiling of plant tissue extracts has been
32
33 reviewed by several authors (Colquhoun, 2007; Verpoorte *et al.*, 2007; Ward *et al.*,
34
35 2007). Reproducibility is the most important criteria for developing a metabolomics
36
37 technology platform, and NMR is the most suited choice even though its sensitivity is
38
39 not as high as MS (1 μM – 1 mM in NMR tube) (Colquhoun, 2007; Verpoorte *et al.*,
40
41 2007). The simple and fast sample preparation, short measurement time, the availability
42
43 of less automatization and advanced data analysis methods, plus the possibility to
44
45 elucidate structures of known or unknown compounds in a complex mixture using
46
47 advanced two-dimensional (2D) NMR methods, are further advantages (Colquhoun,
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49 2007; Verpoorte *et al.*, 2007).
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54 55 56 **4.2. Data processing** 57 58 59 60

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Data resulting from metabolomics based work are typically high dimensional data which require multivariate data analysis (MVDA) methods for interpretation. Most metabolomics data analysis methods are based on the classification of samples into different groups (e.g. treatment, genotype), both by supervised (e.g. discriminant function analysis, artificial neural networks) or unsupervised type data analysis method e.g. principal component analysis (PCA), hierarchical cluster analysis (HCA) (Nobeli and Thornton, 2006). It is also possible to use MVDA to make a regression modeling between two blocks of data, usually denoted as X and Y. In metabolomics based NP studies, X may represent signals from different metabolites present in plant extracts which are sampled at regular time intervals, and Y the responses (e.g. quality of product, bioactivity, yield). The model then can be used to predict Y from X which is obtained from new observations. The most common MVDA method for this modeling is partial least square (PLS) method (Wold *et al.*, 2001a). Some extended versions of PLS to improve the predictability power were developed recently, such as orthogonal signal correction (OSC), orthogonal-PLS (OPLS), and orthogonal-orthogonal-PLS (O2PLS) (Gabrielsson *et al.*, 2006; Wold *et al.*, 2001b)

5. Various applications of metabolomics in bioactivity screening of natural product

5.1. Metabolomics to improve identification and dereplication steps

The reductionist approach as described above is still widely applied in many NP drug discovery projects, i.e. in bioassay guided isolation work. In this approach, the rapid dereplication of known actives and identification of novel actives is essential. Addressing this issue, some papers have described the way in which a metabolomics approach consisting in the use of various analytical methods followed by appropriate

