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Dorte Glintborg, Hanne Mumm, David Hougaard, Pernille Ravn, Marianne

Andersen

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Ethnic differences in Rotterdam criteria and metabolic risk factors in a multiethnic group of women with PCOS studied in Denmark

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Title page

Full title: Ethnic differences in Rotterdam criteria and metabolic risk factors in a multiethnic group of women with PCOS studied in Denmark

Short title: Ethnicity and PCOS

Authors:

Dorte Glintborg MD, PhD (1)

Hanne Mumm MD (1)

David Hougaard MD (3)

Pernille Ravn MD, Dr Med (2)

Marianne Andersen MD, PhD (1)

1. Department of Endocrinology and Metabolism, Odense University Hospital, 5000 Odense C, Denmark.

2. Department of Gynecology and Obstetrics, Odense University Hospital, 5000 Odense C, Denmark.

5000 Odense C, Denmark.

3. Statens Serum Institut, 2300 Copenhagen S, Denmark

Corresponding author: Dorte Glintborg, Kløvervænget 6, 3rd floor, 5000 Odense C. <u>d.glintborg@gmail.com</u> Fax: 00 45 66 11 33 71.

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Abstract

Objective: Clinical manifestations and metabolic risk factors may differ in ethnic subgroups of patients with PCOS.

Design: Retrospective trans-sectional study.

Patients: 1002 premenopausal women with the diagnoses hirsutism or PCOS were divided according to

ethnicity: Caucasian (CA, n=784), Middle East (ME, n=190), Asian (n=14), other (n=14).

Measurements: Clinical evaluation (hirsutism, BMI, waist, blood pressure), hormone analyses (testosterone, sex hormone binding globulin, prolactin, lipids, insulin, glucose), and transvaginal ultrasound. Oral glucose tolerance tests (OGTT) (n = 499) and ACTH tests (n = 434) were performed in a subgroup of patients. **Results (CA vs. ME women):** CA women were older [32(25-37) vs. 25(18-32) years, median (quartiles)] and had increased BMI compared to ME women. After correcting for age and BMI CA women were less hirsute, but had increased testosterone levels compared to ME women. The Rotterdam criteria were fulfilled in 56% of both populations but PCO was diagnosed in 47% CA vs. 29% ME women, p<0.01. CA women had increased blood pressure and smoked at a higher frequency (40 vs. 23%), whereas area under the curve for insulin during OGTT was decreased, all p<0.001. Prolactin levels were significantly lower in CA women compared to ME women [7(5-10) vs. 9(6-12) µg/l] and were inversely associated with smoking status. **Conclusion:** CA women had a more adverse cardiovascular profile than ME women, whereas insulin sensitivity was higher. The prevalence of the individual Rotterdam criteria differed significantly in the two study populations.

Introduction

Hirsutism affects 10% females in reproductive age and is caused by increased androgenicity in the pilosebacceus gland (1). Hirsutism is the result of high androgen concentrations often in combination with increased androgen sensitivity (2). Hirsutism is most often caused by polycystic ovary syndrome (PCOS) or idiopathic hirsutism (IH), whereas less than 5% hirsute patients are diagnosed with rare endocrine disorders (3). PCOS can be defined according to the Rotterdam criteria: 1. Irregular/no ovulations, 2. Clinical/biochemical hyperandrogenaemia, and 3. Polycystic ovaries (4). Two out of the three criteria need to be fulfilled and other causes of hyperandrogenaemia should be excluded. In contrast, the previously used PCOS criteria as suggested by the National Institute of Health (NIH) include only the first two criteria: Irregular/no ovulations and clinical/biochemical hyperandrogenaemia. The manifestations of PCOS may differ among different ethnic populations (5). It is recognized that Asian women are rarely hirsute (5). The enzyme 5α -reductase is responsible for the conversion of testosterone to the more potent and rogen dihydrotesterone (DHT) (1). Inherited variations in 5α -reductase activity may explain differences in the prevalence of hirsutism among diverse ethnic populations. Individual variations in dermal 5α -reductase activity may furthermore explain why hirsute patients often have near-normal testosterone levels and that testosterone levels are poorly associated with clinical hirsute manifestations (1). We are not aware of previous studies reporting differences of the individual diagnostic criteria between different ethnic populations with PCOS.

