

## Serological, Pathological and Polymerase chain reaction studies on mycoplasma hyopneumoniae infection in the wild boar

M. Sibila, G. Mentaberre, M. Boadella, E. Huerta, E. Casas -Díaz, J. Vicente, C. Gortázar, I. Marco, S. Lavín, J Segalés

### ▶ To cite this version:

M. Sibila, G. Mentaberre, M. Boadella, E. Huerta, E. Casas -Díaz, et al.. Serological, Pathological and Polymerase chain reaction studies on mycoplasma hypopneumoniae infection in the wild boar. Veterinary Microbiology, 2010, 144 (1-2), pp.214. 10.1016/j.vetmic.2009.12.019. hal-00597834

### HAL Id: hal-00597834 https://hal.science/hal-00597834

Submitted on 2 Jun2011

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

### Accepted Manuscript

Title: Serological, Pathological and Polymerase chain reaction studies on mycoplasma hyopneumoniae infection in the wild boar

Authors: M. Sibila, G. Mentaberre, M. Boadella, E. Huerta, E. Casas -Díaz, J. Vicente, C. Gortázar, I. Marco, S. Lavín, J Segalés



PII:	S0378-1135(09)00610-5
DOI:	doi:10.1016/j.vetmic.2009.12.019
Reference:	VETMIC 4716
To appear in:	VETMIC
Received date:	22-9-2009
Revised date:	9-12-2009
Accepted date:	11-12-2009

Please cite this article as: Sibila, M., Mentaberre, G., Boadella, M., Huerta, E., -Díaz, E.C., Vicente, J., Gortázar, C., Marco, I., Lavín, S., Segalés, J., Serological, Pathological and Polymerase chain reaction studies on mycoplasma hyopneumoniae infection in the wild boar, *Veterinary Microbiology* (2008), doi:10.1016/j.vetmic.2009.12.019

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1	SHORT COMMUNICATION
2	
3	SEROLOGICAL, PATHOLOGICAL AND POLYMERASE CHAIN REACTION STUDIES ON
4	MYCOPLASMA HYOPNEUMONIAE INFECTION IN THE WILD BOAR
5	
6	M. Sibila <sup>1</sup> , G. Mentaberre <sup>2</sup> , M. Boadella <sup>3</sup> , E. Huerta <sup>1</sup> , E. Casas -Díaz <sup>2</sup> , J. Vicente <sup>3</sup> , C. Gortázar <sup>3</sup> ,
7	I. Marco <sup>2</sup> , S. Lavín <sup>2</sup> , J Segalés <sup>1,4</sup>
8	
9	<sup>1</sup> Centre de Recerca en Sanitat Animal (CReSA), UAB-IRTA, Campus de la Universitat Autònoma de
10	Barcelona, 08193 Bellaterra, Barcelona, Spain. <sup>2</sup> Servei d'Ecopatologia de Fauna Salvatge,
11	Departament de Medicina i Cirurgia Animals, Universitat Autònoma de Barcelona, 08193
12	Bellaterra, Barcelona, Spain. <sup>-3</sup> Instituto de investigación en Recursos Cinegéticos, IREC (CSIC-
13	UCLM-JCCM), Ciudad Real, Spain. <sup>4</sup> Departamente de Sanitat i d'Anatomia Animals, Facultat de
14	Veterinària, Universitat Autònoma de Barcelona, 08193 Bellaterra, Barcelona, Spain
15	Email: marina.sibila@cresa.uab.cat
16	
17	
18	
19	
20	
21	* Corresponding author at: Centre de Recerca en Sanitat Animlal (CReSA), UAB-IRTA, Campus de la
22	Universitat Autònoma de Barcelona, 08193 Bellaterra, Barcelona, Spain.
23	E-mail address: marina.sibila@cresa.uab.cat (M. Sibila)
24	
25	

