

Low Rate of Virological Failure and Maintenance of Susceptibility to HIV-1 Protease Inhibitors with First-Line Lopinavir/Ritonavir-Based Antiretroviral Treatment in Clinical Practice

Mattia Cf Prosperi, Maurizio Zazzi, Grazia Punzi, Laura Monno, Grazia Colao, Paola Corsi, Simona Di Giambenedetto, Genny Meini, Valeria Ghisetti, Stefano Bonora, et al.

▶ To cite this version:

Mattia Cf Prosperi, Maurizio Zazzi, Grazia Punzi, Laura Monno, Grazia Colao, et al.. Low Rate of Virological Failure and Maintenance of Susceptibility to HIV-1 Protease Inhibitors with First-Line Lopinavir/Ritonavir-Based Antiretroviral Treatment in Clinical Practice. Journal of Medical Virology, 2010, 82 (12), pp.1996. 10.1002/jmv.21927 . hal-00589435

HAL Id: hal-00589435 https://hal.science/hal-00589435

Submitted on 29 Apr 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Journal of Medical Virology

Low Rate of Virological Failure and Maintenance of Susceptibility to HIV-1 Protease Inhibitors with First-Line Lopinavir/Ritonavir-Based Antiretroviral Treatment in Clinical Practice

Journal:	Journal of Medical Virology
Manuscript ID:	JMV-10-1726.R2
Wiley - Manuscript type:	Research Article
Date Submitted by the Author:	08-Jul-2010
Complete List of Authors:	Prosperi, Mattia; Catholic University of Sacred Heart, Clinic of Infectious Diseases Zazzi, Maurizio; University of Siena, Molecular Biology Punzi, Grazia; Bari University Hospital, Microbiology and Virology Monno, Laura; Bari University Hospital, Clinical Infectious Diseases Colao, Grazia; Careggi University Hospital, Microbiology and Virology Corsi, Paola; Careggi University Hospital, Unit of Infectious Diseases Di Giambenedetto, Simona; Catholic Universisty of Sacred Heart, Clinic of Infectious Diseases Meini, Genny; University of Siena, Molecular Biology Ghisetti, Valeria; A. Savoia Hospital, Microbiology and Virology bonora, stefano; university of Torino, infectiuos diseases Pecorari, Monica; Modena University Hospital, Microbiology and Virology Gismondo, Maria Rita; L. Sacco University Hospital, Microbiology and Virology Carli, Tiziana; Grosseto General Hospital, Unit of Infectious Diseases De Luca, Andrea; Sacro Cuore Catholic University, Institute of Infectious Diseases; University Hospital of Siena, Infectious Diseases
Keywords:	human immunodeficiency virus type 1, lopinavir/ritonavir, first-line antiretroviral therapy, genotypic resistance, virologic failure



|--|

			>50 cop	>50 copies/ml			>500 copies/ml		
factor			RH	95% CI	p-value	RH	95% CI	p-value	
year of birth (pe	er 1-year incre	ease)	0.994	(0.958-1.03)	0.725	1.024	(0.983-1.066)	0.264	
		male vs. female	0.649	(0.298-1.415)	0.277	0.575	(0.248-1.336)	0.198	
gender		unknown vs. female	1.041	(0.128-8.463)	0.970	1.374	(0.17-11.079)	0.765	
		homo/bisexual vs. heterosexual	1.264	(0.524-3.046)	0.602	1.368	(0.53-3.534)	0.517	
		intravenous drug user vs. heterosexual	0.512	(0.105-2.498)	0.408	1.519	(0.432-5.346)	0.515	
mode of HIV tra	Insmission	other/unknown vs. heterosexual	1.236	(0.558-2.737)	0.602	1.156	(0.477-2.802)	0.748	
calendar year o	f HAART initi	ation (per more recent)	0.919	(0.719-1.174)	0.497	0.978	(0.733-1.304)	0.878	
	abacav	ir+lamivudine vs. tenofovir+emtricitabine	0.362	(0.079-1.65)	0.189	0.197	(0.025-1.533)	0.12	
	zidovuo	dine+lamivudine vs. tenofovir+emtricitabine	1.142	(0.47-2.773)	0.769	0.991	(0.375-2.619)	0.986	
backbone	other b	ackbone vs. tenofovir+emtricitabine	1.592	(0.403-6.299)	0.507	1.936	(0.402-9.323)	0.410	
HAART	tenofov	ir+lamivudine vs. tenofovir+emtricitabine	0.275	(0.056-1.361)	0.114	0.754	(0.206-2.767)	0.67	
baseline CD4+ cell count cells/mm3 (per log higher)		0.844	(0.656-1.087)	0.190	0.892	(0.679-1.173)	0.414		
baseline HIV-RNA load cp/ml (per log10 higher)		0.844	(0.529-1.348)	0.479	1.790	(1.097-2.921)	0.020		
lopinavir GRS (per 10 points higher)		0.975	(0.912-1.042)	0.450	0.945	(0.89-1.003)	0.064		
backbone GRS (per 10 points higher)		1.002	(0.979-1.026)	0.849	1.038	(1.017-1.06)	0.0003		

 Table 1b.
 Multivariable Cox regression analysis: predictors of virological failure beginning from month 3 after treatment initiation (n=548).

