



HAL
open science

MILK THISTLE IN LIVER DISEASES: PAST, PRESENT, FUTURE

Ludovico Abenavoli, Raffaele Capasso, Natasa Milic, Francesco Capasso

► **To cite this version:**

Ludovico Abenavoli, Raffaele Capasso, Natasa Milic, Francesco Capasso. MILK THISTLE IN LIVER DISEASES: PAST, PRESENT, FUTURE. *Phytotherapy Research*, 2010, 24 (10), pp.1423. 10.1002/ptr.3207 . hal-00599834

HAL Id: hal-00599834

<https://hal.science/hal-00599834>

Submitted on 11 Jun 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

**MILK THISTLE IN LIVER DISEASES: PAST, PRESENT,
FUTURE**

Journal:	<i>Phytotherapy Research</i>
Manuscript ID:	PTR-10-0101.R2
Wiley - Manuscript type:	Review
Date Submitted by the Author:	01-Apr-2010
Complete List of Authors:	Abenavoli, Ludovico; University Magna Graecia, Department of Experimental and Clinical Medicine Capasso, Raffaele; University Federico II Milic, Natasa; University of Novi Sad Capasso, Francesco; University Federico II
Keyword:	Milk thistle , silybin, silipide, silymarin, liver diseases, reactive oxygen species



Review

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

MILK THISTLE IN LIVER DISEASES: PAST, PRESENT, FUTURE

Ludovico Abenavoli^{a,*}, Raffaele Capasso^b, Natasa Milic^c, Francesco Capasso^b

^aDepartment of Experimental and Clinical Medicine, University "Magna Græcia",
Catanzaro, Italy

^bDepartment of Experimental Pharmacology, University Federico II, Naples, Italy

^cDepartment of Pharmacy, University of Novi Sad, Novi Sad, Serbia

Author for correspondence:

*Ludovico Abenavoli M.D.

Department of Experimental and Clinical Medicine,

University "Magna Græcia", Viale Europa, 88100, Catanzaro, Italy

Tel.: +39 0961 3697113; Fax: +39 0961 754220

l.abenavoli@unicz.it

Abstract

Silybum marianum or Milk Thistle (MT) is the most well-researched plant in the treatment of liver disease. The active complex of MT is a lipophilic extract from the seeds of the plant and is composed of three isomer flavonolignans (silybin, silydianin, and silychrstine) collectively known as silymarin. Silybin is a component with the greatest degree of biological activity and makes up 50 to 70% of silymarin. Silymarin is found in the entire plant but it is concentrated in the fruit and seeds. Silymarin acts as an antioxidant by reducing free radical production and lipid peroxidation, has anti-fibrotic activity and may act as a toxin blockade agent by inhibiting binding of toxins to the hepatocyte cell membrane receptors. In animals, silymarin reduces liver injury caused by acetaminophen, carbon tetrachloride, radiation, iron overload, phenylhydrazine, alcohol, cold ischemia, and *Amanita phalloides*. Silymarin has been used to treat alcoholic liver disease, acute and chronic viral hepatitis and toxin-induced liver diseases.

Key words: Milk Thistle, silybin, silipide, silymarin, liver diseases, reactive oxygen species.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

The Past: History of Milk Thistle

Silybum marianum (L.) Gaernt. (= *Cardus marianum* L.) is an annual to biannual plant of the family *Asteraceae* growing to 1.5 m (Rambaldi et al., 2005). “Silybum” is the name Dioscorides gave to edible thistles and “marianum” comes from the legend that the white veins running through the plant's leaves were caused by a drop of the Virgin Mary's milk. While looking for a place to nurse the infant Jesus when leaving Egypt, Mary could only find a shelter in a bower formed from the thorny leaves of the Milk Thistle (MT) (Morazzoni and Bombardelli, 1995). According to this story the folk belief that the plant was good for nursing mothers was born. Other names that have been attributed to MT include Marian thistle, Mary thistle, St. Mary's thistle, Our Lady's thistle, Holy thistle, sow thistle, Blessed Virgin thistle, Christ's crown, Venue thistle, heal thistle, variegated thistle and wild artichoke. *S. marianum* is native to southern Europe, southern Russia, Asia Minor and northern Africa and is naturalized in North and South America as well in South Australia. The plant grows in warm, dry soil and blooms in July-August. The crude drug consists of the obliquely obovoid fruits (achenes) from which the silvery pappus is removed. Each fruit is about 5-7mm long, up to 2-3 mm wide and 1.5 mm thick, with a glossy, brownish black to greyish, brown husk. The commercial drug originates principally from the cultivated sources, partly from Germany, but primarily from China, Argentina, Romania, Hungary and from a few Mediterranean countries (Figure 1). The freshly milled fruits have a cocoa-like odour and an oily and bitter taste (Capasso et al., 2003).

MT has been used since the time of ancient physicians and herbalists to treat a range of liver and gallbladder disorders, including hepatitis, cirrhosis and jaundice, and to protect the liver against poisoning from chemical and environmental toxins, including snakebites, insect stings, mushroom poisoning, and alcohol (Rambaldi et al., 2005).

1
2
3 One of the earliest records of MT is found in the Bible (Genesis 3:18). In this verse,
4
5 God told Adam and Eve when they were banished from the Garden of Eden that "*thorns*
6
7 *also and thistles shall it bring forth to thee*".
8
9

10 Some of the earliest people to use and write about MT were ancient Greek and
11
12 Roman physicians and herbalists, each of whom seemed to have their own name for the
13
14 herb. Dioscorides called it "sillybon", Pliny the Elder called it "sillybum", and Theophrastus
15
16 called it "pternix". Dioscorides' use of MT is one of the oldest known references of the
17
18 medicinal use of this plant. He suggested preparing it in a tea "for those that be bitten of
19
20 serpents". Another famous ancient herbalist, Pliny the Elder, wrote that mixing the juice of
21
22 the plant with honey was good for "carrying off bile" (Ross, 2008).
23
24
25
26

27 More recently it has been discovered that Indian and Chinese medicines used MT in
28
29 clinical practice (Abenavoli et al., 2008). Available records do not seem to offer us the
30
31 insight how (i.e., by what anecdotal or empirical evidence) MT came to be advised for liver
32
33 and gallbladder problems. Although MT is the most often associated with treating liver
34
35 disorders, physicians have tried to apply its curative properties to other ailments, including
36
37 the stimulation of breast-milk production and bile secretion, treatment of depression, the
38
39 protection against the poisonous mushroom *Amanita phalloides* and other environmental
40
41 toxins (Pradhan and Girish, 2006)
42
43
44
45
46
47

48 **The Present use of Milk Thistle**

49 *Biochemistry*

50
51
52 The crude drug contains 15-30% lipids, in the form of triglycerides [linoleic (about
53
54 60%), oleic (about 30%) and palmitic (about 9%) acid]; about 30% proteins, sugars
55
56 (arabinose, rhamnose, xylose, glucose); tocopherol, (0.038%) sterols (0.063%) with
57
58 cholesterol, campesterol and stigmasterol, and flavonoids including quercetin, taxifolin,
59
60 eriodictyol and chrysoeriol (Table 1). However, the constituents responsible for the activity

1
2
3 are flavanolignans (flavanone derivatives) initially isolated as a mixture of addition
4 products of a coniferyl alcohol, phenylpropanoid alcohol, and a 2,3-dihydroflavonol,
5
6
7 taxifolin (Wu et al., 2009). This mixture, known as silymarin, represents 1.5-3 % of the dry
8 drug weight and consists of (Figure 2): silybin (=silybinin, silibinin) (approximately 50 to
9 60%), isosilibyn (about 5%), silychristin (about 20%) and silydianin (about 10%), as well
10 as silimonin, isosilychristin, isosilibinin, etc. The drug can be identified for its microscopic
11 characteristics by Thin Layer Chromatography (TLC) or High Performance Liquid
12 Chromatography (HPLC); spectrophotometry can also be used.
13
14
15
16
17
18
19
20
21
22
23

24 *Pharmacokinetics*

25
26
27 About 20-50% of silymarin is absorbed following an oral administration in humans
28 and about 80% of the dose is excreted in the bile while about 10% enters the
29 enterohepatic circulation. However, the pharmacokinetic studies have principally been
30 performed on silibinin, the main component of silymarin.
31
32
33
34
35

