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Regulatory role of cannabinoid receptor 1 in stress-induced excitotoxicity and neuroinflammation

Silvia Zoppi¹, MD; Beatriz G. Pérez Nievas¹, PhD; José L.M. Madrigal¹, PhD; Jorge Manzanares² PhD; Juan C. Leza¹ MD, PhD and Borja García-Bueno, PhD¹.

¹ Dpt. of Pharmacology. Fac. Medicine Univ.Complutense de Madrid (UCM), Centro de Investigación Biomédica en Red de Salud Mental (CIBERSAM) and Instituto de Investigación Sanitaria Hospital 12 de Octubre. Madrid.

² Instituto de Neurociencias, Universidad Miguel Hernández-CSIC, Avda, Ramón y Cajal s/n, San Juan de Alicante, 03550 Alicante.

Correspondence to:

Dr. Borja García-Bueno. Dpt. of Pharmacology. Fac. Medicine Univ.Complutense
28040 - Madrid (Spain)

bgbueno@med.ucm.es

Tel. +34 91 394 1478; Fax. +34 91 394 1464 / 65

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ABSTRACT

Exposure to stress elicits excitotoxicity and neuroinflammation in the brain, contributing to cell death and damage in stress-related neurological and neuropsychiatric diseases. The endocannabinoid system is present in stress-responsive neural circuits and has been proposed as an endogenous neuroprotective system activated in some neuropathological scenarios to restore homeostasis. In order to elucidate the possible regulatory role of CB1 in stress-induced excitotoxicity and neuroinflammation, both genetic and pharmacological approaches were used alternatively: 1) wild type (WT) and CB1 knockout mice (CB1-KO) were exposed to immobilisation/acoustic stress (2h/day for 4d), 2) to specifically activate CB1, the selective CB1 agonist ACEA (2.5 mg/kg) was i.p. administered daily to some groups of animals.

Stress exposure increased CB1 mRNA and protein expression in the prefrontal cortex of WT mice in a mechanism related to NMDA glutamate receptor activation. Daily ACEA pre-treatment prevented stress-induced: 1) upregulation of CB1 mRNA and protein, 2) decrease in glutamate uptake and glutamate astroglial transporter EAAT-2 expression, 3) increase in consecutive proinflammatory molecules, such as cytokines (TNF- α and MCP-1), nuclear factor kappa B (NF- κ B) and enzymatic sources, such as inducible nitric oxide synthase (NOS-2) and cyclooxygenase-2 (COX-2), 4) increase in lipid peroxidation; while having no effect on plasma corticosterone. Interestingly, a possible related mechanism could be the positive ACEA modulation of the antiinflammatory pathway deoxyprostaglandin/peroxisome proliferator activated receptor gamma (15d-PGJ₂/PPAR γ).

Conversely, KO animal experiments indicated that a lack of CB1 produces HPA axis dysregulation and exacerbates stress-induced excitotoxic/neuroinflammatory responses.

These multifaceted neuroprotective effects suggest that CB1 activation could be a new therapeutic strategy against neurological/neuropsychiatric pathologies with HPA axis dysregulation and an excitotoxic/neuroinflammatory component in their pathophysiology.

INTRODUCTION

Uncontrolled excitotoxicity and neuroinflammation contribute to cell death and damage in neurological and neuropsychiatric diseases, including some that are related to stress exposure (neurodegenerative diseases, depression, post-traumatic stress disorder and schizophrenia) (McLeod et al., 2001; Madrigal et al., 2006). Indeed, previous studies indicate that long-lasting stress affects synaptic plasticity, dendritic morphology, and neurogenesis in animals (reviewed in Kim and Yoon 1998) and induces both clinical and anatomical features of neurotoxic damage in humans (Bremner et al 1995). Moreover, the relationship between stressful events and the onset, evolution and resolution of inflammatory-related diseases (cardiovascular, neurological, digestive, or immunological) is well documented (Baum and Posluszny 1999; Slimmer et al 2001). The precise molecular and cellular events induced by stress that are responsible for the brain damage found in these conditions are still a matter of debate. In this way, previous studies using stress models (physical, psychological or mixed) reported a consistent brain response that is mainly characterized by excitotoxicity and a resulting activation of several consecutive inflammatory mediators (García-Bueno et al., 2008a). Briefly, stress exposure elicits excitotoxicity due to the massive release of the excitatory amino acid, glutamate, in some brain areas (e.g. the prefrontal cortex) (Moghaddam, 1993), inducing the release of proinflammatory cytokines such as tumour necrosis factor- α (TNF- α) or interleukin 1 β (Madrigal et al., 2002). Stress also activates the nuclear transcription factor-kappa B (NF- κ B) pathway in a TNF- α dependent mechanism (Bierhaus et al., 2003; Madrigal et al.,

2002). NF- κ B activation elicits the expression and activity of pro-inflammatory enzymatic sources, such as inducible nitric oxide (NO) synthase (NOS-2) and cyclooxygenase-2 (COX-2), among others (Madrigal et al., 2001a, 2003). The result of this sequence of events is the accumulation of oxidative and nitrosative mediators, which can attack membrane phospholipids and cause cell damage in a process known as lipid peroxidation (McEwen, 1998, Madrigal et al., 2001b) (see Figure 9 for a schematic view of this stress-activated pro-inflammatory pathway). However, there remain unresolved issues concerning the mechanisms, regulation, and functional significance of the excitotoxic/inflammatory processes induced by stress in the brain.

In spite of stress-induced production and accumulation of potentially cytotoxic and/or pro-inflammatory mediators like glutamate, NO, peroxynitrite anion (ONOO⁻) or prostaglandin E₂ (PGE₂), various authors have discussed the possibility that some of the many changes caused by stress response effectors are not damaging to the neurons but in fact predominantly beneficial to their structure and function (McEwen, 1998). One possible mechanism involved as a possible neuroprotective pathway that has received considerable experimental attention is led by some COX-2-derived lipid mediators, such as prostaglandin 15d-PGJ₂ and its peroxisome proliferator-activated nuclear receptor gamma (PPAR γ). This COX derived sub-pathway is activated in the brain in response to stress and other neuroinflammatory stimuli, constituting a possible endogenous anti-inflammatory defence mechanism against excessive excitotoxicity and neuroinflammation (García-Bueno et al., 2008b) (see Figure 8 for a schematic view of this stress-activated antiinflammatory pathway).

The study of alternative/related antiinflammatory pathways deserves further attention in order to understand pathophysiological inflammatory alterations as well as to develop possible new therapeutic approaches for the treatment of stress-related neuropathologies. Currently, the endocannabinoid system (ECS) is considered as an endogenous homeostatic system activated by different immune challenges, restoring

brain balance at different levels (Mechoulam and Shohami, 2007) in different experimental settings (Galve-Roperh et al., 2008; Wolf et al., 2008; Bambico et al., 2009; Cabral and Griffin-Thomas, 2009). ECS refers to a group of endogenous arachidonate-based lipids (i.e. anandamide and 2-arachidonoylglycerol), widely known as “endocannabinoids”; their cannabinoid G protein-coupled receptors, namely CB1 and CB2, and the enzymes fatty acid amide hydrolase and monoacylglycerol lipase that are responsible for their degradation and/or reuptake.

Interestingly, several epidemiological studies have identified that cannabinoids affect anxiety and stress responsivity (Ledent et al., 1999), and the presence of the ECS in stress-responsive neural circuits suggests that it may play a critical role in regulating Hypothalamic/Pituitary/Adrenal (HPA) axis activation and the resulting excitotoxic/neuroinflammatory response. Although it is widely assumed that this response is mainly mediated by the CB2 receptor, which is expressed by a wide range of immune cells (Arévalo-Martín et al., 2003), recent evidence suggests that pharmacological enhancement of CB1 protects neurons from acute brain injury, demonstrating a potentially attractive alternative therapeutic strategy against CNS pathologies that deserves further investigation (Cabral and Griffin-Thomas, 2009).

Taking into account all this background, the aim of the present study was to explore the effect of sub-chronic stress exposure on CB1 expression in the prefrontal cortex in mice and the possible regulatory role of CB1 in conditions of stress-induced excitotoxicity and neuroinflammation, using alternatively pharmacological (activation of CB1 with the selective agonist ACEA) and genetic (CB1 receptor-deficient mice) strategies.

MATERIAL AND METHODS

Animals

Adult male cannabinoid receptor 1 double mutant CB1 (-/-) mice and wild-type CB1 (WT) littermates (ICR (Swiss) strain) were used in all the experiments. The generation of mice lacking the CB1 receptor has been described previously (Ledent et al., 1999). WT and CB1 (-/-) mice used in a given experiment originated from the same breeding series and were matched for age and weight (age = 2–3 months; weight = 25–30 g). All experimental protocols adhered to the guidelines of the Animal Welfare Committee of the Universidad Complutense in accordance with European legislation (DC 86/609/CEE). The mice were housed individually with standard temperature and humidity conditions and in a 12-hour light/dark cycle (lights on at 8:00 am) with free access to food and water. All the animals were maintained under constant conditions for 4 days before the exposure to stress.

Stress protocol

Mice were exposed to sub-chronic immobilisation and acoustic stress using an ultrasound bath (2h from 13:00h to 15:00h for 4 days) as previously described (Kiank et al., 2006; Depke et al., 2008; Olfe et al., 2010). The restraint/acoustic stress model is classified as both physical and psychological, predictable, and inescapable, suitable for studying the neurochemical effects of stress on brain function. Some authors have evaluated restraint stress and acoustic stress both separately and in combination and found that larger and more predictable responses were obtained with the combined stressors (Qiu et al., 1999). Stressed animals were put down immediately after the restraint (whilst still in the restrainer) using sodium pentobarbital (320 mg/kg ip). Control animals were not submitted to stress but were handled at 13:00 h for a few seconds and food and water were removed. They were put down at the same time as

the stressed animals (15:00h). Blood for plasma determinations was collected by cardiac puncture and anticoagulated in the presence of trisodium citrate (3.15% (wt/vol), 1 vol citrate per 9 vol blood). After decapitation, the brain was removed from the skull, and after careful removal of the meninges and blood vessels, the prefrontal cortical areas from both brain hemispheres were excised and frozen at -80°C until assayed. Mouse brain prefrontal cortex was chosen because of its high levels of CB1 receptor, pro-inflammatory (NOS-2, prostaglandin E2) and antiinflammatory (PPAR γ , L-PGDS) mediators, its susceptibility to excitotoxic/neuroinflammatory process elicited by inflammatory/immune stimulus (Garcia-Bueno et al., 2008b) and finally because this brain area is an important neural substrate for the regulation of the Hypothalamic/Pituitary/Adrenal (HPA) axis response to stress (Radley et al., 2006).

