

Enhanced increase of omega-3 index in response to long-term n-3 fatty acid supplementation from triacylglycerides vs. ethyl-esters

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1 Enhanced increase of omega-3 index in response to long-term n-3 fatty

2 acid supplementation from triacylglycerides vs. ethyl-esters

3 Running title: Omega-3 index: superiority of n-3 fatty acid triacylglycerides vs. ethyl-esters

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- The study supplements used in this study were a gift from Dr. Loges + Co. GmbH, Winsen,
- 45 Germany.

ABSTRACT

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eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) are absorbed in an identical

Background: There is a debate currently about whether different chemical forms of

- 50 way. The objective of this study was to investigate the response of the omega-3 index, the
- 51 percentage of EPA + DHA in red blood cell membranes, to supplementation with two
- 52 different n-3 FA formulations in humans.
- 53 **Design:** The study was conducted as a double-blind placebo-controlled trial. 150 volunteers
- were randomly assigned to one of three groups: 1) fish oil concentrate with EPA + DHA
- 55 (1.01g + 0.67g) given as re-esterified triacylglycerides (rTAG-group); 2) corn oil (placebo-
- group); or 3) fish oil concentrate with EPA + DHA (1.01g + 0.67g) given as ethyl-esters
- 57 (EE-group). Volunteers consumed four gelatine-coated soft capsules daily over a period of
- six months. The omega-3 index was determined at baseline (t_0) , after three months (t_3) and
- at the end of the intervention period (t_6) .
- 60 **Results:** The omega-3 index increased significantly in both groups treated with n-3 FA
- from the baseline to t_3 and t_6 (P<0.001). The omega-3 index increased to a greater extend in
- 62 the rTAG-group than in the EE-group (t₃: 186% vs. 161% [*P*<0.001]; t₆: 197% vs. 171%
- 63 [*P*<0.01]).
- 64 **Conclusion:** A six month supplementation of identical doses of EPA + DHA led to a faster
- and higher increase in the omega-3 index when consumed as triacylglycerides than when
- 66 consumed as ethyl-esters.
- 67 **KEYWORDS**
- 68 n-3 fatty acids, triacylglycerides, ethyl-esters, omega-3 index, eicosapentaenoic acid,
- 69 docosahexaenoic acid

71 **Introduction**

72 Cardiac societies recommend an increased intake of the long-chain n-3 fatty acids (FAs) 73 eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA), especially for patients with 74 known cardiovascular disease (CVD) or for individuals at high risk of CVD (Fruchart et al., 75 2008; Graham et al., 2007; Smith et al., 2006). Although most of the large intervention 76 studies on which these recommendations are based were conducted with EPA + DHA ethyl-77 ester (EEs), nutrition and cardiac societies recommend increasing the intake of n-3 FAs by 78 consuming at least two servings of (preferably oily) fish per week, providing 400-500 mg/d of 79 EPA + DHA, mainly esterified as triacylglycerides (TAGs), but also in membrane 80 phospholipids (PLs). For the portion of the population reluctant to eat fish, the societies 81 recommend concentrated fish oil preparations which typically contain EPA and DHA as 82 TAGs or EEs. These recommendations presume that different chemical forms of EPA + DHA 83 have an identical efficacy. 84 The majority of previous studies which compared the bioavailability of EPA + DHA from 85 TAGs or rTAGs vs. EEs (El Boustani et al., 1987; Lawson & Hughes, 1988 [a & b]; 86 Beckermann et al., 1990; Dyerberg et al., 2010; Luley et al., 1990; Nordøy et al., 1991; 87 Krokan et al., 1993) were short-term with a maximal length of seven weeks. The focussed 88 endpoints in these trials - FA composition of plasma PLs, TAGs, chylomicron TAGs, or 89 serum PLs – similarly reflect short-term rather than long-term intake (Katan et al., 1997; Cao 90 et al., 2006). Moreover, larger doses (> 2 g/d EPA and DHA) were used in many of these 91 studies, which is in contrast to the recommendations of the societies mentioned. 92 We compared the bioavailability of a moderate dose of n-3 FA-TAGs vs. n-3 FA-EEs in 93 comparison to a placebo as part of a six-month randomized controlled study, where we 94 likewise assessed the effects of the two n-3 FA formulations on the lipid profile in 95 hyperlipidemic subjects. The omega-3 index, the percentage of EPA + DHA in red blood cell 96 (RBC) membranes, reflects the long-term intake and n-3 FA status of an individual (Harris & 97 von Schacky, 2004; Harris, 2007; Harris, 2008; Sun et al., 2007).

