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Short-term adaptation of conditioned fear responses through endocannabinoid signaling in the central amygdala

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Running title: Endocannabinoid signaling in central amygdala

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Abstract

Both, the cannabinoid receptor type 1 (CB1) and the central nucleus of the amygdala (CeA) are known to play crucial roles in the processing of fear and anxiety, whereby they appear to be especially involved in the control of fear states. However, in contrast to many other brain regions including the cortical subregions of the amygdala, the existence of CB1 in the CeA remains enigmatic. Here we show that CB1 is expressed in the CeA of mice and that CB1 in the CeA mediates short-term synaptic plasticity, namely depolarization-induced suppression of excitation (DSE) and inhibition (DSI). Moreover, the CB1 antagonist AM251 increased both excitatory and inhibitory postsynaptic responses in CeA neurons. Local application of AM251 in the CeA in vivo resulted in an acutely increased fear response in an auditory fear conditioning paradigm. Upon application of AM251 in the basolateral nucleus of the amygdala (BLA) in an otherwise identical protocol no such acute behavioral effects were detected, but CB1 blockade resulted in increased fear responses during tone-exposures on the subsequent days. Moreover, we observed that the efficacy of DSE and DSI in the CeA was increased on the day following fear conditioning, indicating that a single tone-shock pairing resulted in changes in endocannabinoid signaling in the CeA. Taken together, our data show the existence of CB1 proteins in the CeA, and their critical role for ensuring short-term adaptation of responses to fearful events, thereby suggesting a potential therapeutic target to accompany habituation-based therapies of posttraumatic symptoms.

Keywords: Fear, anxiety, conditioning, extinction, amygdala, synaptic transmission

Introduction

The therapy of anxiety disorders benefits strongly from a profound understanding of the biological mechanisms that control the intensity and temporal course of fear reactions. Among those mechanisms, accumulating evidence suggests that the endocannabinoid system may play a major role because of its anxiolytic-like properties (Navarro *et al*, 1997; Patel and Hillard 2006; for review see Viveros *et al*, 2005; Wotjak 2005) and its involvement in the extinction of aversive memories (Marsicano *et al*, 2002; Suzuki *et al*, 2004; Chhatwal *et al*, 2005; Kamprath *et al*, 2006; for review see Chhatwal and Ressler 2007). Endocannabinoids are fatty acid derivatives, which are synthesized and released on demand, e.g. upon postsynaptic depolarization or activation of metabotropic glutamate receptors, and bind to presynaptically located G-protein coupled receptors, the cannabinoid receptors. The cannabinoid receptor type 1 (CB1) ranks among the most abundant G-protein coupled receptors in the mammalian brain and its activation results in the reduction of transmitter release (e.g. GABA, glutamate or serotonin) from the respective presynapse (for review see Wilson and Nicoll 2002; Freund *et al*, 2003; Di Marzo 2009). Recent data obtained in humans show that certain promoter variants of the cannabinoid receptor 1 gene (*CNR1*) and the serotonin transporter gene lead to extremely low or high synaptic serotonin concentrations and, in turn, to an anxious phenotype (Lazary *et al*, 2009). Another recent study reported a reduction of otherwise treatment-resistant nightmares in PTSD patients upon treatment with a synthetic endocannabinoid receptor agonist (Fraser 2009), which is supported by an analysis of

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single nucleotide polymorphisms in the *CNR1* gene suggesting a possible association with PTSD (Lu *et al*, 2008).

The prominent role of CB1 in fear and anxiety is reflected by its expression pattern in the mammalian brain: CB1 is highly abundant in brain regions involved in fear-related processing including the amygdala, the hippocampus, and the neocortex among others (Marsicano and Lutz 1999; Marsicano and Kuner 2008). Among those brain regions, the amygdala plays a key role in fear memory and, thus, in various anxiety disorders (for review see LeDoux 2000; Pape and Pare 2010). Among the different nuclei of the amygdala, the lateral (LA), basolateral (BLA), and central nucleus (CeA), represent the key substrates of fear memory (LeDoux 2000; Pape and Pare 2010). In the last decades the CeA was mainly considered as “output” station of the amygdala relaying information from the LA/BLA to different brain areas responsible for distinct aspects of fear behavior (LeDoux *et al*, 1988). However, recent studies indicate that the CeA comprises inhibitory circuits enabling the processing of information received by its inputs from LA and BLA as well as from thalamic and cortical areas, which project to the CeA either directly or via intercalated cells (for review see Ehrlich *et al*, 2009). Thus, neural circuits within the CeA eventually determine the strength and nature of the fear response during different stages of fear learning and unlearning.

Although it seems conceivable that a neuromodulatory system like the endocannabinoid system, which controls the release of other neurotransmitters including GABA and glutamate, would be ideally positioned in a brain structure with gating functions such as the CeA to exert its effects on fear and anxiety, the existence of CB1 in the CeA remains to be proven. A detailed analysis of CB1 expression in the amygdala by

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immunohistochemistry revealed high levels of CB1 in lateral and basal nuclei, whereas it appeared absent in the central and medial nuclei of the amygdala (Katona *et al*, 2001). In the same study the authors failed to detect cannabinoid effects in the CeA by electrophysiological means (Katona *et al*, 2001), whereas another recent electrophysiological study reported that endocannabinoids regulate inhibitory transmission in the CeA (Roberto *et al*, 2010). In line with this finding, other studies detected a faint immunopositive signal in the CeA (Tsou *et al*, 1998; McDonald and Mascagni 2001; Patel *et al*, 2005a).

Therefore we thought it timely to investigate the possible role of CB1 in CeA. The strategy was to (i) demonstrate CB1 expression immunocytochemically, (ii) investigate the endocannabinoid-related electrophysiological phenomena of depolarization-induced suppression of excitation (DSE) and inhibition (DSI) *in vitro*, (iii) use a behavioral fear conditioning paradigm proven to recruit endocannabinoid signaling combined with local pharmacological blockade of CB1 in amygdala *in vivo*, and (iv) test for specificity of observed effects through use of CB1 null mutant mice.

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Material and Methods

Animals

Male CB1 null mutant mice (CB1^{-/-}; n=22)(Marsicano *et al*, 2002), male wild-type littermates (CB1^{+/+}; n=36) and male C57BL/6NCrl mice (n=67) at an age of 6 weeks were single housed under a 12h:12h light-dark cycle. All experiments were carried out in accordance with the European Committees Council Directive (86/609/EEC). Protocols were approved by the Bezirksregierung Münster (AZ50.0835.1.0, G53/2005).

Drugs

The AMPA receptor antagonist 2,3-Dioxo-6-nitro-1,2,3,4-tetrahydrobenzo[f]quinoxaline - 7-sulfonamide disodium salt (DNQX); the NMDA receptor antagonist DL-2-Amino-5-phosphonopentanoic acid (DL-AP5), the GABA_A receptor antagonist 6-Imino-3-(4-methoxyphenyl)-1(6H)-pyridazinebutanoic acid hydrobromide (Gabazine), the GABA_B receptor antagonist (2S)-3-[[[(1S)-1-(3,4-dichlorophenyl)ethyl]amino-2-hydroxypropyl] (phenylmethyl) phosphinic acid (CGP55845) and the CB1 antagonist (N-(Piperidin-1-yl)-5-(4-iodophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (AM251) were purchased from Biotrend (Tocris), Köln, Germany.

Immunocytochemistry

Animals were deeply anesthetized with pentobarbital and trans-cardially perfused with PBS containing heparin (5U/ml), followed by 4% PFA solution. Brains were isolated and post-fixed for 24h at 4°C in 4% PFA solution, treated with 30% sucrose/PBS solution for

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48h at 4°C and stored at -80°C until use. Thirty µm thick brain sections were prepared on a Microm HM560 cryostat, and stored at -20°C in cryoprotection solution (25%glycerin, 25%ethylenglycol and 50%PBS) until use. All incubation steps were performed in 48-well plates on a wave shaker (Heidolph) at RT if not stated otherwise. Sections were first rinsed in PBS (10min) and then pre-incubated in blocking solution (5% normal donkey serum, 2.5% BSA, 0.3% Triton X-100 in PBS) for 1h. After blocking, the sections were treated o/n at 4°C with a polyclonal antibody against CB1 (rabbit anti-CB1; 1:500; AF380 from Frontier Science) diluted in low concentrated blocking solution (1% normal donkey serum, 0.1% BSA, 0.3% Triton X-100 in PBS). On the next day, the sections were washed in PBS-T (PBS/0.1% Triton X-100) five times for 10min and then incubated for 2h with the secondary antibody (Cy3 labeled anti-rabbit-IgG from goat, 1:250, Jackson Immuno Research) diluted in the low concentrated blocking solution. The incubation was followed by three 10min washing steps in PBS. Sections were counterstained with Hoechst 33258 (2µg/µl). After the counterstaining the sections were washed twice for 10min in PBS, followed by 2min in distilled water. Sections were mounted on glass slides to dry for 2-4 h at 37°C. The remaining salt was washed off by dipping the slides for 2s into distilled water. Finally the sections were dried overnight in a dust free environment at RT, covered with MOWIOL and coverslips. Sections were viewed on a laser-scanning confocal microscope (Nikon eC1 plus, Düsseldorf, Germany) with a 40x oil immersion objective (HCX PL APO 40X /1.25-0.75, Leica Microsystems, Wetzlar, Germany). Results presented here were collected from CB1^{+/+} and CB1^{-/-} mice (n=4 for total experiment).