			>50 cop	>50 copies/ml			>500 copies/ml		
Factor			RH	95% CI	p-value	RH	95% CI	p-value	
year of birth (pe	er 1 year incre	ase)	1.018	(0.991-1.045)	0.2012	1.032	(1.002-1.063)	0.036	
		male vs. female	1.233	(0.679-2.239)	0.4915	0.918	(0.504-1.67)	0.778	
gender		unknown vs. female	1.497	(0.339-6.605)	0.5941	0.468	(0.06-3.63)	0.467	
		homo/bisexual vs. heterosexual	0.875	(0.468-1.634)	0.6751	0.827	(0.407-1.678)	0.598	
		intravenous drug user vs. heterosexual	0.202	(0.046-0.892)	0.0348	1.065	(0.433-2.621)	0.890	
mode of HIV transmission other/unknown vs. heterosexual		0.935	(0.517-1.69)	0.8228	1.223	(0.673-2.225)	0.509		
calendar year of HAART initiation (per more recent)		0.974	(0.805-1.178)	0.7859	1.101	(0.903-1.343)	0.340		
	abacav	ir+lamivudine vs. tenofovir+emtricitabine	0.255	(0.077-0.849)	0.0259	0.111	(0.015-0.824)	0.03	
	zidovuc	line+lamivudine vs. tenofovir+emtricitabine	0.895	(0.467-1.716)	0.7386	1.138	(0.585-2.212)	0.704	
backbone HAART	other ba	ackbone vs. tenofovir+emtricitabine	1.546	(0.505-4.737)	0.4457	2.015	(0.646-6.291)	0.22	
	tenofov	ir+lamivudine vs. tenofovir+emtricitabine	0.897	(0.397-2.029)	0.7948	1.199	(0.485-2.967)	0.693	
baseline CD4+ cell count cells/mm3 (per log higher)		0.875	(0.732-1.047)	0.145	0.967	(0.795-1.176)	0.73		
baseline HIV-RNA load cp/ml (per log10 higher)			1.127	(0.799-1.589)	0.4968	1.901	(1.312-2.754)	0.00	
lopinavir GRS (per 10 points higher)			0.997	(0.958-1.039)	0.9007	0.999	(0.966-1.032)	0.93	
backbone GRS (per 10 points higher)			1.001	(0.983-1.02)	0.8793	1.021	(1.005-1.037)	0.01	

RH = relative hazard; HAART = highly active antiretroviral therapy; GRS = Stanford's 6.0.1 genotypic resistance score

Table 2. Evolution of protease and reverse transcriptase resistance mutations under lopinavir/ritonavir plus 2 nucleoside/tide reverse transcriptase inhibitors pressure

 in 36 previously naive patients with available baseline and follow-up HIV-1 genotype. Substitutions emerging during treatment are in bold.

patient	nrti ^a	Baseline HIV-RNA load Log10 copies/ml	Baseline genotype ^b	Follow-up genotype ^b	LPV/rtv susceptbility (baseline/follow-up)°
1	3TC TDF	6.76	PR: 63P RT:	PR: 63P RT: 41L 70R 101Q 103N 181C 210W 215F 215Y 219E 221Y 225H 238T	S/S
2	FTC TDF	5.72	PR: 63P RT:	PR: 63P RT: 184 I	S/S
3	3TC TDF	5.18	PR: 10F 20T 46I 54V 63P 71T 73T 84V 90M RT: 41L 70R 74V 103N 118I 184V 215F 219Q 228H	PR: 10F 20T 46I 54V 63P 71T 73T 84V 90M RT: 41L 70R 74V 103N 118I 184V 215F 219Q 228H	1/1
4	FTC TDF	5.53	PR: 16E 20R 36I RT:	PR: 16E 20R 36I RT:	S/S
5	3TC AZT	5.12	PR: 36I 60E RT: 101Q	PR: 36I 60E RT: 101Q 184V	S/S
6	FTC TDF	5.70	PR: 20M 77I RT:	PR: 77I RT:	S/S
7	FTC TDF	4.93	PR: 13V 63P 77I RT:	PR: 13V 63P 77I RT:	S/S
8	FTC TDF	5.86	PR: 10I 11I 20V 32I 33F 36I 43T 63P 71V 73S 82A 89V 90M RT: 215D	PR: 10I 20V 32I 33F 36I 43T 46I 47V 54V 63P 71V 73S 82A 89V 90M RT: 44D 118I 184V 215Y	R/R
9	FTC TDF	6.25	PR: 36I 63P RT:	PR: 63P RT:	S/S
10	3TC AZT	5.17	PR: 63P 71T RT:	PR: 63P 71T RT:	S/S
11	FTC TDF	5.70	PR: 10V 13V 16E 20M 35G 36L RT:	PR: 10V 13V 16E 20M 35G 36L RT:	S/S
12	FTC TDF	4.77	PR: 821 RT:	PR: 361 821 RT:	S/S
13	FTC TDF	5.70	PR: 63P 77I RT: 333D	PR: 63P 77I RT: 333D	S/S
14	3TC AZT	4.17	PR: 36I 93L RT:	PR: 36I 63P 93L RT:	S/S
15	3TC AZT	4.18	PR: 10I 13V 36I 58E 60E RT: 103N 184I 184V	PR: 10I 13V 36I 58E 60E RT: 103N 184V	S/S
16	D4T DDI	5.17	PR: 10I 46I 60E 63P 73T 77I 84V 90M 93L RT: 41L 67N 101E 181C 190A 215Y 219Q 228H	PR: 10F 20I 36I 46I 54V 60E 63P 73T 84V 90M 93L RT: 41L 67N 101E 108I 118I 181C 190A 210W 215Y 219R 221Y	1/1