SUBJECTS AND METHODS

Subjects

150 subjects between 30 and 75 years old were recruited via newspaper advertisements in four different German cities (Munich, Hamburg, Hannover, and Goslar) between April and October, 2008. Since the study was conducted to assess the effects of the two fish oil formulations on the lipid profile in hyperlipidemic subjects, the inclusion criterion was hyperlipidemia solely treated with HMG-CoA-reductase inhibitors (statins). The exclusion criteria were serious illness (type 1 diabetes, cancer, coronary heart disease, bleeding disorders); body mass index > 35 kg/m²; gastrointestinal disorders; medications known to affect lipid metabolism; daily fish consumption or ingestion of dietary n-3 FA; and plant sterol supplements. Written informed consent was obtained from all participants. The trial was conducted with respect to GCP Guidelines and the protocol was approved by an ethics commission.

Study design

manner. Randomization was conducted using a computer-generated randomization scheme. Codes were kept in a remote secure location by an independent third party. All participants, as well as medical, laboratory and clinical trial staff and investigators assessing the endpoints, were blinded to the randomization until all study data had been collected, checked and verified. Each of the study products was given two codes to further facilitate the masking of the capsules. The capsules were provided in numbered containers.

Qualified participants were randomly assigned to one of three groups in a double masked

The three different types of gelatine-coated soft capsules contained (1) n-3 FAs as re-esterified TAGs (rTAG-group; n=52); (2) corn oil (placebo-group; n=49); or (3) n-3 FA-EEs (EE-group; n=49), and were provided by Dr. Loges and Co. GmbH (Winsen, Germany),

a pharmaceutical company. Re-esterified TAGs are made from fish body oil to concentrate specific FAs in the resulting products. Therefore, FAs from the natural TAGs are transferred to ethanol, forming EEs. After removing undesirable FAs, EEs are enzymatically reconverted into TAGs, now called re-esterified TAGs (rTAGs). The FA composition of the two different supplements was identical (**Table 1**). The daily intake of EPA and DHA in both n-3 FAgroups was 1008 mg, and 672 mg, respectively. The placebo capsules contained corn oil and were outwardly identical to the n-3 FA-capsules in all aspects.

Participants were instructed to ingest four capsules of their assigned study supplement daily together with food, two in the morning and two in the evening, and to maintain their usual exercise and dietary habits throughout the intervention time of six months.

The subjects' height and weight were measured, and fasting blood samples were collected by venipuncture into K-EDTA tubes at the baseline (t_0) , after three months (t_3) and at the end of the intervention period (t_6) . Subjects additionally completed a questionnaire to obtain information about lifestyle habits (e.g. usual fish intake, physically activity) as well as tolerability of the two different n-3 FA formulations.

Volunteers' compliance was assessed by count of left-over capsules between the three investigation dates.

RBC membrane fatty acid analysis

RBC membrane FA composition was analyzed according to the omega-3 index methodology as described previously (Harris & von Schacky, 2004). FA methyl-esters were generated from RBCs by acid transesterification and analyzed by gas chromatography using a GC2010 Gas Chromatograph (Shimadzu, Duisburg, Germany) equipped with a SP2560, 100-m column (Supelco, Bellefonte, PA), using hydrogen as the carrier gas. FAs were identified by comparison with a FA standard mixture characteristic for RBCs. Results for the omega-3 index are given as EPA + DHA expressed as a percentage of the total identified FAs after

response factor correction. The coefficient of variation for EPA + DHA was 5%. Quality was assured according to DIN ISO 15189.

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Statistics

SPSS 17 (SPSS Inc., Chicago IL, USA) was used for statistical analysis. The results are presented as mean±SD. The sample size (n = 50 per group) was calculated for differences in serum TAG levels (primary endpoint) in order to be sufficient to prove a slightly more than medium-sized superiority (standardized difference of means $(\mu_1 - \mu_2)/\sigma = 0.566$) of verum over placebo with a 5% significance level and a power of 80%. If the test for difference of all three groups resulted in significance, the three pairwise tests could be done without adjustment of the significance level following the Closed Test Procedure. The increase of the omega-3 index was a secondary endpoint, therefore P < 0.05 was considered significant. Statistical comparisons were based on per protocol population, defined as subjects completing all visits and not developing any of the exclusion criteria during the intervention period. An intention to treat analysis would have included non-compliant subjects from both groups, and would have obscured rather than clarified the effects of n-3 FA treatment. Testing for normality (KS-Test) revealed normal distribution for arachidonic acid (AA), EPA, DHA, and omega-3 index at the baseline in each intervention group. Baseline markers were compared among groups using one way ANOVA. Scheffe tests of contrast were performed where appropriate. Changes in values of the different variables observed at baseline and t3 or t6 were evaluated within groups by Student's t test for dependent samples (**Table 2**). To compare differences of RBC membrane FAs among groups, corresponding baseline values were used as covariates to diminish possible effects caused by diverse t₀ values.