PCOS and IH is characterized by hyper-inflammation and is associated with insulin resistance and increased risk for cardiovascular disease and diabetes (6;7). It is therefore recommended that PCOS patients are screened for the metabolic syndrome and cardiovascular risk factors (6;7). The metabolic disturbances of PCOS seem to be more pronounced in hyperandrogen patients compared to patients with no hyperandrogenaemia (8). The prevalence of metabolic and cardiovascular risk factors may furthermore vary in different ethnic populations with PCOS. Several studies documented that immigrants in western countries may be at increased risk for life style diseases such as obesity and diabetes (9). Increased risk for diseases

may be attributable to genetic predisposition or to environmental influences such as different exercise or eating habits. It remains to be documented whether some ethnic PCOS populations are at increased risk for diabetes and therefore need closer follow up. In addition, some ethnic subgroups may be relatively protected against the long term side effects of PCOS.

Prolactin levels are routinely measured in hirsute patients to exclude prolactinomas as a cause of menstrual irregularity. Increased prolactin levels could be associated with irregular menses, but we are not aware of previous studies evaluating associations between prolactin and manifestations of PCOS in population studies.

The aim of the present study was to evaluate differences in clinical manifestations, metabolic risk factors, and diabetes risk in a multiethnic group of women with PCOS studied in Denmark



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Material

Inclusion criteria: All premenopausal women referred to the outpatient clinic at the Department of Endocrinology during 1997 –2008 with the referral diagnoses hirsutism and/or PCOS were included. The patients should fulfill the Rotterdam criteria for PCOS or have clinical/biochemical hyperandrogenemia. As part of the clinical practice protocol of the department patients suspended oral contraceptives for at least three months prior to evaluation and metformin treatment was paused for at least one month prior to evaluation. Seventeen patients in the study population were on antihypertensive treatment. All included patients had normal thyroid function. All presented data are pretreatment values.

Exclusion criteria: Women younger than 15 years were referred to the department of Pediatrics at Odense University Hospital and patients with a primary complaint of infertility were referred to the local Fertility Clinic. Patients not attending for the measurement of hormonal or metabolic parameters were excluded. Patients with serious endocrine diseases were excluded as previously described (3).

The study was designed as a retrospective observational audit and we did therefore not search for approval from the local Ethics Committee.

Methods

Routine evaluation included medical history, clinical examination, transvaginal ultrasound (US), and fasting blood samples.

Medical history: Cycle history was recorded in all patients using a standard scheme prepared for hirsute patients. Ancestry was recorded by using an evaluation scheme where the doctor marked the patient as originating from Caucasia, Middle East, Africa or Asia and the country of origin was specified. In few cases the originating country was not specified in the patient reports (Table 1). Only patients where both parents originated from the same continent were included in the study population. All participating patients were first or second generation immigrants. Marital status was not routinely recorded.

Clinical evaluation: Hirsutism was assessed by the Ferriman Gallwey score (10). Waist circumference was measured to the nearest cm in a standing position midway between the lower costal margin and the iliac crest. The BMI was calculated as the weight in kilograms divided by the square height in meters.

Transvaginal US were performed at the Department of Gynecology, Odense University Hospital and polycystic ovaries were defined according to Adams (11). Ovarian volume was not routinely measured in the present study. All patients were offered transvaginal US but a small group of patients did not attend for the examination. Transvaginal US was performed in 840/974 patients.

Blood samples: Fasting blood samples was drawn during the morning in follicular phase (cycle days 2-8) in patients with a cycle length shorter than three months, patients with cycle length > 3 months had the blood samples drawn on a random cycle day. Blood tests included androgens (total testosterone, free-testosterone, sex hormone binding globulin (SHBG), dehydroepiandrosterone sulfate (DHEAS), 17-hydroxyprogesterone (170HP)), LH, FSH, prolactin, and lipid profile.