36 The bioavailability of silibinin is low and seems to depend on several factors such as
37 (i) the content of accompanying substances with a solubilising character such as other
38 flavonoids, phenol derivatives, aminoacids, proteins, tocopherol, fat, cholesterol and other
39 substances found in the preparation and (ii) the concentration of the preparation itself
40 (Voinovich et al., 2009). The systemic bioavailability can be enhanced by adding
41 solubilising substances to the extract (Saller et al., 2001). The bioavailability of silybinin
42 can also be enhanced by the complexation with phosphatidylcholine or β -cyclodextrin and
43 possibly by the choice of the capsule material (Morazzoni et al., 1992; Arcari et al., 1992).
44 The variations in the content, dissolution and (oral) bioavailability of silybinin between
45 different commercially available silymarin products - despite the same declaration of
46 content - are significant (Schulz et al., 1995). Therefore, the comparisons between studies
47 should be carried out with the caution, considering the differences between the analytical
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 methods used (TLC vs HPLC) and whether free, conjugated or total silybinin is the object
4
5 of measurement. Systemic plasma concentrations are usually measured, even though the
6
7 site of action of silymarin is the liver, as they provide an estimate on the quantity of the
8
9 drug being absorbed from the gastrointestinal tract. The adequate bioavailability accounts
10
11 for the dose-related oral activity of silymarin in the liver (Saller et al., 2001).
12
13
14

15 In male volunteers, after single oral administration of a standardised dose of silibinin
16
17 100 to 360mg, plasma silibinin C_{max} was reached after approximately two hours and
18
19 ranges between 200 and 1400 µg/L, of which approximately 75% was present in the
20
21 conjugated form (Weyhenmeyer et al., 1992; Gatti and Perucca, 1994). For total silibinin,
22
23 an elimination half-life of approximately six hours was estimated (Saller et al., 2008).
24
25 Between 3% and 8% of an oral dose was excreted in the urine, while 20 to 40% was
26
27 recovered from the bile as glucuronide and sulfate conjugates. The remaining part was
28
29 excreted via faeces (unchanged, not absorbed). Silibinin concentrations in bile reached
30
31 approximately 100 times those found in serum (10⁻⁵ to 10⁻⁴ mol/L of silibinin in bile) with
32
33 peak concentrations reached within two to nine hours (Saller et al., 2008).
34
35
36
37

38 Barzaghi and co-workers investigated the pharmacokinetics of silybin
39
40 phosphatidylcholine complex (Id B1016) and detected an increase in the oral bioavailability
41
42 of silybin in healthy human subjects, probably by a facilitatory role of the drug complex on
43
44 the passage of the drug across the gastrointestinal tract (Barzaghi et al., 1990). The
45
46 bioavailability of Id B1016 was shown to be several times higher than that of silymarin in
47
48 the patients with hepatic cirrhosis (Orlando et al., 1990).
49
50
51
52
53
54

55 *Pharmacological studies*

56
57 Several pharmacological studies have been carried out on the active components of
58
59 MT, silymarin and silybinin. It has been found that these substances exert hepato-
60
protective, anti-oxidant, anti-inflammatory and anti-fibrotic properties; in addition, they

1
2
3 stimulate protein biosynthesis and liver regeneration, increase lactation and possess
4 immuno-modulation activity. The following is the synthesis of the selected studies on this
5
6
7
8 subject.

9 10 *Anti-inflammatory and immuno-modulation activity*

11
12 Silymarin shows anti-inflammatory and immuno-modulation activity in various
13 structures and pathways of the cell. The tumour necrosis factor- α receptor (TNF α -R)
14
15 super-family, located in the cell membranes, contains several members with homologous
16
17 cytoplasmic domains known as death domains (DD), important in initiating apoptosis and
18
19 other signalling pathways following ligand binding by the receptors (Saller et al., 2008).
20
21
22
23

24
25 A pre-clinical study showed that silibinin prevented the effect of TNF- α induced
26
27 with α -amanitin in hepatocytes (El-Bahay et al., 1999), possibly by reactive oxygen species
28
29 (ROS)-dependent mechanisms (Chovolou et al., 2003). In mice treated with 750 mg/kg/day
30
31 silymarin by gavage and 2.25 mg/kg Fumonisin B1 (inhibitor of ceramide synthase),
32
33 silymarin prevented the Fumonisin B1-induced increases in the TNF receptor 1 (TNF-R1)
34
35 expression, TNF-R1-associated apoptosis (by caspases pathways) and the induction of
36
37 lymphotoxin β and interferon (IFN)- γ (He et al., 2004).
38
39
40

41
42 Some evidence suggested that silymarin inhibited the expression of adhesion
43
44 molecules, such as E-selectin (Kang et al., 2003), another family of transmembrane
45
46 molecules, expressed particularly on the surface of leukocytes, involved in inflammatory
47
48 pathways. In human cells, silymarin caused "*in vitro*" a reduction of lectin-dependent and
49
50 natural killer (NK) cell mediated cytotoxicity but not of antibody dependent cell-mediated
51
52 cytotoxicity (ADCC) (Deák et al., 1990).
53
54

55
56 The inhibition of the 5-lipoxygenase pathway in the cytoplasm, in particular
57
58 leukotriene B4 (LTB4), at the silibinin concentrations achieved "*in vivo*", could well
59
60 represent a pivotal pharmacological property of silymarin. The study which evaluated the
action of silibin in isolated Kupferr cells, indicated a strong inhibitory effect on LTB4

1
2
3 formation with the IC₅₀ of 15 μmol/L silibinin (Dehmlow et al., 1996a). Similar results were
4
5 found in porcine basilar arteries (IC₅₀ = 100 μmol/L), human blood cells and human
6
7 omentum endothelial cells (Dehmlow et al., 1996b) This selective inhibition of leukotriene
8
9 formation by Kupffer and possibly other cells could at least partly account for the
10
11 hepatoprotective properties of silibinin.
12
13

14
15 Silymarin anti-inflammatory nuclear DNA/RNA mediated effects involve nuclear
16
17 factor kappa B (NF-κB), a ubiquitous rapid response transcription factor in inflammatory
18
19 cells. Also, many genes encoding the proteins of the hepatic acute phase response are
20
21 under the control of the transcription factor NF-κB (Saliou et al., 1998). Inactive NF-κB is
22
23 present in the cytoplasm complexed with an inhibitory protein I-κB. NF-κB is activated by a
24
25 number of incoming signals from the cell surface. Released from I-κB inhibition, NF-κB
26
27 translocates into the nucleus and binds to the kappa B motif of the target gene. The NF-κB
28
29 activation process can be inhibited by pharmacologic agents at each activation step; that is
30
31 glucocorticoids, cyclosporine, tacrolimus, and antioxidants (Lee and Burckart, 1998).
32
33
34
35

36
37 *In vitro* silymarin was found to suppress both the kappa B motif of NF-κB DNA
38
39 binding activity and its dependent gene expression in hepatoma cells. In addition, silymarin
40
41 blocked the translocation of NF-κB p65 protein (through phosphorylation) to the nucleus
42
43 without affecting its ability to bind the DNA (Manna et al., 1999). Silymarin also inhibited
44
45 the TNF-α- induced activation of mitogen-activated protein kinase and c-Jun N-terminal
46
47 kinase and abrogated TNF-α-induced cytotoxicity and caspase activation (Saller et al.,
48
49 2008). Kang et al. reported that silymarin had exerted an inhibitory effect on nitric oxide
50
51 (NO) production and inducible nitric oxide synthase (iNOS) gene expression in
52
53 macrophages, although at relatively high concentrations of 12.5–25 μg/ml (Kang et al.,
54
55 2002). The oral administration at the dosage of 100 mg/kg attenuated NO production by
56
57 peritoneal macrophages in lipopolysaccharide (LPS)-treated mice. Silymarin, also
58
59 concentration-dependent, suppressed the LPS-induced production of NO in isolated
60

1
2
3 mouse peritoneal macrophages. Moreover, iNOS mRNA and its protein expression were
4 completely abrogated by silymarin in LPS-stimulated murine RAW 264.7 cells. These
5 results suggest that silymarin inhibits NO production and iNOS gene expression by
6 inhibiting NF- κ B/Rel activation. This conclusion was subsequently confirmed by Schümann
7 and co-workers (Schümann et al., 2003). The authors tested silibinin in the mouse model
8 of concanavalin A (ConA)-induced, T cell-dependent hepatitis. Silibinin significantly
9 inhibited ConA-induced liver disease. Silibinin proved to be an immune-response modifier
10 *in vivo*, inhibiting intrahepatic expression of tumor necrosis factor, interferon-g, interleukin
11 (IL)-4, IL-2, and iNOS, and augmenting synthesis of IL-10. In addition, silibinin inhibited
12 intrahepatic activation of NF- κ B. The major mechanism of this hepatoprotective activity
13 appears to be the inhibition of intrahepatic NF- κ B activation, which prevents the
14 subsequent synthesis of TNF, IFN-g, IL-2, and iNOS. Furthermore, while the synthesis of
15 IL-10 was augmented, the production of IL-4 was inhibited within the liver.
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33