Pharmacological tools

Various groups of animals were injected with the potent and highly selective CB1 receptor agonist ($K_i = 1.4 \text{ nM}$; >1400-fold CB1 selectivity over CB2) Arachidonyl-2'-chloroethylamide (ACEA), using a dose of 2.5 mg/kg administered intraperitoneally (i.p.) daily or its vehicle (a solution of 5% ethanol, 5% dextrose and 5% Tween 80 in sterile saline). The dose of ACEA was used based on preceding studies in neuroinflammation-related models (Arévalo-Martín et al., 2003). To block N-methyl-D-aspartate (NMDA) glutamate receptor-dependent effects, two groups of animals were i.p. injected with dizocilpine [(+)-5-methyl-10,11-dihydroxy-5H-dibenzo (a,d) cyclohepten-5,10-imine; MK-801], a specific non-competitive NMDA antagonist (MK; 0.1 mg/kg). This low dose was chosen to avoid possible neurotoxic, hallucinogenic, and HPA axis disturbing effects from this compound when it is used at higher doses and was based on preceding studies from our laboratory and others (García-Bueno et al., 2008; Jezova et al., 1995). ACEA and vehicle were injected at the onset of the stress (13:00h) and MK-801 and its vehicle (saline) were respectively i.p. injected 1h before the onset of the stress (12:00h). None of the parameters studied were modified

in the two different vehicle-treated groups of mice when compared with the non-injected animals. To simplify figures, both groups were unified into one vehicle-injected group in control (CWT) and stress (SWT) conditions. Each experimental group included at least six animals.

Preparation of cytosolic and nuclear extracts

A modified procedure based on the method of Schreiber et al (1989) was used. Tissues (cerebral cortex) were homogenized with 300 μ L of buffer (10 mmol/L N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid, pH 7.9; 1 mmol/L ethylenediamine tetraacetic acid (EDTA), 1 mmol/L EGTA, 10 mmol/L KCl, 1 mmol/L DTT), 0,5 mmol/L phenylmethylsulfonyl fluoride, .1 μ g/mL aprotinin, 1 μ g/mL leupeptin, 1 μ g/mL N α -p-tosyl-L-lysine-chloromethyl ketone, 5 mmol/L NaF, 1 mmol/L NaVO₄, .5 mol/L sucrose, and 10 mmol/L Na₂MoO₄). After 15 min, Nonidet P-40 (Roche) was added to a 0.5% concentration level. The tubes were gently vortexed for 15 sec, and nuclei were collected by centrifugation at 8000g for 5 min. Supernatants were taken as a cytosolic fraction. The pellets were resuspended in 100 μ L of buffer supplemented with 20% glycerol and 0,4 mol/L KCl and gently shaken for 30 min at 4 μ C. Nuclear protein extracts were obtained by centrifugation at 13,000g for 5 min, and aliquots of the supernatant were stored at -80 μ C. All steps of the fractionation were carried out at 4 μ C.

Western Blot Analysis:

After determining and adjusting protein levels, homogenates of prefrontal cortex tissue, once centrifuged (12000 g, 20 min at 4 μ C) were mixed with Laemmli sample buffer (Bio Rad, Hercules, CA, USA) (SDS 10%, distilled H₂O, glycerol 50%, Tris HCl 1M pH 6,8, dithiothreitol and blue bromophenol) with beta mercaptoethanol (50 μ L per mL of Laemmli) and 20 μ L (2 μ g/ μ L) were loaded into an electrophoresis gel. Once separated by molecular weight, proteins from the gels were blotted onto a

polyvinylidene difluoride membrane (Millipore, Bedford, MA, USA) or nitrocellulose membrane (Amersham Ibérica, Spain) with a semi-dry transfer system (Bio-rad) and were incubated with specific antibodies: a) rabbit polyclonal Cannabinoid Receptor I, raised against the synthetic peptide MSVSTDTSAEAL, corresponding to C terminal amino acids 461-472 of Human Cannabinoid Receptor I in a dilution of 1: 1000 in 5% bovine serum albumin (BSA) in TBS-Tween (Abcam, Cambridge, UK); b) rabbit polyclonal EAAT-2 raised against an epitope corresponding to amino acids 1-85 mapping at the N-terminus of EAAT-2 (excitatory amino acid transporter 2) of human origin in a dilution of 1:1000 in 5% BSA in TBS-Tween (sc-15317) (Santa Cruz Biotechnology, CA, USA); c) specific rabbit polyclonal PPAR γ (epitope corresponding to amino acids 8-106 mapping at the N-terminus of PPAR γ 1 of human origin) in a dilution of 1: 1000 in 5% BSA in TBS-Tween (sc-7196) (Santa Cruz Biotechnology); d) rabbit polyclonal I κ B α (epitope mapping at the C-terminus of I κ B- α of human origin) in a dilution of 1: 1000 in 5% Skimmed milk in BSA (Santa Cruz Biotechnology) e) rabbit polyclonal NF- κ B p65 (epitope mapping within the N-terminus of NF κ B p65 of human origin) in a dilution of 1:500 in BSA 2% (sc-109) (Santa Cruz Biotechnology); f) NOS-2 rabbit polyclonal antibody from Santa Cruz Biotechnology raised against a peptide mapping at the amino terminus of NOS-2 of human origin in a dilution of 1:1000 in TBS-Tween (sc-651); g) COX-2 goat polyclonal antibody from Santa Cruz Biotechnology raised against a peptide mapping at the C-terminus of COX-2 of human origin in a dilution of 1:750 in 5% BSA in TBS-Tween (sc-1745); h) lipocalin Prostaglandin D synthase (L-PGDS) rabbit polyclonal antibody which recognizes human L-PGDS amino acids 30-41 (VQPNFQPDKFLG) in a dilution of 1: 500 in 5% BSA in TBS-Tween (Cayman Chemical Europe, Tallinn, Estonia). Proteins recognised by the respective horseradish peroxidase-linked secondary antibodies (in a dilution of 1:1000 in 2-5% BSA in TBS-Tween) and visualised on X-ray film by chemiluminescence (ECL) following manufacturer's instructions (Amersham). Autoradiographs were quantified by densitometry (program Image J, NIH), and several

time exposures were analysed to ensure the linearity of the band intensities. All densitometries are expressed in arbitrary units (AU). In all the Western blot analyses, the house keeping gene β -actin was used as a loading control (the blots are shown in the respective figures).

mRNA analysis

Total RNA was extracted from cells using TRIZOL reagent (Invitrogen, Barcelona, Spain); aliquots were converted to cDNA using random hexamer primers. Quantitative changes in mRNA levels were estimated by real time PCR(Q-PCR) using the following cycling conditions: 35 cycles of denaturation at 95°C for 10 s, annealing at 58–61°C for 15 s depending on the specific set of primers, and extension at 72°C for 20 s. Reactions were carried out in the presence of SYBR green (1:10000 dilution of stock solution from Molecular Probes, Eugene, OR, USA), carried out in a 20- μ L reaction in a Corbett Rotor-Gene (Corbett Research, Mortlake, NSW, Australia).

The primers used for CB1 were: forward 5'-AAC TGT GCC AGG CAA TCT GTT GTG-3' and reverse 5'-GGC CTT CCC ATT GAA GCA CAA ACT-3'; for TNF- α : forward ACC ACT CTC CCT TTG CAG AAC TCA and reverse TCT CAT GCA CCA CCA TCA AGG ACT and for tubulin were: forward 5'-CCC TCG CCA TGG TAA ATA CAT-3' and reverse 5'-ACT GGA TGG TAC GCT TGG TCT-3'. Relative mRNA concentrations were calculated from the take-off point of reactions using included software, and tubulin levels used to normalize data.

Preparation of Synaptosomes

After decapitation, half of the forebrain was dissected on ice. All subsequent steps were performed at 4°C. Cortical tissue was immediately homogenized in 25 vols of 0.32M sucrose in a glass homogenizer fitted with a Teflon pestle. The homogenate

was centrifuged at 200g for 10 min, and the supernatant was then collected and centrifuged at 20 000g for 20 min. The pellet was resuspended in 0.32M sucrose and centrifuged at 20 000g for 20 min. The crude synaptosomal pellet was finally resuspended in 1ml of 0.32M sucrose. Synaptosomal preparations, consisting of pre- and post-synaptic elements of neurons, and associated astrocytic end feet, have provided insights into the regulation of a variety of synaptic functions including neurotransmitter release and reuptake, energy metabolism and ion transport systems in physiological, neurological, and neurodegenerative disorders (Begley et al., 1998).

[³H]Glutamate Uptake by Synaptosomes

Sodium-dependent glutamate uptake in synaptosomes was measured according to the procedure previously described by Robinson et al (1991) with some modifications. In brief, 25-ml aliquots of synaptosomes were added to 250 ml of incubation buffer (5mM Tris, 10mM HEPES, 2.5mM KCl, 1.4M NaCl, 1.2mM CaCl₂, 1.2mM MgCl₂, 1.2mM KH₂PO₄, and 10mM dextrose, pH 7.4) containing L-[³H]glutamic acid 0.125 mM (1mCi/ml; Amersham Biosciences Europe GmbH, Freiburg, Germany) and incubated for 3 min at 37°C in a shaking bath. The reaction was terminated using 1ml of ice-cold choline buffer (an incubation buffer in which an equimolar concentration of choline chloride was substituted for NaCl), and the samples were centrifuged at 10 000g for 2 min to recover the synaptosomes. The ³H-bound radioactivity was measured using a liquid scintillation counter.

Plasma corticosterone levels

Plasma was obtained from blood samples by centrifuging them at 1000g for 15 min immediately after the stress exposure. All plasma samples were stored at -20°C before assay with a commercially available radioimmunoassay kit (¹²⁵I-labeled corticosterone; DPC, Los Angeles, California). A γ counter was used to measure

radioactivity of the samples. The values obtained in the control animals (100 ± 34.22 ng/mL) matched the kit manufacturer's expected values in adult mice at the time of blood extraction (approximately 15:00h).

MCP-1 measurement

MCP-1 levels in the prefrontal cortex homogenates were detected using a specific ELISA kit for MCP-1, according to manufacturer's instructions (R&D systems, Abingdon, UK). The assay detection limits were 31.3-2000 pg/ml. To determine tissue levels of MCP-1, brain prefrontal cortices were removed and homogenised by sonication for 10 seconds at 4°C in PBS containing Complete Protease Inhibitor Cocktail (Roche Diagnostics). The homogenates were centrifuged ($13,000 \times g$, 10 min, 4°C), and supernatants were used for determinations using the ELISA kit. Protein content was determined by Bradford's method using bovine serum albumin as standard.

Lipid peroxidation

Lipid peroxidation was measured by the thiobarbituric acid test for malondialdehyde following the method described by Das and Ratty (1987) with some modifications. Cerebral cortex was sonicated in 10 volumes of 50 mmol/L phosphate buffer and deproteinized with 40% trichloroacetic acid and 5 mol/L HCl, followed by the addition of 2% (wt/vol) thiobarbituric acid in 0.5 mol/L NaOH. The reaction mixture was heated in a water bath at 90°C for 15 min and centrifuged at 12000g for 10 min. The pink chromogen was measured at 532 nm in a Beckman DU-7500 spectrophotometer (Beckman). The results were expressed as nmol/mg of protein.