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4 MVDA can be used to shorten the reductionist bioassay driven isolation route,
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6 especially in the identification and dereplication step.
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9 In one study, the hierarchical cluster analysis of the chromatographic profiles of 500
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11 lipidic extracts of bacteria, resulted in the identification of 2 new
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13 oligoprenylsquaresquiterpenes and 2 new N-(2-arylethyl) amides, though the bioactivity
14
15 profile was not reported here. HCA was chosen instead of PCA since the number of
16
17 principal components representing more than 50% of the variance was much too high
18
19 for PCA. PCA is more suitable for a large data set of similar compounds, which was not
20
21 the case in this project. The use of this clustering method was able to advance the
22
23 dereplication step and to avoid redundant analyses for selection of strains containing
24
25 new NP (Böröczky *et al.*, 2006).
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30 The most time consuming step in metabolomics based work is the identification of
31
32 metabolites in a prefractionated mixture. The availability of a compound spectral library
33
34 is crucial for rapid identification. A free web-based tool containing data related to
35
36 primary and secondary metabolites from various species based on NMR or MS data, the
37
38 Madison Metabolomics Consortium Database ((MMCD) was developed (Cui *et al.*,
39
40 2008). The sensitivity of MS and the feasibility of NMR spectroscopy, including the
41
42 application of two dimensional (2D) NMR followed by MVDA application, make these
43
44 techniques particularly well suited for the characterization of crude unfractionated NP.
45
46 The analysis based on double quantum filtered correlation spectroscopy (DQF-COSY)
47
48 was introduced to simplify the analysis of 2D NMR spectra and has demonstrated its
49
50 utility to detect previously unreported terpenoid indole alkaloids from *Tolypocladium*
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52 *cylindrosporum* grown in different media. Graphical analysis of the obtained spectra
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54 through multiplicative stacking and graphical manipulation of the bitmap spectra
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4 allowed signals only present in a specific media to be distinguished. Further structure
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6 elucidation was then performed by heteronuclear single quantum correlation (HSQC)
7
8 and heteronuclear multiple bond correlation (HMBC) spectra analysis. This method is
9
10 considerably more efficient than spectra subtraction since results in a clear color
11
12 alteration and partially obsolescence of a common signals present in two spectra even if
13
14 those are present in a distinct different concentration (Schroeder *et al.*, 2007). The big
15
16 size of data limits the application of this method. Three different clustering methods, i.e.
17
18 tree clustering analysis (TCA), K-means clustering and multidimensional scaling, were
19
20 used to cluster together partially purified plant and marine organism extracts which
21
22 contain the same bioactive compound or groups of compounds. These clustering
23
24 methods were chosen to overcome limitations of PCA to handle the data with a large
25
26 number of clusters or subsets within the data (Pierens *et al.*, 2005). Only a few of the
27
28 secondary metabolites from *Myxococcus xanthus* have been investigated, in particular
29
30 some antibiotics such as myxovirescin, saframycin, and althiomycin. However, only
31
32 one metabolite unique to *M. xanthus* was known, that is DKxanthanes. Intraspecific
33
34 diversity in the metabolite profiles of 98 *Myxococcus xanthus* strains originated from
35
36 78 locations worldwide was rapidly explored by means of fast chromatographic
37
38 separation (UPLC)-coupled ESI-TOF-MS and PCA. As a result, 37 candidates for novel
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40 compounds were identified but their bioactivities were not reported (Krug *et al.*, 2008).
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51 **5.2. Metabolomics for quality control of phytomedicines**

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53 At present, the most popular application of metabolomics in NP research is for the
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55 quality control of phytomedicines. Different metabolite profiles may occur due to
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57 variations between species or variety, adulteration, environmental changes during
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4 growth and harvesting, post harvesting treatment, extraction and method of preparation.
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7 These factors may significantly alter the efficacy of phytomedicines prepared with
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9 these herbs (Wang *et al.*, 2005). The application of ^1H NMR combined with
10
11 multivariate data analysis to detect variations between different herbal preparations has
12
13 been reported. The most common methods that have been applied for these ends are
14
15 unsupervised principal component analysis (PCA) and supervised partial least square
16
17 analysis/ partial least square analysis with discriminant analysis (PLS/PLS-DA). These
18
19 methods were applied to differentiate various chamomile (*Matricaria recutita* L.)
20
21 preparations made with plants grown in different locations, percentage of the desired
22
23 plant part, and extraction method (Wang *et al.*, 2004). Discrimination between different
24
25 *Cannabis sativa* cultivars based on the content of Δ^9 -tetrahydrocannabinolic acid
26
27 (THCA) and cannabidiolic acid (CBDA) was achieved (Choi *et al.*, 2004), as well as
28
29 discrimination between 11 *Ilex* species including 3 varieties (Choi *et al.*, 2005).
30
31 Similarly, discrimination between *Ephedra* species (Kim *et al.*, 2005), and commercial
32
33 feverfew (*Tanacetum parthenium*) preparations from different suppliers (Bailey *et al.*,
34
35 2002) was obtained. The discrimination of commercial preparations of St. John's Wort
36
37 (*Hypericum perforatum*) from different batches of the same supplier, including variation
38
39 of flavonoid content which have been previously related to the antidepressant activity
40
41 was also reported (Rasmussen *et al.*, 2006). An example of application of GC and GC-
42
43 MS combined with multivariate data analysis to correlate artemisinin concentration with
44
45 the growth stage of *Artemisia annua* L. and to distinguish between transgenic and wild
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47 type has been reported (Ma *et al.*, 2008).
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56 Metabolomics can also be used to detect adulterations of herbal preparations with
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58 similar species but with low levels of the desired active compound(s). ^1H NMR and
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PCA was used to verify that the anti-malaria capsules produced by one specific company did not contain *Artemisia annua* as labeled but *Artemisia afra*, which has never been reported to contain the active compound artemisinin or any of its derivatives (van der Kooy *et al.*, 2008). Instead of ^1H NMR, the use of HPLC fingerprints followed by application of PCA and PLS-DA was applied to examine the authenticity of the traditional Chinese medicines *Pericarpium Citri Reticulatae* and *Pericarpium Citri Reticulatae Viride* which contain *Citrus reticulata* ‘Chachi’, *Citrus reticulata* ‘Dahongpao’ and *Citrus erythrosa Tanaka* as well that of commercial preparations containing a mixture of many tangerine peels which had been made with new tangerine mutations of uncertain quality (Yi *et al.*, 2007).