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Preparation of amygdala slices

Mice were anesthetized with isoflurane (5%) and decapitated, their brains were rapidly removed and put into oxygenated (95% O₂, 5% CO₂) ice-cold artificial cerebrospinal fluid (ACSF) containing (in mM): 120 NaCl, 2.5 KCl, 1.25 NaH₂PO₄, 22 NaHCO₃, 2 MgSO₄, 2 CaCl₂ and 15 glucose; pH was 7.35. Coronal slices (250- μ m-thick) containing the amygdalar complex were cut using a Leica VT-1200S microtome (Leica Microsystems, Wetzlar, Germany) at 4°C. Slices were incubated at 34°C for 20min and then allowed to equilibrate for at least 1h at room temperature before commencement of recording.

Recording from visualized neurons

Recordings were obtained from visually identified neurons in the amygdala as described previously (Jungling *et al*, 2008; Sosulina *et al*, 2010). In brief, central amygdala (CeA) neurons were identified using differential interference contrast infrared videomicroscopy (S/W-camera CF8/1, Kappa, Gleichen, Germany; Stuart *et al*, 1993). Whole cell patch clamp-recordings were made from CeA neurons at 30°C, using a patch clamp amplifier (EPC-7, List Medical Systems, Darmstadt, Germany). Slices were superfused continuously (2.0ml/min) with an oxygenated solution identical to one used for cutting the slices. During recording, the majority of cells were filled with biocytin (1%) and then processed by the conventional immunoperoxidase method. The exact location of the cells was determined by light microscopy. Immediately after breaking through for whole-cell patch-clamp, neurons were characterized by electrotonic and electrogenic parameters as reported before (Sosulina *et al*, 2010). Access resistance was evaluated

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during recordings upon a 3mV, 10ms pulse applied 600ms after each evoked response. Correction for liquid junction potential changes (10mV) was applied. Switching frequency was 30kHz, gain 1.0-1.2. Records were filtered at 3kHz (eight-pole Bessel filter), sampled at 10kHz using pClamp9 software (Axon Instruments, Union City, CA, USA), and analyzed off-line using the programs MATLAB 7.0-R14 (The Math Works, Natick, MA) and Clampfit 10.0 (Molecular Devices Corporation).

Electrophysiological Recordings

Coronal slices containing the amygdala were prepared as described above and placed in a submerged chamber, continuously superfused with ACSF at 30°C. Whole cell recordings were obtained with micropipettes filled with (in mM): K-Gluconic acid 100, KCl 50, CsCl 10, HEPES 10, EGTA 0.2, MgCl₂ 1, Mg-ATP 1, Na-GTP 0.3, pH7.3 adjusted with 1M KOH. EGTA-concentration was kept low because of the Ca²⁺-dependence of DSE or DSI (Lenz and Alger 1999). Access resistance was continuously monitored and ranged between 4-9.5MΩ. Neurons were recorded in the medial section of the CeA (CeAM; S1; Fig. S1, 2A, 3A), and those displaying resting membrane potential positive to -60mV or changes in capacitive transient >10% during the course of the experiment, were discarded. Extracellular stimuli (100μs, 50-600μA) were delivered through a bipolar stainless-steel electrode placed in the BLA or lateral part of the CeA (CeAL). Glutamatergic and GABAergic components of synaptic responses were isolated by addition of gabazine (1μM) and CGP55845 (50μM), and DNQX (10μM) and AP-5 (50μM), respectively.

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Depolarization-induced suppression of excitation (DSE) and inhibition (DSI)

DSE or DSI tests were performed every 5min, and consisted of 30 microstimuli (50 to 600 μ A at 0.25Hz) applied to either BLA or CeAL before, and 75 stimuli after postsynaptic depolarization from -70 to 0mV. Stimuli were set at 20-30% of maximal synaptic response amplitude, and duration of depolarization was varied between 0.1s-10s. For each neuron at least three tests of DSE or DSI were made, and different durations were tested in a pseudo-randomized fashion. The degree of suppression was calculated using the mean of 15 evoked responses just before the depolarization (*amp baseline*) and three evoked responses just after depolarization (*amp test*) using the formula(modified from Wilson and Nicoll 2001): $\% \text{ of baseline} = 100 * (\text{amp test} / \text{amp baseline})$. For statistical analysis the amplitudes of every three evoked postsynaptic currents in control and after depolarization were averaged and normalized against mean amplitude in control. These normalized values were then averaged over groups of cells. Duration of DSE or DSI was calculated from the sliding average of the amplitude of every three responses plotted as a function of time. Plots were adjusted with a one-phase association equation. Duration of DSE or DSI was defined as the time between the end of the depolarization and the time when evoked currents amplitude returned to control. Some slices were pre-incubated with the CB1 antagonist AM251 (2 μ M) at least one hour prior to testing DSE or DSI.

Surgery and local drug application

Mice were implanted bilaterally with a 26 gauge stainless steel guide cannula ending 1.0mm above CeA (stereotaxic coordinates: 1.0mm anterior, 2.8mm lateral, and 3.0mm

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ventral from brain surface) or BLA (stereotaxic coordinates: 1.0mm, 3.1mm, 3.3mm, respectively) under deep pentobarbital anesthesia (75mg/kg i.p.), as described previously (Jungling *et al*, 2008). After one week recovery from surgery, vehicle (6,66% dimethylsulfoxide, 6,66% Tween-80 in saline) or AM251 (1ng or 1µg in 0.5µl vehicle) were infused with a 33 gauge beveled needle injector at 0.1µl/min (0.5µl each side) under isoflurane anesthesia. Placement of the infusion cannulae was histologically verified, and only animals with proper injection sites in the CeA or in the BLA were included in the analysis (Fig. S2, for examples please note the histological controls therein).

Fear conditioning/extinction

Mice were placed in a fear training apparatus (TSE, Bad Homburg, Germany), as described previously (Kamprath *et al*, 2006; Plendl and Wotjak 2010; Sangha *et al*, 2009). Three minutes later, a 20-s tone of 9kHz and 80dB (conditioned stimulus, CS+) was presented, which co-terminated with a 2-s scrambled electric foot shock of 0.7mA (unconditioned stimulus). On the next day (day1/d1), mice were locally treated with AM251 or vehicle, 30min later transferred to a novel context, and exposed to CS+ for 200s. For fear extinction, CS+ exposure was repeated on days 2 and 3 (d2, d3; Fig.5A). Behavioral responses were assessed by a trained observer unaware of the treatment conditions. Freezing was defined as the absence of all movements except for those related to respiration. During analysis, the freezing response was either summarized over the complete 200s of tone presentation or it was subdivided into ten distinct 20-s intervals, and freezing response was expressed as a percentage of the respective time

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('freezing time' per interval/total interval time). In two series of experiments, slices were prepared following fear conditioning, either 24 hours after fear training (FC-d1) or 24 h after the second series of CS+ exposure (FC-d3), and DSE and DSI were tested as described above.

Elevated Plus maze.

The plus maze consisted of two open (30 x 5cm) and two lall-enclosed armas (30 x 5 x 25 cm) connected by a central platforms (5 x 5 cm) elevated 75 cm above the floor.