Brain PGE₂ and 15d-PGJ₂ levels

PGE₂ and 15d-PGJ₂ brain levels were measured by enzyme immunoassay (EIA) using reagents in kit form (Prostaglandin E₂ EIA Kit-Monoclonal; (Cayman Chemical) and 15-deoxy- $\Delta^{12,14}$ - Prostaglandin J₂ ELISA Kit (DRG Diagnostics, Marburg, Germany), respectively. Samples were sonicated in 300 μ l homogenization buffer (0.1M phosphate buffer, pH7.4, 1mM EDTA and 10 μ M indomethacin) and purified by incubation in ethanol at 4x sample volume for 5 min at 4° C and then centrifugation at 3,000 x g for 10 min. They were acidified with glacial acetic acid to pH 3.5 and PGE₂ was extracted using SPE (C-18) columns (Amersham Biosciences, Buckinghamshire, UK) rinsed with methanol and water. After the application of samples, columns were washed with water and hexane and PGE₂ and 15d-PGJ₂ were eluted with ethyl acetate. Samples were then evaporated to dryness under nitrogen and resuspended in EIA buffer. Levels of PGE₂ and 15d-PGJ₂ were measured in a 96-well plate and read at 405 nm following manufacture's instructions. The sensitivity of the assay for PGE₂ was 15 pg/ml; intra- and interassay coefficients of variation were 6.6% and 15.5%, respectively, at 62.5 pg/ml and for 15d-PGJ₂ the sensitivity was 36.8 pg/ml; intra- and interassay coefficients of variation were 7.4% and 13%, respectively, at 4.4pg/ml.

Protein assay

Protein levels were measured using Bradford method based on the principle of protein-dye binding (Bradford, 1976).

Chemicals and statistical analyses

Unless otherwise stated, the chemicals were from Sigma Spain, Madrid. Data in text and figures are expressed as mean \pm SEM. For multiple comparisons, a one-way ANOVA followed by the Newman-Keuls post hoc test to compare all pairs of means between groups was made. A p value < 0.05 was considered statistically significant.

RESULTS

1.- CB1 expression in the prefrontal cortex of control and stressed WT mice.

Glutamate related mechanism.

Our first study was aimed to elucidate the possible effect of stress exposure on CB1 expression in the prefrontal cortex in mice. Western Blot (WB) studies and subsequent densitometric analysis of the band of interest (Fig. 1A) revealed a significant increase in CB1 protein expression (≈ 60 kDa band) elicited by stress. As expected, this band is absent in CB1 knockout mice in control and stress conditions, validating the specificity of the antibody used. This observed increase in protein expression is in agreement with CB1 mRNA up-regulation, quantified by Q-PCR (Fig. 1B).

Pre-stress daily ACEA administration blocked stress-induced CB1 mRNA up-regulation (Fig. 1B) and protein expression (Fig. 1C) in the prefrontal cortex in mice. In control conditions, neither ACEA nor MK treatments were able to modify CB1 protein and mRNA expression (data not shown to simplify figures 1B-C).

Stress exposure leads to the massive release of glutamate into the synaptic cleft, reaching excitotoxic levels in some brain areas (i.e. hippocampus, prefrontal cortex) (Moghaddam et al., 1993). In order to elucidate whether stress induced CB1 up-regulation is related to this massive glutamate release, a group of animals was i.p. injected with the NMDA selective blocker MK-801 1h before the onset of the stress exposure. Interestingly, daily administration of MK-801 partly inhibited stress induced CB1 mRNA up-regulation (Fig. 1B) and protein over-expression (Fig. 1C) in the prefrontal cortex of WT mice.

2.- CB1-dependent effects on stress-induced changes in synaptosomal glutamate uptake and expression of glutamate transporters.

The Endocannabinoid system has been shown to confer neuroprotection inhibiting glutamatergic excitotoxic neurotransmission by mechanism/s still not completely understood. We decided to explore the possible effects of CB1 on glutamate transport in brain cortical synaptosomes from WT and CB1 KO mice submitted to stress.

As it has been previously described (García-Bueno et al., 2007), stress exposure leads to a marked decrease in glutamate uptake compared to what was seen in the control group (Fig 2A). Pre-treatment with ACEA fully prevented this stress-induced impairment in glutamate uptake function (Fig 2A). Conversely, CB1 KO mice showed dramatically reduced glutamate transport in cortical synaptosomes in both control and stress conditions (Fig 2A).

We also assessed the possibility that these effects might be due to expressional changes in the main brain glutamate transporter, EAAT-2 (astrocytic). Western blot analysis revealed that stress exposure reduced EAAT-2 expression (Fig. 2B), and that treatment with ACEA prevented this stress-induced decrease. As in the case of synaptosomal glutamate transport, both stressed and non stressed CB1 KO mice showed reduced EAAT-2 expression in brain cortical homogenates (Fig. 2B).

3.- Antiinflammatory effects elicited by CB1 activation. Mechanisms involved

In general, ECS has been proposed as an endogenous protective system against excessive inflammatory/immune responses in multiple CNS pathologies. Our following studies were aimed at clarifying the particular role of CB1 as a possible regulator of stress-induced inflammatory response.

3.1: Effects on HPA axis activity. Plasma corticosterone levels

Our first study evaluated CB1 effects on glucocorticoid synthesis, the main stress hormones, widely known as classical regulators of the inflammatory/immune response in brain and periphery.

The quantification by RIA of plasma corticosterone levels at the time of blood extraction (approximately 15:00h) in all groups studied revealed an expected increase in corticosterone in all groups of stressed animals compared to their respective controls (Fig. 3); however, this increase is statistically higher in KO mice than in WT animals (Fig. 3). Interestingly, daily ACEA administration did not change plasma corticosterone levels after stress at the time of blood extraction (Fig. 3), suggesting that the mechanism(s) of this compound in modulating stress response could be independent of peripheral corticosterone levels.

3.2: Brain pro-inflammatory cytokines and chemokines

Pro-inflammatory cytokines, such as TNF- α or MCP-1, are the first pro-inflammatory mediators activated in the brain by the massive release of glutamate after stress exposure. Q-PCR studies showed a consistent increase in TNF- α mRNA in WT mouse prefrontal cortex after stress, an effect that is blocked by ACEA (Fig. 4A). Conversely, stressed CB1 KO animals presented a reliable increase in the mRNA of the pro-inflammatory cytokine, suggesting a potentially deleterious chronic inflammatory status in brain. A very similar profile is maintained for the chemoattractant chemokine MCP-1 also studied: stress increased MCP-1 levels in the prefrontal cortex of WT mice and pharmacological stimulation of CB1 decreased them. CB1 KO mice showed no differences from WT in non-stressed animals. However, stress increased MCP-1 levels in brain in SKO group compared with SWT group (Fig. 4B).

In this set of neuroinflammation-related experiments (Figs. 4-7) there were no differences between CWT and CWT+ACEA groups and the data is not shown to simplify figures.

3.3: NF κ B

The release of TNF- α after stress also accounts for NF- κ B activation and we also tested the possible involvement of CB1 in the level of NF- κ B expression.

Western Blot studies of the inhibitory protein of NF- κ B (I κ B α) in cytosolic extracts (Fig 5A) and pro-inflammatory NF κ B subunit (p65) in nuclear extracts (Fig 5B) revealed that CB1 regulated NF- κ B activation after stress. As it can be observed, stress exposure induced a decrease in the expression of I κ B α in brain extracts from WT mice, which was reversed by ACEA (Fig. 5A). Levels of nuclear p65 in the different groups studied (Fig 5B) mirrored the expression of its inhibitory factor. CB1 KO animals presented lower levels of I κ B α in cytosolic extracts than their respective controls (CWT) (Fig. 5A) and a consequent increase in p65 protein in nuclear extracts (Fig 5B), suggesting a state of chronic NF- κ B activation in these animals. Interestingly, stress exposure did not affect I κ B α /NF- κ B expression in the prefrontal cortex of CB1 KO mice.

3.4: Pro-inflammatory enzymes (NOS-2 and COX-2)

NF- κ B regulates the expression of genes involved in the accumulation of oxidative/nitrosative and inflammatory mediators after stress exposure. Among others, two main sources of these mediators dependent on NF- κ B are inducible NO synthase (NOS-2) and cyclooxygenase-2 (COX-2). Their major products (NO and PGE₂, respectively) are potent oxidant/pro-inflammatory molecules that have been widely connected with damage and even cellular death in multiple CNS pathologies. We decided to explore CB1 effects on the expression and/or activity of these enzymes.

NOS-2 expression increased in the cerebral cortex after stress exposure in WT mice (Fig. 6A), although no changes in COX-2 expression were found after this stress

protocol (Fig. 6B, inset). However, the presumed brain COX-2 derived product, PGE₂, appeared in an increased quantity after the stress exposure (Fig. 6B).

CB1 receptor activation with ACEA was able to completely block NOS-2 up-regulation produced by stress exposure (Fig. 6A). As in the case of other pro-inflammatory mediators, CB1 KO mice both in control and stress conditions showed consistent NOS-2 up-regulation compared with their WT control groups, and a higher increase after stress (Fig. 6A).

The interaction between CB1 receptor and COX-2 is more complex, as suggested by our results. PGE₂ cortical levels, a presumed COX-2 product in different tissues including the brain, are affected by CB1 activation. As it can be observed in Fig 6B, ACEA reduced PGE₂ tissue levels compared with those in the SWT animals, whereas CB1 SKO animals presented insignificant differences in PGE₂ content compared with the SWT group.

3.5: Lipid peroxidation

As a final index of stress-induced cellular damage that could be affected by CB1 selective modulation, we measured the accumulation of the lipid peroxidation marker malondialdehyde (MDA) in the brain prefrontal cortex of the different groups of mice. Daily treatment with ACEA prevented MDA over-accumulation observed after stress exposure in WT mice (Fig. 6C). Conversely, CB1 KO mice presented higher levels of MDA after stress than the SWT group (Fig. 6C).

4.- CB1-dependent modulation of the antiinflammatory pathway L-PGDS/15d-PGJ₂/PPAR γ

So far, our results had shown that the daily administration of the selective CB1 receptor agonist ACEA prevented stress-induced excitotoxicity and pro-inflammatory

mediator accumulation in the prefrontal cortex of WT mice, whereas CB1 KO mice presented an aggravated excitotoxic/neuroinflammatory process after stress exposure. Our final series of studies were aimed to elucidate possible alternative antiinflammatory mechanisms related to CB1 activation. One candidate of interest is the anti-inflammatory pathway L-PGDS/15d-PGJ₂/PPAR_γ, that is activated after inflammatory/immune stimuli (stress included) and whose activation (even by some cannabinoids) has been shown to confer neuroprotection after stress exposure and in diverse CNS pathologies with an inflammatory component in their pathophysiology.

Western Blot studies and subsequent densitometric analysis (Fig. 7A) revealed a significant increase in the 15d-PGJ₂ enzymatic source L-PGDS in the prefrontal cortex homogenates of mouse brains daily treated with ACEA compared with the SWT group. On the other hand, L-PGDS protein levels were reduced in the brain of CB1 KO mice in control conditions and after stress exposure (Fig 7A).

Consequently, 15d-PGJ₂ levels, quantified by ELISA in brain prefrontal cortex samples, were increased by the administration of ACEA in WT mice compared to their controls (Fig 7B). Both in WT and CB1 KO mice, stress exposure produced a marked decrease in 15d-PGJ₂ levels but there were no differences between WT and KO animals.