During 1997-2002 patients routinely underwent OGTT and ACTH tests. These data on Caucasian patients have previously been published (3;12). During 2002- 2009 OGTT were only performed in high risk patients as recommended by international guidelines (13). Similarly, ACTH tests were only performed in patients with increased baseline 17OHP levels as previously discussed (12).

The 0.25 mg ACTH test (Synachten 0.25 mg/ml, Novartis Healthcare, Copenhagen, Denmark) was performed between 08.00 h and 10.00 h during follicular phase. Cortisol and 17OHP were measured at baseline and at 30 and 60 min. ACTH tests were performed in 434 individuals (1997-2002, n=416). OGTT was performed at 08.00 on a random day of the cycle. Insulin, C-peptide, and capillary blood glucose (CBG) were measured at baseline and at 30, 60, and 120 minutes after oral ingestion of 75 g glucose dissolved in water. OGTT was performed in 499 patients (1997-2003, n=390) excluding 33 patients with an established diabetes diagnosis.

Assays

LH, FSH, cortisol, insulin, and C-peptide were analyzed by time-resolved flouroimmunoassay using commercial kits (AutoDELFIA, Wallac Oy, Turku, Finland). Intra-assay variations were: FSH 1.0 - 1.4%, LH 1.8 - 9.4%, cortisol 2.7 - 3.6%, insulin 2.1 - 3.7%, and C-peptide 3.1 - 5.0%. Inter- assay variations were: FSH 2.1 - 3.7%, LH 2.0 - 3.9%, cortisol 0.8 - 1.9%, insulin 3.4 - 4.0%, and C-peptide 1.9 - 3.0%.

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17OHP was analyzed by radioimmunoassay using a commercial kit (Coat-A-Count, Diagnostic Products Corporation, Los Angeles). Intra- and inter-assay variations were 3.5 - 7.1% and 5.0 - 11%, respectively. Serum total testosterone and sex hormone binding globulin (SHBG) were analyzed using a specific radioimmunoassay (RIA) after extraction as previously described (14). In this method, testosterone, dihydrotestosterone and androstendione are extracted before applying RIA and overestimation of testosterone levels is avoided. The method shows a close correlation with the determination of testosterone levels by using mass spectrometry. The intra-assay CV for total testosterone was 8.2% and for SHBG it was 5.2%. The inter-assay CV for total testosterone was 13.8% and for SHBG it was 7.5%. Free testosterone levels were calculated from measurements of total testosterone and SHBG. DHEAS was measured by RIA on diluted serum using specific antibodies. The inter-assay variation for DHEAS was 10.7%. Plasma total cholesterol, high-density lipoprotein (HDL), cholesterol, and triglyceride (TG) were analyzed by enzymatic colorimetric reactions (Modular P, Roche), and LDL cholesterol was calculated using the Friedewald equation. Blood glucose was measured on capillary ear blood using Hemo Cue. We calculated HOMA-r = fasting insulin * fasting CBG / 22.5 (15), reference ≤ 15 pmol mmol 1-². Area under the curve (AUC) for insulin during OGTT was calculated using the trapezium rule.

Statistic methods

All presented data represent pre-treatment values. *Pakistan is most often referred to as part of the South Asian continent but may according to recent definitions also be included as part of the Middle East. Due to the limited number of subjects we chose to omit Asian patients. In order not to exclude the 14 patients from Pakistan we included these patients as part of the Middle Eastern patient group. Omitting the 14 Pakistani women from the calculations did not change the significant differences between ME and CA patients.* Blood pressures measured in patients using antihypertensive treatment were not included in the dataset. Most parameters were not normally distributed and were therefore described using medians and quartiles. The Mann Whitney test was used to compare differences between two groups. The Chi squared test was used to test for differences in frequencies between groups. We performed multiple regression analyses to take into account that the two study populations differed significantly in age and BMI. These analyses were performed on log data. In regression analyses metabolic and clinical characteristics were entered as the dependent variable and age, BMI, and ethnicity (Caucasian =1, Middle East =2) were entered as the explanatory variables. In tables, p-values for the relevant explanatory variable are presented. We used SPSS version 13 (SPSS, Chicago, IL) in our calculations. P-values < 0.05 were considered significant.