34 35 36 *Enhanced protein synthesis*

37
38
39 Regeneration of liver cells is necessary for hepatic recovery from acute or chronic
40 conditions. In chronic disease, fibrosis occurs simultaneously with cellular regeneration.
41 The ultimate outcome is determined by the type of the process which dominates.
42 Sonnenbichler and Zetl reported that the intra-peritoneal administration of silibinin had
43 caused a marked increase in the synthesis of ribosomal RNA (increased polymerase I) in
44 rat liver. The same authors found that silibinin had stimulated the synthesis of DNA in
45 partially hepatectomised rats, but neither in healthy controls nor in hepatoma or other
46 neoplastic cells (Sonnenbichler and Zetl, 1986; Pradhan and Girish, 2006). The exact
47 mechanism of the action, probably involving polymerase I activation, is unknown. Silybin
48 appeared to stimulate ribonucleic acid (RNA) polymerase-I and ribosomal RNA as it was
49 found in several preclinical studies. This effect leads to more rapid formation of ribosomes,
50
51
52
53
54
55
56
57
58
59
60

1
2
3 which in turn increases protein synthesis. This action has important therapeutic
4
5 implications in repairing the damaged hepatocytes and restoring the normal liver functions.
6
7

8 *Antifibrotic activity*

9
10 Liver fibrosis can result in remodelling of liver architecture leading to hepatic
11
12 insufficiency, portal hypertension and hepatic encephalopathy. These processes involve
13
14 complex interplay of cells and mediators (Gebhardt, 2002). In the initial phase the
15
16 proliferation of hepatic parenchymal cells is developed. The conversion of hepatic stellate
17
18 cells (HSC) into myofibroblast is considered as the central event in fibrogenesis. Silymarin
19
20 inhibits NF-kB and also retards HSC activation. It also inhibits protein kinases and other
21
22 kinases involved in signal transduction and may interact with the intracellular signalling
23
24 pathways. The evidence for anti-fibrotic activity comes largely from animal studies. Favari
25
26 and Perez-Alvez reported that orally administered silymarin (50 mg/kg) to the rats with
27
28 chronic carbon tetrachloride (CCl₄) liver damage could reduce the collagen content in the
29
30 liver of these animals up to 55% (increased with CCl₄ approximately 4-to 6-fold as
31
32 compared to controls) (Favari and Perez-Alvarez, 1997). In similar experiments, there was
33
34 a reduction of collagen and pro-collagen III contents after biliary obstruction in the rat by
35
36 30% with 50 mg/kg/day although not at the level of 25 mg/kg/day of silymarin (Mourelle et
37
38 al., 1989; Boigk et al., 1997). Silymarin suppressed the expression of pro-fibrogenic pro-
39
40 collagen- α 1(I) and TIMP-1 most likely via down-regulation of TGF- β 1 mRNA in the rats
41
42 with biliary fibrosis (Jia et al. 2001). Silymarin, administered during three years, retarded
43
44 the development of alcohol-induced hepatic fibrosis in baboons (Lieber et al., 2003).
45
46
47
48
49
50
51
52
53

54 *Antioxidant effects*

55
56 Free radicals, including the superoxide radical, hydroxyl radical (OH), hydrogen
57
58 peroxide (H₂O₂), and lipid peroxide radicals were implicated in liver diseases (Urtasun et
59
60 al., 2008). The mechanism of free radical damage included ROS-induced peroxidation of

1
2
3 the polyunsaturated fatty acid in the bilayer cell membrane, which caused the chain
4
5 reaction of lipid peroxidation, thus damaging the cellular membrane and causing further
6
7 oxidation of membrane, lipids and proteins (Nagata et al., 2007). The antioxidant
8
9 properties of silibinin were evaluated by studying the ability to react with relevant biological
10
11 ROS or oxidants such as superoxide anion radical (O_2^-), hydrogen peroxide (H_2O_2),
12
13 hydroxyl radical (HO^\cdot) and hypochlorous acid ($HOCl$) (Detaille et al., 2008). Several pre-
14
15 clinical studies reported that MT was not a good scavenger of O_2^- and no reaction with
16
17 H_2O_2 was detected ($IC_{50\%} > 200 \mu\text{mol/L}$). However, it reacted rapidly with HO^\cdot radicals in
18
19 free solution at approximately diffusion-controlled rate ($IC_{50\%} = 1.2\text{--}7 \mu\text{mol/L}$). It was
20
21 reported that in vitro incubation with silymarin in the concentration equivalent to the usual
22
23 therapeutic dosage markedly increased the expression of superoxide dismutase (SOD) in
24
25 lymphocytes in the patients with alcoholic cirrhosis (Feher et al., 1988; Pradhan and
26
27 Girish, 2006). Silymarin had no direct effect on ethanol metabolism and had no role in
28
29 reducing ethanol levels or the rate at which ethanol was removed from the body. In fact,
30
31 there is no evidence of the interaction of silymarin or silibinin with cytochrome P450–2E1,
32
33 thus suggesting that these antitoxic effects are due to its antioxidant and free radical
34
35 scavenging properties (Miguez et al., 1994).
36
37
38
39
40
41
42

43 The phenolic conformation of silymarin is thought to permit the formation of stable
44
45 compounds from hydroxylic and oxygen radicals (Das and Vasudevan, 2006). The *in vivo*
46
47 studies in rats indicate that silymarin can reduce the free radical load. The rats exposed to
48
49 acetaminophen at toxic doses, had increased the levels of reduced glutathione and
50
51 superoxide dismutase when treated with silymarin compared with the levels in controls
52
53 (Singh et al., 2009). Furthermore, the other studies demonstrated that silymarin could
54
55 inhibit cell lysis as measured by changes in alanine aminotransferase levels when
56
57 exposing isolated hepatocytes to carbon tetrachloride and galactosamine (Tsai et al.,
58
59
60 2008).

1
2
3 The *in vitro* experiments with simian kidney cells damaged by paracetamol, cisplatin
4 and vincristin demonstrated that administration of silibinin before or after the drug-induced
5 injury could lessen or avoid the toxic effects. In the recently published study, the rats
6 received cisplatin (3 mg/kg) intraperitoneally and silymarin (50 mg/kg) two hrs before or
7 after cisplatin injection, during five days (Karimi et al., 2005). The pre-treatment with
8 silymarin prevented the nephrotoxic effects of cisplatin while silymarin administered after
9 the chemo-therapeutic agent reduced them to some extent.
10
11
12
13
14
15
16
17
18
19
20
21

22 *Toxin blockade*

23
24 Silymarin is a suitable candidate to treat iatrogenic and toxic liver diseases. It has a
25 regulatory action on cellular and mitochondrial membrane permeability in association to an
26 increase in membrane stability against xenobiotic injury. It can prevent the absorption of
27 toxins into the hepatocytes by occupying the binding sites as well as inhibiting many
28 transport proteins at the membrane. The phalloidin-transporting system, belonging to the
29 hepatocyte-specific organic anion uptake transporters OATP2 (Fehrenbach et al., 2003), is
30 inhibited in a competitive way by silymarin with no influence on membrane fluidity
31 (Sebastian et al., 2004). The OATP2 may represent the part of a hepatic equipment which
32 clears portal blood from bile acids, lipophilic hormones or xenobiotics. The transporting
33 system incorporates phallotoxins into the hepatocytes leading to the death of these cells.
34 Phalloidin (a bicyclic heptapeptide) and antamanide (a monocyclic decapeptide from
35 *Amanita phalloides*) interact with bile-salt-binding polypeptides of the hepatocyte
36 membrane. Similar or possibly identical transport mechanism inhibited by silibinin was
37 described for amanitin (Wellington and Jarvis, 2001). Therefore, silymarin is able to reduce
38 the cellular absorption of noxious xenobiotics other than mushroom poisons, thereby
39 exerting cell-protection.
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

MT and lactation

Traditionally MT has been used by nursing mothers for stimulating milk production and it was recently reported that MT increased the lactation in cows (Tedesco et al., 2004) and women (Di Pierro et al., 2008). However, the MT mode of action has not been established yet. Prolactin is the principal lactogenic hormone and it was recently shown that MT significantly increased circulating prolactin levels in female rats (Capasso et al., 2009). This effect seems to involve, at least partly, dopamine D₂ receptors. In the light of these results MT (or silymarin) could be considered a good candidate for the treatment of lactation insufficiency.