WB analysis of PPAR_γ protein revealed a significant increase of this 15d-PGJ₂ nuclear receptor in prefrontal cortex homogenates of mouse brains daily treated with ACEA compared with their controls after stress exposure (SWT) (Fig 7C). CKO mice presented significant lower levels of PPAR_γ than the CWT group but stress exposure did not affect PPAR_γ expression in the brains of CB1 KO animals (Fig 7C).

5.- Validation of ACEA CB1-dependent antiexcitotoxic/antiinflammatory effects.

Finally in order to elucidate if ACEA actions are CB1-mediated, we examined its effects in CB1 KO mice on some representative excitotoxic (EAAT-2 expression) and

inflammatory parameters (NOS-2 expression and MDA levels) after sub-chronic stress exposure.

As has been previously shown (see Fig 2), stress exposure down-regulates EAAT-2 expression in mouse prefrontal cortex and this effect is completely prevented by ACEA pre-treatment in WT mice, however this ACEA anti-excitotoxic effect is absent in CB1 KO mice (SKO+ACEA) (Fig 8A), suggesting the direct involvement of CB1 receptor. A similar pattern occurs with the pro inflammatory parameter NOS-2 expression (Fig 8B) and MDA levels (Fig 8C) in which ACEA neuroprotective effects are lost in CB1 KO mice compared with WT. These results suggest that ACEA antiinflammatory effects are also dependent on CB1 in our model.

DISCUSSION

Our studies show for the first time to our knowledge, CB1 up-regulation in mouse prefrontal cortex exposed to sub-chronic restraint/acoustic stress in a mechanism dependent on NMDA glutamate receptors. In addition, daily ACEA administration prevented stress-induced upregulation of CB1 mRNA and protein, and presented an anti-excitotoxic profile, regulating glutamate transport and uptake after stress exposure, at least in part by restoring protein expression of EAAT-2. CB1 activation also produces antiinflammatory effects, regulating stress-induced pro-inflammatory response at multiple levels. Notably, a possible related mechanism is the positive modulation of the antiinflammatory pathway L-PGDS/15d-PGJ₂/PPAR_γ, although the precise mechanism/s responsible for this relationship needs to be further investigated (see figure 9 for a schematic view of the regulatory role of CB1 in stress-induced excitotoxicity and neuroinflammatory balance).

Stress exposure is a major contributing factor to cell death and damage in neurological and neuropsychiatric diseases by three interrelated mechanisms: HPA axis dysregulation, excitotoxicity and neuroinflammation. However, stress exposure

also activates alternative mechanisms aimed at correctly resolving this response. In this way, the CB1 up-regulation demonstrated here could be a mechanism related to this protective response of the brain. Similarly, endocannabinoids are released to negatively regulate HPA axis activation (Manzanares et al., 1999; Patel et al., 2004) and brain SEC activation has been considered as a protective mechanism aimed at reducing both neurodegenerative and inflammatory damage after an immune/inflammatory challenge (Centone et al., 2008).

In particular, our data is in agreement with several studies that propose a role for CB1 as a main regulator of glutamate neurotransmission and metabolism in multiple pathological scenarios (Parmentier-Batteur et al., 2002; Marsicano et al., 2003; Veldhuis et al., 2003; Fernández-López et al., 2006; Kim et al., 2006; Gilbert et al., 2007; Loría et al., 2010). In relation to our findings, stress-induced CB1 up-regulation could represent a compensatory response aimed at controlling the massive release of glutamate into the synaptic cleft that might produce cellular damage and even death by excitotoxicity after stress exposure (Lowy et al., 1993; Stein-Behrens et al., 1994; García-Bueno et al., 2007). As reported here, CB1 receptor activation with ACEA enhances the main glutamate uptake mechanism, by restoring protein expression of the glutamate transporter EAAT-2. Astrocyte uptake of glutamate is a high affinity process regulated by EAAT-2 (Hertz et al., 1983); the fact that stress decreases EAAT-2 expression in the cerebral cortex (García-Bueno et al., 2007) indicates a specific damaging effect at this level, and makes the effect of ACEA more interesting. CB1 effects on EAAT-2 have also been demonstrated in other experimental settings, such as an *in vitro* model of multiple sclerosis (MS) (Loría et al., 2010) or after prenatal exposure to WIN 55,212-2 (Castaldo et al., 2007). It has been widely demonstrated that presynaptic CB1 inhibits glutamate release (Freund et al., 2003; Chevaleyre et al., 2006; Steiner and Wotjak, 2008), but our data suggests that glutamatergic neurotransmission is also regulated by CB1 at the level of postsynaptic reuptake clearance mechanisms.

A possible explanation of how neuronal CB1 activation enhances glutamate glial uptake may be via neuron-astrocyte communication that regulates PPAR γ activity in astrocytes. Notably, EAAT-2 glutamate transporter is a target gene of PPAR γ in excitotoxic-related models, such as stress exposure (García-Bueno et al., 2007) or brain ischaemia (Romera et al., 2007). As well, a recent study identified endocannabinoids as PPAR γ agonists affording neuroprotection targeting EAAT-2 in an *in vitro* model of MS (Loría et al., 2010).

Regarding stress-induced neuroinflammation, although cannabinoid-dependent antiinflammatory and antioxidant actions have been commonly related with CB2 receptor activation in brain immune cells (Cabral and Griffin-Thomas, 2009; Ashton and Glass, 2007; Arévalo-Martín et al., 2008), or even with receptor-independent mechanisms (Hampson et al., 2000; Marsicano et al., 2002)), our and other studies revealed a significant role for CB1 as well in inflammatory/immune responses in animal models of MS (Arévalo-Martín et al., 2003, Pryce et al., 2003; Maresz et al., 2007) and brain ischaemia (Fernández-López et al., 2006).

Here we demonstrated that CB1 activation interacted with the proinflammatory response elicited by stress at multiple levels. In the case of cytokines, CB1 activation has been shown to reduce TNF α and IL-1 β synthesis in neurons, astrocytes and microglia (Fernández-López et al., 2006, Maresz et al., 2007, Marchalant et al., 2009). Notably, the opposite relationship, in which cytokines presumably up-regulate cannabinoid receptor expression, has also been suggested in some immune/inflammatory scenarios (Jean-Gilles et al., 2010), stress included (Lee et al., 2006).

Regarding other stress-induced proinflammatory mediators, like the key transcription factor NF- κ B, our results coincide with others showing a role for CB1 in NF- κ B regulation in injured mouse brains (Panikashvili et al., 2005); however regulation of NF- κ B by cannabinoids has been commonly related with CB2 in some immune cells such as macrophages or monocytes (Jeon et al., 1996; Rajesh., 2008); and even with

CB1/CB2 receptor independent mechanisms, as is the case of cannabidiol in LPS activated microglia (Kozela et al., 2010) or 2-arachidonyl glycerol through PPAR γ (Rockwell et al., 2006).

Two main NF- κ B-dependent enzymatic sources of oxidative/nitrosative mediators after stress in the brain are NOS-2 and COX-2. NOS-2 expression is down-regulated by selective CB1 activation in neurons and astrocytes both in *in vitro* and *in vivo* inflammatory-related models (Fernández-López et al., 2006; Esposito et al., 2006; Sheng et al., 2005), as it is also reported in our study. However, cannabinoid-cyclooxygenase interactions may be more intricate. In our stress model, CB1 regulates the levels of the presumed COX-2 product PGE₂ without altering enzyme expression. These results suggest possible effects of ACEA on the catalytic activity or protein stability of COX-2, or a possible effect on the activity of tissue-specific prostaglandin E₂ synthases, which is yet to be explored. Some authors have reported opposite COX-2/CB1 interactions: protection afforded by 2-Arachidonoylglycerol and THC by down-regulation of COX-2 expression and activity in cultured neurons and LPS stimulated J774 macrophages (Zhang and Chen, 2008; Chang et al., 2001), or THC neurotoxicity in hippocampal neurons in a mechanism attributable to COX-2 activity and the concomitant generation of free radicals (Chan et al., 1998).

Our results show expression changes in the antiinflammatory pathway L-PGDS / 15d-PGJ₂ / PPAR γ after CB1 pharmacological and genetic manipulations; although still speculative, the involvement of this antiinflammatory pathway in CB1 neuroprotective effects is an interesting idea that deserves further investigation. This potential relationship is especially relevant since some pharmacological and genetic studies using CB1/CB2 KO mice indicate possible CB1/CB2 alternative mechanisms of action. PPARs could be excellent candidates, taking into account that diverse cannabinoids are potent ligands of PPARs (O'Sullivan, 2007). Thus, the main endocannabinoids, anandamide and 2-arachidonoylglycerol and also ajulemic acid, a THC structural analogue, have anti-inflammatory properties mediated by PPAR γ (Sun

and Bennett, 2007). In addition, the endogenous acylethanolamines, oleoylethanolamide and palmitoylethanolamide have neuroprotective effects mediated through PPAR α . The possibility of these related acylethanolamines modulating the stress response via interaction with PPAR α remains unexplored.

Our results show a significant upregulation of L-PGDS / 15d-PGJ₂ after ACEA administration. This up-regulation has been also demonstrated after the administration of ajulemic acid to human synovial cells (Stebulis et al., 2008), inducing an antiinflammatory profile of eicosanoids. PPAR γ regulation is not yet completely understood, but one of the main mechanisms may be ligand availability (Daynes and Jones, 2002). This kind of regulation could be important in our stress model because we also found increased 15d-PGJ₂ levels after ACEA administration. Other authors have proposed alternative mechanisms by which cannabinoids can potentially lead to PPARs activation. For example, CB1 binding could stimulate MAPK pathways, as it occurs in other scenarios (Lépicier et al., 2006), which regulates PPAR γ transcriptional activity by direct phosphorylation on serine residues (Burns et al., 2007).

Contrasting ACEA neuroprotective effects in stress conditions, we and others have showed a dysregulated HPA axis activity in basal and after stress exposure in CB1 KO mice (Barna et al., 2004; Cota et al., 2008; Aso et al., 2008). In addition, a status of excessive excitotoxicity and neuroinflammation is also presented, increasing their susceptibility to experience damage and even cellular death. Similarly, in an animal model of Parkinson's disease, other authors have also found increased vulnerability to 6-hydroxydopamine lesion in CB1 KO mice, presenting a higher index of lipid peroxidation and NOS-2 expression in selected brain areas (Pérez-Rial et al., 2009).

In addition to HPA axis dysregulation, chronic excitotoxicity and neuroinflammation, CB1 KO mice present defective adult neurogenesis (Jin et al., 2004), increased anxiety (Urigüen et al., 2004; Mikics et al., 2009) and deficiency in neuronal plasticity by decreasing BDNF levels in the hippocampus (Aso et al., 2008),

suggesting a possible role of CB1 in the patho-physiology of depression and its pharmacological manipulation as a potential therapeutic strategy.

In summary, the multifaceted neuroprotective effects here described suggest that CB1 activation is an attractive therapeutic strategy against diverse neuropsychiatric pathologies with HPA axis dysregulation and an excitotoxic/neuroinflammatory component in their patho-physiology.