Results

Clinical and biochemical characteristics

1206 patients were referred for evaluation. 1002 patients fulfilled the inclusion criteria and 204 patients were excluded. **Table 1** shows the geographic distribution of included patients. The patients were divided according to ethnicity: Caucasian (CA, n=784), Middle East (ME, n=190), Asian (n=14), various (n=14). **Table 2** presents clinical and biochemical characteristics in CA and ME patients. CA patients were significantly older and had higher BMI compared to ME patients. *After correcting for differences in age and BMI between the two study populations, clinical hyperandrogenicity was lower in CA women and SHBG and androgen levels were higher*. Prolactin levels were lower in CA women. CA women had increased blood pressure and smoking prevalence, but no significant differences were found in lipid profiles (**Table 2, Figure 1**). The risk of diabetes and IGT was comparable in the two study populations, but CA women had decreased AUC insulin and 2 hour glucose during OGTT and decreased frequency of high fasting insulin (**Table 3**). Basal 170HP was increased in CA women but no significant differences were detected in adrenal activity during ACTH tests.

PCOS criteria

Rotterdam criteria: A total of 557 patients fulfilled the Rotterdam criteria for PCOS: 56.2% (448/784) CA vs. 56.8% (109/190) ME women. The prevalence of the individual criteria differed significantly between the two study populations, **Figure 1**. 47.0% (314/668) CA women vs. 29.0% (50/172) ME women had PCO and 89.1% (699/684) vs. 93.2% (179/190) had clinical and/or biochemical hyperandrogenism (both p<0.05).

Table 4 presents clinical and biochemical characteristics of the 557 patients fulfilling the Rotterdam criteria

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divided according to BMI. Overweight patients were significantly more insulin resistant and had a more adverse cardiovascular risk profile than normal weight patients within both ethnic subgroups of patients. *The different prevalence of hyperandrogenism and PCO remained significant between CA and ME women fulfilling the Rotterdam criteria (data not shown).*

NIH criteria: The NIH criteria were fulfilled in 439 patients, 44.5% (349/784) CA vs. 47.3% (90/190) ME women. *Differences in metabolic and hormonal parameters between CA and ME women were not significantly affected when NIH criteria were applied instead of the Rotterdam criteria (data not shown).* Idiopathic hirsutism defined as regular menses, testosterone levels within reference interval and US with normal ovaries was diagnosed in 232 patients (CA n=183, ME n=49).

Multiple regression analysis

Ethnicity as an independent predictor of testosterone levels and metabolic risk factors is shown in **Table 5.** ME women had increased AUC insulin and 2h glucose and decreased androgen and SHBG levels after correcting for BMI and age.

Multiple regression analyses were performed as described above with prolactin entered as the dependent variable. HDL (B= 0.21, p= 0.03), LDL (B= - 0.17, p= 0.02), 17OHP 0 (B= 0.12, p= 0.004), and smoking status (B= - 0.05, p= 0.007) were independent predictors of prolactin levels after correcting for ethnicity, age, and BMI. Prolactin levels were 7(5-10) vs. $6(5-9) \mu g/l$ in non-smoking vs. smoking patients (p<0.001). No significant associations were found between prolactin and sex hormones, cycle history, or PCO status.