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RESULTS

Of the 150 eligible subjects who started the study, 21 were excluded from subsequent statistical analyses. The number of subjects lost during the study and the number available for analysis are summarized in **Figure 1**. Five subjects failed one or more of the inclusion criteria at the baseline visit. Four more subjects decided to discontinue participation before the t_3 visit and three before the t_6 visit. Three subjects failed to attend the t_3 visit because of illness which was not related to the study product, and two additional subjects had relevant changes in lipid metabolism affecting medication (medical treatment of thyroid dysfunction). Four more participants were excluded from subsequent statistical analysis because of the lack of compliance. Hence, 129 participants completed the study protocol and were included in the analysis.

Subject characteristics

The three groups were normally distributed with respect to age, gender, body weight, and body mass index (BMI), and did not show any differences between the groups (**Table 3**). There were no significant changes in BMI during the trial (data not shown). 19 Volunteers reported side effects occurring during intervention time. While eight subjects from both active treatment-groups reported eructation, three subjects from the EE-group reported flatulence.

RBC membrane fatty acids

RBC membrane FAs for $t_0/t_3/t_6$ are shown in **Table 2**. The baseline values of AA, EPA and DHA did not differ between groups.

Between t_0 , t_3 and t_6 , there were significant changes in RBC membrane FAs in both n-3 FA treated groups for AA, EPA and DHA: AA decreased, while EPA and DHA increased (P<0.001). Significant changes were also seen in the placebo-group, showing an increase in

AA and DHA between t_0 and t_6 , and a decrease in EPA between t_0 and t_3 (P<0.001), as well as

200 between t_0 and t_6 (P<0.05) (**Table 2**).

Three months of n-3 FA treatment resulted in significant increases of EPA, DPA (docosapentaenoic acid) and DHA content in RBC membranes compared with the placebo-

group, whereas the AA contents significantly decreased (P<0.001). In a similar way, the

observed tendencies were constant between t3 and t6 with a slight, although not significant,

further increase for EPA and DHA. The decrease between t₃ and t₆ for AA was significant in

both n-3 FA-groups compared with the placebo-group (rTAG-group, -0.43%; EE-group, -

207 0.58%; *P*<0.001).

208 The increase in the omega-3 index was significantly higher in the rTAG-group compared to

the EE-group: performing an ANCOVA to compare absolute values of the omega-3 index

between the three groups revealed that the omega-3 index was significantly higher after

three and six months in both n-3 FA treated groups compared to the placebo-group ($\Delta t_3 t_0$:

212 P< 0.001; $\Delta t_6 t_0$: P<0.001). Similarly, the omega-3 index was significantly higher after

three and six months in the rTAG-group compared to the EE-group ($\Delta t_3 t_0$: P<0.001; $\Delta t_6 t_0$:

214 P<0.01) (**Figure 2**).

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DISCUSSION

217 To our knowledge, this is the first long-term randomized study comparing EPA + DHA

rTAGs with EEs with regard to their incorporation into RBC membranes, i.e. the omega-3

index in humans. EPA + DHA consumed as rTAGs resulted in a higher increase of the

omega-3 index than when consumed as EEs over a six-month period.

221 Previous studies, which compared the effects of n-3 FA-administration from TAGs or rTAGs

vs. EEs on incorporation of EPA and DHA into plasma lipids (e.g. TAGs, chylomicron

TAGs, PLs), revealed heterogeneous results (El Boustani et al., 1987; Lawson & Hughes,