Light intensity on the open arms was 250 lux. At the beginning of the experiment, the mouse was placed in the central area facing a closed arm and was then allowed to freely explore the maze for 5 min. Mice were exposed to the maze for a second trial 24 h later. Local drug application was performed 30 min before the first plus maze exposure only. The behavior on the plus maze was monitored by a video motility system (Video-Mot II, TSE, Bad Homburg, Germany), which automatically quantified the total distance moved. Entries and time spent in the open and closed arms, respectively, were scored by a trained observer unaware of the genotype or treatment condition with the criterion for an arm entry defined as all four paws in an arm of the maze. Percent entries into open arms was expressed as number of open arm entries/total arm entries x 100. Exclusion criteria embraced freezing over the full time of the test and stereotypic behavior. None of the animals tested met any of the behavioral exclusion criteria.

Statistics

Data were analyzed by Student's t-test, 1- or 2-way ANOVA for repeated measurements, using STATISTICA 8.0 (StatSoft Inc., Tulsa, OK, USA), GraphPad

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Prism 5.0 (GraphPad Software Inc., San Diego, CA, USA), or PASW Statistics 18 (SPSS, Inc., Chicago, IL, USA). Newman-Keuls test was used as post-hoc test. Data are presented as mean \pm SEM. Statistical significance was accepted if $p < 0.05$.

Results

Immunolocalization of CB1 in the CeA

Confocal inspection revealed high levels of CB1 immunofluorescence in BLA, whereas signals were sparse in CeA (Fig.1A), where fiber-like elements bearing beaded protrusions formed CB1 immunoreactive mesh-like patterns mostly in medioventral parts of CeA, with denser networks in the CeAM compared to CeAL (Fig.1,A1,A2). CB1 immunopositive reactions were absent in CB1^{-/-} (Fig. 1B, B1).

Suppression of synaptic transmission mediated via CB1 in CeAM in vitro

To probe functional CB1 in CeA *in vitro* we used depolarization-induced suppression of synaptic transmission, whereby strong postsynaptic depolarization triggers endocannabinoid release and subsequent decrease in transmitter release via activation of presynaptic CB1 (for review see Chevalleyre *et al*, 2006). Recordings were obtained from CeAM neurons possessing electrophysiological properties as described previously (Fig. S1; Dumont *et al*, 2002). Excitatory (EPSCs) and inhibitory postsynaptic currents (IPSCs) were evoked upon local electrical microstimulation of the BLA and CeAL, respectively (Ehrlich *et al*, 2009). High failure rates at low stimulation intensities and constant response latencies indicated the monosynaptic nature of the evoked responses. EPSCs were sensitive to ionotropic glutamatergic antagonists (NBQX: 10 μ M and DL-AP5: 50 μ M; tested in n=6 cells), and IPSCs could be blocked by the GABA_A receptor antagonist gabazine (1 μ M; n=3). For DSE, neurons were depolarized from -70mV to 0mV through direct current injection (duration 10s and 5s), and the effects on

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EPSCs evoked upon BLA stimulation were examined (Fig.2A). In all neurons tested (n=9) DSE was significant upon 10-s depolarization, in that EPSCs were reduced from an average amplitude of -62.3 ± 7.6 pA at baseline to $60\% \pm 3.2\%$ after postsynaptic depolarization (Fig.2B; $p=0.001$; 1-way ANOVA). EPSC amplitudes recovered to baseline within 70.3 ± 6.3 s. By contrast, 5-s depolarization was not sufficient to induce significant DSE in the same cells ($90.8 \pm 8.8\%$ compared to baseline; n=9), and the maximal change in EPSC amplitude differed significantly following 5-s and 10-s depolarization ($p=0.0137$, unpaired t-test; Fig.2B,D). In age-matched CB1^{-/-}, 10-s depolarization did not induce measurable DSE ($97.1 \pm 2.5\%$ of baseline; n=10; Fig.2C,D). Moreover, DSE was not detected in CeAM slices from CB1^{+/+} that had been pre-incubated with the CB1 antagonist AM251 (2 μ M; $100.8 \pm 2\%$ of baseline; n=10; Fig.2C,D), and bath application of AM251 blocked the previously established DSE in CeAM neurons ($67.3 \pm 4.4\%$ before, $101.4 \pm 2.6\%$ after AM251; n=5; $p<0.05$; paired t-test; Fig.2C,D; Fig.4A,B).

Next, DSI was investigated using IPSCs evoked upon CeAL stimulation, and testing the effects of postsynaptic depolarization (-70mV to 0mV; Fig.3A). IPSCs displayed a mean amplitude of -136.6 ± 23.2 pA at $V_h = -70$ mV (n=7), which was significantly reduced upon both 10-s and 5-s depolarization in all neurons tested (n=4; $65.1 \pm 5.7\%$, and $48.9 \pm 6.3\%$ of baseline, respectively; $p<0.05$; 1-way ANOVA; Fig.3B,D), indicating effective DSI. Interestingly, short duration depolarization (0.1s, 1s, 5s) tested in a separate group of cells (n=4) were also sufficient to induce DSI (0.1s: $77.6 \pm 5.7\%$, 1s: $70.5 \pm 14.4\%$ and 5s: to $70.6 \pm 7\%$ of baseline; $p<0.01$; unpaired t-test; n=4; Fig.3D). Differences in DSI

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magnitude were only different between shortest and longest depolarization tested (0.1s vs. 10s; $p=0.016$, unpaired t-test, Fig.3D). Significant DSI was not detected in CeAM neurons in $CB1^{-/-}$ ($97.1\pm 3.3\%$ of baseline, $n=4$; Fig.3C,D), and application of the CB1 antagonist AM251 ($2\mu M$) blocked established DSI in $CB1^{+/+}$ in all CeAM neurons tested ($n=4$; $54.9\pm 5\%$ vs. $106.4\pm 7.5\%$ of baseline; $p<0.05$; 1-way ANOVA; Fig.3C,D; Fig4C,D). Taken together, these data indicate that CB1 is functional in CeA and capable of suppressing both excitatory and inhibitory synaptic inputs to CeAM neurons.

Notably, application of AM251 ($2\mu M$) increased the amplitudes of both EPSCs (by $52\pm 5\%$, $p<0.05$; paired t-test; $n=5$; Fig.4A) and IPSCs (by $42\pm 6.6\%$, $p<0.05$; paired t-test; $n=4$; Fig.4C). When the suppression after 10s depolarization was plotted against the change produced by AM251 application, a strong correlation between both parameters either on DSE ($R=0.8984$) or DSI ($R=0.8905$) (Fig.4B,D, respectively) was found, indicating the presence of an endocannabinoid tone.

Short term effects on conditioned fear behavior via CB1 in CeA

In an auditory fear conditioning paradigm $CB1^{-/-}$ displayed increased freezing responses in contrast to $CB1^{+/+}$ (Fig. S3), with a 2-way ANOVA revealing significant genotype and interval effects at all non-reinforced tone presentations, i.e. at day 1 (*Genotype*: $p=0.0081$; *Interval*: $p<0.0001$), day 2 (*Genotype*: $p<0.0001$; *Interval*: $p<0.0001$), and day 3 (*Genotype*: $p<0.0001$; *Interval*: $p<0.0001$), which is in line with previous reports indicating impaired fear extinction in $CB1^{-/-}$ (Marsicano *et al*, 2002; Cannich *et al*, 2004; Kamprath *et al*, 2006; Plendl and Wotjak 2010). To identify the possible contribution of

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CB1 in CeA, the CB1 antagonist AM251 (1ng, n=7; 1µg, n=10) or vehicle (n=10) was injected bilaterally into the CeA in C57BL/6NCrI mice 30min before the first non-reinforced tone presentation following fear conditioning (Fig.5A,B). A 1-way ANOVA of the freezing response revealed a significant drug effect (*Drug*: p=0.013) between the 1-µg AM251 and vehicle group (p=0.0165; Newman-Keuls Posthoc test), therefore this dose of AM251 was used for all following experiments. Detailed analyses showed that injection of AM251 into the CeA resulted in acutely increased fear responses at day 1 (*Drug*: p=0.0032; *Interval*: p=0.008, Fig.5C; 2-way ANOVA), but not at day 2 or day 3 (Fig.5C; statistics not shown). By comparison, local application of AM251 bilaterally into the BLA in the same experimental paradigm, but in a separate batch of animals (n=15; Fig.5D), did not result in significant differences in conditioned fear at day 1, but led to an increased freezing response during the drug-free testing at day 2 (*Drug*: p=0.0071) and day 3 (*Drug*: p=0.0028; 2-way ANOVA, Fig.5D). This phenotype differs substantially from that obtained upon injections of AM251 into the CeA indicating that CB1 might play different roles in the CeA and BLA in fear memory.