Discussion

Limited data are available on ethnic differences in the manifestations of PCOS in North European study populations. In the present study we included data from a large well characterized Danish population of premenopausal PCOS women. CA women were significantly older than ME women and had significantly higher BMI. *After correcting for differences in age and BMI, ME women were characterized by increased androgenicity, which could be explained by decreased SHBG, and increased diabetes risk whereas a more unfavourable cardiovascular risk profile was found in CA women.*

It has been reported that African-American and Hispanic women with PCOS are more insulin resistant and have increased diabetes risk compared to Caucasian women (5;16). Our findings of increased metabolic risk factors in ME women compared to CA women are in agreement with studies in populations without PCOS (9;17;18), whereas few studies have been performed in PCOS populations. Wijeyaratne et al included 47 south Asian women with PCOS (including 40 women from Pakistan) and 40 Caucasian women with PCOS (19). In agreement with our study, south Asian patients were younger with decreased BMI and increased FGscore compared to Caucasian patients (19). Asian women had significantly decreased insulin sensitivity compared to CA women, whereas no significances were found in fasting glucose levels (19). We and Wijeyaratne et al (19) reported significantly increased SHBG levels in Caucasian women, which may support higher insulin sensitivity (19:20). Sixty-four percent Asian women vs. 30% Caucasian women with PCOS had a family history of diabetes, which may have affected results (19). The effects of race and family history on diabetes risk were studied by Ehrmann et al in a mixed ethnic study population of 408 PCOS patients of primarily black and white race (16). Ethnicity was found to influence parameters of insulin resistance and body composition independent of family history (16). Intra-family marriages are more prevalent in ME women than in CA women (21). In the present study we did not have data on family marriages. In a Pakistan population of PCOS women, consanguineous marriage was associated with insulin resistance and glucose intolerance and a family history of diabetes and cardiovascular disease was observed more often in the group of women marrying within family (21).

CA women in the present study had increased blood pressure, smoked at a higher frequency, and tended to have a more adverse lipid profile than ME women. These findings suggest that CA women are at increased

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cardiovascular risk than ME women. In populations without PCOS, the data on cardiovascular risk profile in ME vs. CA populations are conflicting (9;17). A meta-analysis found a tendency to improved lipid profile in ME women compared to CA women whereas this could not be reproduced in other studies (9). In an American population, the blood pressure in Asian PCOS patients was comparable to CA women (22). The present data are in agreement with the findings of Wijeyaratne *et al* reporting similar lipid profile in Asian vs. CA women with PCOS but significantly decreased blood pressure in Asian women (systolic 114 vs. 120 mmHg and diastolic 72 vs. 76 mmHg) (19).

ACTH stimulated 17OHP levels did not differ between ME and CA women in the present study suggesting comparable adrenal androgen production. In agreement with this finding, the age and BMI corrected DHEAS and cortisol levels were similar in the two study populations. In contrast to these findings, two studies found decreased DHEAS levels in Black and Mexican American vs. white women with PCOS (23;24). Using race specific cut offs, black women with PCOS had increased prevalence of high DHEAS levels (23) suggesting that adrenal activity may differ between some ethnic groups.

An unexpected finding of the present study was the higher prevalence of polycystic ovaries in CA women compared to ME women. We are not aware of previous studies reporting similar differences in the prevalence of PCO between different ethnic populations with PCOS. In the present study PCO was defined according to the criteria suggested by Adams and ovarian volume was not systematically recorded. The Rotterdam criteria suggest that increased ovarian volume >10 ml alone can be used to define PCO (4). It is therefore possible that the prevalence of PCO may be higher than estimated in the present study. In the study by Wijeyaratne, there was a trend to increased ovarian volume in Asian PCOS patients, but this did not reach statistical significance (19). Welt *et al* reported similar prevalence of PCO between Caucasian PCOS patients from Boston and Iceland but Icelandic patients had smaller ovaries with decreased follicle number supporting that ovarian morphology may vary in different study populations (25).