17	3TC D4T	4.56	PR: 10F 33I 35G 36I 46I 63P 71V 73S 83D 84V 90M RT: 41L 44D 67N 74I 74V 103N 118I 181C 210W 215Y 219R	PR: 10F 20V 36I 46I 54V 63P 71V 73S 84V 90M RT: 41L 44D 67N 74V 101Q 118I 184V 210W 215Y 219R	1/1
			PR: 13V 16A 58E 60E 77I	PR: 13V 16A 58E 60E 77I	
18	3TC AZT	5.31	RT: 179D	RT: 179D	S/S
			PR: 60E 63P 71T 77I	PR: 60E 63P 71T 77I	
19	3TC AZT	5.26	RT: 106I 138G 179I	RT: 1061 138G	S/S
			PR: 10F 35D 36I 54V 63P 71I 84V 93L	PR: 10F 35D 36I 46I 54V 63P 71I 84V 93L	
20	D4T TDF	5.17	RT: 41L 67G 74V 101P 184V 210W 215Y	RT: 41L 67G 74V 101P 210W 215Y	1/1
21	3TC AZT	5.69	PR: 71T 77I	PR: 35D 71T 77I	S/S
21	STC AZT	5.69	RT:	RT:	3/3
00	D4T TDF	5.46	PR: 10F 63P 77I	PR: 63P	S/S
22	D41 IDF	5.40	RT:	RT:	5/5
00	070 477	F 47	PR: 63P 77I 93L	PR: 63P 77I 93L	0/0
23	3TC AZT	5.17	RT:	RT:	S/S
~ /			PR: 36I 63P	PR: 36I 63P	2/2
24	3TC TDF	5.03	RT: 103R	RT: 103R	S/S
			PR: 201 361	PR: 10F 13V 20I 20M 36I 63P 77I 82I	0.10
25	3TC TDF	5.70	RT:	RT: 184V 215C 215S 215Y	S/S
			PR: 13V	PR: 13V	
26	3TC AZT	5.00	RT:	RT:	S/S
			PR: 10V 20V 36I 47V 54V 63P 71V 73S 82A 89V 90M	PR: 10V 20V 36I 47V 54V 63P 71V 73S 82A 89V 90M	
27	3TC TDF	5.56	RT: 215F 215S	RT: 215F 215S 215Y	R/R
			PR: 20R 35D 36I	PR: 20R 35D 36I	
28	3TC AZT	4.59	RT:	RT:	S/S
			PR: 13V 20I 35N 36L 60E 63P	PR: 10I 13V 20I 36L 60E 63P	
29	3TC ABC	5.85	RT:	RT:	S/S
			PR: 63P	PR: 63P	
30	FTC TDF	5.31	RT:	RT:	S/S
			PR:	PR:	
31	3TC AZT	4.20	RT: 67N 219Q	RT: 67N 219Q	S/S
32	FTC TDF	5.44	PR: 63P 93L	PR: 63P 93L	S/S
	-		RT:	RT:	
33	FTC TDF	7.00	PR: 10V 13V	PR: 10V 13V	S/S
			RT:	RT:	
34	FTC TDF	5.81	PR: 60E 63P	PR: 60E 63P	S/S
			RT:	RT:	
35	FTC TDF	5.17	PR: 63P 71V 93L	PR: 71V 93L	S/S
50		0.17	RT: 69A 179I	RT:	0,0
36	FTC TDF	4.91	PR: 10I 46L 93L	PR: 10I 46L 93L	S/S
00		7.31	RT: 118I 333E	RT: 118I	5,5

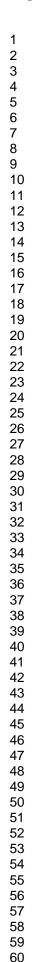
Journal of Medical Virology

^a3TC, lamivudine; ABC, abacavir; AZT, zidovudine; D4T, stavudine; DDI, didanosine; FTC, emtricitabine; TDF, tenofovir.

^bPR, protease; RT, reverse transcriptase.

°S, susceptible (Stanford's score <=14); I, intermediate resistance (Stanford's score from 15 to 59); R, high-level resistance (Stanford's score >=60).

Figure 1. Kaplan-Meier curves of survival rates with respect to a virological failure at or after 3 or 6 months using different HIV-RNA thresholds (>1000, >500 and >50 cp/ml), to a Stanford's lopinavir genotypic resistance score >14 (intermediate/full resistance), to an accumulation of at least one major IAS resistance mutation to lopinavir/ritonavir, and to a therapy discontinuation (at any time for any reason) end-point.



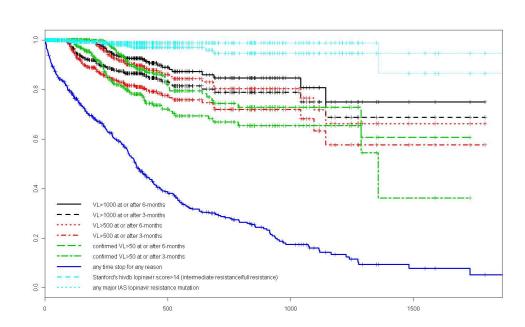


Figure 1. Kaplan-Meier curves of survival rates with respect to a virological failure at or after 3 or 6 months using different HIV-RNA thresholds (>1000, >500 and >50 cp/ml), to a Stanford's lopinavir genotypic resistance score >14 (intermediate/full resistance), to an accumulation of at least one major IAS resistance mutation to lopinavir/ritonavir, and to a therapy discontinuation (at any time for any reason) end-point.

370x244mm (96 x 96 DPI)

Low Rate of Virological Failure and Maintenance of Susceptibility to HIV 1 Protease Inhibitors with First-Line Lopinavir/Ritonavir-Based Antiretroviral Treatment in Clinical Practice

Mattia CF Prosperi¹, Maurizio Zazzi², Grazia Punzi³, Laura Monno⁴, Grazia Colao⁵, Paola Corsi⁶, Simona Di
 Giambenedetto¹, Genny Meini², Valeria Ghisetti⁷, Stefano Bonora⁸, Monica Pecorari⁹, Maria Rita
 Gismondo¹⁰, Patrizia Bagnarelli¹¹, Tiziana Carli¹², and Andrea De Luca^{1,13}, for the ARCA Collaborative Group.

- 12 ⁸ 13 9 ¹Clinic of Infectious Diseases, Catholic University of Sacred Heart, Rome, Italy
- 14 10 ²Department of Molecular Biology, University of Siena, Siena, Italy
- 15 11 ³Microbiology and Virology, Bari University Hospital, Bari, Italy
- ¹⁶ 12 ⁴Clinical Infectious Diseases, Bari University Hospital, Bari, Italy
- ¹⁷ 13 ⁵Microbiology and Virology, Careggi University Hospital, Florence, Italy
- 19 14 ⁶Unit of Infectious Diseases, Careggi University Hospital, Florence, Italy
- 20 15 ⁷Microbiology and Virology, A. Savoia Hospital, Turin, Italy
- 21 16 ⁸Unit of Infectious Diseases, A. Savoia Hospital, Turin, Italy
- ²² 17 ⁹Microbiology and Virology, Modena University Hospital, Modena, Italy
- ²³ 18 ¹⁰Microbiology and Virology, L. Sacco University Hospital, Milan, Italy
- ²⁴ 19 ¹¹Microbiology and Virology, Ancona University Hospital, Ancona, Italy
- 20 12 Unit of Infectious Diseases, Grosseto General Hospital, Grosseto, Italy
- ¹³Infectious Diseases Unit, Siena University Hospital, Siena, Italy
 22
- 29 23 Correspondence to:

12

3

4

5

6 7

8

9

10

11

36 2937 30

38

- 30 24 Mattia Prosperi, PhD
- ³¹ 25 Clinic of Infectious Diseases
- 32 26 Catholic University of Sacred Heart
- 33 27 Largo F. Vito, 1 00168 Rome, Italy
- 3528E-mail: ahnven@yahoo.it
 - 31 **Running head:** First-line lopinavir/ritonavir HAART

Abstract

Protease inhibitor (PI)-resistant HIV-1 has hardly ever been detected at failed boosted PI-based first-line antiretroviral regimens in clinical trials. However, this phenomenon has not been investigated in clinical practice. To address this gap, data from patients starting a first-line lopinavir/ritonavir (LPV/rtv)-based therapy with available baseline HIV-1 RNA load, a viral genotype and follow-up viral load after 3 months and 6 months of treatment were extracted from the Italian Antiretroviral Resistance Cohort Analysis (ARCA) observational database. Based on survival analysis, 39 (7.1%) and 43 (7.8%) of the 548 examined patient cases had an HIV-1 RNA >500 and >50 copies/ml, respectively, after 6 months of treatment. Cox proportional hazard models detected baseline HIV-1 RNA (RH 1.79, 95%CI 1.10-2.92 per 1-log₁₀ increase, P = 0.02) and resistance to the nucleoside backbone (RH 1.04, 95%CI 1.02-1.06 per 10-point increase using the Stanford HIVdb algorithm, P < 0.001) as independent predictors of HIV-1 RNA at >500 copies/ml, but not at the >50 copies/ml cutoff criteria. Higher baseline viral load, older patient age, heterosexual route of infection and use of tenofovir/emtricitabine were predictors of failure at month 3 using the 50-copy and/or 500-copy threshold. Resistance to LPV/rtv did not occur or increase in any of the available 36 follow-up HIV-1 genotypes. Resistance to the nucleoside backbone (M184V) developed in four cases. Despite the likely differences in patient population and adherence, both the low rate of virological failure and the lack of development of LPV/rtv 50 51 resistance documented in clinical trials are thus confirmed in clinical practice.

Key Words

60 55 human immunodeficiency virus type 1; boosted protease inhibitor; lopinavir/ritonavir; first-line antiretroviral therapy; antiretroviral drug resistance; virologic failure.

Introduction

7

Ritonavir-boosted protease inhibitors (PIs) have been used successfully for years and still remain a key component of highly active antiretroviral therapy (HAART). A major advantage of boosted PIs in first-line therapy is the very low risk of selecting drug-resistant HIV-1 variants at virological failure, as shown in multiple clinical trials [Bartlett et al., 2006; Gupta et al., 2008; Lima et al., 2008; Molina et al., 2008; Ortiz et al., 2008; Mills et al., 2009]. This feature is, in fact, the main argument made by experts favouring boosted PI over non-nucleoside reverse transcriptase inhibitor (NNRTI)-based first-line therapy because NNRTIs select typically for NNRTI-resistant virus at failure [Riddler et al., 2008]. In addition, resistance to the backbone nucleoside reverse transcriptase inhibitors (NRTIs) occurs less frequently with a boosted PI compared to an NNRTI-based therapy [Soulié et al., 2009].

While the latest treatment guidelines recommend other boosted PIs for first-line HAART [DHHS, 2009], lopinavir/ritonavir (LPV/rtv) has been the most widely used PI owing to its long-standing availability, unique heat-stable co-formulation, efficacy and safety profiles [Oldfield et al., 2006]. In order to evaluate virological response to first-line LPV/rtv-based therapy and confirm its low propensity for selecting resistance mutations in HIV-1 protease in clinical practice, a targeted analysis was carried out using the Italian Antiretroviral Resistance Cohort Analysis (ARCA) observational database. ARCA (www.hivarca.net) is a nationwide initiative started in 2002 which 50 78 retrospectively and prospectively collates clinical and virological data from HIV-1-infected patients followed at 105 centres. The data include demographics, hepatitis B and C status, AIDS-defining 55 80 events, antiretroviral treatment, viral load, CD4+ T cell counts, HIV-1 genotype. The oldest HIV-1 genotype entry is dated back to 1991. At the time of this study, data from 19,984 patients were 60 82 available.

Patients and Methods

First-line treatments comprised of LPV/rtv plus two NRTIs were extracted from the ARCA database. All patients with available HIV-1 genotype and HIV-1 RNA load at baseline were included and all HIV-1 RNA load data available from the treatment start date to the latest follow-up time were collected. The rate of treatment failure and development of drug resistance were evaluated by survival analyses using the cases with at least one follow-up HIV-1 RNA load measurement obtained after 3 months and after 6 months of uninterrupted treatment. Information on HIV-1 genotype obtained between 30 days after treatment start and 90 days after LPV/rtv discontinuation was also included, if no other PI was administered after LPV/rtv.

The Stanford HIVdb version 6.0.1 (http://hivdb.stanford.edu/) algorithm was used to calculate genotypic resistance scores (GRS). HIVdb scores below 10 are classified as full drug 32 97 susceptibility, from 10 to 14 as potential low-level resistance, from 15 to 29 as low-level resistance, from 30 to 59 as intermediate resistance and >59 as high-level resistance. This five-level system was converted to a three-category classification by grouping scores up to 14 as susceptibility, 15 to 59 as intermediate resistance and >59 as complete resistance. A continuous ⁴²101 numeric GRS scale was used in the multivariable analysis in order to minimise the number of 45 102 parameters to be optimised: results are showed per 10 points higher, which roughly corresponds 48 103 to a one-category increase in the Stanford HIVdb categorisation.

50 104 Survival analysis using the Kaplan-Meier method and Cox proportional hazard models was ⁵² 105 performed using the following outcome measures: having an HIV-1 RNA load >500 and >50 copies/ml at 3 months after treatment initiation; an HIV-1 RNA load >500 and >50 copies/ml at 6 58 107 months after treatment initiation; intermediate or full LPV/rtv resistance defined as a GRS >14; and the presence of at least one major LPV/rtv resistance mutation according to the 2009 IAS-USA list (i.e. V32I, I47A/V or V82A/F/S/T) [Johnson et al., 2009]. Cases not reaching the specific

2 110 endpoint were censored at the regimen discontinuation date or at the latest available HIV-1 RNA or HIV-1 genotype if the regimen was not discontinued. After adjusting for the other explanatory variables, the Cox model provides an estimate of the treatment effect on the remaining data with no outcome event. In addition, the model allows estimating the outcome event hazard for each individual, based on his/her baseline variables.