Short term effects on anxiety-related behavior mediated via CB1 in CeA

The previous behavioral experiments show a differential involvement of CB1 in CeA *versus* BLA in fear adaptation processes. While CB1 in BLA appears to be involved in long-term extinction learning processes, CB1 in CeA seems to relate to acute fear adaptation rather than long-term fear learning and thus might play a role in general anxiety tasks. To test this hypothesis we exposed a new batch of C57BL/6NCrI mice to the elevated plus maze (EPM) 30 min after local injection of either 1µg AM251 (n=13) or vehicle (n=12) bilaterally into the CeA (i.e. a similar treatment protocol as used for the

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fear conditioning experiments). However, no significant effects on anxiety-related parameters (percent time spent on open arms, entries into open arms) could be detected, neither at day 1 nor on a drug-free second exposure to the EPM at day 2 (Fig.6A,B; statistics not shown). Similarly, the locomotion-related parameters (total distance moved, total entries) did not reveal any significant effects, neither at day 1 nor at day 2 (Fig.6C,D; statistics not shown).

Altered DSE and DSI in CeA upon fear conditioning

Next, slices were prepared *ex vivo* from CB1^{+/+} at day 1 after fear conditioning (FC-d1), and DSE and DSI were assessed in CeAM. While DSE upon 10-s depolarization did not differ significantly in conditioned (55.4 ±8.4% of baseline; n=5 neurons from 3 animals) *versus* naïve mice (61.9±2.6%; n=11/5 mice), 5-s depolarization in the same cells resulted in a significant DSE in conditioned animals (61.9±6.4%; p<0.01, unpaired t-test; n = 5/3mice), but failed to evoke DSE in naïve controls (90.8±8.8%; n=9; Fig.7A,B). Testing DSI revealed a significant (p=0.022, unpaired t-test) increase in suppression of inhibition upon 0.1-s depolarization in conditioned (60.1±5.6%; n=6/3mice) compared to naïve controls (77.63±5.7%;n=4/3mice) (Fig.7C), whereas DSI upon longer depolarization tested was not different between groups of animals (Fig.7D). Slices prepared from fear-conditioned mice 24 hours after the second series of CS+ exposure (FC-d3) revealed no differences in DSE and DSI compared to naïve mice. Of note, depolarization protocols were used in these experiments proven to reveal significant differences in DSE and DSI 24 hours after fear conditioning (FC-d1). The magnitude of DSE upon 5-s depolarization in the FC-d3 group (91.2±7.9% of baseline; n=5 neurons

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from 2 animals) was not different from that in naïve mice ($p=0.9737$), but significantly smaller compared to FC-d1 ($p<0.01$; unpaired t-test; Fig. 7B). A 0.1-s depolarization induced effective DSI ($79.8\pm 4.8\%$ of baseline; $n=5$ neurons from 2 animals; Fig. 7B) in the FC-d3 group, similar to that in naïve mice ($P=0.7701$), and significantly different compared to the FC-d1 group ($p<0.01$, unpaired t-test).

Discussion

Both DSI and DSE have been described in BLA projection neurons, and have been associated with CB1 activation in axons of GABAergic interneurons (Zhu and Lovinger 2005; Patel *et al*, 2009) and putative glutamatergic cortical input fibers (Kodirov *et al*, 2010), respectively. In the present study, DSI and DSE were observed in CeAM neurons for putatively monosynaptic responses evoked upon stimulation of CeAL and BLA, respectively. Mediation through CB1 was shown by sensitivity towards a CB1 antagonist and absence in CB1^{-/-}. In view of glutamatergic neurons projecting from BLA to CeAM, and a strong GABAergic projection from CeAL to CeAM(Ehrlich *et al*, 2009), it seems reasonable to suggest that CB1 are located on the respective axonal terminals. Involvement of fibers traversing the BLA and CeAL cannot be excluded, although this possibility would be difficult to reconcile with the local nature of CB1-containing interneuronal axons, the lack of CB1 in intercalated GABAergic neurons(Geracitano *et al*, 2007), and the observed regional specificity of evoked EPSCs and IPSCs in CeAM. In keeping with this, endocannabinoids spread over only micrometer distances(Wilson and Nicoll 2001), indicative of a local effect in CeAM. While the strongest suppression was found upon a 10-s depolarization for both DSI and DSE, shorter depolarization times abolished DSE, but not DSI. This might relate to differences in CB1 expression, since GABAergic neurons express CB1 at higher levels compared to glutamatergic neurons(Marsicano and Lutz 1999; Ohno-Shosaku *et al.*, 2002), and differences in endocannabinoid production/availability(Straiker and Mackie 2005; Straiker and Mackie 2009). Moreover, blockade of CB1 by AM251 resulted in an increased amplitude of

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evoked EPSCs and IPSCs, indicating that tonic endocannabinoid signaling (Roberto *et al*, 2010) or constitutive activity of CB1 (Losonczy *et al*, 2004) dampens synaptic transmission in CeAM. A similar increase in evoked IPSCs in CeAM upon application of CB1 antagonists has recently been reported, and been related to retrograde endocannabinoid signaling based upon paired-pulse analysis and the blocking effect of postsynaptic Ca^{2+} -chelators (Roberto *et al*, 2010).

Adaptation to a stressor resulted in increased late DSI in the BLA (Patel *et al*, 2009), letting us hypothesize that the experience of fear conditioning induces changes in endocannabinoid signaling in CeA, which might be reflected in DSE or DSI. A single tone-shock pairing indeed resulted in more efficient DSE and DSI twenty-four hours later, in that shorter postsynaptic depolarization was sufficient to induce near-maximal suppression of synaptic transmission as compared to naïve animals. Of note, the enhanced DSE and DSI had normalized 3 days after fear conditioning, supporting the notion of a short-term adaptation mediated via endocannabinoid signaling in CeAM. Possible underlying mechanisms include increases in endocannabinoid availability, for instance through regulated *de novo* synthesis or degradation (for review see Hillard and Jarrahian 2003), or increases in CB1 sensitivity or activity. In fact, endocannabinoid levels notably vary in different behavioral tasks, depending on task specificity, stage of adaptation, brain region and prevailing endocannabinoid (Marsicano *et al*, 2002; Kamprath *et al*, 2006; Patel *et al*, 2005b; Hill *et al*, 2005; Rademacher *et al*, 2008).

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Mice were subjected to an auditory fear conditioning/extinction protocol previously proven to recruit endocannabinoid signaling during the non-reinforced exposures to the conditioned tone, but not during acquisition of conditioned fear (Marsicano *et al*, 2002; Kamprath *et al*, 2006; Plendl and Wotjak 2010; Niyuhire *et al*, 2007; see Fig. S3). Using this protocol, but local infusion of the CB1 antagonist into the CeA before the first tone exposure, yielded a similar increase in subsequent conditioned freezing as previously described with systemic CB1 antagonist application (Marsicano *et al*, 2002; Kamprath *et al*, 2006; Plendl and Wotjak 2010), which was not observed upon drug-free testing on later days. Despite histological verification of infusion sites in the CeA, the pharmacologically active substance might have reached the BLA, which expresses CB1 at high levels. Voting against this possibility is the finding that local application of AM251 bilaterally into the BLA had no acute effect on freezing to conditioned tone at day 1, but resulted in increased freezing upon drug-free testing on later days. These findings indicate a specific role of regionally distributed CB1 in the amygdala in fear memory, with CB1 in BLA being involved in extinction processes at the long-term range, whereas CB1 in CeA contributes to acute fear expression and/or within-session extinction. In this behavioral paradigm, first tone exposure has been previously found to decrease phosphorylation of ERK p44 and calcineurin levels in the BLA of CB1^{-/-} as compared to wildtypes, whereas levels were increased in the CeA (Cannich *et al*, 2004), which may relate to the regionally specific role of endocannabinoid signaling for short- and long-term regulation of conditioned fear. Tests of anxiety-related behavior in the elevated plus maze test revealed anxiogenic-like effects of CB1 agonist application in CeA (Zarrindast *et al*, 2008), whereas CB1 antagonists tested over a wide concentration range had no

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effect(Zarrindast *et al*, 2008; present study), making it unlikely that endocannabinoids are endogenously recruited in this test of anxiety. These conclusions are in line with previous findings indicating that repeated homotypic stress produces temporally and anatomically distinct changes in endocannabinoid signaling components within stress-responsive brain regions (Patel and Hillard, 2008). These signals appear to be essential for the process of stress adaptation (Hill et al., 2010), particularly for habituation to the aversive content of the sensory stimulus (Kamprath et al., 2006). Noteworthy, stress adaptation has been found to differentially involve the two endocannabinoid ligands, N-arachidonyl ethanolamine (anandamide; AEA) and 2-arachidonoylglycerol (2-AG), such that the AEA content is persistently decreased throughout the corticolimbic stress circuit, whereas 2-AG is exclusively elevated within the amygdala (Hill et al., 2010).