5.3. Metabolomics to link chemical profile and bioactivity pattern of phytomedicines

As previously described, a reductionist approach focusing on a single active compound is necessary in some cases, but in case of St. John’s Wort extract, where no single compound or even group of compounds have been found to be responsible for its activity, the possibility of synergism and pro-drugs is evident, and a holistic *in vivo* approach would be more valuable to guarantee its therapeutic efficacy (Ulrich-Merzenich *et al.*, 2007; Verpoorte *et al.*, 2005). Animal experiments and clinical trials are two classical ways to conduct *in vivo* experiments, but the time required and costs involved make this option practically non viable. Furthermore, if these tests are performed with extracts of uncertain composition, results will invariably be unreliable or inconsistent. Recently new approaches are evolving which might help to get a better view of the mode of action of herbal or natural products and lead to the possibility of

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4 obtaining proof and guarantee of their pharmacological activity over different batches.
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6 These approaches are based on considering the NP holistically, i.e. on the bulk of
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8 chemical constituents rather than focusing on certain individual components or groups
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10 of compounds. On the other hand, the changes caused by a sample on the transcriptome,
11
12 proteome and/or metabolome patterns can be compared with those observed after
13
14 treatment with known drugs rather than looking only at certain specific changes.
15
16 Genomics, proteomics, and metabolomics platforms will be crucial since they allow the
17
18 identification of detectable protein/genes or compounds which might relate to the
19
20 biological activity of complex extract (Ulrich-Merzenich *et al.*, 2007). This may shift
21
22 the paradigm in the development and application of complex plant/phytocompound
23
24 mixtures in modern medicine (Shyur and Yang, 2008). Moreover, by measuring the
25
26 effect of a complex extract administered to a living organism, either *in vitro* or *in vivo*, it
27
28 is possible to identify the compound or combination of compounds correlated with
29
30 activity. Besides, new or unsuspected mode(s) of action and targets might also be
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32 revealed in this approach (Verpoorte *et al.*, 2005).
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40 Several papers discussed a metabonomics approach to study various aspects of
41
42 natural products extracts such as bioavailability, toxicity, and proof of efficacy.
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44 Metabonomics is defined as ‘the quantitative measurement of the dynamic
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46 multiparametric metabolic response of living systems to pathophysiological stimuli or
47
48 genetic modification’ (Nicholson *et al.*, 2002). This term is in general equivalent to the
49
50 NMR spectroscopic analysis of biofluids to identify changes in the concentrations and
51
52 patterns of endogenous metabolites induced by exogenic compounds (Nobeli and
53
54 Thornton, 2006) but mostly used in medicinal or toxicological studies although
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56 sometimes the term metabolomics is also used.
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4 A summary on the implementation of metabolomics, particularly NMR based
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metabolomics coupled to multi-variate data analysis tools to detect links between the chemical profile of a herbal preparation and its bioactivity is provided in Figure 1. A quick review of the literature reflects the enormous amount of studies that are carried out nowadays using this technology, some representative examples of which are summarized below.