#### 26 Abstract

27 There are evidences that wild boar and domestic pig populations share the vulnerability to certain 28 pathogens and, in consequence, the risk of pathogen transmission. Based on serological evidences, it is 29 known that Mycoplasma hyopneumoniae (Mhyo) is able to infect the wild boar. However, if this 30 infection causes lesions and disease in this species is still unknown. The objectives of the present study 31 were to assess the seroprevalence to Mhyo, as well as the rates of detection of Mhyo (by nested 32 polymerase chain reaction, nPCR) in upper and lower respiratory tract of the wild boar. Moreover, 33 presence of enzootic pneumonia (EP)-like gross and microscopic lesions in these animals was also 34 assessed. Antibodies against Mhyo were detected in 92 out of 428 (21%) serum samples tested. 35 Moreover, Mhyo DNA was detected by nPCR in 17 out of 85 (20%) nasal swabs and in 12 out of 156 36 (8%) lung samples. No gross EP-like lesions were observed in any of the studied pigs. Besides, presence 37 of EP-like microscopic lung lesions was observed in 18 out of 63 (29%) animals. Mhyo DNA was 38 detected at lung level in 2 out of these 18 (11%) animals. Presence of EP-like microscopic lesions was 39 significantly related (p<0.05) with Mhyo detection at nasal swab swab but not (p>0.05) with its 40 detection in lung samples; such lesions were also linked (p<0.05) with presence of Metastrongylus spp-41 like parasite structures. Results of the present study confirm that Mhyo is able to infect and might be 42 able to cause EP-like microscopic lesions in wild boar but, apparently, with a subclinical impact.

- 43
- 44 Key words:
- 45 *Mycoplasma hyopneumoniae*, wild boar, seroprevalence, infection, enzootic pneumonia
- 46
- 47

48

#### 49 **1. Introduction**

50 Populations of the Eurasian wild boar (Sus scrofa) have been expanding throughout Europe 51 during last decades (Acevedo et al., 2006). Consequently, probabilities of interaction between wild boar 52 and domestic pigs as well as pathogen transmission are also increased. Therefore, the study of the 53 prevalence of pathogens in wild boar populations and the role of these animals as pathogen reservoirs is 54 of importance for public health and pig producers (Ruiz-Fons et al., 2008; Wyckoff et al., 2009). There 55 are evidences that domestic and wild boar share the vulnerability to certain viral and bacterial pathogens (Ruiz-Fons et al., 2008; Wyckoff et al., 2009). However, little information on the prevalence and impact 56 57 of Mycoplasma hyopneumoniae (Mhyo), one of the most economically important pathogens for the pig 58 industry, is available for the wild boar.

59 Mhyo is the etiologic agent of enzootic pneumonia (EP), a chronic respiratory disease with high 60 morbidity and low mortality. The most significant clinical sign of EP is a non-productive cough 61 (particularly in finishing pigs) and the main gross associated lesion is a craneo-ventral pulmonary 62 consolidation (Caswell and Williams, 2007). Histologically, EP-like lesions are characterized by a 63 broncho-interstitial pneumonia with broncho-alveolar lymphoid tissue (BALT) hyperplasia (Caswell and 64 Williams, 2007).

Up to now, evidences of infection (Marois et al., 2006) and detection of antibodies against Mhyo (Marois et al., 2006, Vengust et al., 2006, Shcherbakov et al., 2007) have been reported in different countries. However, it is unknown if this pathogen is able to cause EP-like lesions to the wild boar. Therefore, the aims of this study were firstly to gain insight into Mhyo seroprevalence, infection and excretion in the Spanish wild boar population and, secondly, to assess the potential presence of EP-like lesions in these animals.

#### 72 **2. Materials and methods**

#### 73 **2.1.** Animals and samples

74 Number of samples tested and techniques performed are summarised in table 1. A total of 428 75 wild boar serum samples (males, n=185; females n=231; non-recorded gender, n=12) were included in 76 this study. Tested sera were obtained from piglets (<7 months of age, n=3), weaners (7-12 months, 77 n=95), juveniles (12-24 months, n=87) and adult wild boars (>24 month, n=234). Age was unknown for 78 9 animals. Age classification was based on tooth eruption patterns and weight (Sáenz De Buruaga et al., 79 1991). From these 428 samples, 225 sera were collected during the hunting season between years 2000 80 and 2008 in the South-Central part of Spain. Samples from this region were collected from hunter-81 harvested wild boar reared in high-wire fenced hunting estates. Rest of the samples (n=203) were 82 collected from wild boars in North-Eastern Spain between 2004 and 2005. Samples from this region 83 were collected from hunted free-living wild boars in national parks. Fresh lung samples were also taken 84 from 156 (102 from South-Central and 54 from North-Eastern Spain) out of these 428 wild boars, and 85 used to assess Mhyo infection at the lower respiratory tract. Moreover, different portions of lung were 86 taken from 63 (all of them from South-Central Spain) out of these 156 animals, and fixed by immersion 87 in formalin to perform histopathological studies. None of these 63 animals showed gross lesions 88 attributable to Mhyo (pulmonary cranio-ventral consolidation). Finally, presence of this pathogen in the 89 upper respiratory tract (nasal swabs) was also assessed in 85 (all of them from South-Central Spain) out 90 of these 156 animals.