In conclusion, the localization of CB1 in the CeA puts the endocannabinoid system in a near-ideal position to control both inhibitory and excitatory synaptic transmission to the major station of fear-related outputs in the amygdala. This intra-amygdaloid feedback system may complement extra-amygdaloid influences, such as prefrontal cortical inputs regulating inhibitory synaptic signals to the CeA during fear extinction(Amano *et al*, 2010). Since recruitment of endocannabinoids appears to require highly aversive conditions (e.g., a strong shock intensity)(see Kamprath *et al*, 2009; also Haller *et al*, 2004; Naidu *et al*, 2007), it may take an outstanding position to ensure proper adaptation after traumatic events.

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Conflict of Interest. The authors declare no conflict of interest.

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Supplementary information is available at the *Neuropsychopharmacology* website.

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Figure Legends

Figure 1. CB1 immunofluorescence in the amygdala of CB1^{+/+} (A) and CB1^{-/-} (B) mice. (A) In the amygdala of CB1^{+/+} mice we detected high levels of CB1 immunoreactivity in the BLA compared to the LA and CeA, with area displayed at higher magnification in (A1 and A2) as indicated. Note higher density of CB1-positive fibers in medial (A2) compared to lateral (A1) CeA. CB1 immunoreactivity shows up as bead cord-like structures forming a mesh-like pattern. (B) In CB1^{-/-} mice no CB1 immunofluorescence could be detected. Only a low level background-fluorescence could be observed. B1) Area indicated in B at higher magnification.

Figure 2. CB1-mediated DSE in CeAM. (A) Recording (R) and stimulation sites (S) in an amygdalar slice. (B) Summarized data of DSE in 9 neurons upon 10-s and 5-s depolarization. In this and subsequent panels, the bar indicates the time when the depolarization was applied. Averaged EPSCs before (n=15 traces) and after (n=3 traces) (a) 10-s and (b) 5-s depolarization. Dotted line indicates the latency. (C) Summarized data of DSE (10s) in control, CB1^{-/-} and in AM251. (a) Traces taken from AM251 and (b) CB1^{-/-}. (D) Summary of the change in EPSC amplitudes (baseline=100%) for every experimental manipulation. Time of depolarization is depicted. For this and following figures the number in brackets is the number of neurons recorded. The number in square corresponds to the same neurons. # compared to baseline; * in between groups; *p<0.05; ###,***p<0.001.

Figure 3. CB1-mediated DSI in CeAM. (A) Recording (R) and stimulation sites (S) in an amygdalar slice. (B) Same DSI induced by 10-s and 5-s depolarization, averaged from recordings in 4 CeAM neurons. Averaged traces of IPSCs are depicted from one neuron before (n=15 traces averaged) and after (n=3 traces) (a) 10-s and (b) 5-s depolarization. Dotted line indicates the latency. (C) Summarized data of DSI (10s) in control, CB1^{-/-} and AM251. (D) Summary of the change in IPSC amplitudes (baseline=100%) for every experimental manipulation.

Figure 4. Effects of the CB1 antagonist AM251 on DSE and DSI in CeAM. A) Time course of a single DSE experiment. Each dot represents one evoked EPSC, tests of DSE (10-s depolarization) are indicated by arrows, colours mark examples of EPSCs depicted in (a) and (b). Application of AM251 (2 μ M) is indicated by horizontal bar. (a) Averaged traces of EPSCs before (black, n=15) and immediately after (red; n=3) the (10-s) depolarization in control. (b) Same as in a) but during AM251 application (black: before, green: after 5 s-depolarization). Note the blockade of DSE and increase in EPSC amplitude with no change in latency during action of AM251. B) Summary of DSE and the change in EPSC amplitudes (baseline=100%), from 5 CeAM neurons. a) Averaged degree of DSE under control conditions (acsf) and 30 minutes after application of AM251 (2 μ M). b) Plot of the degree of suppression after 10-s depolarization against the change in EPSC amplitude at 30 minutes of AM251 application, revealing a linear correlation between these two parameters. Each dot represents data from one CeAM neuron, the square represents the mean from 5 neurons. C) Same as in A) but for DSI. Note the blockade of DSI and increase in IPSC amplitude with no change in latency during action of AM251. D) Summary of the change in DSI (baseline=100%), from 4 CeAM neurons (a) and its relationship with the increase in IPSC amplitude and DSI after AM251 application (b). ***p<0.01

Figure 5. Effects of local application of AM251 in CeA (B, C) and BLA (D) on conditioned freezing. Behavioral protocol is depicted in (A). (B) Effects of local AM251 application in the CeA on CS+-induced freezing at day 1 summarized over the complete 200-s tone presentation. (C) Course of CS+-induced freezing responses at days 1, 2, 3 (each analyzed in 20-s intervals), upon application of vehicle (open circles) or AM251 (black circles) in CeA. (D) Same as (C) upon application of vehicle (open circles) or AM251 (black circles) in BLA. *p<0.05; **p<0.01.

Figure 6. Effects of local application of AM251 in CeA on elevated plus maze behavior. Panels (A) and (B) show the effects on the anxiety-related parameters with percent time spent on open arms (A) and percent entries into open arms (B). Panels (C) and (D)

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show the effects on the locomotion-related parameters with total distance moved (C) and total entries (D).

Figure 7. DSE and DSI in CeAM slices prepared from naïve animals 1 and 3 days after fear conditioning (FC-d1 and FC-d3 respectively). (A) DSE upon 5-s depolarization. (B) Summary of the change in EPSC amplitudes upon 5-s and 10-s depolarization (baseline=100%). Note enhancement of DSE upon 5-s but not 10-s depolarization in FC-d1, but not FC-d3. (C) DSI upon 0.1-s depolarization. (D) Summary of the change in IPSC amplitudes at depolarization of different duration (baseline=100%). Note enhancement of DSI at 0.1-s depolarization in FC-d1 but not in FC-d3. [#] compared to baseline; * in between groups; *p<0.05; ##p<0.01; ###,***p<0.001.