5.3.1. Identification of active compounds and quantitative prediction of bioactivity

Chemometric and ^1H NMR analyses were used to discriminate *Artemisia annua* L samples obtained from different sources. PCA assignment showed that artemisinin is the main discriminant factor for clustering. PLS analysis was applied to predict the quantitative anti-plasmodial activity (IC_{50}) and toxicity (ToxEd_{50}) value of different *A. annua* extracts. The predictive values were in close agreement with the actual experimental values (Bailey *et al.*, 2004). The same approach was used to demonstrate the correlation between the spectral pattern and opioid receptor binding properties of St. John's Wort (*Hypericum perforatum*) extracts prepared with different solvents (Roos *et al.*, 2004). GC-MS-based metabolic profiling coupled with anti-inflammatory activity data of ginger (*Zingiber officinale*) from different sources and of some other *Zingiber* species showed that two other closely related *Zingiber* species had similar anti-inflammatory activity to *Z. officinale*, but there was lack of correlation between the content of gingerol, a compound specific to *Z. officinale*, and the observed total anti-inflammatory activity. Therefore, it was concluded that other compounds, unidentified to date which contributed to this activity had to be detected and quantified in order to guarantee the bioactivity of a particular *Zingiber* sample (Jiang *et al.*, 2006). The application of ^1H NMR and PCA analysis allowed the differentiation of six samples of

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4 *Galphimia glauca*, a Mexican herbal used to treat central nervous disorders, collected
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6 from six different areas. Galphimines and 1,3,4,5-tetra-*O*-galloylquinic acid contents
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8 were found to be the main discriminating factors for PCA separation. Further analysis
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10 with PLS-DA confirmed that the level of both compounds was correlated with sedative
11
12 and anxiolytic effects in mice (Figure 2) (Cardoso-Taketa *et al.*, 2008). This technique
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14 was also applied successfully to distinguish thrips-resistant and –susceptible F2 hybrids
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16 of *Senecio jacobaea* and *Senecio aquaticus*. Pyrrolizidine alkaloids, kaempferol and
17
18 jacaranone were found to be higher in resistant plants. This is of interest since the last
19
20 two compounds have positive effects on human health as anti-cancer agents (Leiss *et*
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22 *al.*, 2009).

23 24 25 26 27 28 **5.3.2. Proof of efficacy and mode of action identification of herbal medicines**

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30 The *in vitro* cellular effects of *Cannabis* from different cultivars and preparations
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32 were analyzed using a proteomic and transcriptomic platform (Wang *et al.*, 2005). In
33
34 another case, the *in vivo* effect of *Ginkgo* on rats was studied by applying a ¹H-NMR
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36 and supervised MVDA method to measure metabolites in their urine (Wang *et al.*,
37
38 2005). The effect of three traditional Chinese medicines on insulin resistance APOE*3-
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40 Leiden transgenic mice which are highly responsive in their plasmatic lipids to dietary
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42 treatments was also studied. LC-MS measurement combined with MVDA was applied
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44 to three TCM preparations to discover which was most effective TCM but had the
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46 lowest toxicity (Wang *et al.*, 2005).

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51 The use of metabolomic profiling to study the mode of action of active natural
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53 products recently emerged as well, predominantly in antimicrobial activity analysis. As
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55 an example, HPLC/DAD/ESI-MS application followed by PCA confirmed that
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57 dihydrocurbitacin F-25-*O*-acetate was the main component having an antimicrobial
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4 activity in *Hemsleya pengxianensis*. The possible antibacterial mode of action on
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7 *Staphylococcus aureus* CCTCC AB9105 was predicted by comparing the metabolite
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10 profile of *S. aureus* treated by the plant extract and commercial antibacterials with
11
12 known mode of action (Biao-Yi *et al.*, 2008).