91

### 92 **2.2.** Serology

93 Sera were tested by a monoclonal blocking ELISA (Laboratorios HIPRA<sup>®</sup>, Girona, Spain) to 94 detect Mhyo antibodies. The inhibition percentage (IP) was calculated taking into account the optical 95 densities (OD) of each sample as well as the one of the negative control (negative value). ELISA result

96 interpretation, based on the percentage of inhibition (IP) values, was as follows: IP < 50%, negative and 97 IP > 50%, positive.

- 98
- 99

#### 2.3. DNA extraction and PCR methods

100 Nasal swabs were suspended in 1 ml of sterile PBS and vigorously vortexed. Nasal swab 101 suspensions and fresh lung tissue were stored at -80°C until further processing. DNA was extracted using 300 µl of nasal swab suspensions (Nucleospin<sup>®</sup> Blood, Macherey-Nagel GmbH & Co KG Düren, 102 Germany) and from 25 mg (approximately) of fresh lung sample (Nucleospin<sup>®</sup> Tissue, Tissue Macherey-103 104 Nagel GmbH & Co KG Düren, Germany), according to manufacturer's instructions. To assess for 105 potential contaminations during the extraction procedure, a negative control was included using PBS as 106 extraction substrate in each group of extracted samples. Presence of Mhyo DNA was studied using a 107 previously described nested PCR (nPCR) (Calsamiglia et al., 1999). In each nPCR reaction, a negative 108 control (consisting of millipore water instead of extracted DNA) was added every four test samples. 109 DNA extracted from a pure culture of Mhyo was used as positive control and included in the last 110 position of the gel.

111

#### 112 2.4. Histopathology

113 Since no gross EP-like lesions were evident, portions of the cranial part of the apical, medium 114 and diaphragmatic lobes were collected and fixed by immersion in 10% buffered formalin. Lung 115 samples were processed routinely for histopathology (haematoxylin&eosin stain). Presence of 116 microscopic lung lesions were graded from 0 to 4, following previously described criteria (Calsamiglia 117 et al., 2000). Lesions scoring 3 and 4 were considered compatible with EP. Presence (or absence) of 118 *Metastrongylus spp-*like parasites (eggs, larvaes and/or adult individuals) (Caswell and Williams, 2007) 119 in lungs sections was recorded.

#### 121 **2.5.** Statistical analyses

122 Bivariate analyses using contingency tables (Chi-square statistics) were used to compare: 1) 123 serology, Mhyo DNA detection by nPCR (nasal swab and lung tissue) and presence of EP-like 124 microscopic lesions (histopathological scores 3 or 4) taking into account origin, sex and age of the 125 animals, 2) serology versus nPCR detection (nasal swab and lung tissue) and EP-like microscopic 126 lesions, and 3) EP-like microscopic lesions versus nPCR detection (nasal swab and lung tissue), and 4) 127 presence of *Metastrongylus spp-like parasites* against the rest of studied parameters. To analyze the 128 relationship between serology and nPCR results versus lung lesions, animals were divided in non-EP-129 like (histopathological score 0 to 2) or EP-like (scoring 3 or 4) microscopic lung lesions. Relationship 130 between nPCR detection at nasal swabs and lung samples was assessed by means of the McNemar test 131 and the Kappa value. Statistical analyses were performed with the SAS system for Windows version 9.0 (SAS Institute Inc, Cary, North Carolina, USA). Statistical significance level was set at  $\alpha$ =0.05. 132

133

#### 134 **3. Results**

Number of tested samples per technique and number of samples that yielded positive results aredetailed in table 1.