224 1988 [a & b]; Beckermann et al., 1990; Dyerberg et al., 2010; Luley et al., 1990; Nordøy et 225 al., 1991; Krokan et al., 1993; Hansen et al., 1993; Reis et al., 1990). While most trials 226 reported a faster and higher mean relative bioavailability of DHA and EPA from TAGs or 227 rTAGs compared to EEs (El Boustani et al., 1987; Lawson & Hughes, 1988 [a & b]; 228 Beckermann et al., 1990; Dyerberg et al., 2010), other findings suggest a comparable 229 bioavailability (Luley et al., 1990; Nordøy et al., 1991; Krokan et al., 1993) or were 230 heterogeneous in their outcomes (Hansen et al., 1993; Reis et al., 1990). In their recent 231 publication, Dyerberg et al. (2010) clearly demonstrated in a short-term study over two weeks 232 that EPA + DHA from rTAGs show a superior bioavailability compared to EEs. Harris et al. 233 (1988) compared the differences of EPA + DHA incorporation into plasma PLs between n-3 234 FA-TAGs and methyl-esters (ME), which are assumed to be equivalent to EEs. While no 235 differences in EPA incorporation could be detected between ME and TAG, the DHA 236 incorporation was slightly elevated after ME administration. 237 However, short-term bioavailability – as measured by plasma levels – does not necessarily 238 reflect the incorporation into tissues. We chose the omega-3 index (RBC membranes EPA + 239 DHA) as the endpoint because it reflects the incorporation of EPA and DHA into tissues, 240 including cardiac tissue (Metcalf et al., 2007; Harris et al., 2004; Harris, 2008), and is thus a 241 reasonable reflection of a person's EPA + DHA status. After six months, but not after seven 242 weeks, of supplementation, a steady state is reached in RBC membranes (Arterburn et al., 243 2006). Our approach contrasts with previous studies mentioned in the INTRODUCTION which 244 had a maximum duration of seven weeks, and analyzed FA compartments reflecting short-245 term rather than long-term intake. 246 The omega-3 index increase was significantly higher after consumption of n-3 FA as rTAGs 247 than after EEs. The mechanisms explaining this observation are unclear at present. Therefore, 248 we discuss the following possible explanations:

- Differences in amount of ingested study medication: Capsule counts did not reveal any
- differences between the two relevant groups (percentage of ingested capsules was 95% in the
- rTAG-group vs. 96% in the EE-group).
- 252 Age: In a previous study, older subjects exhibited a stronger response to DHA
- supplementation than younger subjects (Vandal et al., 2008). In the present study, however,
- age was not different between groups.
- 255 Matrix effects: There are conflicting results in RBC membrane FA composition after
- administering n-3 FA-EEs in capsules or n-3 FA-TAGs as fish. While Visioli et al. (2003)
- observed a higher uptake of n-3 FAs in the fish-meal group than in the capsule group, Harris
- 258 et al. (2007) found similar elevations in omega-3 index produced by both products. However,
- 259 the study conducted by Visioli *et al.* was a small pilot study, including only 4-8 volunteers.
- Additionally, their study was retrospective, not randomized and, moreover, n-3 FA-intake
- 261 from the two sources was not matched. Nevertheless, absorption of n-3 FAs from EE was
- three-fold higher in the presence of a high-fat meal compared to a low-fat meal, but still not as
- 263 high as from n-3 FA-TAGs (Lawson & Hughes, 1988 [a & b]). In another study, no
- 264 differences were seen in n-3 FA-rTAG and -EE absorption after administering these FAs in
- meals with similar fat contents (Nordøy et al., 1991). However, the study was conducted with
- very high doses of EPA + DHA (~22-24g) and with five volunteers a comparatively low
- number of cases. In regard to the results of Visioli et al. (2003) and Lawson & Hughes (1988)
- 268 [a & b]), matrix effects and different nutritional patterns could be held responsible for
- different findings.
- Differences in digestion: n-3 FA-TAGs, but not n-3 FA-EEs, have a glycerol backbone
- 271 (Carlier et al., 1991). In the small intestine, pancreatic lipase hydrolyzes FAs from alcohol
- backbones and free FAs, and monoacylglycerols are taken up by enterocytes. *In vitro* studies
- 273 conducted by Yang et al. (1990 [a]) showed that the FA-ethanol bond is hydrolysed 10 to 50
- times more slowly by pancreatic lipase compared to hydrolysis of TAGs. This might result in