13
14 The use of metabonomics to prove the efficacy of herbal medicines was reported in
15
16 some papers. The evaluation of the anti-ageing effects of the total flavone content of
17
18 Herba Epimedii (TFE), a famous Chinese herbal medicine used to treat some age-
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20 related diseases was conducted by ¹H NMR analysis and PCA assignment. Ten
21
22 metabolites were identified as ageing markers in the rat urine profile: creatinine, lactate,
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24 alanine, acetate, acetone, succinate, allantoin, methylamine, dimethylamine and
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26 trimethylamine-N-oxide. It showed that after TFE intervention, the 24-month-old rats
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28 urine profile was shifted towards that of 18-months-old rats (Wu *et al.*, 2008). In
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30 another case, the study of *Epimedium brevicornum* Maxim, was undertaken. This herbal
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32 medicine, one of the most popular in China is used for toning the kidney and
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34 strengthening bones, but the kind of constituents responsible for its pharmacological
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36 effect, and the mechanism through which it affects the entire body metabolism was
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38 unknown. The plasma metabolites of rats from three different treatments (pre-
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40 hydrocortisone intervention, post-hydrocortisone intervention, and *Epimedium*
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42 *brevicornum* Maxim. treated) were analyzed using UPLC-MS followed by PCA. The
43
44 distinct increase of the level of three metabolites was found after hydrocortisone
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46 intervention, one of them identified as ethylindole-3-crylic acetate. The result showed
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48 that after *Epimedium brevicornum* Maxim treatment, the plasma metabolite profile was
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50 restored closer to that of the pre-hydrocortisone intervention. Of the four compounds of
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52 this herbal preparation detected in plasma (epimedin C, icariin, icariside II, and 2''-O-
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4 rhamnosoyl icaride), epimedin C and icariin were also found in urine, and it was
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6 concluded that those might be the active ones (Famei Li *et al.*, 2007).
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10 The efficacy and the mechanism of action of the traditional Chinese medicine
11 preparation known as Xindi soft capsules (consisting of sea buckthorn flavonoids and
12 sea buckthorn berry oil) used to treat blood stasis, was studied by UPLC combined with
13
14 quadrupole time-of-flight tandem mass spectrometry (UPLC-QTOF-MS) followed by
15
16 PCA and PLSDA analysis. Blood stasis includes hematological disorders such as
17
18 haemorrhage, nasal congestion, thrombosis, and local ischemia (microclots).
19
20 The comparison of the urinary metabolite pattern obtained for 5 groups of rats (healthy
21 control group, acute blood stasis model group, low dose group of Xindi soft capsule,
22 middle dose group of Xindi soft capsule, high dose group of Xindi soft capsule),
23 showed that the treated groups were located between the acute blood stasis model group
24 and the healthy control group. Confirmation by hemorheological analysis proved that
25 within the treated group, especially in the case of the highest dose group, the urine
26 metabolite pattern tends toward that of the control group. Some potential biomarkers
27 were identified as well, such as cholic acid, phenylalanine and kynurenic acid (Zhao *et*
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5.3.3. Bioavailability and fate of natural compounds assessment

The metabonomics approach was successfully applied to study the bioavailability and fate of natural compounds. A traditional Chinese medicine formulation, Dangguibuxue decoction (ODD) consisting of huangqi (*Radix astragali*)-danggui (*Radix angelicae sinensis*), is widely used as a tonic. ODD is a very complex mixture containing hundreds of components. More than 70 compounds have currently been isolated and identified but only a few constituents were reported to be responsible for its

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4 activity. Metabolic fingerprinting by LC/DAD-MS was suitable for analysis of the
5
6 bioavailability of the metabolites of the ODD components *in vivo* by comparing
7
8 LC/DAD-MS fingerprint of the ODD formula with the metabolite fingerprint of plasma
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10 of rabbits treated with ODD. Identification of detected compounds was done by MS and
11
12 UV spectra comparison with references from literature. Some flavonoids, phtalides, and
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14 triterpene saponins, which were known as ODD active compounds were detected,
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16 showing that they are well absorbed into the body (Wang *et al.*, 2007).
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20 21 **5.3.4. Safety and toxicity of herbal medicines assessment** 22