A total of 428 serum samples were analyzed by ELISA. Ninety two out of the 428 (21.5%) sera were Mhyo seropositive. No statistically significant differences (p>0.05) between number of Mhyo seropositive from North-Eastern (36 out of 203, 18%) and South-Central (56 out of 225, 25%) parts of Spain were found. The highest number of Mhyo seropositive animals were observed in the adult population (n=60 [25%], p<0.05) (table 2). No statistically significant differences (p>0.05) between numbers of Mhyo seropositive males (41%) and females (53%) were observed (table 3).

Mhyo DNA was detected by nPCR in 12 (8%) out of the 156 analyzed lung samples. No statistically significant differences (p>0.05) between number of nPCR positive lung samples from animals hunted in North-Eastern (4 out of 54, 7%) and South-Central (8 out of 102, 8%) parts of Spain

were found. Number of Mhyo seropositive animals with an nPCR positive lung tissue (n=10 [83%]) was significantly higher (p<0.05) than the number of seronegative ones (n=2 [17%]). No statistically significant differences in the number of nPCR positive lung samples regarding age and gender of tested animals were found.

From the 85 animals from which nasal swab and lung sample were available, Mhyo was detected by nPCR in lung only in 2 (2%) animals, in nasal swab only in 13 (15%) animals, and in both samples in 4 (5%) animals (McNemar test p<0.004, Kappa= 0.27).

Number of Mhyo seropositive animals with an nPCR positive nasal swab (n=11 [65%]) was significantly higher (p<0.05) than the number of seronegative ones (n=6 [35%]). No statistically significant differences in the number of nPCR positive nasal swabs regarding age (table 2) and gender (table 3) were found among tested animals.

Eighteen out of the 63 (29%) lung samples evaluated by histopathology showed EP-like 157 158 microscopic lesions (10 lungs scored 3 and 8 scored 4). Nasal swab was available in 15 out of these 18 159 animals showing EP-compatible lung lesions; Mhyo DNA was detected in 9 out of these 15 (60%) nasal 160 swabs (p<0.05). On the other hand, Mhyo was detected at lung level only in 2 out of the 18 (11%) 161 animals showing EP-like microscopic lesions. Mhyo was detected in nasal swab and lung at the same 162 time in these 2 animals. No statistically significant differences between seropositive (6 out of 18) and 163 seronegative (12 out of 18) animals showing EP-like microscopic lesions (p>0.05) were found. Also, no 164 statistically significant differences in the number of animals showing EP-like microscopic lesions 165 regarding age and gender of the tested animals were found.

Presence of *Metastrongylus* spp.-like parasite structures were observed in 6 (33.3%) out of these 18 animals showing EP-compatible lung lesions (p<0.05). On the contrary, parasites were observed in only 2 (4%) out of the 45 animals that did not show these lesions. All these 8 wild boar harbouring parasites were negative by Mhyo nPCR at lung level and only 2 (28.5%, p>0.05) were positive by nPCR in nasal swabs (available in 7 out of these 8 animals). No statistically significant differences in the

number of animals showing presence of *Metastrongylus* spp.-like parasite structures regarding age and
gender of the tested animals were found.

From the 54 animals from which all the samples were available, one (2%) wild boar was Mhyo seropositive, nPCR positive (at nasal as well as at lung levels) and had EP-like microscopic lesions with no evidence of parasitosis.

176

#### 177 **4. Discussion**

The present study gives further light on the role of Mhyo infection in the wild boar. Presence of 178 179 antibodies against this pathogen, together with the detection of its DNA and presence of 180 histopathological lesions compatible with EP suggest that, besides the infection, Mhyo is capable of 181 causing broncho-interstitial pneumonia in wild boar. However, lack of consistent gross lesions 182 compatible with EP in studied animals points out that the effect of Mhyo probably remains subclinical in 183 this species. Taking into account that a number of wild boar had the pathogen in the nose, its potential 184 role as a reservoir for the domestic pig can not be ruled out. Although wild boars specifically included in 185 this study came from areas in which the contact with domestic pigs was unlikely, the putative 186 transmission between wild boars and domestic pigs should not be considered negligible and an especial 187 risk for Mhyo seronegative farms. These type of farms tend to be high health herds, usually isolated from high density pig rearing areas (Muirhead and Alexander, 1997) and, therefore, close to areas where 188 189 wild boar live.