275 a delay in, or even in less, n-3 FA entering the enterocyte when EPA + DHA are consumed as 276 EEs. However, such in vitro experiments cannot mimic the in vivo situation; e.g. the extreme 277 excess of pancreatic lipase vs. substrate in the normal gut is not considered. 278 - Differences in absorption: Free FAs must be reconverted in the enterocyte to TAGs for 279 further transport. Glycerol molecules or 2-monoacylglycerols are needed for this. n-3 FA-280 TAG, but not n-3 FA-EE, has its own glycerol or monoacylglycerol substrate in storage. Yang 281 et al. (1990 [b]) suggested that slower absorption of FAs observed in rats fed with EEs might 282 be attributed to a lower efficiency of the phosphatidic acid pathway, which is required in the 283 absence of dietary 2-monoacylglycerols. 284 - Stereo specificity: As reported by Visioli et al. (2003), EPA and DHA are esterified mainly 285 in the sn-2 position in TAG molecules, and this position is to a large extent prevented from 286 hydrolysis during digestion and absorption, which might facilitate digestion and absorption of 287 TAGs. However, recent investigations by Dyerberg et al. (2010) showed that the 288 stereochemistry of FAs in acylglycerols has no influence on the bioavailability of EPA and 289 DHA. 290 Taken together, although we clearly cannot delineate a specific mechanism, we propose that 291 differences in digestion and in absorption between EEs and TAGs, which might partly depend 292 on the nutritional matrix, are the most likely mechanisms to explain our findings. 293 In keeping with previous publications (Katan et al., 1997; Belluzzi et al., 1994; Harris et al., 294 2007; Barceló-Coblijn et al., 2008), RBC membrane baseline levels of EPA were lower than 295 baseline levels of DHA. Both increased upon supplementation, with increases in EPA being 296 more pronounced, corresponding to the larger amount of EPA than DHA in the capsules. As 297 observed previously (Arterburn et al., 2006; Belluzzi et al., 1994; Krokan et al., 1993), most

omega-6 FAs were significantly reduced at t₃ and t₆.

The relative increases in EPA RBC membrane content between t₀ and t₆ is 286% for the rTAG-group and 215% for the EE-group. These observations are in agreement with findings from Katan et al. (1997), presenting a change of around 260% for EPA RBC membrane content in consequence of 1g EPA/d as TAGs after six, and similarly twelve months. Referring to these findings, it can be concluded that EPA values would not rise much higher in the rTAG-group of the present study if supplementation were to be continued. Contrary to EPA, our data shows that DHA is still accumulating in RBC membranes between month three and six in both the rTAG-group (Δ -t₆ t₃: 19%) and the EE-group (Δ -t₆ t₃: 21%). DHA is located in the inner leaflet of the PL bilayer and becomes integrated in membranes during RBC development (von Schacky & Weber, 1985). Since the first investigation timepoint takes place approximately 90 days after the baseline, and RBC lifespan is approximately 120 days (~four months), this further accumulation of DHA is not surprising. DHA reaches a steady state in RBC membranes after four to six months (Arterburn et al., 2006). As also described by Marangoni et al. (1993), accumulation of EPA in RBC membranes is faster in comparison to DHA. This observation might be caused by the preferential distribution of EPA in the outer layer of RBC membranes, which allows a more rapid exchange with plasma lipoproteins (Cartwright et al., 1985; Popp-Snijders et al., 1984). In respect to previous findings (Tynan et al., 1995; Katan et al., 1997; Arterburn et al., 2006), we could expect that differences in the omega-3 index will persist after the six-month intervention, provided the supplementation dosage remains stable. Additionally, the daily intake of four capsules of both n-3 FA study supplements for six months (1.01 g EPA + 0.67 g DHA per day) caused a boost in the mean omega-3 index up to 13.2% and 12.2% for the rTAG-group and EE-group, respectively. We calculated the

hypothetical number of n-3 FA-EE capsules which are needed to yield the same rise in the

omega-3 index obtained from four n-3 FA-rTAG capsules intake. Hence, subjects would have

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to consume at least five capsules of n-3 FA-EEs daily to achieve the same effect in the omega-3 index. Nevertheless, the goal for treatment with EPA + DHA – an omega-3 index of >8% – is highly exceeded after consumption of four or five capsules of our study medication. In any case, with respect to the strength of different increases in the omega-3 index between n-3 FA given as rTAGs or as EEs, we expect higher increases in RBC membrane EPA + DHA content even for lower dosages of our tested rTAG-capsules as compared with EEcapsules. Omega-3 index baseline levels of our study participants were slightly higher than expected (rTAG-group: 7.00±1.90%; placebo-group: 7.24±1.44%, EE-group: 7.42±1.76%). Previously, in Germany, we found a mean omega-3 index 6.14±1.83% in 1000 unselected, non-Supplementers, whereas it was 5.94±1.41% in 190 atherosclerotic subjects (von Schacky, 2010). Preliminary data indicated slight differences between the participating centers: Hannover, Hamburg, Goslar, and Munich (not shown). This should be evaluated more systematically. However, our trial aimed at comparing the magnitude of change in the omega-3 index in the three study groups, not at measuring absolute levels. Therefore, we consider it more important that baseline omega-3 index levels in our trial were comparable among the three groups. In conclusion, our findings underscore that the intake of both n-3 FA study supplements resulted in an enhanced incorporation of EPA and DHA into tissues, as evidenced by changes in RBC membrane FA content. Since the resulting omega-3 index was significantly higher after n-3 FA-rTAG administration compared to n-3 FA-EE, the results indicate that n-3 FArTAG is superior to n-3 FA-EE in view of the EPA + DHA tissues incorporation following a long-term administration. However, whether this difference would result in differences in clinical outcomes (i.e. reduction in serum TAG-levels, reductions in coronary heart disease