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FINANCIAL DISCLOSURES

The authors declare that there is **NO** conflict of interest to disclose

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FIGURE LEGENDS

Figure 1.- CB1 up-regulation is elicited by stress exposure in mouse prefrontal cortex in a glutamate related mechanism. (A) Western blot and densitometric analysis of CB1 band in the prefrontal cortex of control (CWT) and stressed during 4d (SWT) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. The densitometric data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT. One-way ANOVA (ow-ANOVA) following Newman-Keuls post-test. AU: arbitrary units. **(B)** Q-PCR analysis of CB1 mRNA in the prefrontal cortex of control (CWT), stressed during 4d (SWT), stressed+ ACEA (SWT+ACEA) and stressed+ MK-801 (SWT+MK) wild type mice. Data is normalized by tubulin and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT. One-way

ANOVA (ow-ANOVA) following Newman–Keuls post-test. AU: arbitrary units. **(C)** Western blot and densitometric analysis of CB1 band in the prefrontal cortex of control (CWT), stressed during 4d (SWT), stressed+ACEA (SWT+ACEA) and stressed+ MK-801 (SWT+MK) wild type mice. Data is normalized by β -actin and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units.

Figure 2.- CB1 effects on synaptosomal glutamate uptake and transporters. (A)

Glutamate uptake in forebrain synaptosomes of control (CWT), control+ACEA (CWT+ACEA), stressed during 4d (SWT) and stressed+ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. The data represents the mean \pm SEM of six mice. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). **(B)** Western blot and densitometric analysis of glial glutamate transporter EAAT-2 band in the prefrontal cortex of control (CWT), control+daily ACEA (CWT+ACEA), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units.

Figure 3.- CB1-related effects on plasma corticosterone levels.

Corticosterone plasma levels (ng/mL) at the time of blood extraction (15:00h) of control (CWT), control+daily ACEA (CWT+ACEA), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data represents the mean \pm SEM of six mice. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; & $p < 0.05$ vs CKO (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test).

Figure 4.- CB1 antiinflammatory effects on brain I. Proinflammatory cytokines and chemokines. A) Q-PCR analysis of the proinflammatory cytokine TNF- α mRNA in the prefrontal cortex of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by tubulin and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT. One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test. AU: arbitrary units. **B)** MCP-1 levels ($\mu\text{g}/\text{mg}$ prot.) in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data represents the mean \pm SEM of six mice. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; & $p < 0.05$ vs CKO (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test).

Figure 5.- CB1 antiinflammatory effects on brain II. NF- κ B. A) Western blot and densitometric analysis of the NF- κ B inhibitory protein I κ B α in prefrontal cortex cytosolic extracts of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units. **B)** Western blot and densitometric analysis of the NF- κ B proinflammatory subunit p65 in prefrontal cortex nuclear extracts of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units.

Figure 6.- CB1 antiinflammatory effects on brain III. Proinflammatory enzymes and lipid peroxidation. A) Western blot and densitometric analysis of the

proinflammatory enzyme NOS-2 in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units. **B)** Proinflammatory PGE₂ levels (index of COX-2 activity) in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. The data represent the mean \pm SEM of six mice. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; & $p < 0.05$ vs CKO (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). **Inset:** COX-2 expression by Western blot in all groups studied. **C)** MDA levels in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data represents the mean \pm SEM of six mice. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; & $p < 0.05$ vs CKO (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test).

Figure 7.- CB1 interactions with the antiinflammatory pathway L-PGDS/15d-PGJ₂/PPAR γ . **A)** Western blot and densitometric analysis of the enzyme L-PGDS in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units. **B)** Antiinflammatory 15d-PGJ₂ levels in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. The data represents the mean \pm SEM of six mice. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; & $p < 0.05$ vs CKO (One-way ANOVA (ow-ANOVA)

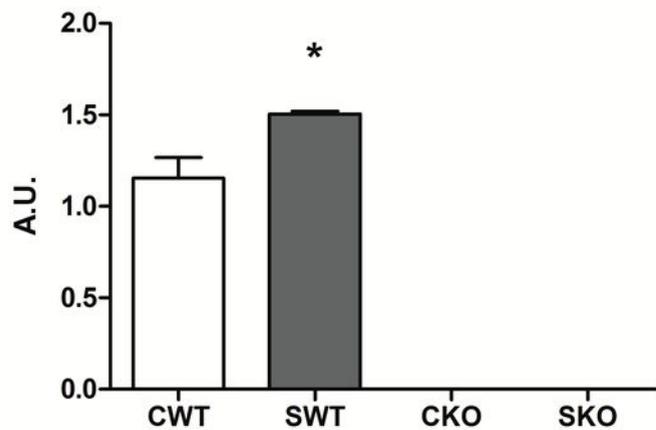
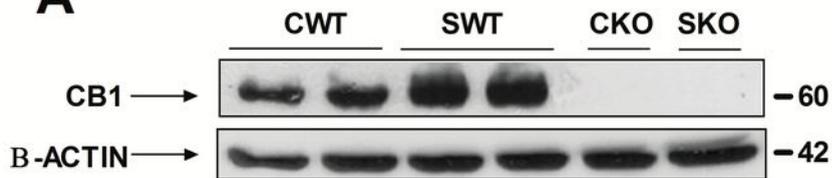
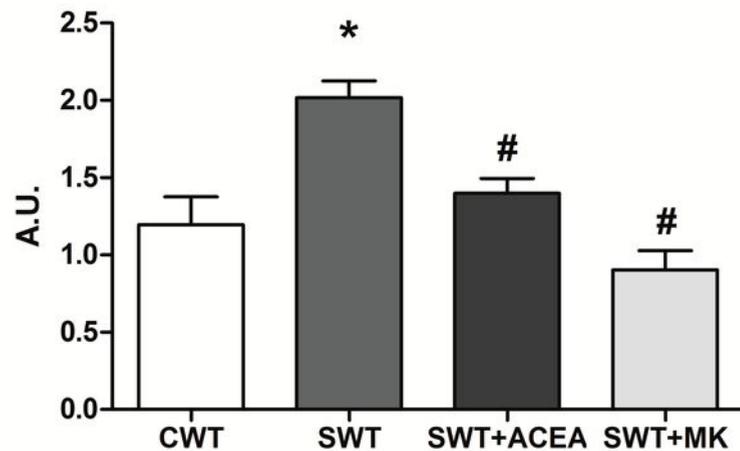
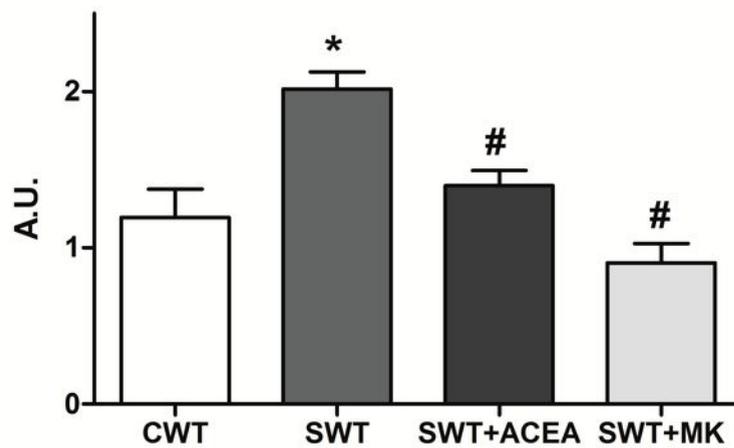
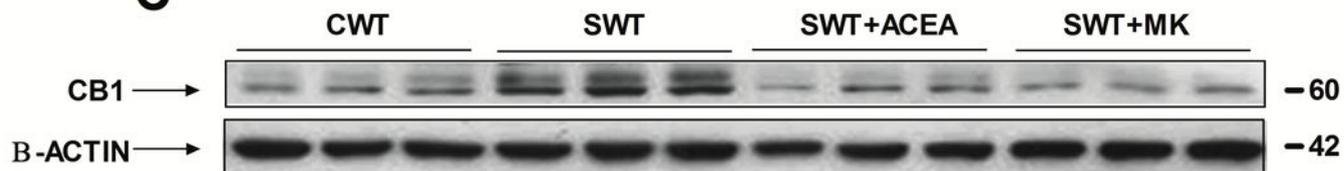
following Newman–Keuls post-test) **C)** Western blot and densitometric analysis of the nuclear receptor PPAR γ in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and control (CKO) and stressed (SKO) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units.

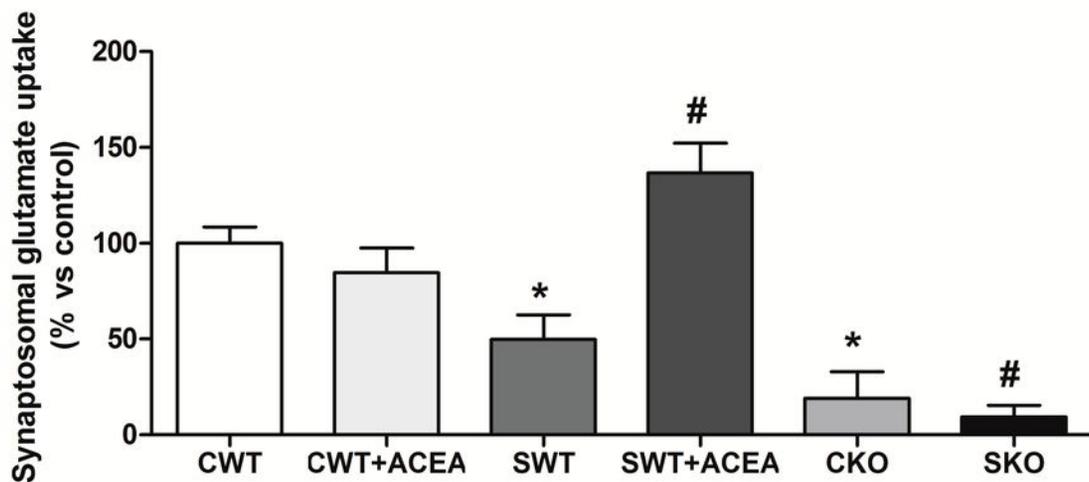
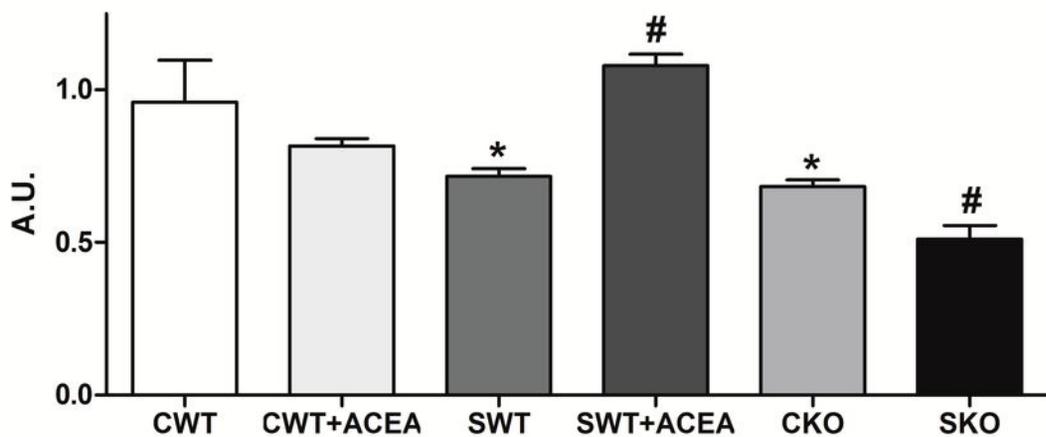
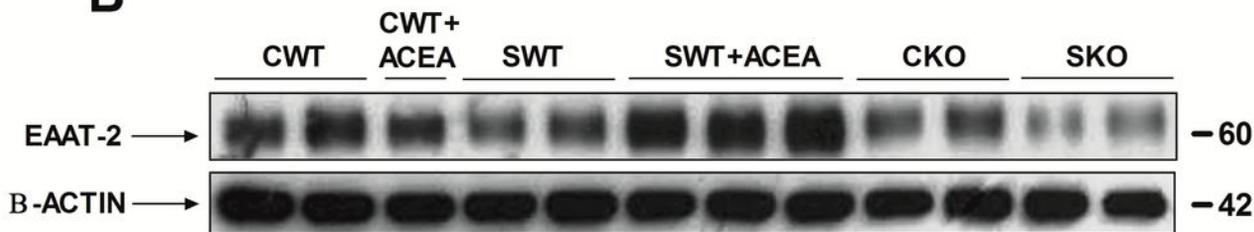
Figure 8. Validation of CB1-mediated ACEA effects.