Results

1 2 117

The ARCA database contained 1,141 patients who were treated with first line LPV/rtvcontaining HAART regimens. Of these, 549 (48%) did not have a baseline genotype and were excluded from the analysis. The remaining 592 patients had baseline median (IQR) CD4+ T cell counts and HIV-1 RNA load of 127 (48-255) cells/mm³ and 5.20 (4.80-5.60) log₁₀ copies/ml, respectively. The median (IQR) calendar year of LPV/rtv initiation was 2006 (2004-2007). The NRTI backbones employed most frequently were tenofovir plus emtricitabine (37.5%), zidovudine plus lamivudine (34.3%), abacavir plus lamivudine (12.2%), and tenofovir plus lamivudine (10.6%).

At baseline, 15 (2.5%) and 1 (0.2%) patients presented with a LPV/rtv intermediate and resistant GRS, respectively. The prevalence of genotypes carrying at least one major IAS mutation associated to LPV/rtv resistance was 7.3% (n=43). Intermediate and full resistance to at least one NRTI were detected in 5.4% and 2.5% of the cases, with 1.8% showing intermediate and 5.7% full resistance to at least one NNRTI; and 1.8% and 2.0% to at least one PI, respectively. When looking at the GRS of the treatment provided, the prevalence of regimens with all fully active drugs (i.e. no drug with a GRS >14) was 93%, while 2.9% of regimen were composed by either at least one drug with full resistance or two drugs with intermediate resistance. Ten (1.7%) genotypes were found with both intermediate-level resistance to LPV/rtv and to at least one of the other drugs in the backbone.

Forty-four (7.4%) of the 592 patients had not have a follow-up HIV-1 RNA measurement after 3 or 6 months hence were excluded from subsequent incidence and survival analyses. Following 3- and 6 months therapy, 74 (13.5%) and 39 (7.1%) patients had an HIV-1 RNA >500 copies/ml, respectively. Using the more stringent criterion, the number of cases with a confirmed HIV-1 RNA >50 copies/ml after 3 and 6 months were 77 (14%) and 43 (7.8%), respectively.

1

According to Kaplan-Meier analysis (Fig. 1) the estimated proportion of patients with an HIV-1 RNA below 500 copies/ml was 0.90 (95% Cl 0.86-0.94) at one year and 0.80 (0.74-0.87) at two years, with an incidence rate of 0.23 (0.17-0.32) per 1,000 person-years of follow-up. With the 50-copy threshold, the probability to remain virological failure-free was 0.90 (0.85-0.93) at one year and 0.73 (0.65-0.81) at two years, with an incidence rate of 0.26 (0.19-0.35) per 1,000 personyears of follow-up. The incidence rate of therapy discontinuation for any reason was 1.8 (1.7-2.1) per 1,000 person-years of follow-up. The probability of continuing the first-line LPV/rtv-based regimen was 0.51 (0.46-0.56) at one year and 0.28 (0.24-0.34) at two years. Notably, 26% of the 324 LPV/rtv therapy discontinuations at any follow-up time were owing to toxicity, 17% to treatment simplification, 3% to adherence problems, whereas 33% were on other grounds and 21% were stopped owing to unspecified reasons.

Multivariable Cox analysis showed that at 6 months a higher baseline HIV-1 RNA load and a higher NRTI backbone GRS were independent predictors of having viremia at >500 copies/ml, but not at >50 copies/ml (Table 1a). Factors predictive of virological failure at 3 months using the 500copy and/or the 50-copy threshold were higher baseline viral load, older patient age, use of tenofovir/emtricitabine vs. abacavir/lamivudine and heterosexual vs. intravenous route of infection (Table 1b). However, drug users had less frequent visits, thus viral load monitoring, compared to other transmission groups (not shown). This may explain the apparently lower risk of short-term failure for drug users.

Follow-up of HIV-1 genotype was available for a total of 36 patients. Mutations associated with decreasing susceptibility to NRTIs, NNRTIs and PIs were detected at baseline in seven, five and six patients, respectively, with three patients harboring triple-class resistant virus. Overall, genotyping was carried out after a median of 220 days (IQR 102-348) from LPV/rtv initiation. Median HIV-1 RNA and CD4+ T cell count at the time of follow-up genotype were 3.64 (IQR 2.56-4.67) log₁₀ copies/ml and 207 (120-432) cells/mm³. Eight (22.2%) viral genotype tests were Page 17 of 22

Journal of Medical Virology

performed after LPV/rtv discontinuation. Paired analysis of the baseline and follow-up genotypes revealed very limited selection of protease mutations (Table 2). There were no changes in Stanford HIVdb LPV/rtv susceptibility category in any case. Among the 30 patients with a LPV/rtv-sensitive baseline genotype, only one was found with more than one additional minor PI mutation (patient 25: L10F, I13V, L63P, V77I, V82I) but this did not have any impact on the predicted LPV/rtv susceptibility. In some other cases, the baseline and follow-up genotype differed for one minor mutation, either acquired or lost. Resistance mutation I54V, contributing to resistance to LPV/rtv, was observed in three cases with pre-existing LPV/rtv resistance. In one of the four cases harboring a virus populations with intermediate resistance to LPV/rtv at baseline (patient 20), the major PI mutation M46I was identified at the follow-up stage. Moreover, major LPV/rtv mutations M46I and I47V were found in one of the two patients (patient 8) with a virus that was already highly resistant to LPV/rtv at baseline. There were four cases with NRTI resistance mutations, namely M184I/V with or without a more extensive set of NRTI resistance mutations. In one of these patients, the follow-up HIV-1 genotype also contained NNRTI resistance mutations as a result of NNRTI therapy following discontinuation of LPV/rtv. Overall, 30 of the 36 (83.3%) followup genotypes were susceptible to LPV/rtv and to at least another PI, 33 (91.7%) were susceptible to NNRTIs and to at least two NRTIs.