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348 events) is unclear and needs further investigations. Nevertheless, these obvious differences 349 between rTAGs and EEs should be considered in the n-3 FA intake recommendations. 350 351 Acknowledgements 352 Gratuitous supply of the study supplements from Dr. Loges + Co. GmbH, Winsen, Germany 353 is gratefully acknowledged. We specially thank Heike Kohrs for her technical assistance in 354 the laboratory. 355 356 **Declaration** 357 C. von Schacky received a speaker's honoraria from Solvay, and grant support from Sanofi-358 Aventis and Smartfish. He founded Omegametrix, a company offering fatty acid analyses. A. 359 Hahn and J. P. Schuchardt worked as consultants for companies which also produce and 360 merchandise fatty acid supplements. The authors are solely responsible for the design and 361 conduct of the study; collection, management, analysis, and interpretation of the data; as well 362 as preparation of the manuscript. 363 364 365

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TABLES

Table 1

Composition of the two supplements per capsule and overall daily consumption (four

484 capsules)

	n-3 fatty acids as triacylglycerides or as ethyl-esters		
Nutrient	per capsule	per day (four capsules)	
total n-3	504 mg	2016 mg	
20:5n-3 (EPA)	252 mg	1008 mg	
22:6n-3 (DHA)	168 mg	672 mg	
α-Tocopherol	6 mg	24 mg	

Table 2487 Red blood cell membrane fatty acid composition (% of total fatty acids)* at baseline (t₀), three (t₃) and six months (t₆)*