A) Western blot and densitometric analysis of the astroglial glutamate transporter EATT-2 in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and stressed+daily ACEA (SKO+ACEA) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; \$ $p < 0.05$ vs SWT+ACEA (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units. **B)** NOS-2 protein expression analysis by Western blot and densitometric analysis of the band of interest in prefrontal cortex samples of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and stressed+daily ACEA (SKO+ACEA) CB1 KO mice. Data is normalized by β -actin (lower band) and is representative of 3 experiments. * $p < 0.05$ vs CWT; # $p < 0.05$ vs SWT; \$ $p < 0.05$ vs SWT+ACEA (One-way ANOVA (ow-ANOVA) following Newman–Keuls post-test). AU: arbitrary units. **C)** MDA levels (lipid peroxidation index) in prefrontal cortex homogenates of control (CWT), stressed during 4d (SWT) and stressed+daily ACEA (SWT+ACEA) wild type mice and stressed+daily ACEA (SKO+ACEA) CB1 KO mice. Data represents the mean \pm SEM of six mice.

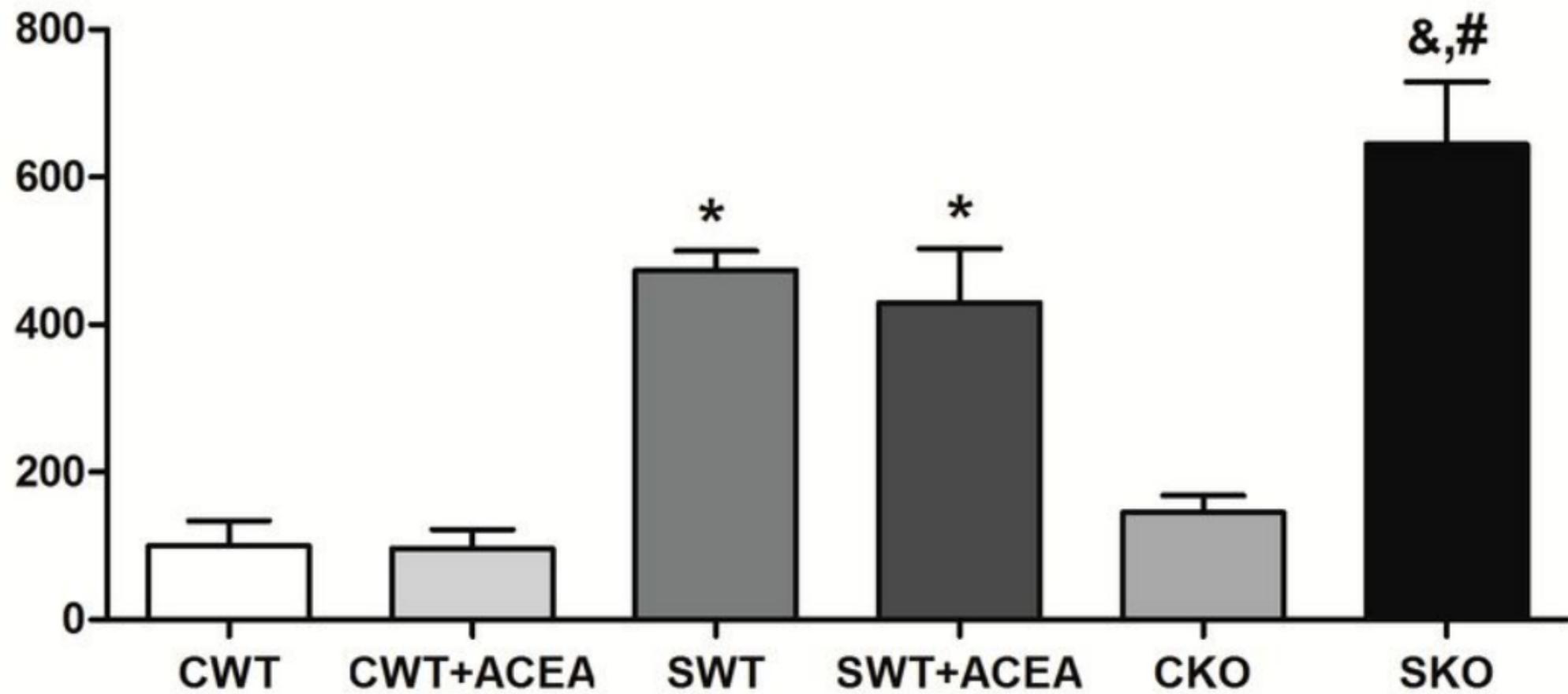
Figure 9. Schematic view of the regulatory role of CB1 on stress-induced excitotoxicity and neuroinflammatory balance.

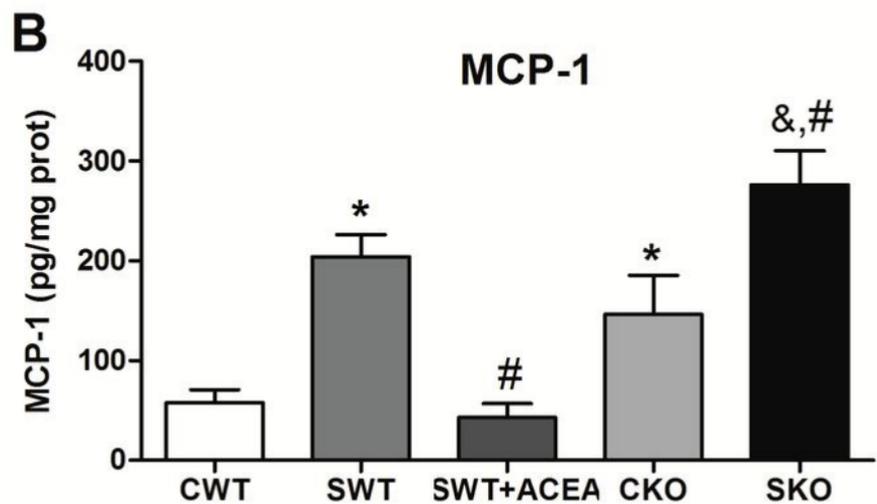
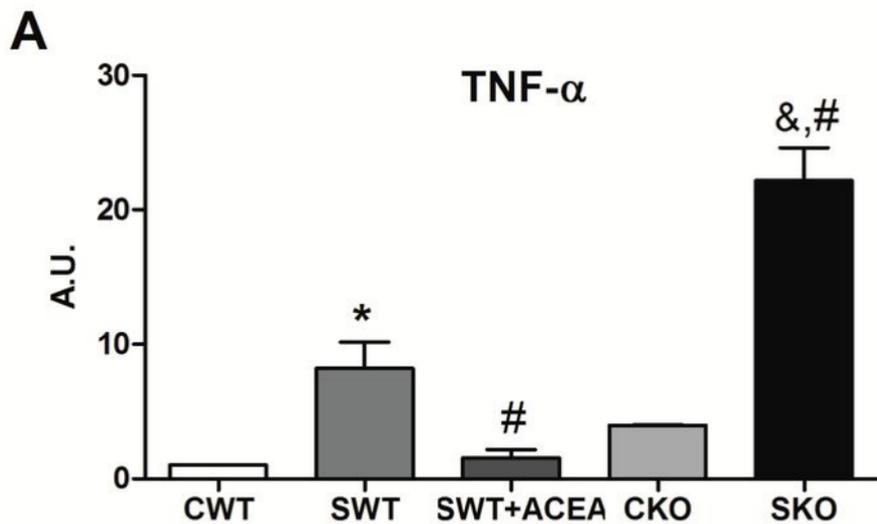
NMDA: N-Methyl-D-Aspartate receptor; EAAT-2: excitatory aminoacid transporter-2; CB1: cannabinoid receptor 1; NFκB: nuclear factor kappa B; NOS-2: nitric oxide synthase type II; COX-2: cyclooxygenase type II; L-PGDS: lipocalin prostaglandin D synthase; 15d-PGJ₂: 15d-Prostaglandin J₂; PPARγ: peroxisome proliferators activated receptor gamma; (-): inhibition; (+): stimulation; ?: alternative mechanisms. Black: proinflammatory mediators. Grey: antiinflammatory mediators