184 Discussion

1 2

3 4

8 9

11

21

31

186 The use of boosted PIs as first-line HAART has been shown to be a valuable treatment 10¹⁸⁷ option in clinical trials with virological failure being comparable or slightly higher than with NNRTI-12188 based therapy [Riddler et al., 2008; Soriano et al., 2009; Daar et al., 2010]. However the residual 13 14 15¹⁸⁹ number of effective drugs at failure has been generally far higher with boosted PIs [Bartlett et al., 16 17 190 2006; Gupta et al., 2008; Lima et al., 2008]. Although the underlying mechanism is not completely 18 19 20 191 understood, it has been acknowledged that not only boosted PI failures devoid any major protease ²²192 mutations but also resistance to the NRTI backbone is typically minimized by an apparently 23 24 25193 increased genetic barrier to resistance. Notwithstanding the excellent performance in clinical 26 27 194 trials, evaluation of virological response and development of drug resistance with first-line 28 29 ₃₀195 boosted PI treatment in the different context of clinical practice is advisable. Larger heterogeneity ³²196 and lower adherence rate of the patient population may introduce variables affecting the success 33 34 35 197 of a treatment regimen based mostly on a high genetic barrier. Moreover, genotypic resistance 36 37₁₉₈ assays in clinical trials are performed typically at the first sign of virological failure, while they may 38 39 40 199 be delayed in clinical practice owing to practical constraints. 41

⁴²200 The analysis of a large multicentric observational cohort confirms both a very low rate of 43 44 45201 virologic failure, which is in some cases driven by pre-existing resistance, and a negligible selection 46 47 48²⁰² of drug resistance in patients undergoing a LPV/rtv-based first-line regimen. The results shown 49 50203 with respect to the 6-month follow-up, which is more relevant in the current clinical settings, and 51 ⁵²204 for the 3-month end-point, which could be of interest in the evaluation of the predictors of early 53 54 55205 failure. Although the number of follow-up genotypes was limited, clinical trials of drug-naive 56 57 58²⁰⁶ patients have generated comparably low numbers of genotypic information at failure [Gupta et 59 60207 al., 2008]. In addition, patients with transmitted PI-resistant virus are generally excluded from 208 boosted PI-based first-line treatment trials while such patients are encountered in clinical practice

Journal of Medical Virology

[Geretti, 2007] and may benefit from boosted PI therapy. Interestingly, it was observed that decreased susceptibility to the NRTI backbone appeared to compromise treatment more than decreased susceptibility to LPV/rtv. This highlights the potency of this high-genetic barrier PI, 10²¹² although the low number of cases with resistance at baseline may have hampered the detection of minor effects. According to the genotypic susceptibility profile at failure, for the vast majority of patients a fully active regimen can be still based on a boosted PI or consisting of two NRTI plus one NNRTI. 20216 While novel drug classes are being considered for convenient and potentially less toxic first-line treatment, boosted PIs, particularly LPV/rtv, maintain an excellent and hardly achievable efficacy record of in terms of suppression of virus replication and limitation of drug resistance evolution. In 27 219 addition, recent trial data suggest better tolerability and/or higher potency for the currently recommended boosted PIs compared to LPV/rtv [Madruga et al., 2007; Molina et al., 2008]. While 32₂₂₁ this strengthens the importance of the PI class as a cornerstone in HAART, future analysis of large 35²²² cohort data will be required to confirm such advantages in clinical practice. ³⁸223

224 Acknowledgements

16234

1 2

226 This work was supported, in part, by an unrestricted educational grant from Abbott Virology to 6 7 227 ARCA. The sponsor played no role in study design, data analysis and manuscript revision. ARCA is 8 228 supported by educational grants by Abbott, Boehringer-Ingelheim, Bristol-Myers Squibb, 9 10229 GlaxoSmithKline, Gilead Sciences, Janssen-Cilag Tibotec division. The study was supported partly 11230 by the European Community's Seventh Framework Programme (FP7/2007-2013) under the project 12231 "Collaborative HIV and Anti-HIV Drug Resistance Network (CHAIN)" - grant agreement number ¹³232 223131. We are very grateful to Andrea Petroczi (Kingston University, UK) for critical reading of 14 15233 the manuscript.