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24 There are also many reports on the application of the metabonomics approach to
25
26 solve the two other major issues related to the quality control of herbal medicines -
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28 safety and toxicity. The mechanism of the toxicity of aristolochic acid was studied by
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30 analyzing the urine of rats submitted to intra-peritoneal injection of aristolochic acid and
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32 several toxins with known modes of action. The PCA, confirmed by biochemical
33
34 parameter analysis, allowed researchers to conclude that aristolochic acid induced the
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36 damage of renal proximal tubules and medulla, accompanied with a slight hepatic lesion
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38 (Zhang *et al.*, 2006). Similarly, the toxicity of Hei-Shun-Pian (the processed lateral root
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40 of *Aconitum carmichaelii* Debx., Ranunculaceae) was evaluated using the ¹H NMR
41
42 spectral metabolic profile of urine and plasma samples of rats after 5 days oral
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44 treatment with the decoction of Hei-Shun-Pian at different doses. Hei-Shun-Pian is a
45
46 traditional Chinese medicine with analgesic, anti-pyretic, anti-rheumatoid arthritis and
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48 anti-inflammatory effects. The C-19 diterpenoid alkaloid aconitine and a series of its
49
50 derivatives are suggested to be the active principles, but some of those are also very
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52 toxic and have narrow therapeutic windows (LD50/ED50). Studying the PCA and PLS
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54 analysis of ¹H NMR data, it was found that the primary differences between the plasma
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4 of treated and the control groups were the increase in the concentration of lipids and the
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6 decrease in the concentration of phosphatidylcholine (PtCho) and *O*- and *N*-acetyl
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8 glycoproteins, while taurine, acetate, creatinine, 2-oxoglutarate, dimethylamine and
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10 hippurate levels in urine contributed to the distinction between high and medium dose
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12 groups and control group (Li *et al.*, 2008a).
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18 **Conclusions and perspectives**