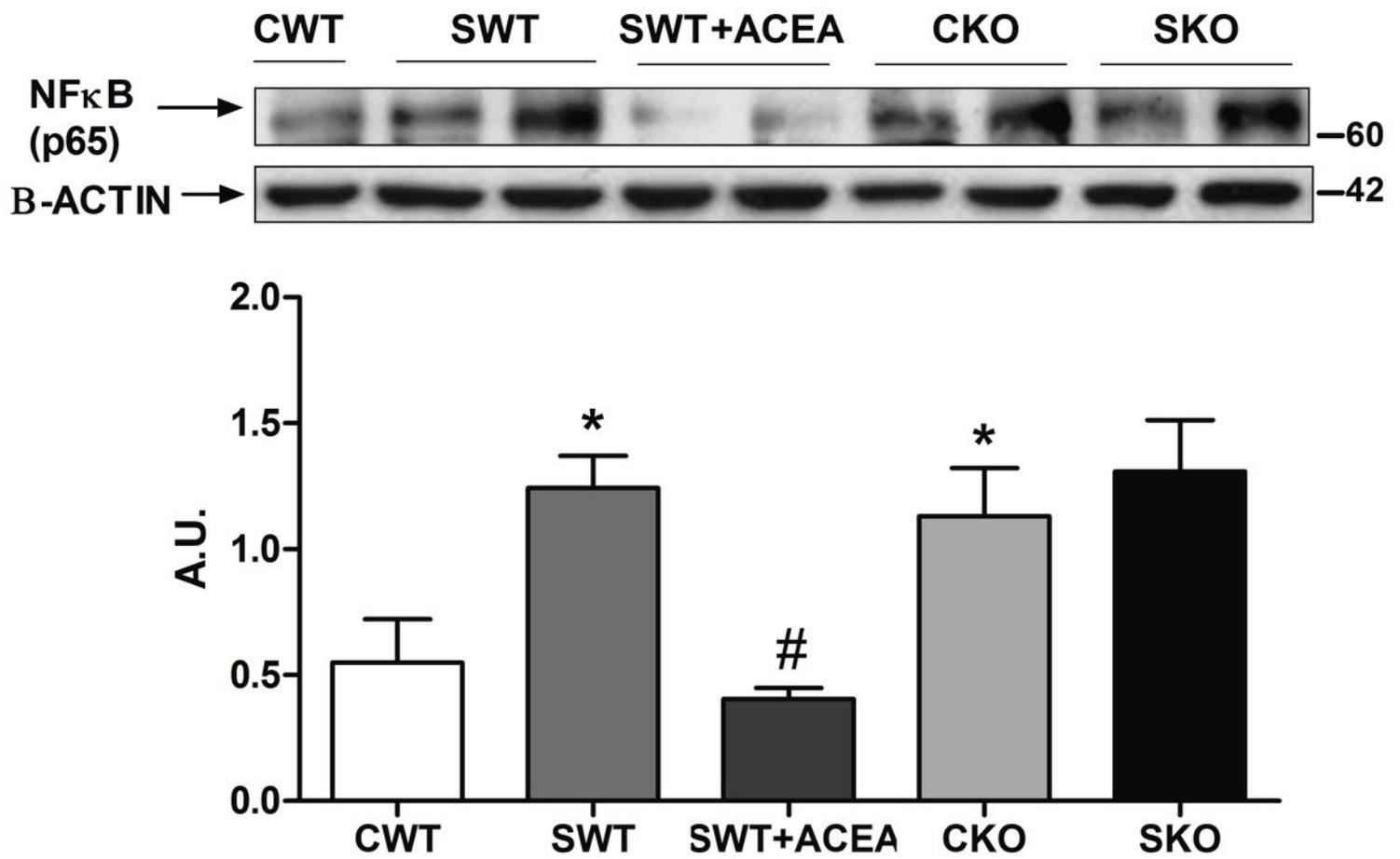
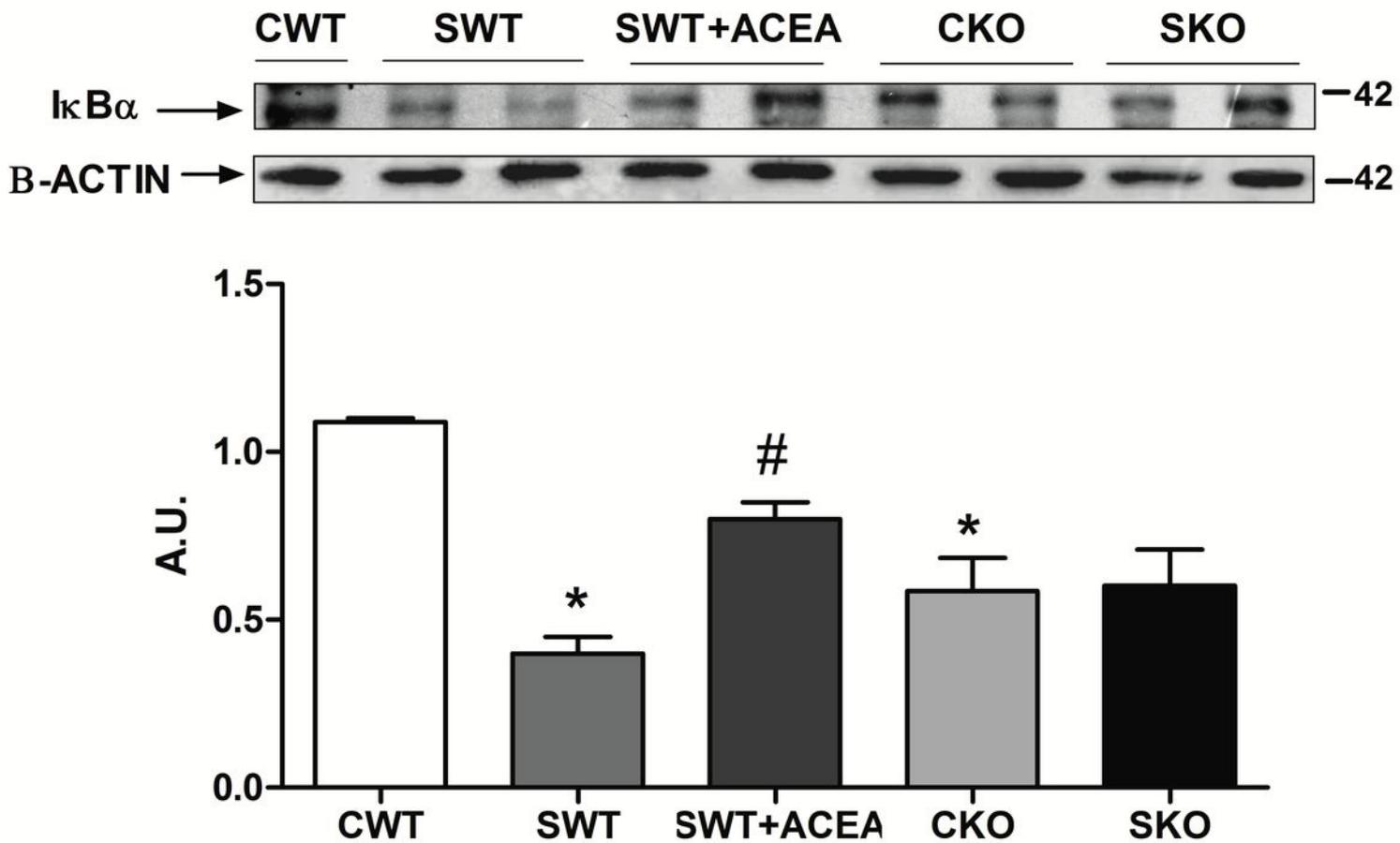
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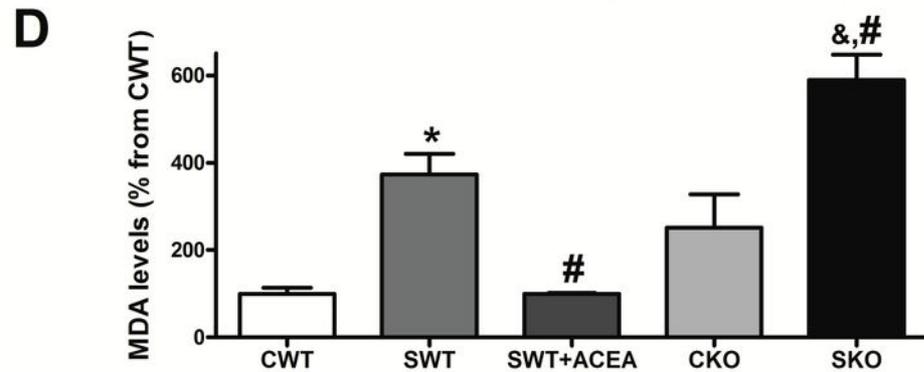
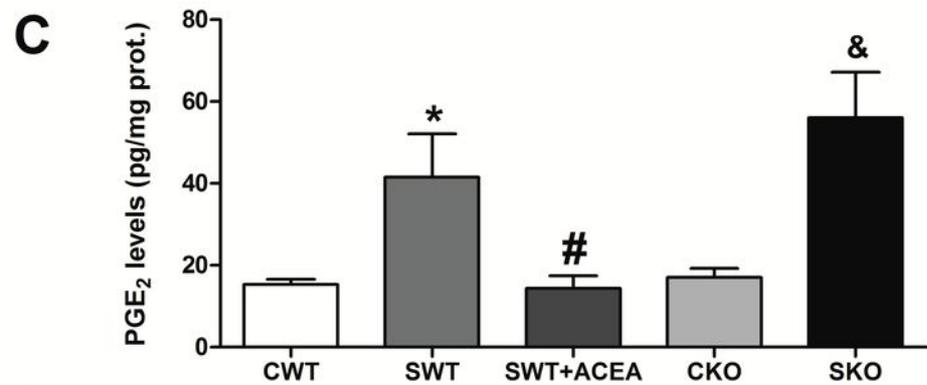
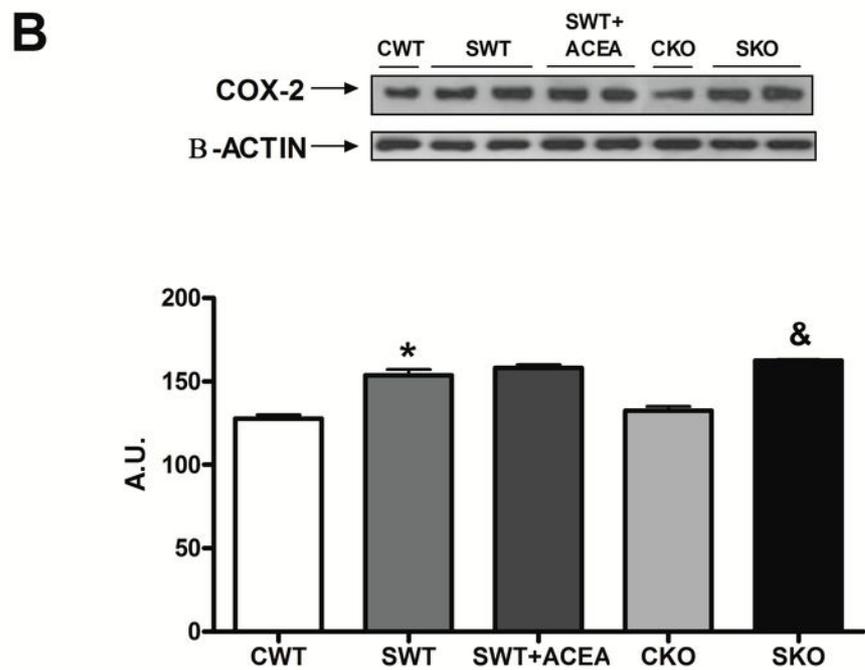
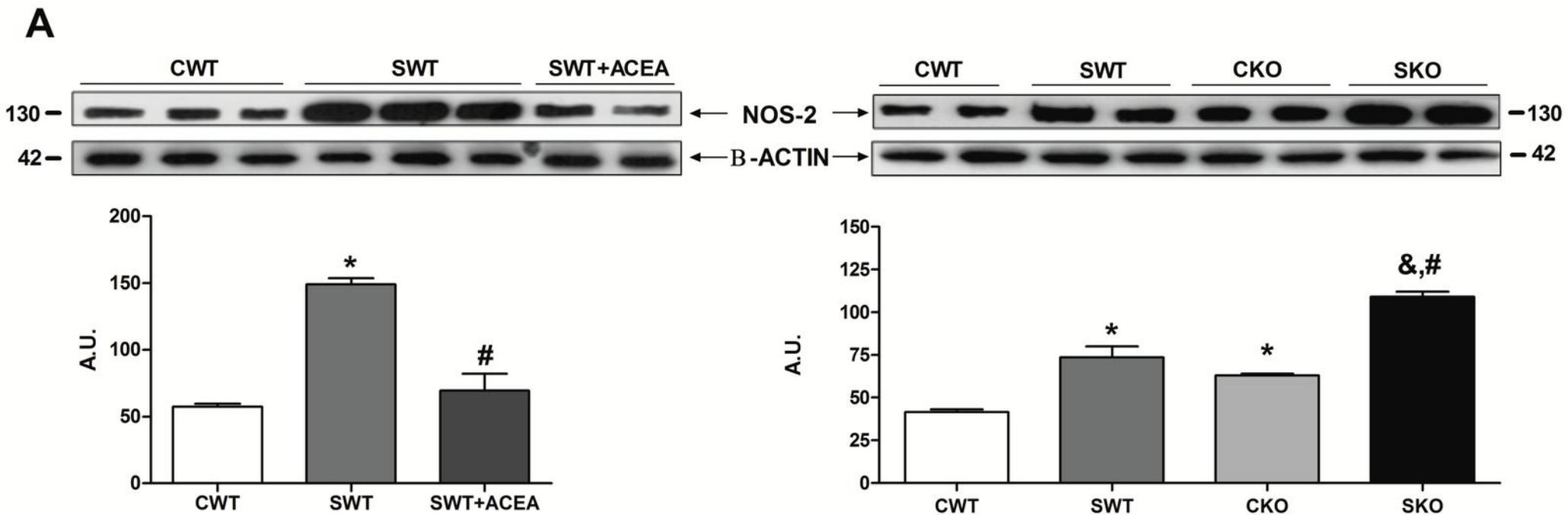
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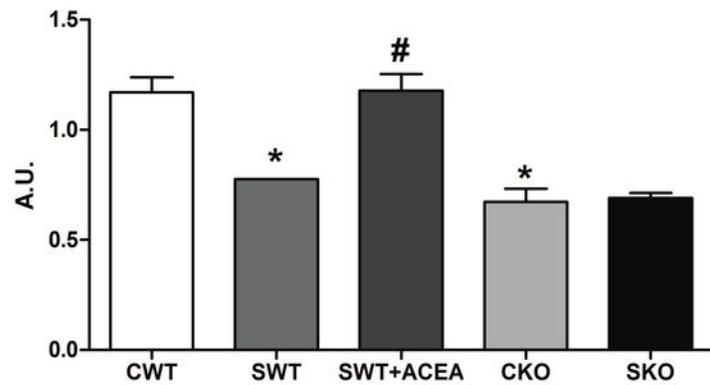
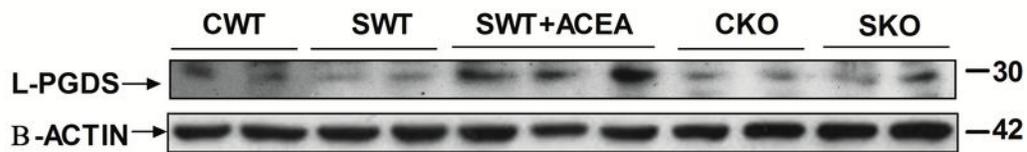
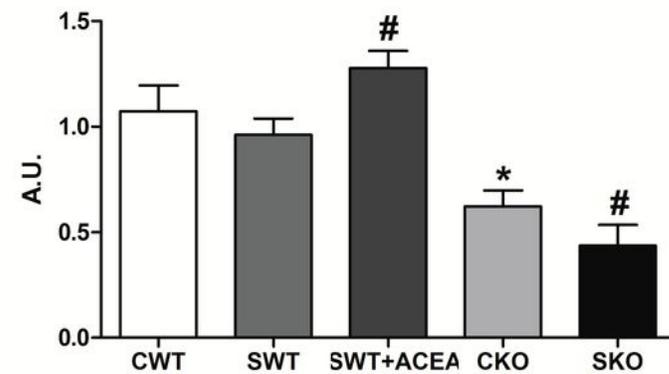
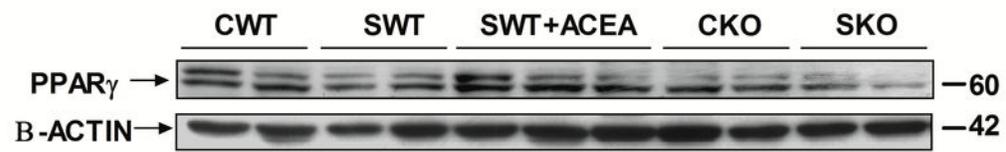
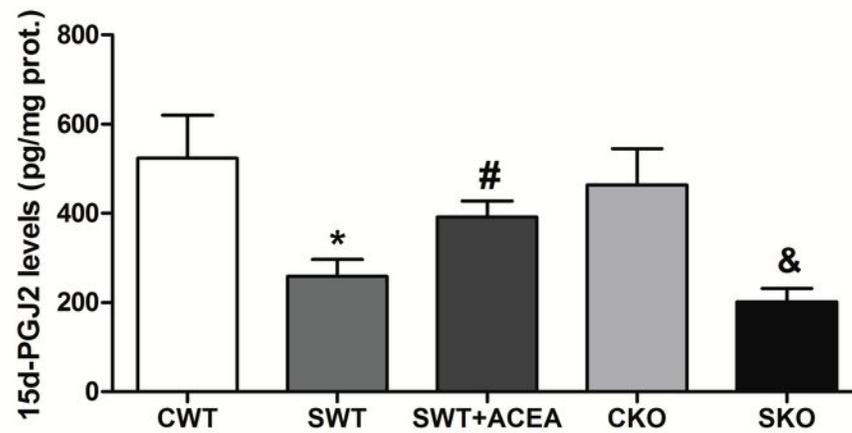
Plasma Corticosterone (ng/mL)