(3)	(0)			
		rTAG-group	Placebo-group	EE-group
		(n=41)	(n=43)	$(n=45)^{T}$
	t_0	0.53±0.58	0.45±0.16	0.44±0.22
C14:0	t_3	0.43 ± 0.16	0.41 ± 0.13	0.42 ± 0.15
	t_6	0.35 ± 0.15	0.42 ± 0.21	0.31 ± 0.11^{-3}
	t_0	21.44±2.54	21.56±1.75	21.36±1.72
C16:0	t_3	21.69±1.33	21.33±1.51	21.44 ± 1.76
	t_6	20.72±1.33 a	9.99 ± 1.45^{3}	20.34 ± 1.37^{3}
	t_0	0.90 ± 2.28	0.57±1.61	0.55 ± 1.62
C16:1n-7t	t_3	0.21 ± 0.13	0.22 ± 0.16	0.21 ± 0.09
	t_6	0.15±0.14 ¹	0.15 ± 0.12	0.12 ± 0.09
	t_0	0.65±0.32	0.75±0.33	0.58±0.28
C16:1n-7	t_3	0.60 ± 0.31	0.72 ± 0.27	0.56 ± 0.31
	t_6	$0.45\pm0.22^{3 \text{ a}}$	0.60 ± 0.28^{3}	0.42 ± 0.19^{3} a
	t_0	15.76±1.80	15.01±1.85	15.49±1.78
C18:0	t_3	14.99 ± 2.26^{-1}	15.06±1.69	14.97±2.10
	t_6	14.72 ± 1.81^{-2}	14.08 ± 2.05^{-1}	14.61 ± 1.42^{2}
	t_0	0.54±0.30	0.51±0.25	0.49±0.12
C18:1t	t_3	0.49 ± 0.16^{a}	0.57 ± 0.25	0.55 ± 0.28
	t_6	0.35 ± 0.16^{3}	0.41 ± 0.36	0.32 ± 0.17^{3}
	t_0	15.53±1.38	15.62±1.35	15.61±1.14
C18:1n-9	t_3	14.62±1.23 ^{3 a}	15.40±1.57	14.73±1.15 ^{3 a}
	t_6	14.96±1.16 ^{1 a}	15.56 ± 1.49	15.39±1.17
	t_0	0.26±0.22	0.23±0.13	0.24±0.12
C18:2n-6tt	t_3	0.22 ± 0.09	0.19 ± 0.07	0.24±0.09 a
	t_6	0.19 ± 0.08	0.18 ± 0.09	0.20 ± 0.11
	t_0	0.17±0.32	0.19±0.35	0.15±0.21
C18:2n-6ct	t_3	0.11 ± 0.13	0.07 ± 0.07^{-1}	0.07 ± 0.08^{-1}
	t_6	0.04±0.03 ¹	0.05 ± 0.05^{2}	0.04 ± 0.09^{2}
	t_0	0.26±0.34	0.22±0.32	0.21±0.28
C18:2n-6tc	t_3	0.19 ± 0.11	0.19 ± 0.09	0.24 ± 0.28
	t_6	0.14 ± 0.07^{-1}	0.18 ± 0.32	0.11 ± 0.04^{2}
LA	t_0	11.69±1.62	12.23±2.20	12.41±1.98
C18:2n-6	t_3	10.95±1.65 ^{1 a}	12.74±1.75	11.58±1.83 ^{2 a}
010.211 0	t ₆	10.90±1.66 ^{2 a}	12.96 ± 2.36^{-1}	11.86±1.98 ^{1 a}
	t_0	0.20±0.11	0.20±0.10	0.18±0.07
C18:3n-6	t ₃	0.12 ± 0.05^{3} a	0.16 ± 0.06^{-1}	0.12 ± 0.09^{3} a
C10.511 0	t_6	0.09 ± 0.04^{3}	0.13 ± 0.06^{3}	0.09 ± 0.04^{3}
	t_0	0.24±0.08	0.24±0.08	0.24±0.06
C20:1n-9	t_3	0.22 ± 0.06	0.22 ± 0.04	0.24 ± 0.09
020.111)	t_6	0.19 ± 0.05^{-1}	0.25±0.15	0.21 ± 0.06^{2}
ALA	t_0	0.26 ± 0.27	0.22±0.15	0.25±0.20
C18:3n-3	t_3	0.20±0.27 0.22±0.09	0.21 ± 0.07	0.22±0.08
C10.311-3	t_6	0.22 ± 0.09 0.20 ± 0.08	0.21 ± 0.07 0.22 ± 0.08	0.25±0.13
		0.20±0.08 0.27±0.16	0.25±0.14	0.23±0.13 0.27±0.12
C20:2n 6	t_0	0.27 ± 0.16 0.20 ± 0.06^{-1} a	0.23 ± 0.14 0.23 ± 0.05	0.27 ± 0.12 0.24 ± 0.13
C20:2n-6	t_3	U.∠U±U.U0	U.∠3±U.U3	U.24±U.13

	t_6	$0.20\pm0.05^{\ 2}$	0.25±0.10	0.20 ± 0.06^{3}
C20:3n-6	t_0	1.62 ± 0.32	1.66 ± 0.34	1.64 ± 0.32
	t_3	1.62 ± 0.97	$1.87\pm0.53^{\ 2}$	1.40 ± 0.36^{3} a
	t_6	2.22 ± 1.45^{-1}	2.63 ± 1.25^{-3}	2.17±1.48 ¹
AA	t_0	15.40±1.92	16.00±1.95	15.39±1.66
C20:4n-6	t_3	13.62 ± 1.74^{3} a	16.19 ± 1.82	14.14 ± 1.52^{3} a
	t_6	13.20±1.99 ^{3 a}	16.89 ± 2.16^{-1}	13.56±1.68 ^{3 a}
	t_0	0.81 ± 0.86	0.62 ± 0.51	0.64 ± 0.50
C24:0	t_3	0.51 ± 0.22^{-1}	0.56 ± 0.30	0.49 ± 0.23
	t_6	0.80 ± 0.38	0.70 ± 0.34	0.72 ± 0.35
EPA	t_0	1.34 ± 0.63	1.45 ± 0.50	1.45±0.57
C20:5n-3	t_3	$4.36\pm1.13^{3 a b}$	1.20 ± 0.33^{3}	$3.88\pm0.98^{3 a b}$
	t_6	$4.52\pm1.26^{3 a b}$	1.29 ± 0.55^{-1}	$4.10\pm1.30^{3 a b}$
C24:1n-9	t_0	0.66 ± 0.29	0.54±0.22	0.65±0.37
	t_3	0.55 ± 0.25	0.62 ± 0.31	0.57 ± 0.29
	t_6	0.96 ± 0.34^{3}	0.91 ± 0.35^{-3}	0.89 ± 0.42^{-1}
	t_0	2.62±0.66	2.42±0.57	2.52±0.56
C22:4n-6	t_3	1.82 ± 0.50^{3} a	2.59 ± 0.53^{-1}	$1.92\pm0.46^{3 a}$
	t_6	1.59±0.47 ^{3 a}	2.66 ± 0.64^{-1}	1.75±0.52 ^{3 a}
C22:5n-6	t_0	0.49 ± 0.17	0.47±0.17	0.44±0.13
	t_3	0.33 ± 0.11^{3} a	0.52 ± 0.15^{-1}	0.39±0.27 ^a
	t_6	0.31 ± 0.10^{3} a	0.54 ± 0.17^{2}	0.34 ± 0.16^{2a}
C22:5n-3	t_0	2.71±0.38	2.79±0.41	2.82±0.47
	t_3	$3.85\pm0.44^{3 \text{ a}}$	2.87 ± 0.39	3.80 ± 0.37^{3} a
	t_6	$4.04\pm0.57^{3 \text{ a}}$	2.78 ± 0.45	3.86 ± 0.50^{3} a
DHA	t_0	5.67±1.34	5.79±1.09	5.97±1.28
C22:6n-3	t_3	$8.08\pm1.04^{3 a b}$	5.87 ± 1.05	$7.58\pm0.94^{3 a b}$
	t_6	$8.73\pm1.32^{3 a b}$	6.16 ± 1.13^{2}	8.14±1.27 ^{3 a b}
	t_0	7.00±1.90	7.24±1.44	7.42±1.76
omega-3 index	t_3	$12.44\pm1.98^{3 a b}$	7.06 ± 1.29	$11.46\pm1.74^{3 a b}$
	t_6	$13.25\pm2.36^{3 a b}$	7.45±1.57	12.24 ± 2.33^{3ab}