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23 In response to the renewed interest in NP, several strategies have been designed to
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25 improve NP lead finding. Strategies aimed at improving the compatibility of natural
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27 products with high throughput screening requirements by improving the NP libraries
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29 shorten the discovery route, using advanced analytical methods and combining them
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31 with online bioassays, and selecting new approaches which can rapidly provide
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33 evidence of efficacy.
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37 The reductionist approach is still widely used in NP drug discovery studies.
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39 However, metabolomics coupled with MVDA can be applied in those cases to speed up
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41 the limiting steps in dereplication and identification of active compounds. This must be
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43 supported by the availability of a high quality open access NP library based on NMR or
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45 MS data. Metabolomics is undoubtedly an important tool for the quality control of
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47 herbal preparations. It captures the differences in the composition due to different
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49 cultivars, growth areas, methods of preparation, and adulterations, all of which might
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51 significantly affect the content of active principles in the preparation.
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56 To explain the chemodiversity offered by Nature in a more intelligent and
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58 innovative way than random studies using HTS can be employed. For this systems
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4 biology based approach, metabolomics is viewed as a crucial tool. Systems biology may
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6 allow the connection between the chemical profile of herbal preparations and the
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8 response given by a living system, either *in vitro* or *in vivo*. Such a holistic approach to
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10 obtain proof of efficacy has emerged as the new paradigm to detect novel NP in drug
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12 discovery. Significant and obvious advantages are that synergistic interaction can be
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14 detected, as well as prodrugs. At the same time metabolomics will provide information
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16 on the mode of action and include toxic side effects. The capability to quickly analyze a