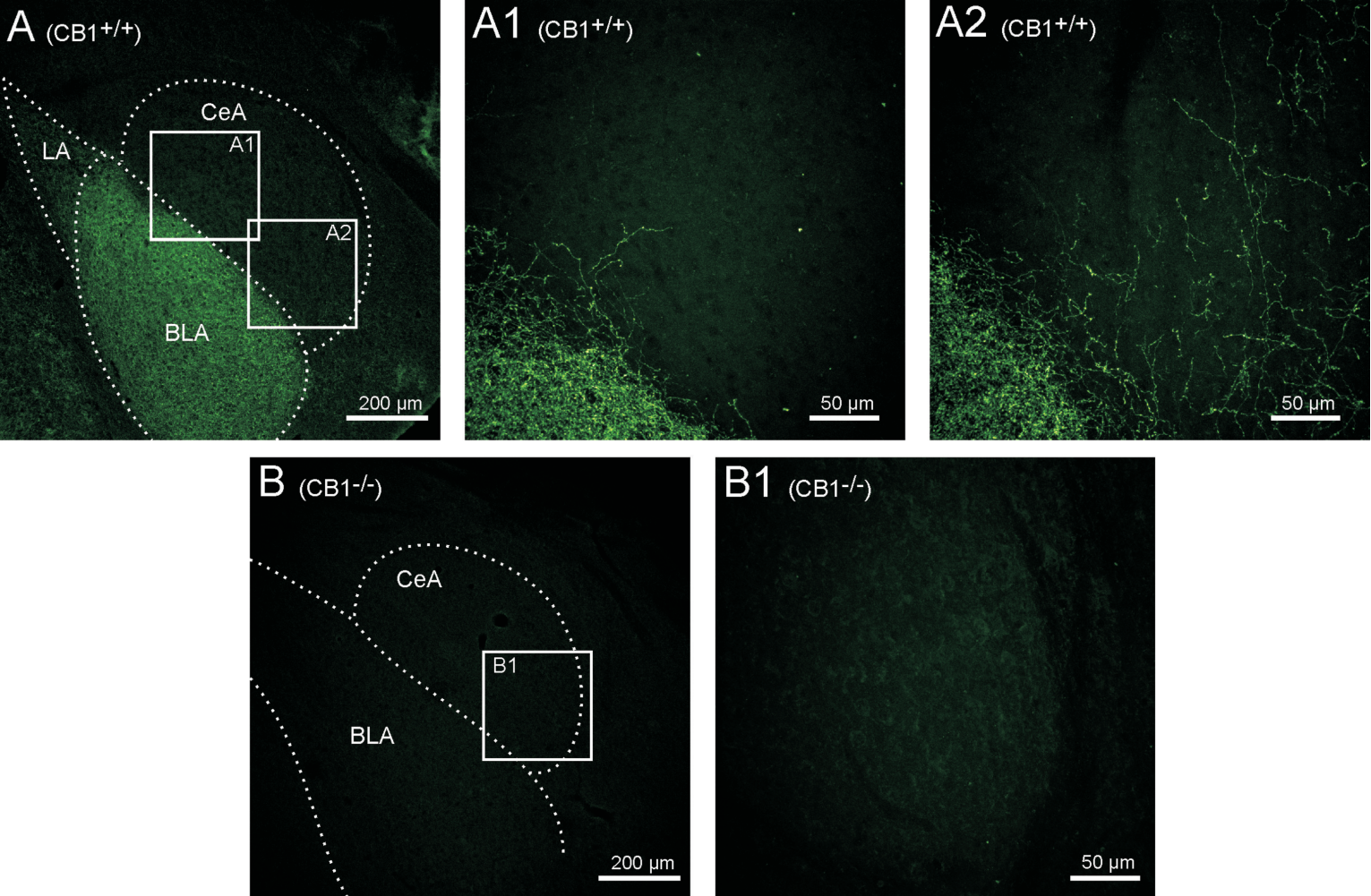
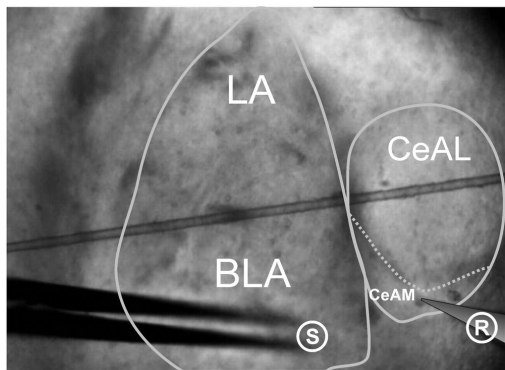
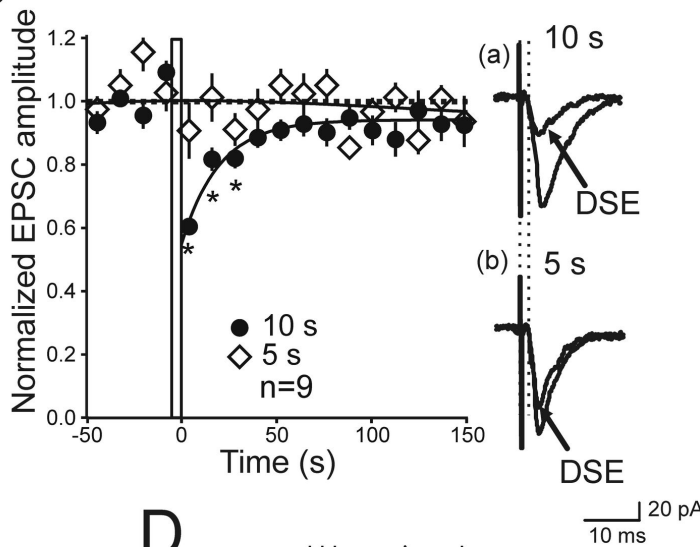
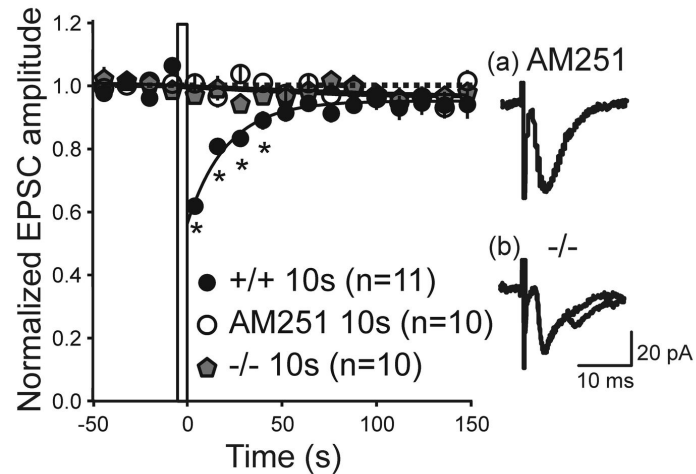
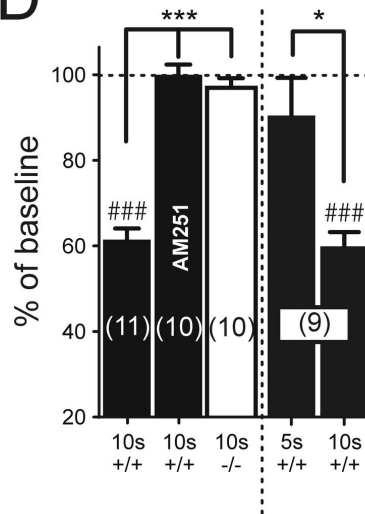


Fig. 1

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A**B****C****D****Fig. 2**

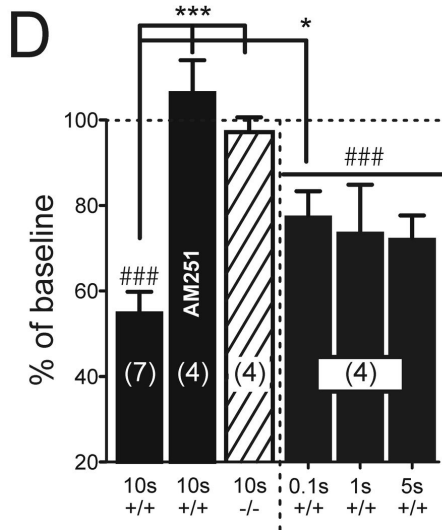
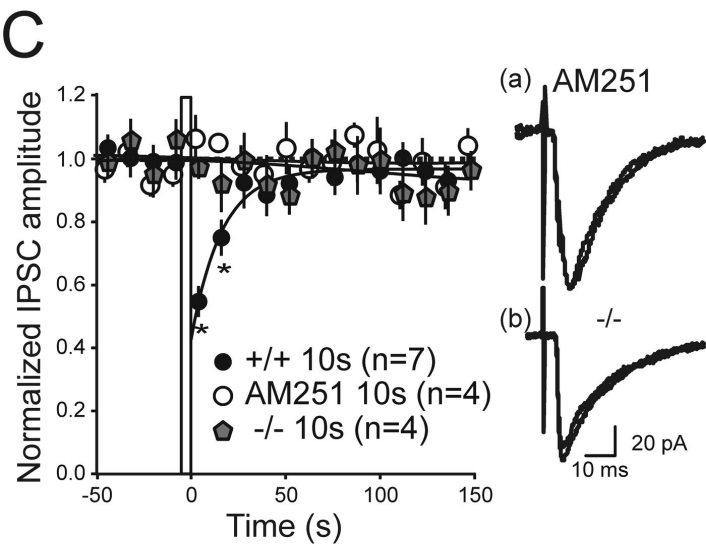
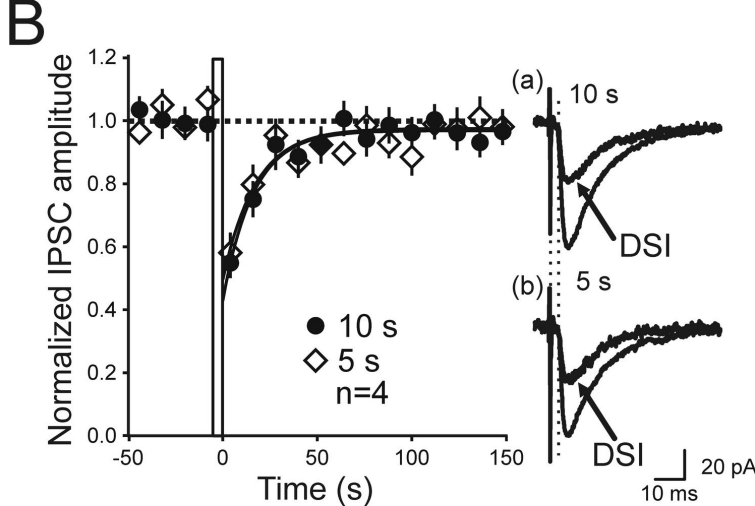
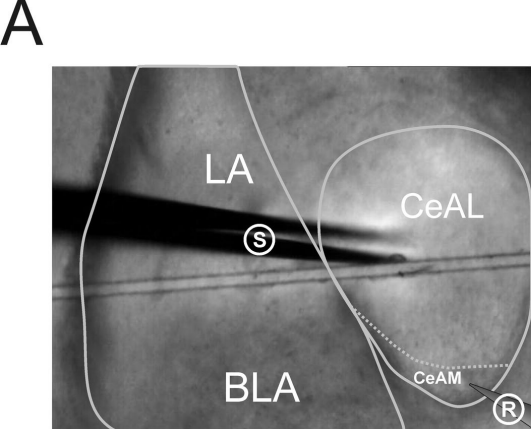