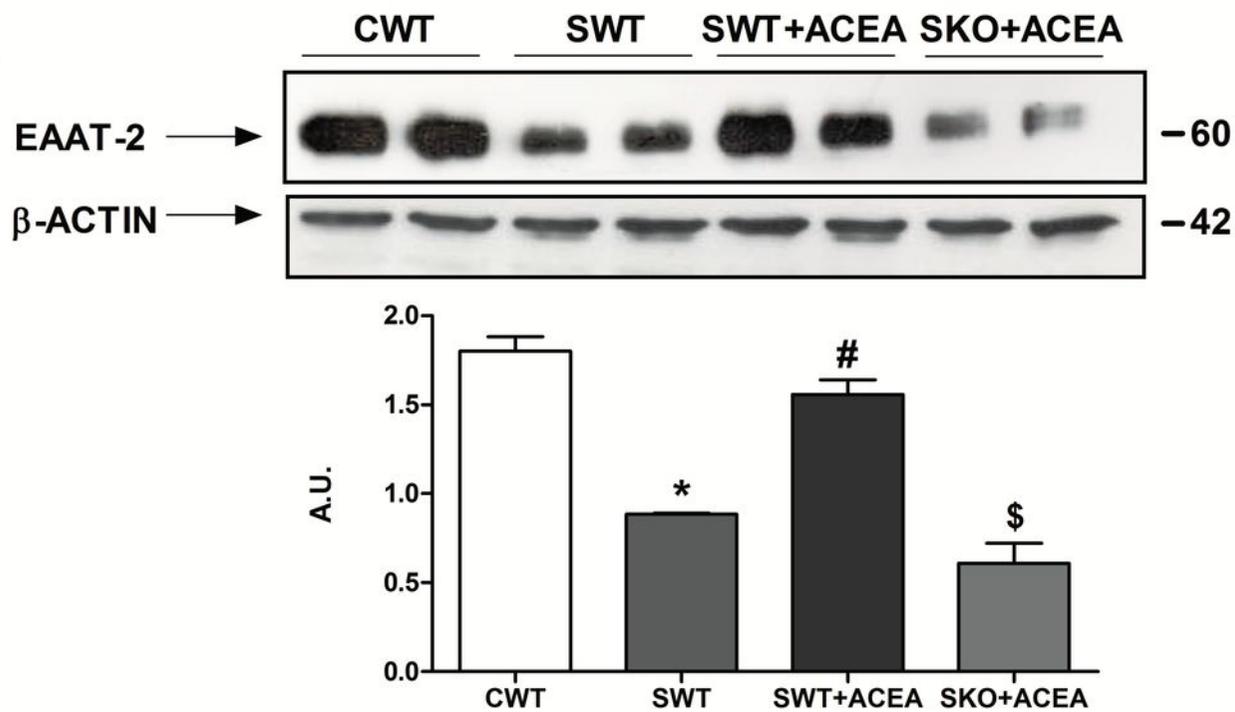
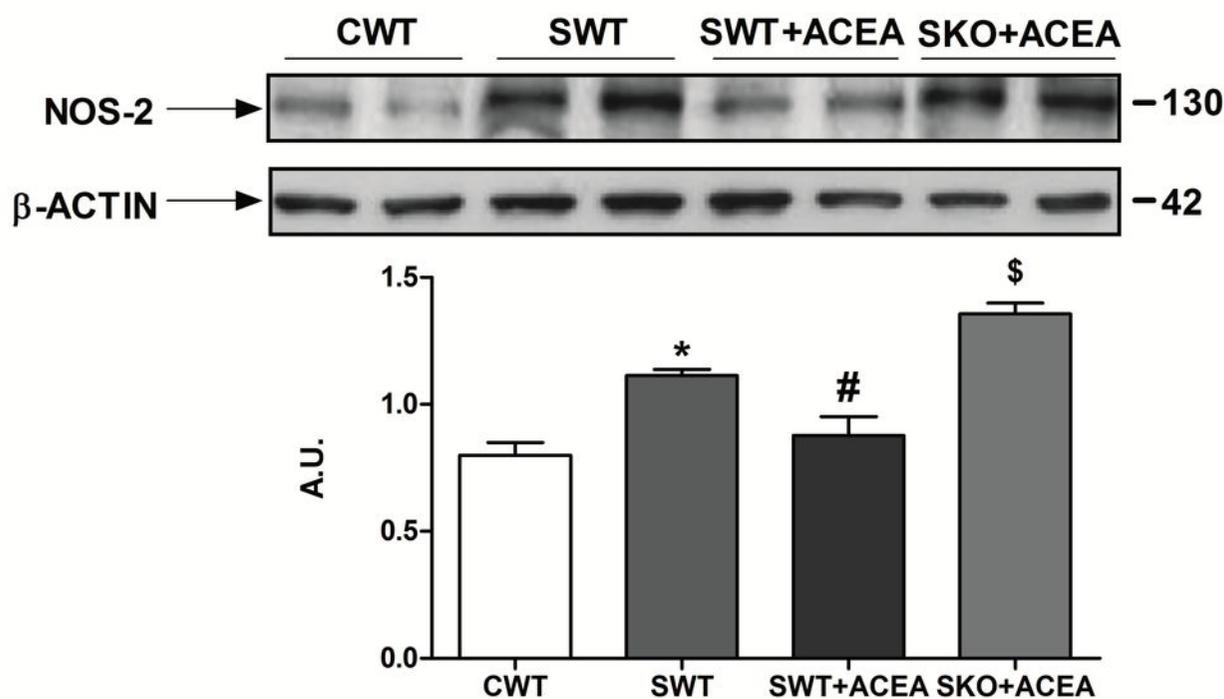




A**B**



A**C****B**

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