17 235 The ARCA collaborative group

18236 Andrea Giacometti (ANCONA - Clinica di Malattie Infettive), Luca Butini (ANCONA - Immunologia Clinica), Romana del 19237 Gobbo (ANCONA - Malattie Infettive), Stefano Menzo (ANCONA - Virologia), Danilo Tacconi (AREZZO - Malattie 20238 Infettive), Giovanni Corbelli (ASCOLI PICENO - Malattie Infettive), Stefania Zanussi (AVIANO - Centro di Riferimento 21 2 39 Oncologico), Laura Monno (BARI - Clinica Malattie Infettive Università), Grazia Punzi (BARI - Virologia), Franco ²²240 23241 Maggiolo (BERGAMO - Malattie Infettive), Annapaola Callegaro (BERGAMO - Microbiologia e Virologia), Leonardo Calza (BOLOGNA - Malattie Infettive S. Orsola), Maria Carla Re (BOLOGNA - UO Microbiologia, Lab. Retrovirus), 23 241 24 242 25 243 26 244 27 245 28 246 29 246 29 246 30 247 30 247 31 248 32 249 Raffaele Pristerà (BOLZANO - Malattie Infettive), Paola Turconi (BRESCIA - Fleming Labs), Antonella Mandas (CAGLIARI - Centro S.I.D.A., Policlinico Universitario), Sauro Tini (CITTA' DI CASTELLO - Medicina Generale), Alessia Zoncada (CREMONA - Malattie Infettive), Elisabetta Paolini (CREMONA - Servizio Immunoematologia e Medcina Trasfusionale), Giorgio Amadio (FERMO - Malattie Infettive), Laura Sighinolfi (FERRARA - Malattie Infettive AOU S. Anna), Giuliano Zuccati (FIRENZE - Centro MTS), Massimo Morfini (FIRENZE - Ematologia CAREGGI), Roberto Manetti (FIRENZE -Immunoallergologia CAREGGI), Paola Corsi (FIRENZE - Malattie Infettive CAREGGI), Luisa Galli (FIRENZE - Malattie Infettive Pediatria Meyer), Massimo Di Pietro (FIRENZE - Malattie Infettive SM Annunziata), Filippo Bartalesi (FIRENZE -Malattie Infettive Università), Grazia Colao (FIRENZE - Virologia CAREGGI), Andrea Tosti (FOLIGNO - Malattie Infettive / 33²⁵⁰ SERT), Antonio Di Biagio (GENOVA - Clinica Malattie Infettive AOU S. Martino), Maurizio Setti (GENOVA - Clinica 34251 Medica Immunologia), Bianca Bruzzone (GENOVA - Laboratorio di Igiene Ospedale S. Martino), Giovanni Penco 35252 (GENOVA - Malattie Infettive Ospedali Galliera), Michele Trezzi (GROSSETO - Malattie Infettive), Anna Orani (LECCO -36253 Malattie Infettive), Riccardo Pardelli (LIVORNO - Malattie Infettive), Michele De Gennaro (LUCCA - Malattie Infettive), 37254 Alessandro Chiodera (MACERATA - Malattie Infettive), Alfredo Scalzini (MANTOVA - Malattie Infettive Ospedale 'C. 38255 Poma'), Loredana Palvarini (MANTOVA - Virologia), Paolo Almi (MASSA - Malattie Infettive), Giovanni Todaro 39256 (MESSINA - Malattie Infettive), Antonella d'Arminio Monforte (MILANO - Clinica Malattie INfettive AO S. Paolo), Paola 40257 Cicconi (MILANO - Clinica di Malattie Infettive Ospedale S. Paolo), Stefano Rusconi (MILANO - Dipart. Scienze Cliniche, 41258 Sez. Malattie Infettive - Università degli Studi), Maria Rita Gismondo (MILANO - Laboratorio Microbiologia Ospedale L. 42259 Sacco (Dipart. Scienze Cliniche, Sez. Malattie Infettive)), Maria Rita Gismondo (MILANO - Laboratorio Microbiologia 43260 Ospedale L. Sacco (Prima Divisione Malattie Infettive)), Valeria Micheli (MILANO - Laboratorio Microbiologia Ospedale 44 261 L. Sacco (Seconda Divisione Malattie Infettive)), Maria Luisa Biondi (MILANO - Laboratorio di diagnostica molecolare 45262 infettivologica AO S. Paolo), Nicola Gianotti (MILANO - Malattie Infettive San Raffaele), Amedeo Capetti (MILANO -46263 Prima Divisione Malattie Infettive Ospedale L. Sacco), Paola Meraviglia (MILANO - Seconda Divisione Malattie Infettive 47 264 48 265 49 266 50 267 51 268 53 269 53 269 54 270 55 271 56 272 57 273 Ospedale L. Sacco), Enzo Boeri (MILANO - Virologia HSR), Cristina Mussini (MODENA - Clinica Malattie Infettive), Monica Pecorari (MODENA - Virologia), Alessandro Soria (MONZA - Malattie Infettive), Laura Vecchi (MONZA - UO Microbiologia AO S. Gerardo), Maurizio Santirocchi (NARNI - SERT), Diego Brustia (NOVARA - Malattie Infettive AO Maggiore), Paolo Ravanini (NOVARA - Virologia), Federico Dal Bello (PADOVA - Virologia), Nino Romano (PALERMO -Centro Riferimento AIDS Università), Salvatrice Mancuso (PALERMO - Servizio Riferimento Regionale Diagnosi AIDS), Carlo Calzetti (PARMA - Divisione Malattie Infettive ed Epatologia Azienda Ospedaliera), Renato Maserati (PAVIA -Ambulatorio Clinica Malattie Infettive S. Matteo), Gaetano Filice (PAVIA - Clinica Malattie Infettive e Tropicali), Fausto Baldanti (PAVIA - Virologia S. Matteo), Daniela Francisci (PERUGIA - Malattie Infettive), Giustino Parruti (PESCARA -Malattie Infettive), Ennio Polilli (PESCARA - Virologia Pescara), Daria Sacchini (PIACENZA - Malattie Infettive), Chiara Martinelli (PISA - Malattie Infettive), Rita Consolini (PISA - Pediatria I Università), Linda Vatteroni (PISA - Virologia), 58274 Angela Vivarelli (PISTOIA - Malattie Infettive), Daniele Dionisio (PISTOIA - Virologia), Alessandro Nerli (PRATO -59275 Malattie Infettive), Lucia Lenzi (PRATO - Virologia), Giacomo Magnani (REGGIO EMILIA - Malattie Infettive), Patrizia 60276 Ortolani (RIMINI - Malattie Infettive RIMINI), Massimo Andreoni (ROMA - Cattedra Malattie Infettive Tor Vergata), 277 Guido Palamara (ROMA - IRCCS S. Gallicano), Caterina Fimiani (ROMA - Immunologia Clinica Umberto I), Lucia 278 Palmisano (ROMA - Istituto Superiore di Sanità), Andrea De Luca (ROMA - Istituto di Clinica Malattie Infettive 279 Cattolica), Simona Di Giambenedetto (ROMA - Laboratorio virologia Cattolica), Vincenzo Vullo (ROMA - Malattie

Infettive e Tropicali La Sapienza - Umberto I), Ombretta Turriziani (ROMA - Medicina Sperimentale e Patologia -3 281 Sezione Virologia - La Sapienza), Marco Montano (ROMA - Virologia per Malattie Infettive Tor Vergata), Giovanni 4 282 Cenderello (SAN REMO - Malattie Infettive), Angela Gonnelli (SIENA - Malattie Infettive), Maurizio Zazzi (SIENA -5 283 Virologia), Michele Palumbo (TERNI - Malattie Infettive), Valeria Ghisetti (TORINO - Laboratorio di Virologia, Ospedale Amedeo di Savoia), Stefano Bonora (TORINO - Malattie Infettive Amedeo di Savoia), Palma Delle Foglie (TRENTO -Malattie Infettive), Cristina Rossi (TREVISO - Malattie Infettive), Federica Poletti (VERBANIA - Malattie Infettive VERBANIA), Vincenzo Mondino (VERBANIA - Virologia), Marina Malena (VERONA - Centro di Medicina Preventiva-ULSS 20), Emanuela Lattuada (VERONA - Malattie Infettive).