The median age upon referral was relatively high in our study population and CA patients were seven years older than ME women. Patients with a primary wish for conception were referred directly to the fertility

clinic and therefore the primary complaints of our population were irregular menses and hirsutism. The median age for first child birth in Denmark is 29-30 years and more than 25% women in reproductive age use oral contraceptives (OC) (26). OC pills efficiently alleviate the symptoms of PCOS and this may explain the relatively high age upon referral in our population. It is possible that a lesser percentage of ME women use OC as birth control and therefore the symptoms of PCOS present in an earlier age. Impaired quality of life due to for example overweight and hirsutism is well described in PCOS and may be the primary reason for referral of the patient (27). *In a previous Austrian study, quality of life regarding infertility, overweight, hirsutism, and menstrual disturbances were significantly impaired in infertile Moslem immigrant women with PCOS compared to Austrian women with comparable clinical or biochemical symptoms of PCOS (28).*

Prolactin levels were routinely measured to exclude prolactinomas as a cause of menstrual irregularity and none of the included patients had prolactin levels above reference interval. To our knowledge we are the first to report increased prolactin levels in ME patients compared to CA patients. Multiple regression analyses suggested that the difference could in part be explained by decreased smoking prevalence in ME women. Increased prolactin levels were furthermore associated with increased HDL, decreased TG and increased 170HP levels after correcting for BMI, race, and age. Our findings are in agreement with previous studies on the effect of nicotine on prolactin (29;30). We are not aware of previous studies documenting associations between prolactin secretion and lipid metabolism. Further studies are needed to evaluate prolactin levels among different ethnic populations with PCOS.

The present study included patients referred to our specialist center from general practitioners and local gynecologists and dermatologists. The evaluated patient population therefore probably represents a selected patient population. Furthermore, the ME patient group included patients from a large geographic area. The patients underwent a standardized evaluation program including check schemes to minimize the inter observer variation but clearly more prospective studies are needed to confirm our study results in well characterized PCOS populations of mixed ethnicity.

Conclusion

ME women with PCOS are referred at a younger age and are characterized by decreased BMI and increased androgenicity compared to CA women with PCOS. Insulin resistance is increased in ME women whereas the cardiovascular risk profile is more beneficial regarding blood pressure and smoking status. The present study support that ethnic groups differ in clinical and biochemical presentation of PCOS and future studies should focus on prospective data in different ethnic populations with PCOS.

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Table 1: Geographic distribution of included patients (n=1002)

Caucasian (n=784)	Middle East (n=190)	Asian (n=14)	Various (n=14)
Albania (n=2)	Afghanistan (n=4)	India (n=1)	African countries (n=6)
Armenia (n=1)	Arabia (n=29)	Korea (n=1)	South America (n=3)
Bosnia (n=4)	Iraq (n=36)	Sri Lanka (n=11)	Not noted (n=6)
Croatia (n=1)	Iran (n=19)	Vietnam (n=1)	
Denmark (n=765)	Jordan (n=2)		
Poland (n=3)	Lebanon (n=12)		
Rumania (n=1)	Pakistan (n=15)		
Russia (n=1)	Palestine (n=3)		
Yugoslavia (n=6)	Syria (n=2)		
	Turkey (n=57)		
	Middle east* (n=11)		
* Exact country not si	pecified in patient reports.	0	