Results of this study demonstrated the presence of Mhyo seropositive animals in more than 400 wild boars from different Spanish regions. Prevalence of Mhyo seropositive pigs obtained in this study (21%) was similar to the one obtained in another serologic survey conducted in Slovenia (21%) (Vengust et al., 2006), but lower to the one observed in France (58%) (Marois et al., 2006). On the contrary, these results are in disagreement with a previous epidemiologic study conducted in South-Central Spain, in which 78 wild boar (hunted between 1998 and 2000) were seronegative against this

respiratory pathogen (Vicente et al., 2002). Differences in geographical origin, year of collection as well as number of samples could explain these discrepancies. However, the fact that the tested samples were collected in two different regions in different years makes difficult to assess potential differences in seroprevalence and rate of infection between the two areas studied.

Detection of Mhyo infection in wild boar was already described by Marois et al. (Marois et al., 200 201 2006), who detected 15 out of 171 lung exudates positive by PCR. However, detection of Mhyo DNA in 202 that study was not successfully confirmed by culture and presence of lung lesions was not assessed. In 203 the present study, infectiousness of Mhyo PCR positive material was not tested; therefore, like in Marois 204 et al. (2006), the achievement of a Mhyo isolate from wild boar is still lacking. However, the present 205 study represents the first description of Mhyo infection in wild boar at different levels of the respiratory 206 tract (upper and lower levels) in animals showing microscopic lung lesions compatible with EP. It 207 should be taken into account, however, that other agents such as viruses (SIV) or parasites 208 (Metastrongylus spp.) might also cause broncho-interstitial pneumonia with BALT hyperplasia (Caswell 209 and Williams, 2007). In fact, some animals had lesions probably attributed to Metastrongylus spp 210 infestation, since parasite structures were observed by histopathology in Mhyo seronegative and PCR 211 negative hosts. Moreover, infestation was related to the presence of broncho-interstitial pneumonia. It 212 can not be ruled out, therefore, that EP-like microscopic lesions in wild boar with no evidence of 213 parasitation were indeed infested with *Metastrongylus* spp. On the other hand, it is worthy to remark the existence of one Mhyo seropositive animal infected with this pathogen at upper and lower respiratory 214 215 tract and with EP-compatible lung lesions and no evidence of parasitation.

In the present study, Mhyo detection in nasal swabs was linked to the presence of EP-like microscopic lesions and to nPCR detection in lung samples, similarly to what has been found for the domestic swine at slaughter (Sibila et al., 2004a). On the contrary, detection of Mhyo DNA by nPCR in lung tissue was not linked to the presence of EP-like microscopic lesions. This result would be in

220 disagreement with previously published works in domestic pigs, in which Mhyo was detected in lung 221 tissues in a high percentage of cases with moderate to severe lung lesions (Cai et al., 2007; Moorkamp et 222 al., 2007). The low detection rate of Mhyo in lung tissues might be attributable to several lung lesions 223 observed were not caused by Mhyo and, therefore, this pathogen was not located in lungs, even it was 224 present in the wild boar environment (it was detected in the nose). Secondly, different portions of tissue 225 were tested by nPCR and histopathology. In fact, none of the 63 animals from which lung samples were 226 available showed gross lung lesions and, thus, the portion of tissue processed by nPCR was taken 227 without any reference of lesion. Finally, presence of EP-like microscopic lesions may depend, among 228 other factors, on the infectious pressure (Sibila et al., 2009). Since wild boar tend to live in open-air 229 systems; therefore, the infectious pressure must be considered lower than the one found in intensively 230 reared pig farms and may be not sufficient to cause typical EP-like gross lesions. On the other hand, 231 absence of macroscopic lesions attributable to Mhyo in domestic pigs showing EP-like microscopic 232 lesions has been already described elsewhere (Sibila et al., 2007) and may indicate a chronic stage of 233 Mhyo infection, where gross lesions had healed but microscopic lesions remained.