r for per period

2 289 References 3

1

290

294

20303

4 291 Bartlett JA, Buda JJ, von Scheele B, Mauskopf JA, Davis EA, Elston R, King MS, Lanier ER. 2006. 5 6 292 Minimizing resistance consequences after virologic failure on initial combination therapy: a 7 293 systematic overview. J Acquir Immune Defic Syndr 41:323-331. 8

9 10²⁹⁵ Daar E, Tierney C, Fischl M, Collier A, Mollan K, Budhathoki C, Godfrey C, Jahed N, Katzenstein D, 11296 Sax P, and ACTG A5202 Study Team. ACTG 5202: Final Results of ABC/3TC or TDF/FTC with either 12297 EFV or ATV/r in Treatment-naive HIV-infected Patients. 17th Conference on Retroviruses and 13 298 14 298 Opportunistic Infections (CROI), 16-19 Feb 2010, San Francisco (USA). Abstract 59LB. 15 299

16300 DHHS Panel on Antiretroviral Guidelines for Adults and Adolescents. Guidelines for the Use of 17301 Antiretroviral Agents in HIV-1-Infected Adults and Adolescents. December 1, 2009. 18 302 19 http://aidsinfo.nih.gov/contentfiles/AdultandAdolescentGL.pdf (last access on 13 Mar 2010).

21304 Geretti AM. 2007. Epidemiology of antiretroviral drug resistance in drug-naïve persons. Curr Opin 22305 Infect Dis 20:22-32. ²³₂₄³⁰⁶

25 307 Gupta R, Hill A, Sawyer AW, Pillay D. 2008. Emergence of drug resistance in HIV type 1-infected 26308 patients after receipt of first-line highly active antiretroviral therapy: a systematic review of clinical 27 3 09 trials. Clin Infect Dis 47:712-722. 28 29³¹⁰

30311 Johnson VA, Brun-Vezinet F, Clotet B, Gunthard HF, Kuritzkes DR, Pillay D, Schapiro JM, Richman 31312 DD. 2009. Update of the drug resistance mutations in HIV-1: December 2009. Top HIV Med 32313 17:138-145. 33 34 314

35 315 Lima VD, Gill VS, Yip B, Hogg RS, Montaner JS, Harrigan PR. 2008. Increased resilience to the 36316 development of drug resistance with modern boosted protease inhibitor-based highly active 37 317 antiretroviral therapy. J Infect Dis 198:51-58. 38

³⁸₃₉318 40319 Madruga JV, Berger D, McMurchie M, Suter F, Banhegyi D, Ruxrungtham K, Norris D, Lefebvre E, 41 3 2 0 de Béthune MP, Tomaka F, De Pauw M, Vangeneugden T, Spinosa-Guzman S; TITAN study group. ⁴²321 2007. Efficacy and safety of darunavir-ritonavir compared with that of lopinavir-ritonavir at 48 43 44³²² weeks in treatment-experienced, HIV-infected patients in TITAN: a randomised controlled phase III 45 3 2 3 trial. Lancet 370:49-58.

47 325 48 326 49 326 Mills AM, Nelson M, Jayaweera D, Ruxrungtham K, Cassetti I, Girard PM, Workman C, Dierynck I, Sekar V, Abeele CV, Lavreys L. 2009. Once-daily darunavir/ritonavir vs. lopinavir/ritonavir in 50327 treatment-naive, HIV-1-infected patients: 96-week analysis. AIDS 23:1679-1688.

52₃₂₉ Molina JM, Andrade-Villanueva J, Echevarria J, Chetchotisakd P, Corral J, David N, Moyle G, 53 53 54 330 Mancini M, Percival L, Yang R, Thiry A, McGrath D; CASTLE Study Team. 2008. Once-daily 55331 atazanavir/ritonavir versus twice-daily lopinavir/ritonavir, each in combination with tenofovir and 56332 emtricitabine, for management of antiretroviral-naive HIV-1-infected patients: 48 week efficacy 57₃₃₃ 58₂₂₄ and safety results of the CASTLE study. Lancet 372:646-655.

59³³⁴

46324

51 3 28

60335 Oldfield V, Plosker GL. 2006. Lopinavir/ritonavir: a review of its use in the management of HIV 336 infection. Drugs 66:1275-1299.

² 338 Ortiz R, Dejesus E, Khanlou H, Voronin E, van Lunzen J, Andrade-Villanueva J, Fourie J, De Meyer S,

3 339 De Pauw M, Lefebvre E, Vangeneugden T, Spinosa-Guzman S. 2008. Efficacy and safety of once-

- 4 340 daily darunavir/ritonavir versus lopinavir/ritonavir in treatment-naive HIV-1-infected patients at
- 6 341 week 48. AIDS 22:1389-1397.
- 7 342
 8 343
 9 343
 9 343
 10 344
 10 344
 11 345
 11 345
 11 345
 12 346
 12 346
 14 Med 358:2095-2106.
- 13₃₄₇

Soriano V, Köppe S, Mingrone H, Lutz T, Opravil M, Andrade-Villanueva J, Antunes F, Di Perri G,
 Podzamczer D, Taylor S, Horban A, Duiculescu D, de Rossi L. Prospective comparison of Nevirapine
 and Atazanavir/ritonavir both combined with Tenofovir DF/Emtricitabine in treatment-naive HIV-1
 infected patients: ARTEN Study week 48 results 5th IAS Conference on HIV Pathogenesis,
 Treatment and Prevention (IAS 2009), July 19 -22, 2009, Poster LBPEB07.

- Soulié C, Assoumou L, Ghosn J, Duvivier C, Peytavin G, Ait-Arkoub Z, Molina JM, Costagliola D, Katlama C, Calvez V, Marcelin AG. 2009. Nucleoside reverse transcriptase inhibitor-sparing regimen (nonnucleoside reverse transcriptase inhibitor + protease inhibitor) was more likely associated with resistance comparing to nonnucleoside reverse transcriptase inhibitor or protease inhibitor + nucleoside reverse transcriptase inhibitor in the randomized ANRS 121 trial. AIDS 2359 23:1605-1608.