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18 broad range of metabolites makes metabolomics important in the growing field of
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20 personalized medicine. It allows the metabolic profile of individual patients to be linked
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22 to their response to specific drug therapy as a base to develop a concept on markers of
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24 the efficacy, toxicity, and clearance of drugs, permitting a better prescription for an
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26 individual patient based on their genetic makeup and phenotype status.
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33 It is evident that metabolomics has shown to be a potent facilitator in the recovery of
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35 NP as excellent sources for novel leads, and even further, as a means to discover novel
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37 targets, as the strength of the “omics” based approaches lies greatly in the fact that
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39 results can be stored for long term data mining.
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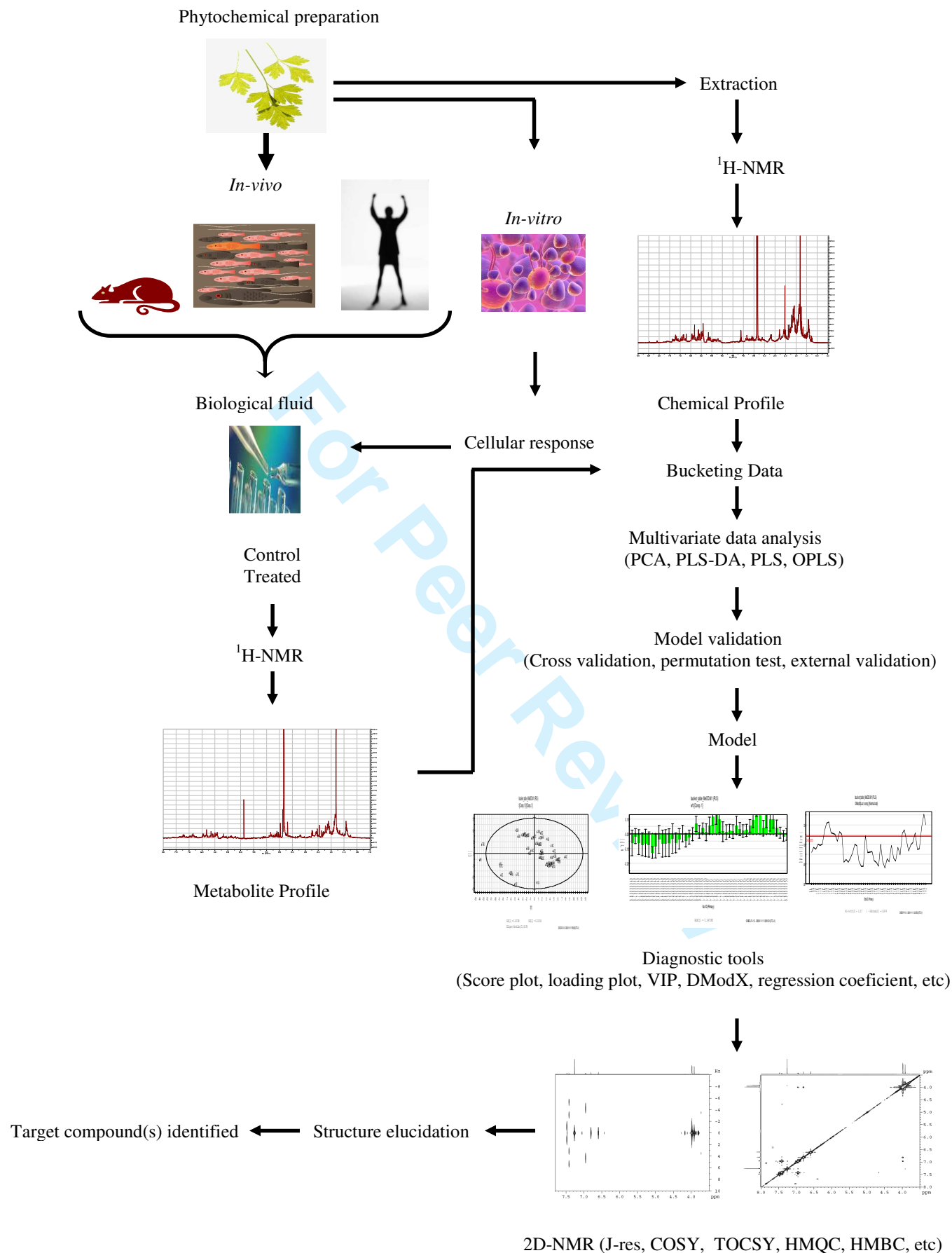
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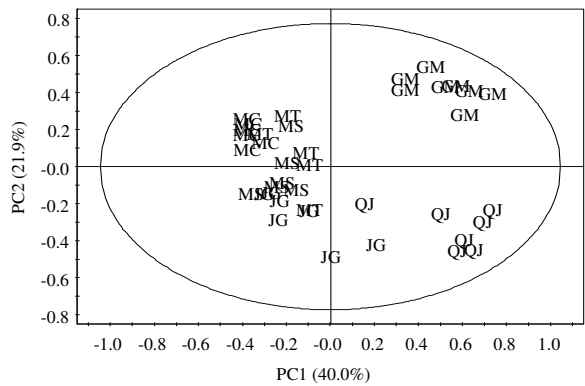
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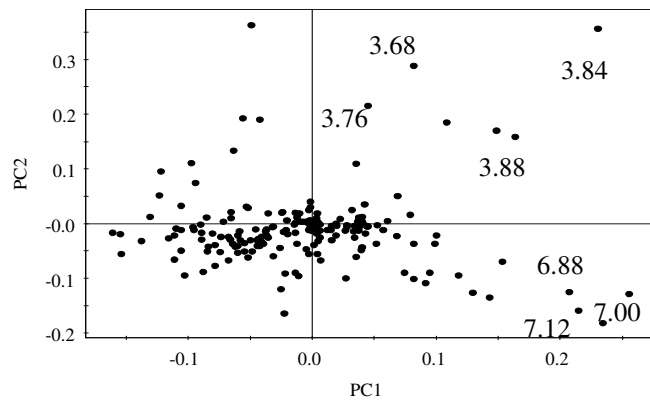
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4 Figure 1. Schematic illustration of metabolomics and multi-variate data application in
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6 bioactivity screening of phytochemical preparation. Multi-variate data analysis permits
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8 the correlation of metabolite profile obtained from bioactivity test with chemical profile
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10 of phytochemical preparation, thus the efficacy involving either single or multi-
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12 synergistically active compounds can be viewed.
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19 Fig. 2. A. Score plot of PCA of *Galphimia glauca* extract from six different locations.
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21 GM and QJ samples were clearly separated from others. B. Signals of the galphimine C-
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23 27 methyl ester proton at δ 3.68 – 3.88 ppm as appeared on the loading plot were the
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25 main factors for the separation C. Partial least square discriminant analysis (PLS-DA)
26
27 allows further separation base on the anxiolytic activity. D. Galphimines and quercetin
28
29 analogs were clearly shown as responsible compounds for the separation (adapted from
30
31 Cardoso-Taketa *et al.*, 2008, with permission from Thieme Medical Publisher).
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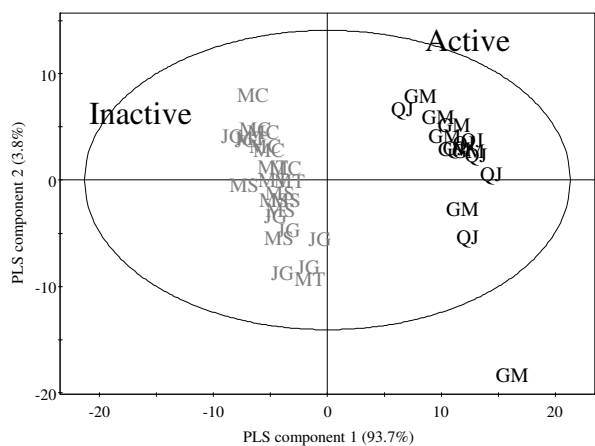




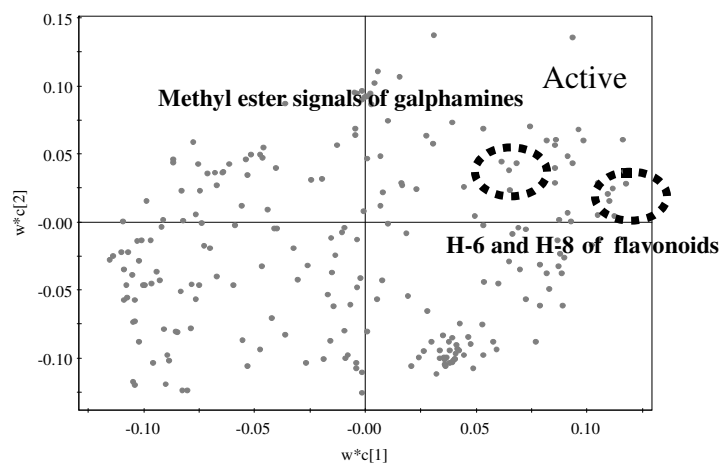
A



B



C



D