In summary, results from this study further expand the existing knowledge on Mhyo infection in the wild boar population, pointing out that Mhyo might be able to cause EP-like microscopic lesions in wild boar, even they seem to have a subclinical impact. Based on the relatively high prevalence of Mhyo in conventional pig farms (Sibila et al., 2004b), it should not be ruled out the fact that domestic pig can be the real reservoir for the wild boar. The wild boar may only represent a potential Mhyo threaten for Mhyo free farms.

240

#### 241 Acknowledgements

The authors are grateful to Christian Font and Mónica Pérez for their help in processing the samples for histopathology.

#### 245 **References**

- Acevedo P., Escudero M. A., Muñoz R., C., G., 2006, Factors affecting wild boar abundance across an
   environmental gradient in Spain. Acta Theriologica 51 327-336.
- Cai, H.Y., van Dreumel, T., McEwen, B., Hornby, G., Bell-Rogers, P., McRaild, P., Josephson, G.,
   Maxie, G., 2007, Application and field validation of a PCR assay for the detection of
   *Mycoplasma hyopneumoniae* from swine lung tissue samples. Journal of Veterinary Diagnostic
- 251 Investigation 19, 91-95.
- Calsamiglia, M., Collins, J.E., Pijoan, C., 2000, Correlation between the presence of enzootic pneumonia
   lesions and detection of *Mycoplasma hyopneumoniae* in bronchial swabs by PCR. Vet Microbiol
   76, 299-303.
- Calsamiglia, M., Pijoan, C., Trigo, A., 1999, Application of a nested polymerase chain reaction assay to
   detect *Mycoplasma hyopneumoniae* from nasal swabs. Journal of Swine Health and Production 7,
   263-268.
- Caswell, J.L., Williams, K.J., 2007, Respiratory System, In: Maxie, M.G. (Ed.) Pathology of Domestic
   Animals.Edinburg, pp. 523-655.
- Marois, C., Tocqueville, V., Le Potier, M.F., Hars, J., Kobisch, M., 2006. Detection of *Mycoplasma hyopneumoniae* in French Wild Boars. In: 19th International Pig Veterinary Society,
   Copenghaguen, Denmark, p. 213.
- Moorkamp, L., Nathues, H., Spergser, J., Tegeler, R., Beilage, E.G., 2007, Detection of respiratory
   pathogens in porcine lung tissue and lavage fluid. Veterinary Journal, doi:
   10.1016/j.tvjl.2007.1001.1008.
- Muirhead, M.R., Alexander, T.J.L., 1997, Managing and treating disease in the weaner, grower and
   finishing periods, In: Alexander, T.J.L. (Ed.) Managing pig health and the treatment of disease.
   5M Enterprises LTd, Sheffield, p. 309.
- Ruiz-Fons, F., Segales, J., Gortazar, C., 2008, A review of viral diseases of the European wild boar:
   effects of population dynamics and reservoir role. Vet J 176, 158-169.
- Sáenz De Buruaga M., Lucio A.J., Purroy J. 1991. Reconocimiento de sexo y edad en especies
   cinegéticas, Diputación Foral de Alava, V., ed.
- Sibila, M., Calsamiglia, M., Segalés, J., Rosell, C., 2004a, Association between *Mycoplasma hyopneumoniae* at different respiratory sites and presence of histopathological lung lesions.
   Veterinary Record 155, 57-58.
- Sibila, M., Calsamiglia, M., Vidal, D., Badiella, L., Aldaz, A., Jensen, J.C., 2004b, Dynamics of
   *Mycoplasma hyopneumoniae* infection in 12 farms with different production systems. Canadian
   Journal of Veterinary Research 68, 12-18.
- Sibila, M., Nofrarias, M., Lopez-Soria, S., Segales, J., Valero, O., Espinal, A., Calsamiglia, M., 2007,
   Chronological study of *Mycoplasma hyopneumoniae* infection, seroconversion and associated
   lung lesions in vaccinated and non-vaccinated pigs. Veterinary Microbiology 122, 97-107.
- Sibila, M., Pieters, M., Molitor, T., Maes, D., Haesebrouck, F., Segales, J., 2009, Current perspectives
   on the diagnosis and epidemiology of Mycoplasma hyopneumoniae infection. Vet J 181, 221 231.
- Vengust, G., Valencak, Z., Bidovec, A., 2006, A serological survey of selected pathogens in wild boar in
   Slovenia. Journal of veterinary medicine. B, Infectious diseases and veterinary public health 53,
   24-27.
- Vicente, J., Leon-Vizcaino, L., Gortazar, C., Jose Cubero, M., Gonzalez, M., Martin-Atance, P., 2002,
   Antibodies to selected viral and bacterial pathogens in European wild boars from southcentral
   Spain. J Wildl Dis 38, 649-652.
- Wyckoff, A.C., Henke, S.E., Campbell, T.A., Hewitt, D.G., VerCauteren, K.C., 2009, Feral swine
   contact with domestic swine: a serologic survey and assessment of potential for disease
   transmission. J Wildl Dis 45, 422-429.