The Present Use of Milk Thistle: Overview of Evidence

The available clinical studies have generally investigated the effects of proprietary products containing silymarin/silybin in patients with hepatic disorders like cirrhosis, acute and chronic hepatitis and liver disease. However, there is also an interest in the use of silymarin in toxin- (mushroom *A. phalloides*) induced hepatitis.

The Agency for Healthcare Research and Quality data, published in 2002, analyzed sixteen prospective placebo-controlled trials. Twelve studies used Legalon®; eight of which used a dosage of 240 to 800 mg/day for the treatment period varying from seven days to six years. No information about doses or duration was reported in four studies. In remaining four of placebo-controlled studies, two used silymarin (no further characterization regarding preparation) and two studies used Silipide®, a silybin phosphatidyl cholin complex (240 mg/day). Seventeen additional trials did not include controls. Silymarin, silybin, silipide and silimarol were used with no standardized dose in nine studies. In the remaining two trials, silymarin was administered to the subjects with no liver disease diagnosed. These trials were probably aimed to determine the prophylactic

1
2
3 use of silymarin in the subjects treated with concomitant anti-tuberculosis drugs and
4
5
6 tacrine, but the results were contradictory, for the absence of hepatic illness.
7

8 A systematic meta-analysis, published in the last years, evaluated nineteen studies,
9
10 eleven double blind and eight single blind (Saller et al., 2008). The authors concluded that
11
12 it was reasonable to employ silymarin in the treatment scheme in the case of *Amanita*
13
14 *phalloides* poisoning. Also, the available evidence supported the therapeutic use of
15
16 silymarin, in view of its excellent safety profile, in alcohol-induced liver diseases, including
17
18 liver cirrhosis (Child-Pugh classification grade A).
19
20
21
22
23

24 *MT in alcohol-induced liver disease*

25
26
27 Randomized, blinded, placebo-controlled studies assessed the effectiveness of MT
28
29 in chronic alcoholic liver disease. The results of these trials might be conflicting and
30
31 confounded because of heterogeneity of the degree of disease severity and alcoholic
32
33 intake or abstinence. In a double-blind controlled study by Salmi and Sarna, one hundred
34
35 and six patients with mild acute and sub-acute liver disease, of which ninety with
36
37 confirmed histological diagnosis, were randomly allocated to either silymarin or placebo
38
39 treatment, for four weeks (Salmi and Sarna, 1982). Alcohol was forbidden during the trial.
40
41 The difference between the two groups was significant in an important decrease in alanine
42
43 aminotransferase (ALT) and aspartate aminotransferase (AST) levels in the silymarin
44
45 group. No difference, instead, in reduction of serum bilirubin in both. Among the subgroup
46
47 with the second histological work-up, the normalization of histological changes occurred
48
49 more often in the silymarin group (eleven out of fifteen) than in controls (four out of
50
51 fourteen).
52
53
54
55
56

57 Feher and co-workers, studied thirty six patients suffering from chronic alcoholic
58
59 liver disease, treating them with an oral dose of 420 mg/day of silymarin or placebo for six
60
months. Serum bilirubin, transaminase values and gamma glutamil transferase (GGT)

1
2
3 activity were normalized in the silymarin group, with an improvement in histology of the
4
5 liver and a decrease of pro-collagen III peptides (a marker of fibrotic progression in liver
6
7 disease), but only a few patients had control histology (Feher et al., 1989).
8
9

10 In a double blind comparative study of 106 patients with histological proven
11
12 alcoholic hepatitis, MT did not significantly influence the liver biopsy findings (Trichet et al.,
13
14 1989).
15
16

17 18 19 *MT in viral hepatitis* 20 21

22 Even though silymarin does not affect viral replication, it might play a beneficial role
23
24 in viral hepatitis by its inhibitory action on inflammatory cascade induced by viral infection.
25
26 Lirussi and Okolicsanyi compared silymarin and ursodeoxicholic acid in a rather
27
28 heterogeneous population of patients with active cirrhosis, the majority were HCV positive
29
30 (Lirussi and Okolicsanyi, 1992). No efficacy was detected in this study. More recently, a
31
32 new silybin-phosphatidylcholine complex was studied in a short pilot study on twenty
33
34 patients with chronic active hepatitis (Buzzelli et al., 1993). Transaminase levels were
35
36 reduced in silymarin group, but without consistent differences in other liver function tests
37
38 (bilirubin, alkaline phosphatase, albumin).
39
40
41
42

43 Rambaldi et al. in the meta-analyses of randomized clinical trials valuated 915
44
45 patients with alcoholic hepatitis and hepatitis B or C (Rambaldi et al., 2005). The primary
46
47 outcome measure was the number of patients dying. The secondary outcome measures
48
49 were the development of clinical symptoms and complications analyzed separately and
50
51 combined with liver biochemistry, liver biopsy findings, as well as the number and type of
52
53 adverse events. The authors found no significant effect of MT on all-cause mortality and
54
55 observed a potential beneficial effect of MT on mortality in patients with alcoholic liver
56
57 disease, but this effect could not be confirmed in two high-quality trials. A potential
58
59
60

1
2
3 beneficial effect of MT on liver-related mortality was observed, but again, this effect could
4 not be demonstrated in three high quality trials.
5
6

7
8 Subsequently, Federico et al. evaluated the antioxidant and antifibrotic activity of
9
10 Realsil® (complex silybin - vitamin E - phospholipids) to improve insulin resistance and
11 liver damage in the patients with non-alcoholic fatty liver disease (NAFLD) and HCV
12 chronic infection (Federico et al., 2006). This study enrolled eighty five patients; fifty nine
13 were affected by primitive NAFLD (group A) and twenty six by HCV related chronic
14 hepatitis C in combination with NAFLD, all HCV genotype-1b, and non-responders to
15 previous antiviral treatment (group B). All the patients with a diagnosed liver disease two
16 years prior to the study, according to histological criteria, were enrolled over six
17 consecutive months and further divided into two subgroups using a systematic random
18 sampling procedure: fifty three [thirty nine NAFLD and fourteen HCV) were treated with
19 four tablets/day of Realsil® (one tablet contained 94 mg of silybin, 194 mg of
20 phosphatidylcholine, and 90 mg of vitamin E) for six months followed by another six
21 months of follow up, while the other thirty two patients (twenty NAFLD and twelve HCV)
22 served as a control group (no treatment). For 0, 6, and 12 months, the outcomes were:
23 body mass index, bright liver by ultrasonography, transaminase and GGT levels, blood
24 glucose and insulin plasma levels with a contemporaneous determination of insulin
25 resistance by Homeostasis Model Assessment (HOMA) test and, as indices of liver
26 fibrosis, plasma levels of transforming growth factor β , hyaluronic acid, and
27 metalloproteinase.
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51

52
53 The results showed the improvement of liver enzyme levels in the treated group, but
54 this only persisted in the group A. Hyperinsulinaemia, present in both groups, was
55 significantly reduced only in the treated patients. The treatment with Realsil® significantly
56 reduced all indices of liver fibrosis in both treated groups, with a persistent effect only in
57 group B.
58
59
60

1
2
3 More recently El-Kamary et al. conducted a randomized controlled trial of silymarin
4 vs placebo with 105 patients with symptoms compatible with acute clinical hepatitis and
5 serum alanine aminotransferase levels > 2.5 times the upper limit of normal (El-Kamary et
6 al., 2009). The patients received 420 mg/day of silymarin (Legalon®) or vitamin placebo for
7 four weeks. The treatment with silymarin significantly reduced the symptoms related to
8 biliary retention (dark urine, jaundice, scleral icterus), while the biomarkers considered
9 (transaminase and direct bilirubin) were not significantly modified.
10
11
12
13
14
15
16
17
18
19
20
21

22 *Liver cirrhosis*

23
24 Clinical trials in liver cirrhosis are complicated by a series of confounding factors,
25 such as concomitant therapies and complications of cirrhosis. The study by Ferenci et al.
26 was performed to determine the effect of silymarin in patients with cirrhosis (Ferenci et al.,
27 1989). It was a randomized controlled study comprising 170 patients with cirrhosis. Eighty
28 seven patients (alcoholic forty, non-alcoholic forty-one) were treated with silymarin 140 mg
29 oral dose, three times daily; eighty three patients (alcoholic forty five, non-alcoholic thirty
30 eight) received a placebo. The mean observation period was forty one months. The
31 survival rate, to four years of the treatment, was 58% in the first group and 39% in the
32 controls. The analysis of subgroups indicated that the treatment was effective in the
33 patients with alcoholic cirrhosis (p=0.01) and in the patients initially rated Child-Pugh A
34 (p=0.03).
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49