	Caucasian	Middle east
	(n=784)	(n=190)
Age (years)	32 (25 - 37)**	25 (18 - 32)
BMI (kg/m ²)	27.0 (22.9 - 32.8)*	25.7 (22.7 - 30.4)
Waist (cm)	90 (78-104)**§§	82 (73 – 98)
FG face	4 (2-5)*§§	4 (3 – 6)
FG total	11 (7-15) **§§	16 (11 – 22)
T-testosterone (nmol/l)	1.72 (1.24 – 2.34)§	1.61 (1.18 – 2.21)
SHBG (nmol/l)	47 (33 – 70)*§	40 (31 - 60)
FTI	0.031 (0.020 - 0.047)§	0.030 (0.019 - 0.048)
DHEAS (µmol/l)	5.8 (4.1 – 8.1)*	6.3 (4.7 – 9.2)
Prolactin (µg/l)	7 (5 – 10)**§§	9 (6 -12)
Systolic BP (mmHg)	127 (117 -138)**§§	115 (10 – 122)
Diastolic BP (mmHg)	80 (72 – 89)**§§	75 (70 - 80)
Cholesterol (mmol/l)	4.8 (4.2 – 5.5) **	4.4 (3.9 – 5.0)
TG (mmol/l)	1.1 (0.8- 1.6)*	0.9 (0.7 – 1.4)
HDL (mmol/l)	1.4 (1.1 – 1.7)	1.4 (1.1 – 1.7)
LDL (mmol/l)	2.8 (2.2 – 3.5)**	2.5 (2.1 – 3.0)
Fasting insulin (pmol/l)	55 (37 – 91)	58 (38 - 92)
AUC insulin (pmol/l)	3.8 (2.4 – 6.6)*§	4.7 (3.0 – 8.3)
HOMA-r (pmol mmol 1 ⁻²)	11.8 (7.9 – 20.3)	12.4 (8.6 – 20.5)
Fasting glucose (mmol/l)	4.8 (4.4 – 5.2)	4.8 (4.5 – 5.2)
2 h. glucose (mmol/l)	6.0 (5.1 – 7.1)§	6.1 (5.2 – 7.2)
Cortisol 0 (nmol/l)	357 (294 – 472)*	326 (263 – 410)
AUC cortisol (nmol/l)	3.4 (3.0 – 3.8)	3.2 (2.9 – 3.8)
17OHP 0 (nmol/l)	4.1 (2.9 – 6.2)§	4.0 (2.8 - 5.7)
AUC 17OHP (nmol/l)	5.6 (4.5 - 7.0)	5.4 (4.3 – 7.2)

Fable 2: Clinical and paraclinical data from	patients divided according to ethnicity (n=974)
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Data presented as median (quartiles)

*<0.05 vs. ME patients

**<0.001 vs. ME patients

 $<\!\!0.05$ vs. ME patients after correcting for age and BMI

§§<0.001 vs. ME patients after correcting for age and BMI

Table 3: Outcome of glucose tolerance tests

	Caucasian	Middle east	
Normal glucose tolerance	344/414 = 83.1 %	69/83 = 83.1 %	
IGT	57/414 = 13.8 %	10/83 = 12.0 %	
Diabetes	13/414 = 3.1 %	4/83 = 4.8 %	
Increased fasting insulin	227/447 = 50.6% §	64/122 = 52.4%	

§<0.05 vs. ME patients after correcting for age and BMI Glucose tolerance defined according to WHO criteria Increased fasting insulin defined as >55pmol/l