Fig 3

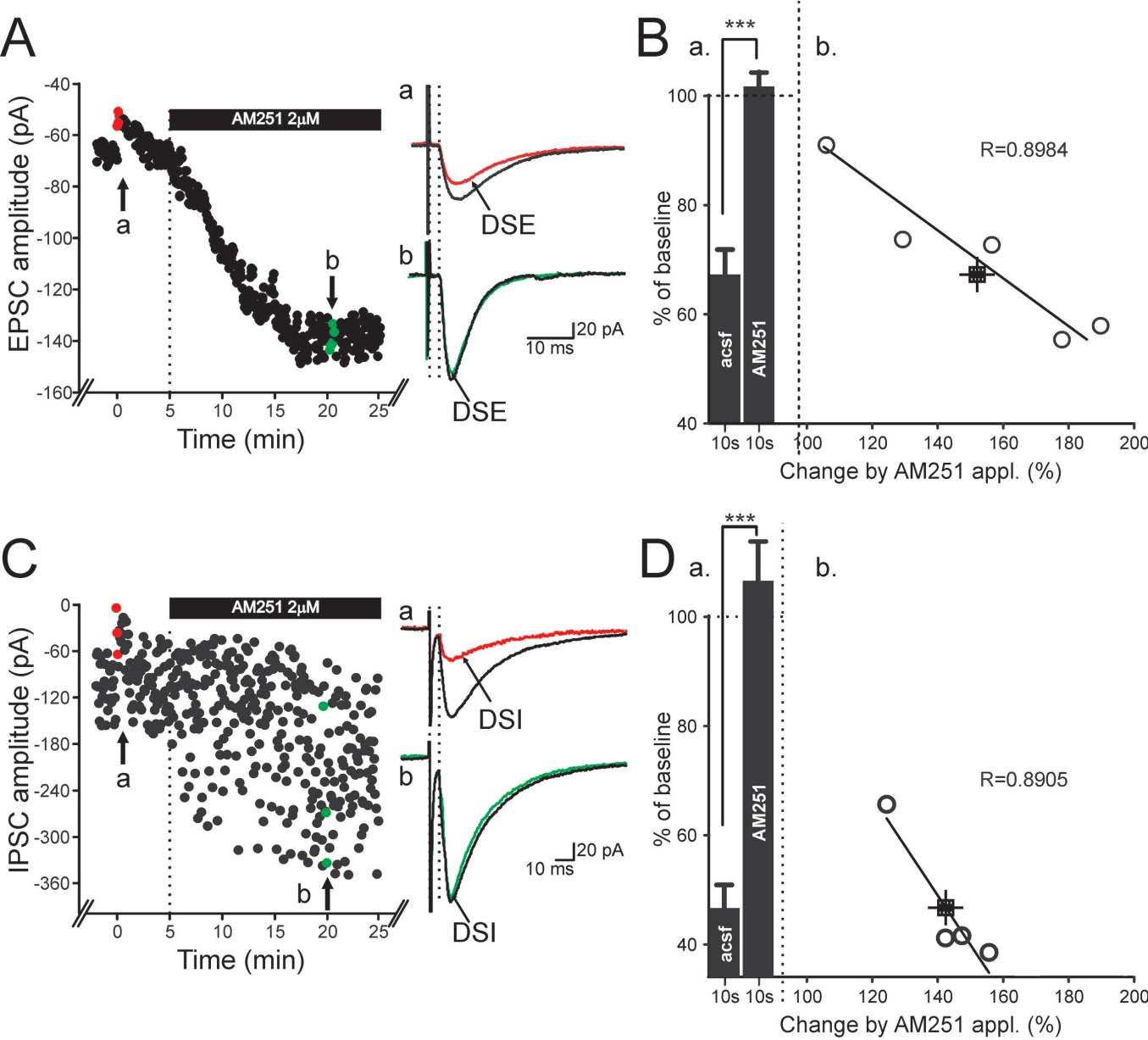


Fig 4

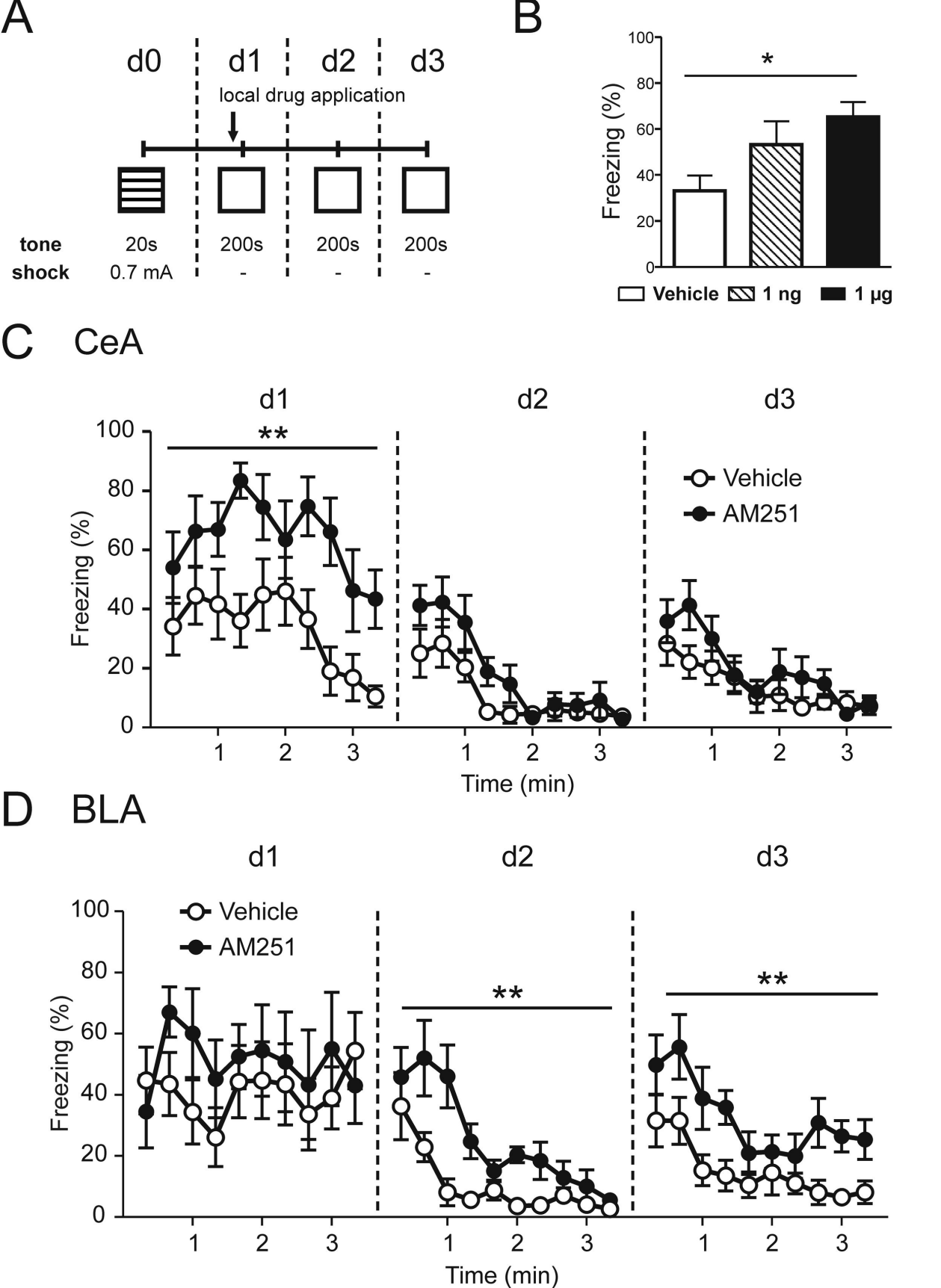


Fig. 5

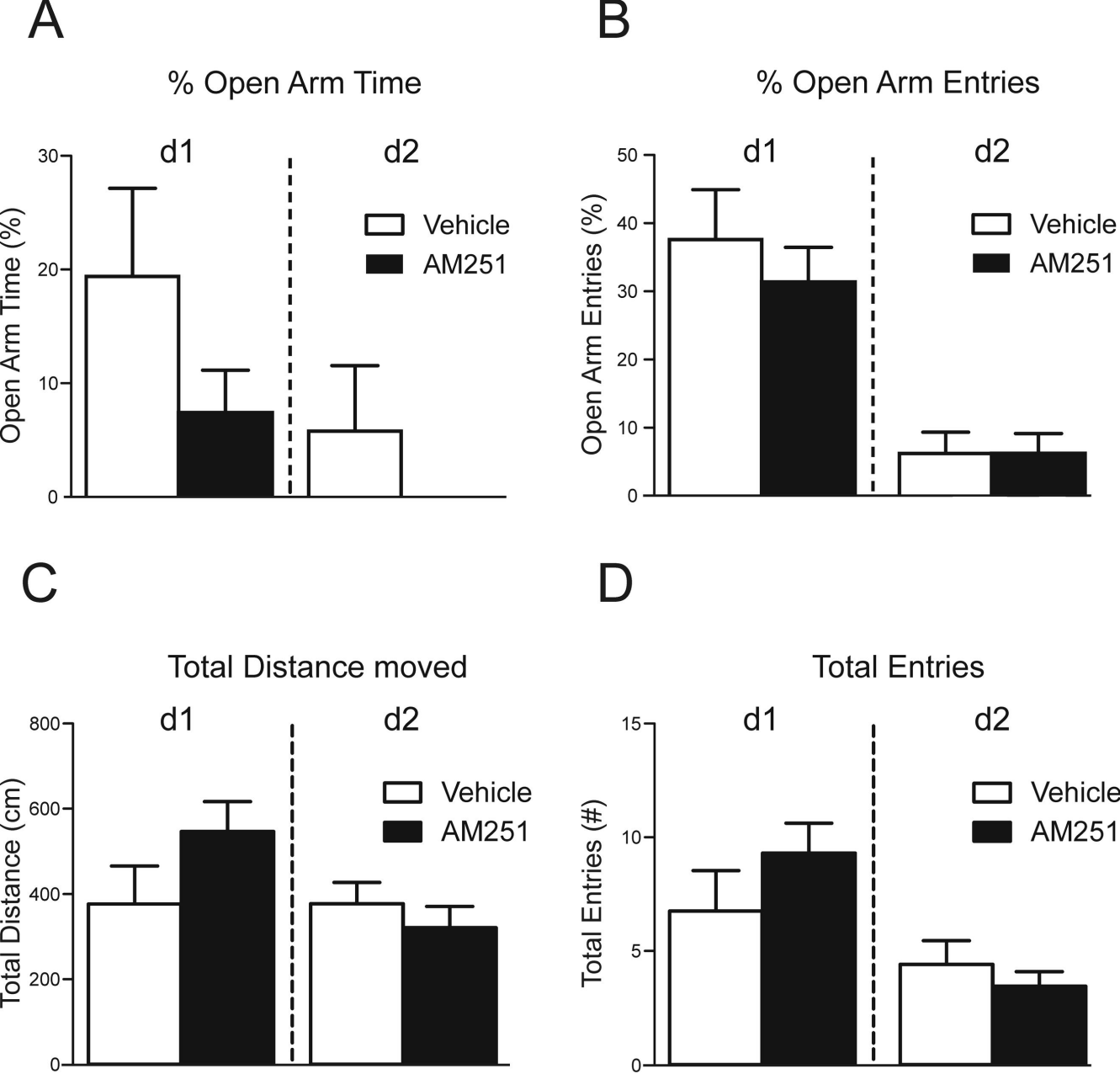
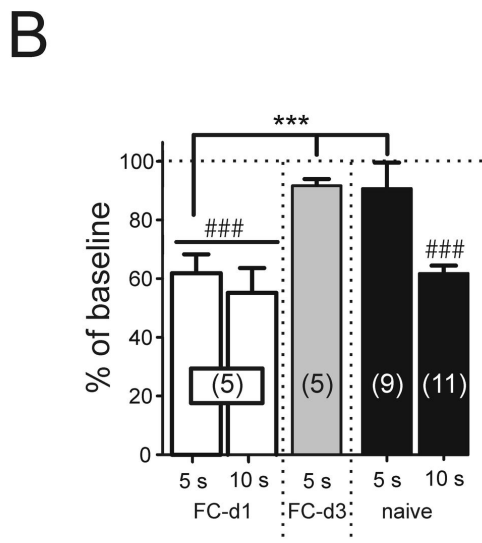