50
51 A French study group, conducted a randomized double-blind trial of silymarin
52 versus placebo in 116 patients with histologically proven alcoholic hepatitis, fifty eight of
53 them with cirrhosis (Trichet et al., 1989). Fifty seven patients received oral silymarin 420
54 mg/day and fifty nine received placebo for three months. The biological parameters were
55 assessed in the serum, and the percutaneous liver biopsy was obtained both at the start of
56 the trial and after three months (control histology available in thirty two silymarin and in
57
58
59
60

1
2
3 thirty five placebo patients). The significant improvement in the score of alcoholic hepatitis
4
5 and serum amino transferase activity was noted in both groups during the trial, irrespective
6
7 of treatment with silymarin or placebo but clearly correlated with abstention from alcohol.
8
9 However, this study is relatively short and half the population was not classified as having
10
11 cirrhosis; since the deaths were not allocated to a diagnostic group, the entire group of the
12
13 patients was considered "at risk". Bunout et al. conducted a placebo-controlled trial with
14
15 silymarin in the patients with alcoholic liver cirrhosis. Seventy-one patients were randomly
16
17 assigned to oral silymarin (n=34) at a relatively low dose of 280 mg/day or an equal
18
19 number of placebo tablets (n=37) (Bunout et al., 1992). Both groups did not differ in their
20
21 initial laboratory assessments and were followed up for an average of fifteen months. Ten
22
23 patients died during the follow up (five in placebo and five in silymarin; liver-related death
24
25 in nine). No details concerning distribution by the therapeutic group were provided. No
26
27 significant differences were observed between these two groups. It was concluded that
28
29 silymarin did not change the evolution in this trial. A subsequent Spanish study was
30
31 specifically aimed to determine the effect of silymarin in the patients with alcoholic liver
32
33 cirrhosis, respect to survival, clinical and laboratory changes (Parés et al., 1998). From
34
35 February 1986 to June 1989, this randomized double-blind multicentre trial, comparing
36
37 silymarin 450 mg/day in three divided doses (n=103) with placebo (n=97) enrolled 200
38
39 alcoholics with histologically- (n=191) or laparoscopically-proven (silymarin six, placebo
40
41 three) liver cirrhosis. The primary outcome was the time of death and the secondary
42
43 outcome was the progression of liver failure. The survival rate was similar in both groups
44
45 and was not influenced by gender, persistence of alcoholic intake, severity of disease or
46
47 by the presence of alcoholic hepatitis in the liver biopsies. Silymarin did not produced any
48
49 significant effect on the progression of disease, but the frequency of complications was
50
51 lower in the treated patients (p=0.06). The study by Bunout had the same end-point,
52
53
54
55
56
57
58
59
60

1
2
3 considered by Ferenci and Parés, however a low dose was used, and it did not state
4
5 whether it was properly blinded.
6
7
8
9

10 *Toxic and iatrogenic liver disease*

11
12 In spite of the large number of pre-clinical studies, the clinical trials in toxic liver
13 disease are scarce and of poor quality. Only one study was set up to assess the ability of
14 silymarin to antagonize or to prevent the hepatotoxic effects of tacrine (Allain et al., 1999).
15 Tacrine is an anticholinesterase drug used for the treatment of Alzheimer's disease. In this
16 randomized double-blind placebo controlled study on twenty three patients suffering from
17 mild to moderate dementia of the Alzheimer type, the silymarin oral dose (420 mg/day)
18 was given for one week, and then tacrine was added, first at 40 mg/day for six weeks, then
19 at 80 mg for six additional weeks. The statistical difference was not observed for serum
20 ALT, but the side effects, especially gastrointestinal disorders, were much less frequent in
21 the silymarin group. Therefore, the co-administration in the initial phases of the treatment
22 may improve tolerability.
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37

38 In the study of prophylactic MT during the treatment for tuberculosis, twenty nine
39 subjects with normal liver function tests received anti-tuberculosis drugs plus Hepabene[®],
40 a mixture of silymarin and *Fumaria officinalis* alkaloids. The differences in the AST and
41 ALT reductions were statistically significant. Therefore, the combination of silymarin with
42 such potentially hepatotoxic drugs may prevent such adverse reactions (Comelli et al.,
43 2007).
44
45
46
47
48
49
50
51
52
53

54 *Mushroom poisoning*

55
56 The *Amanita phalloides* mushroom ("death cap") has been known and feared for at
57 least two millennia and continues to cause serious illness and death. The most important,
58 cytotoxin, out of many cytotoxins produced by mushrooms, is the potent amanitin found in
59
60

1
2
3 some mushrooms belonging to the genera *Amanita* and *Galerina*. Amanitin is a cyclic
4 octapeptide which inhibits RNA polymerase II, thus interfering with protein synthesis.
5
6 Phalloidin, a cyclic heptapeptide that accompanies amanitin and may interfere with actin
7 polymerization, is probably responsible for the initial gastrointestinal symptoms. *Amanita*
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

some mushrooms belonging to the genera *Amanita* and *Galerina*. Amanitin is a cyclic octapeptide which inhibits RNA polymerase II, thus interfering with protein synthesis. Phalloidin, a cyclic heptapeptide that accompanies amanitin and may interfere with actin polymerization, is probably responsible for the initial gastrointestinal symptoms. *Amanita phalloides* intoxications are not very frequent. Unfortunately, there are no controlled studies available and so case-control studies collected over several decades, individual cases reports and "expert opinion" were reviewed (Saller et al., 2008).

Diabetic patients with chronic liver disease

Glucose intolerance in the patients with cirrhosis results from two abnormalities that occur simultaneously: insulin resistance in muscle and inadequate response of β -cells to secrete insulin appropriately in order to overcome the defect in insulin action (Petrides et al., 1994). Diabetes mellitus in the insulin-resistant patients with cirrhosis is the result of the progressive impairment in insulin secretion together with the development of hepatic insulin resistance leading to fasting hyperglycaemia and a diabetic glucose tolerance profile (Younossi and McCullough, 2009).

The prevalence of diabetes mellitus was reported to be higher in the patients with HCV-related cirrhosis (23.6%) than in those with HBV-related cirrhosis (9.4%) (Negro and Alaei, 2009). The prevalence of diabetes mellitus is also associated closely with the Child-Pugh score and increasing age. In a study by Velussi et al., the authors showed the effectiveness of a long-term treatment with MT to reduce lipoperoxidation and insulin resistance in the diabetic patients with cirrhosis (Velussi et al., 1997). A group of patients was enrolled to receive an oral dose of 600 mg/day of silymarin plus standard therapies, while the control group standard therapies alone. The trials outcomes were: fasting blood glucose levels, daily glucosuria, glycosylated haemoglobin (HbA1c) and malondialdehyde levels. There was a significant decrease in fasting blood glucose levels and in fasting

1
2
3 insulin levels (-40%) already after four months of the treatment in the silymarin group, also
4
5 seen for the mean daily blood glucose levels (-14.6%) and daily glucosuria (-32%).
6
7
8 However, the HbA1c levels at the end of twelve months were lowered by 8.8%. In addition,
9
10 there was a significant decrease in the mean exogenous insulin requirements in the
11
12 treated group from 55 ± 5 IU/day to 45 ± 3 IU/day after six months and 42 ± 2 IU/day after
13
14 twelve months, while the untreated group showed the significant increase in fasting insulin
15
16 levels and a stabilized insulin requirement. The authors concluded that these results
17
18 showed that silymarin might reduce the lipoperoxidation of cell membranes and insulin
19
20 resistance, significantly decreasing endogenous insulin overproduction and the need for
21
22 exogenous insulin administration, thus reducing the daily insulin dose required by almost
23
24
25
26
27 25% patients.
28
29
30

31 *Preparations of Milk Thistle for humans*

32
33
34 A water-soluble derivative of silybinin (silybinin dihemisccinate disodium) is
35
36 available in Europe for *Amanita phalloides* poisoning. Madaus Legalon[®] is produced in
37
38 tablets containing 70 or 140 mg silymarin and is given in a dose of one to two tablets up to
39
40 three times daily, with a maximum dosage of 420 mg. The formulation includes extraction
41
42 with alcohol, filtration and evaporation and may also include pressing, heat drying, and
43
44 blending with other compounds. Some brands may add choline, inositol, tumeric extract,
45
46 artichoke extract, whole herb powder, dandelion, licorice, curcuma, boldo, iron, or vitamins
47
48
49
50 A and C.
51

52
53 One formulation is combined with kutkin, the roots and rhizome of *Picrorhiza kurroa*,
54
55 a perennial herb found only in the higher mountains of the north-western Himalayas. Other
56
57 concentrated oral formulations include tablets and softgel capsules. Silipide[®] is a complex
58
59 of one part of silybin and two parts of phosphatidycholine from soybean phospholipids
60
(lecithin), for which the standardization is expressed as silybin equivalents. It was more

1
2
3 bio-available than standardized silymarin after oral ingestion in the normal volunteers,
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

bio-available than standardized silymarin after oral ingestion in the normal volunteers, cirrhotics, and patients after cholecystectomy (Barzaghi et al., 1990; Federico et al., 2006). The bioavailability of silybin in Silipide[®] is approximately tenfold greater than the silybin content of standard MT preparations (Morazzoni and Bombardelli, 1995). Other preparations containing MT standardized to silymarin or to silybin are also found in commerce in different forms (granulates, tablets or coated tablets and drops).