	Caucasian		Middl	e East
	BMI≤25 kg/m ² (n=141)	BMI>25 kg/m ² (n=262)	BMI≤25 kg/m ² (n=38)	BMI>25 kg/m ² (n=66)
Age (years)	27 (23 - 33)**¤¤	31 (25 – 35)**	22 (17 – 27)¤	26 (18 - 33)
Waist (cm)	77 (71 – 81)¤¤	103 (93 – 111)*	75 (70 – 78)¤	98 (87 – 106)
FG face	3 (2 - 4)*§§¤¤	4 (3 – 6)*§§	4 (3 – 6)	5 (3 – 6)
FG total	11 (6-14)**§§	11 (7-16)**§§	19 (11 – 24)	16 (11 - 22)
T-testosterone (nmol/l)	1.89 (1.34 – 2.31)	2.02 (1.43 – 2.73) §	1.59 (1.12 – 2.23)	1.82 (1.31 – 2.47)
SHBG (nmol/l)	57 (42 – 76)**¤¤	37 (24 – 50)	43 (31 – 67)¤	33 (22 - 49)
БТІ	0.030 ¤¤	0.043 §	0.028	0.044
r 11	(0.019 – 0.045)	(0.028 - 0.062)	(0.019 – 0.043)	(0.022 - 0.057)
DHEAS (µmol/l)	5.8 (4.0 - 7.9)	5.8 (4.2 – 7.7)	6.5 (4.5 – 11)	6.2 (4.7 – 9.2)
Prolactin (µg/l)	7 (4 – 10)§	7 (5 - 10)*	11 (7 – 15)	8 (6 – 11)
Systolic BP (mmHg)	120 (110 – 132)*¤¤	129 (120 – 140)**§§	112 (105 – 120)	120 (110 – 126)
Diastolic BP (mmHg)	73 (69 – 81)¤¤	80 (79 – 90)**§§	70 (68 - 80)	75 (70 - 80)
Cholesterol (mmol/l)	4.4 (3.9 – 5.0)¤¤	4.9 (4.3 – 5.7)*§	4.5 (3.7 – 4.8)	4.5 (3.9 – 5.1)
TG (mmol/l)	0.8 (0.6 – 1.1)¤¤	1.4 (0.9 – 1.9)*	0.9 (0.7 – 1.2)	1.1 (0.7 – 1.8)
HDL (mmol/l)	1.6 (1.3 – 1.8)¤¤	1.2 (1.0 – 1.4)	1.5 (1.3 – 1.7)¤¤	1.2 (1.0 – 1.5)
LDL (mmol/l)	2.5 (2.0 – 2.9)¤¤	3.0 (2.4 – 3.7)*§	2.5 (2.0 – 2.7)	2.6 (2.1 – 3.1)
Fasting insulin (pmol/l)	38 (28 – 51)¤¤	82 (51 – 119)	47 (29 – 59)¤¤	77 (49 – 105)
AUCinsulin (10 ⁴ pmol/l)	2.8 (1.7 – 3.5)*¤¤	5.3 (3.2 - 8.3)	3.6 (2.7 – 7.4)	6.0 (3.7 – 8.9)
HOMA (pmol mmol l ⁻²)	8.5 (5.6 – 11.1)¤¤	17.1 (10.8 – 27.8)	10.0 (6.7 – 14.4)¤¤	15.7 (10.9 – 21.9)
F. glucose (mmol/l)	4.6 (4.3 – 4.8)	4.6 (4.2 – 5.0)	4.5 (4.2 – 4.7)	4.7 (4.1 – 4.8)
2 h. glucose (mmol/l)	5.9 (5.1 – 6.5)¤	6.3 (5.1 – 7.4)	6.0 (4.9 - 6.9)	6.4 (5.4 – 7.6)

Table 4: Clinical and paraclinical data from patients with PCOS according to Rotterdam criteria (n=557).

Data presented as median (quartiles). Data on BMI were not available in all cases.

*<0.05 vs. ME patients within BMI group

**<0.001 vs. ME patients within BMI group

§<0.05 vs. ME patients within BMI group after correcting for age

§§<0.001 vs. ME patients within BMI group after correcting for age

¤<0.05 vs. overweight individuals within ethnical group

aa<0.001 vs. overveight individuals within ethnical group

Table 5. Regression analyses on the effect of ethnicity on hormonal and metabolic outcomes (n=974)

Log Dependent		Ethnicity		
variable	Log BMI	Log age	(Cau vs. ME)	R ² model
T-testosterone	0.21 *	-0.32**	-0.06*	0.04**
SHBG	-1.09**	0.22**	-0.05*	0.17**
Free-testosterone	0.82**	-0.50**	-0.06*	0.11**
Fasting insulin	2.07**	-0.46**	0.002	0.47**
AUC insulin	1.73**	-0.31**	0.084*	0.27**
НОМА	2.18**	-0.39**	0.16	0.44**
F. glucose	0.09*	0.085**	0.003	0.05**
2 h. glucose	0.37**	0.10*	0.035*	0.09**

Multiple regression analyses were performed with metabolic outcomes as the dependent variables and

ethnicity (cau=1, ME=2), BMI, and age as explanatory variables.

Metabolic outcomes, BMI, and age were log-transformed.

Data presented as B-value

* p<0.05

** p<0.001







^{*&}lt;0.05 vs. ME patients

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