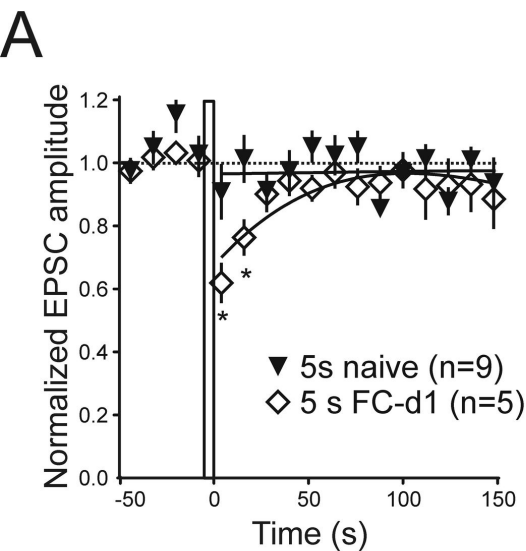


Fig. 6

DSE: BLA → CeAM



DSI: CeAL → CeAM

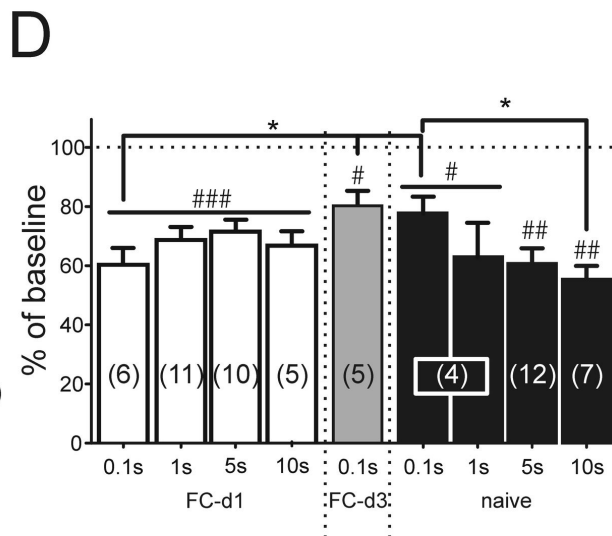
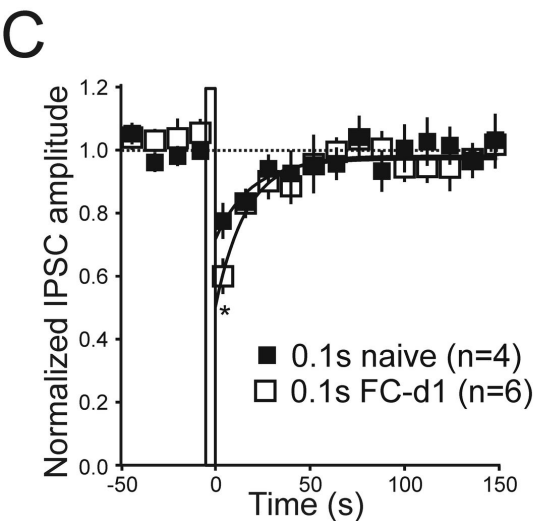


Fig. 7