Complications and adverse effects

Silymarin is reported to have a very good safety profile. No side-effects were observed in volunteers following a single oral dose of silymarin corresponding to 254 mg silybin (Weyhenmeyer et al., 1992). The patients with liver disorders of various origins who received oral silymarin (600-800 mg/day) for six months did not manifest the adverse effects (Velussi et al., 1993; Palasciano et al., 1994). The clinical trials involving about 3500 patients, of whom 2637 with liver disease, treated with Legalon (560 mg/day) for eight weeks reported that the frequency of adverse effects with silymarin was about 1%. The pooled adverse effects were transient gastrointestinal complaints like bloating, nausea, dyspepsia and diarrhea. (Leng-Pschow, 1996).

It was also shown that MT was generally non toxic and without side effects when administered to the adults in an oral dose range of 240-900 mg/day in two or three divided doses. At higher doses of more than 1500 mg/day silymarin may produce a laxative effect with an increased bile flow and secretion. Among the randomized controlled studies neither optimal nor maximal therapeutic doses were defined. In most of the trials the reportedly affective daily doses of silymarin were 420-600 mg (Saller et al., 2008).

However the case reported described an adverse reaction of the MT herbal medication. The symptoms included severe sweating, abdominal cramping, nausea, vomiting, diarrhea and weakness (Adverse Reaction Advisory Committee, 1999). Another

1
2
3 report described an anaphylactic shock in a 54-year-old man with immediate-type allergy
4
5 to kiwi fruit (Geier et al., 1990).
6
7

8 Preclinical data has not documented the acute toxicity for silymarin and silybin.
9
10 Silymarin, in particular, given orally to mice and dogs at doses of 20 or 1 g/kg did not
11
12 cause adverse effects or mortality. However, a long-term oral administration of silymarin
13
14 (100 mg/kg/day) to rats for 22 weeks did not also cause the adverse effects (El-Bahay et
15
16 al., 1999, Saller, 2008). MT preparations are contra-indicated for the individuals with
17
18 hypersensitivity to *Asteraceae*. None of the studies on cirrhosis, carried out with silymarin,
19
20 considered clinical outcomes other than death as the primary end-point. Based on the
21
22 available data, several issues could be partially examined in the above mentioned studies.
23
24 Different issues were usually addressed in different trials; although these issues are
25
26 interrelated, they have to be regarded individually and cannot be consolidated into one
27
28 global picture.
29
30
31
32

33
34 Bleeding oesophageal varices constitute one of the most serious complications of
35
36 cirrhosis. The total incidence of upper gastrointestinal bleedings (UGBs), both as a co-
37
38 factor of death and as "last data", were reported in the two largest trials: 4.6% with
39
40 silymarin vs. 9.6% with placebo, and 6.3% vs. 13.5% 79, respectively, and showed the
41
42 differences in favour of the active treatment [p=0.042; Odds Ratio (OR) 0.44 (95% CI 0.20,
43
44 0.97)] (Ferenci et al., 1989). These findings suggest that the decreased rate of UGBs
45
46 reflects an overall improvement in the patients, also evidenced by the lower liver-related
47
48 mortality rate, rather than being a direct effect of silymarin.
49
50
51
52
53
54

55 **The Future: New Prospective uses of Milk Thistle**

56

57 The cancer chemopreventive role of MT flavonolignans has been reported in recent
58
59 literature (Mazzio and Soliman, 2009). Silymarin modulates imbalance between cell
60
survival and apoptosis through interference with the expressions of cell cycle regulators

1
2
3 and proteins involved in apoptosis. In addition, silymarin also showed anti-inflammatory,
4 anti-angiogenic and anti-metastatic effects (Ramasamy and Agarwal, 2008). Particularly,
5 the protective effects of silymarin and its major active constituent, silibinin, studied in a
6 variety of *in vitro* and *in vivo* cancer models, including liver cancer, suggest that they
7 should be established in therapies as adjuncts in the clinical application in these patients
8 to prevent or reduce chemotherapy as well as radiotherapy-induced toxicity. The molecular
9 mechanisms of silibinin-mediated antiproliferative effects are mainly via receptor tyrosine
10 kinases, androgen receptor, NF- κ B, cell cycle regulatory and apoptotic signalling pathways
11 in various cancer cells (Li et al. 2010). Additional studies are necessary to evaluate
12 chemotherapeutic effects of silymarin.
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33

34 **References:**

35
36
37
38 Abenavoli L., Bardazzi G., Cracolici F *et al.* 2008. Complementary therapies for treating
39 alcoholism First Annual meeting by Complementary Medicine Research Group of the
40 Italian Society for Alcohol Studies-May 5, 2006, Florence, Italy. *Fitoterapia*. **79**: 142-147.
41
42
43
44
45
46
47

48 Adverse reaction Advisory Committee. 1999. An adverse reaction to the herbal medication
49 Milk thistle (*Silybum marianum*). *Med J Aust.* 170, 218-219.
50
51
52
53

54
55 Allain H, Schüick S, Lebreton S *et al.* 1999. Aminotransferase levels and silymarin in de
56 novo tacrine-treated patients with Alzheimer's disease. *Dement Geriatr Cogn Disord.* **10**:
57 181-185.
58
59
60

1
2
3 Arcari M, Brambilla A, Brandt A et al. 1992. A new inclusion complex of silibinin and beta-
4 cyclodextrins: in vitro dissolution kinetics and in vivo absorption in comparison with
5 traditional formulations. *Boll Chim Farm.* **131**: 205-209.
6
7

8
9
10
11
12 Barzaghi N, Crema F, Gatti G et al. 1990. Pharmacokinetic studies on IdB 1016, silybin
13 phosphatidylcholine complex, in healthy human subjects. *Eur J Drug Metab*
14 *Pharmacokinet.* 15: 333-338.
15
16
17

18
19
20
21
22 Boigk G, Stroedter L, Herbst H et al. 1997. Silymarin retards collagen accumulation in
23 early and advanced biliary fibrosis secondary to complete bile duct obliteration in rats.
24 *Hepatology.* **26**: 643-649.
25
26
27

28
29 Bunout D, Hirsch S, Petermann M et al. 1992. Controlled study of the effect of silymarin on
30 alcoholic liver disease. *Rev Med Chil.* **120**: 1370-1375.
31
32
33

34
35
36 Buzzelli G, Moscarella S, Giusti A et al. 1993. A pilot study on the liver protective effect of
37 silybin-phosphatidylcholine complex (IdB1016) in chronic active hepatitis. *Int J Clin*
38 *Pharmacol Ther Toxicol.* **31**: 56-60.
39
40
41

42
43
44
45
46 Capasso F, Gagarella TS, Grandolini G et al. 2003. *Phytotherapy: A Quick Reference to*
47 *Herbal Medicine.* Springer-Verlag, Berlin.
48
49

50
51
52
53 Capasso R, Aviello G, Capasso F et al. 2009. Silymarin BIO-C[®], an extract from *Silybum*
54 *marianum* fruits, induces hyperprolactinemia in intact female rats. *Phytomedicine.* **16**: 839-
55 844.
56
57
58
59
60

1
2
3 Chovolou Y, Watjen W, Kampkotter A *et al.* 2003. Resistance to Tumor Necrosis Factor- α
4 (TNF- α)-induced apoptosis in rat hepatoma cells expressing TNF- α is linked to low
5 antioxidant enzyme expression. *J Biol Chem.* **278**: 29626-29632.
6
7
8
9