 Abbrevations: rTAG-group, intervention group which consumed omega-3 fatty acids as reesterified triacylglycerides; EE-group, intervention group which consumed omega-3 fatty acids as ethyl-esters; LA, linoleic acid; ALA, α -linolenic acid; AA, arachidonic acid; EPA, eicosapentaenoic acid; DHA, docosahexaenoic acid; omega-3 index, EPA + DHA *mean \pm s.d.

^{1:} Significant changes between baseline and month three or baseline and month six; P<0.05

^{2:} Significant changes between baseline and month three or baseline and month six; P < 0.01

³: Significant changes s between baseline and month three or baseline and month six; P < 0.001 ^{1, 2, 3} Calculated within groups by Student's t test for dependent samples.

^a Values are significantly different between rTAG- or EE-group, respectively, and Placebogroup; *P*<0.05

Values are significantly different between rTAG-group and EE-group; *P*<0.05.

^{a,b} Calculated by ANCOVA with corresponding baseline value as a covariate

Table 3
Characteristics of the 129 participants included in the *per protocol* group *

	rTAG-group	Placebo-group	EE-group
Gender	23/18	16/27	25/20
(male/female)			
Age, years	61.2 ± 9.8	61.7±7.9	59.9 ± 9.0
(mean±SD, range)	33 - 75	34 - 73	34 - 72
Body weight	73.3±11.9	76.1 ± 14.2	77.2 ± 12.9
(mean±SD, range)	51.0 - 103.0	55.0 - 111.5	53.0 - 102.0
BMI, kg/m ²	25.6 ± 2.8	26.1 ± 3.4	25.8 ± 2.9
(mean±SD, range)	20.0 - 30.2	19.9 - 34.4	19.0 - 32.2

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Abbrevations: rTAG-group, intervention group which consumed omega-3 fatty acids as reesterified triacylglycerides; EE-group, intervention group which consumed omega-3 fatty acids as ethyl-esters.

*mean \pm s.d.

512 513 **FIGURES** 514 515 Figure 1: Flow chart for enrolment in and progress through the randomized trial. 516 517 518 519 Figure 2: 520 Omega-3 index at baseline (t₀) and after three (t₃) and six (t₆) months of daily 521 supplementation with either omega-3 fatty acids (n-3 FA) given as re-esterified 522 triacylglycerides (rTAGs), as ethyl-esters (EEs) or with a placebo (- - rTAG-group; --- EE-523 group; — placebo-group). Values are given as mean \pm s.d. 524 *** Significant difference in omega-3 index between rTAG- and EE-group at t₃; P<0.001. 525 ** Significant difference in the omega-3 index between rTAG- and EE-group at t₆; P<0.01. 526 ‡ Significant differences in the omega-3 index between rTAG- and EE-group, respectively, 527 and placebo-group at t_3 and t_6 ; P < 0.001.