**Table 1:** Combination of the different techniques performed in the same animal (rows) according to the number of positive samples per technique (rows). nPCR NS (nPCR nasal swabs); nPCR L (nPCR Lung); Histo (Histopathology, animals scored 3 and 4).

			Number of positive samples (percentage) per technique							
		Number of		nl	PCR		Presence of Metastrongylus			
		tested animals	Serology	NS	L	Histo*	spplike structures			
	Serology	272	33 (12)							
I	Serology nPCR L	62	29 (47)		5 (8)	6				
Techniques performed in the same animal	Serology nPCR NS nPCR L	31	8 (26)	3 (10)	2 (6)	2				
lues performed	Serology nPCR L Histo	9	4 (33)	0	1 (11)	3 (33)	1 (11)			
Technic	Serology nPCR NS nPCR L Histo	54	18 (33)	14 (26)	4 (7)	15 (28)	8 (15)			
	Total	428	92 (21,5)	17 (20)	12 (8)	18 (29)	9 (14)			

Age groups         N         Positive           Weaners         95         17         17         5         29         5         10         3         10         2           Juveniles         87         11         20         2         28         0         12         6         12         7         2           Adults         234         60         47         10         9         0         7         2         7         2           Total         428         92         85         17         156         12         63		ELISA		nPCR NS		nPCR L		Histo (score 3 or 4)		Metastrongylus	
Weaners       95       17       17       5       29       5       10       3       10       2         Juveniles       87       11       20       2       28       0       12       6       12       34         Adults       234       60       47       10       90       7       34       7       34       7         Unknown       9       4       1       0       9       0       7       2       7       2         Total       428       92       85       17       156       12       63       18       63       8	_	Ν	Positive	Ν	Positive	N	Positive	N	Positive	N	Positive
Juveniles       87       11       20       2       28       0       12       6       12       34         Adults       234       60       47       10       90       7       34       7       34       7         Unknown       9       4       1       0       9       0       7       2       7       2         Total       428       92       85       17       156       12       63       18       63       8	Piglets	3	0	0	0	0	0	0	0	0	2
Adults       234       60       47       10       90       7       34       7       34       7         Unknown       9       4       1       0       9       0       7       2       7       2         Total       428       92       85       17       156       12       63       18       63       8	Weaners	95	17	17	5	29	5	10	3	10	2
Unknown       9       4       1       0       9       0       7       2       7       2         Total       428       92       85       17       156       12       63       18       63       8	Juveniles	87	11	20	2	28	0	12	6	12	34
Total         428         92         85         17         156         12         63         18         63         8	Adults	234	60	47	10	90	7	34	7	34	7
	Unknown	9	4	1	0	9	0	7	2	7	2
	Total	428	92	85	17	156	12	63	18	63	8

# Table 2: Results of the different techniques in regards the age of the tested wild boar. nPCR NS (nPCR nasal swabs); nPCR L (nPCR Lung); Histo (Histopathology, animals scored 3 and 4).

# Table 3: Results of the different techniques by sex of the tested wild boar. nPCR NS (nPCR nasal swabs); nPCR L (nPCR Lung); Histo (Histopathology, animals scored 3 and 4).

	ELISA		nPCR NS		nPCR L		Histo (score 3 or 4)		Metastrongylus	
Gender	Ν	Positive	Ν	Positive	Ν	Positive	Ν	Positive	N	Positive
Male	185	38	32	6	64	8	28	8	28	2
Female	231	49	49	10	87	4	33	10	33	6
Unknown	12	5	4	1	5	0	2	0	2	0
Total	428	92	85	17	156	12	63	18	63	8