10
11
12 Comelli MC, Mengs U, Schneider C *et al.* 2007. Toward the definition of the mechanism of
13 action of silymarin: activities related to cellular protection from toxic damage induced by
14 chemotherapy. *Integr Cancer Ther.* **6**: 120-129.
15
16
17
18

19
20
21 Deák G, Muzes G, Lang I *et al.* 1990. Effects of two bioflavonoids on certain cellular
22 immune reactions in vitro. *Acta Physiol Hung.* **76**: 113-121.
23
24
25
26

27
28
29 Das SK Vasudevan DM. 2006. Protective effects of silymarin, a milk thistle (*Silybium*
30 *marianum*) derivative on ethanol-induced oxidative stress in liver. *Indian J Biochem*
31 *Biophys.* **43**: 306-311.
32
33
34
35

36
37
38 Dehmlow C, Erhard J, de Groot H. 1996a. Inhibition of Kupffer cell functions as an
39 explanation for the hepatoprotective properties of silibinin. *Hepatology.* **23**:749-754.
40
41
42
43

44
45
46 Dehmlow C, Murawski N, de Groot H. 1996b. Scavenging of reactive oxygen species and
47 inhibition of arachidonic acid metabolism by silibinin in human cells. *Life Sci.* **58** : 1591-
48 600.
49
50
51
52

53
54
55 Detaille D, Sanchez C, Sanz N *et al.* 2008. Interrelation between the inhibition of glycolytic
56 flux by silibinin and the lowering of mitochondrial ROS production in perfused rat
57 hepatocytes. *Life Sci.* **82**: 1070-1076.
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Di Pierro F, Callegari A, Carotenuto D *et al.* 2008. Clinical efficacy, safety and tolerability of BIO-C (micronized Silymarin) as a galactagogue. *Acta Biomed.* **79**: 205-210.

El-Bahay C, Gerber E, Horbach M *et al.* 1999. Influence of tumor necrosis factor-alpha and silibin on the cytotoxic action of alpha-amanitin in rat hepatocyte culture. *Toxicol Appl Pharmacol.* **158**: 253-260.

El-Kamary SS, Shardell MD, Abdel-Hamid M *et al.* 2009. A randomized controlled trial to assess the safety and efficacy of silymarin on symptoms, signs and biomarkers of acute hepatitis. *Phytomedicine.* **16**: 391-400.

Favari L, Perez-Alvarez V. 1997. Comparative effects of colchicine and silymarin on CCl₄-chronic liver damage in rats. *Arch Med Res.* **28**: 11-17.

Federico A, Trappoliere M, Tuccillo C *et al.* 2006. A new silybin-vitamin E-phospholipid complex improves insulin resistance and liver damage in patients with non-alcoholic fatty liver disease: preliminary observations. *Gut.* **55**: 901-902.

Fehér J, Lang I, Nekam K *et al.* 1988. Effect of free radical scavengers on superoxide dismutase (SOD) enzyme in patients with alcoholic cirrhosis. *Acta Med Hung.* **45**: 265-276.

Fehér J, Deák G, Müzes G *et al.* 1989. Liver-protective action of silymarin therapy in chronic alcoholic liver diseases. *Orv Hetil.* **130**: 2723-2727.

1
2
3 Fehrenbach T, Cui Y, Faulstich H et al. 2003. Characterization of the transport of the
4 bicyclic peptide phalloidin by human hepatic transport proteins. *Naunyn Schmiedeberg's*
5
6
7
8 *Arch Pharmacol.* **368**: 415-420.

9
10
11
12 Ferenci P, Dragosics B, Dittrich H et al. 1989. Randomized controlled trial of silymarin
13 treatment in patients with cirrhosis of the liver. *J Hepatol.* **9**: 105-113.

14
15
16
17
18
19
20 Gatti G, Perucca E. 1994. Plasma concentrations of free and conjugated silybin after oral
21 intake of a silybin-phosphatidyl complex (silipide) in healthy volunteers. *Int J Clin*
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Pharmacol Ther. **32**: 614-617.

30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Gebhardt R. 2002. Oxidative stress, plant-derived antioxidants and liver fibrosis. *Planta*
Med. **6**: 289-296.

36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
Geier J, Fuchs T, Wahl R. 1990. Anaphylactic shock due to an extract of *Silybum*
marianum in a patient with immediate-type allergy to kiwi fruit. *Allergologie.* **13**: 387-388.

43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
He Q, Kim J, Sharma RP. 2004. Silymarin protects against liver damage in BALB/c
mice exposed to Fumonisin B1 despite increasing accumulation of free sphingoid bases.
Toxicol Sci. **80**: 335-342.

53
54
55
56
57
58
59
60
Jia JD, Bauer M, Cho JJ et al. 2001. Antifibrotic effect of silymarin in rat secondary biliary
fibrosis is mediated by downregulation of procollagen alpha 1(I) and TIMP-1. *J Hepatol.*
35: 392-398.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Kang JS, Jeon YJ, Kim HM *et al.* 2002. Inhibition of inducible nitric-oxide synthase expression by silymarin in lipopolysaccharide-stimulated macrophages. *J Pharmacol Exp Ther.* **302**: 138-144.

Kang JS, Park SK, Yang KH *et al.* 2003. Silymarin inhibits TNF-alpha-induced expression of adhesion molecules in human umbilical vein endothelial cells. *FEBS Lett.* **550**: 89-93.

Karimi G, Ramezani M, Tahoonian Z. 2005. Cisplatin nephrotoxicity and protection by milk thistle extract in rats. *Evid Based Complement Alternat Med.* **2**: 383-386.

Lee JI, Burckart GJ. 1998. Nuclear factor kappa B: important transcription factor and therapeutic target. *J Clin Pharmacol.* **38**: 981-993.

Leng-Peschlow E. 1996. Properties and medical use of flavonolignans (Sylimarín) from *Sylibum marianum*. *Phytother Res.* **10**: S25-S26.

Li L, Zeng J, Gao Y *et al.* 2010. Targeting silibinin in the antiproliferative pathway. *Expert Opin Investig Drugs.* **19**: 243-255.

Lieber CS, Leo MA, Cao Q *et al.* 2003. Silymarin retards the progression of alcohol-induced hepatic fibrosis in baboons. *J Clin Gastroenterol.* **37**: 336-339.

Lirussi F, Okolicsanyi L. 1992. Cytoprotection in the nineties: experience with ursodeoxycholic acid and silymarin in chronic liver disease. *Acta Physiol Hung.* **80**: 363-367.

1
2
3 Manna SK, Mukhopadhyay A, Van NT et al. 1999. Silymarin suppresses TNF-induced
4 activation of NF-kappa B, c-Jun N-terminal kinase, and apoptosis. *J Immunol.* **163**: 6800-
5
6 6809.
7
8

9
10
11
12 Mazzi EA, Soliman KF. 2009. In vitro screening for the tumoricidal properties of
13 international medicinal herbs. *Phytother Res.* **23**: 385-398.
14
15
16

17
18
19 Miguez MP, Anundi I, Sainz-Pardo LA et al. 1994. Hepatoprotective mechanism of
20 silymarin: no evidence for involvement of cytochrome P450 2E1. *Chem Biol Interact.* **91**:
21
22 51-63.
23
24
25

26
27
28
29 Morazzoni P, Magistretti MJ, Giachetti C et al. 1992. Comparative bioavailability of Silipide,
30 a new flavanolignan complex, in rats. *Eur J Drug Metab Pharmacokinet.* **1**: 39-44.
31
32
33

34
35
36 Morazzoni P, Bombardelli E. 1995. Silybum marianum (Carduus marianus). *Fitoterapia.*
37
38 **66**: 3-42.
39
40

41
42
43 Mourelle M, Muriel P, Favari L et al. 1989. Prevention of CCL4-induced liver cirrhosis by
44 silymarin. *Fundam Clin Pharmacol.* **3**: 183-191.
45
46
47

48
49
50 Nagata K, Suzuki H, Sakaguchi S. 2007. Common pathogenic mechanism in development
51 progression of liver injury caused by non-alcoholic or alcoholic steatohepatitis. *J Toxicol*
52
53 *Sci.* **32**: 453-468.
54
55
56

1
2
3 National Institutes of Health. Agency for Healthcare Research and Quality. 2002. Milk
4
5 Thistle: effects on liver disease and cirrhosis and clinical adverse effects. Evidence
6
7 report/technology assessment: number 21.
8
9

10
11
12 Negro F, Alaei M. 2009. Hepatitis C virus and type 2 diabetes. *World J Gastroenterol.* **15**:
13
14 1537-1547.
15
16

17
18
19 Orlando R, Fragasso A, Lampertico M et al. 1990. Silybin kinetics in patients with liver
20
21 cirrhosis: a comparative study of a silybin-phosphatidylcholine complex (Siliphos®) And
22
23 silymarin. *Med. Sci Res.* **18**: 861-863.
24
25

26
27
28 Palasciano G, Portincasa P, Palmieri V et al. 1994. The effect of silymarin on plasma
29
30 levels of malon-dialdehyde in patients receiving long-term treatment with psychotropic
31
32 drugs. *Curr Ther Res.* **55**: 537-545.
33
34

35
36
37 Parés A, Planas R, Torres M et al. 1998. Effects of silymarin in alcoholic patients with
38
39 cirrhosis of the liver: results of a controlled, double blind, randomized and multi-center trial.
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

59
60 Petrides AS, Vogt C, Schulze-Berge D et al. 1994. Pathogenesis of glucose intolerance
and diabetes mellitus in cirrhosis. *Hepatology.* **19**: 616-627.

Pradhan SC, Girish C. 2006. Hepatoprotective herbal drug, silymarin from experimental
pharmacology to clinical medicine. *Indian J Med Res.* **124**: 491-504.

1
2
3 Ramasamy K, Agarwal R. 2008. Multitargeted therapy of cancer by silymarin. *Cancer Lett.*
4
5 **269**: 352-362.
6
7

8
9
10 Rambaldi A, Jacobs BP, Laquinto G *et al.* 2005. Milk thistle for alcoholic and/or hepatitis B
11
12 or C liver diseases - a systematic Cochrane Hepato-Biliary Group Review with meta-
13
14 analyses of randomized clinical trials. *Am J Gastroenterol.* **100**: 2583-2591.
15
16
17

18
19
20 Ross SM. 2008. Milk thistle (*Silybum marianum*): an ancient botanical medicine for modern
21
22 times. *Holist Nurs Pract.* **22**: 299-300.
23
24
25

26
27 Saliou C, Rihn B, Cillard J *et al.* 1998. Selective inhibition of NF-kappa B activation by the
28
29 flavonoid hepatoprotector silymarin in HepG2. Evidence for different activating pathways.
30
31 *FEBS Lett.* **440**: 8-12.
32
33
34

35
36 Saller R, Meier R, Brignoli R. 2001. The use of silymarin in the treatment of liver diseases.
37
38 *Drugs.* **61**: 2035-2063.
39
40
41

42
43 Saller R, Brignoli R, Melzer J *et al.* 2008. An updated systematic review with meta-analysis
44
45 for the clinical evidence of silymarin. *Forsch Komplementmed.* **15**: 9-20.
46
47
48

49
50 Salmi HA, Sarna S. 1982. Effect of silymarin on chemical, functional, and morphological
51
52 alterations of the liver. A double-blind controlled study. *Scand J Gastroenterol.* **17**: 517-
53
54 521.
55
56
57
58
59
60

1
2
3 Singh D, Singh R, Singh P *et al.* 2009. Effects of Embelin on Lipid Peroxidation and Free
4 Radical Scavenging Activity against Liver Damage in Rats. *Basic Clin Pharmacol Toxicol.*
5 May 26. [Epub ahead of print].
6
7
8
9

10
11
12 Schulz HU, Schürer M, Krumbiegel G *et al.* 1995. The solubility and bioequivalence of
13 silymarin preparations. *Arzneimittelforschung.* **45**: 61-64.
14
15
16
17

18
19
20 Schümann J, Prockl J, Kiemer AK *et al.* 2003. Silibinin protects mice from T cell-
21 dependent liver injury. *J Hepatol.* 39, 333-340.
22
23
24
25

26
27 Sebestian J, Sebestianova SB, Moulisova V *et al.* 2004. Membrane transport without
28 receptors? The role of cyclosporines and silymarines structures for their interactions with
29 lipids of hepatocyte plasma membrane. *Mater Struct.* **11**: 15-17.
30
31
32
33

34
35
36 Sonnenbichler J, Zetl I. 1986. Biochemical effects of the flavonolignan silibinin on RNA,
37 protein, and DNA synthesis in rat liver. *Progr Clin Biol Res.* **213**: 319-331.
38
39
40
41

42
43 Tedesco D, Domeneghini C, Sciannimanico D *et al.* 2004. Silymarin, a possible
44 hepatoprotector in dairy cows: biochemical and histological observations. *J Vet Med A*
45 *Physiol Pathol Clin Med.* **51**: 85-89.
46
47
48
49

50
51
52 Trinchet JC, Coste T, Levy VG *et al.* 1989. Treatment of alcoholic hepatitis with silymarin.
53 A double-blind comparative study in 116 patients. *Gastroenterol Clin Biol.* **13**: 120-124.
54
55
56
57

58
59
60 Tsai JH, Liu JY, Wu TT *et al.* 2008. Effects of silymarin on the resolution of liver fibrosis
induced by carbon tetrachloride in rats. *J Viral Hepat.* **15**: 508-514.

1
2
3
4
5 Urtasun R, Conde de la Rosa L, Nieto N. 2008. Oxidative and nitrosative stress and
6 fibrogenic response. *Clin Liver Dis.* **12** : 769-790.
7
8
9

10
11
12 Velussi M, Cernigoi AM, Viezzoli L, et al. 1993. Silymarin reduces hyperinsulinemia,
13 malondialdehyde levels, and daily insulin need in cirrhotic diabetic patients. *Curr Ther Res.*
14
15
16
17 **53** : 533-545.
18
19

20
21
22 Velussi M, Cernigoi AM, De Monte A, et al. 1997. Long-term (12 months) treatment with an
23 anti-oxidant drug (silymarin) is effective on hyperinsulinemia, exogenous insulin need and
24 malondialdehyde levels in cirrhotic diabetic patients. *J Hepatol.* **26**: 871-879.
25
26
27
28

29
30
31 Voinovich D, Perissutti B, Grassi M et al. 2009. Solid state mechanochemical activation of
32 Silybum marianum dry extract with betacyclodextrins: Characterization and bioavailability
33 of the coground systems. *J Pharm Sci.* **98**: 4119-4129.
34
35
36
37

38
39
40
41 Wellington K, Jarvis B. 2001. Silymarin: a review of its clinical properties in the
42 management of hepatic disorders. *Bio Drugs.* **15**: 465-489
43
44
45
46

47
48 Weyhenmeyer R, Mascher H, Birkmayer J. 1992. Study on dose-linearity of the
49 pharmacokinetics of silibinin diastereomers using a new stereospecific assay. *Int J Clin*
50
51
52
53 *Pharmacol Ther Toxicol.* **30**: 134-138.
54
55

56
57
58 Wu JW, Lin LC, Tsai TH. 2009. Drug-drug interactions of silymarin on the perspective of
59 pharmacokinetics. *J Ethnopharmacol.* **21**: 185-193.
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Younossi ZM, McCullough AJ. 2009. Metabolic syndrome, non-alcoholic fatty liver disease and hepatitis C virus: impact on disease progression and treatment response. *Liver Int.* **29**: S2, 3-12.

For Peer Review

Table 1: Main constituents of Milk Thistle.

THE FRUITS	silybin
	silychristin
	silydianin
	3-deoxyderivatives of silychristin and silydianin (silymonin)
	neosilyhermin A
	neosilyhermin B
	2,3-dehydrosilybin
	taxifolin
	quercetin
	dihydrokaempferol
	kaempferol
	apigenin
	naringin
	eriodyctiol
	chrysoeriol
	5,7-dihydroxy chromone
	dehydroconiferyl alcohol
	silyhermin
	20-30% fixed oil (~60% linoleic acid; ~30% oleic acid; ~9% palmitic acid)
	0.038% tocopherol
0.63% sterols (cholesterol, campesterol, stigmasterol and sitosterol)	
25-30% protein, some mucilage	
flavonoids (apigenin and its 7-O- glucoside, 7-O- glucuronide; 4,7diglucoside, kaempferol and its 7-glucoside and 3-sulphate)	
THE HERB	luteolin and its 7-glucoside
	sitosterol and its glucoside
	triterpene acetate
	fumaric acid
	polyacetylenes

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Figure 1: Distribution of Milk thistle in European countries (in black).



Figure 2: Chemical formulas of Silybin, Silydianin, Isosilybin and Silychristin.

