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Recombinant Deg/HtrA proteases from *Synechocystis* sp. PCC 6803 differ in substrate specificity, biochemical characteristics and mechanism

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ABSTRACT

Cyanobacteria require efficient protein quality control mechanisms to survive under dynamic, often stressful environmental conditions. It was reported that three serine proteases, HtrA, HhoA and HhoB are important for survival of *Synechocystis* sp. PCC 6803 under high light and temperature stresses and might have redundant physiological functions. Here we show that all three proteases can degrade unfolded model substrates, but differ in respect to cleavage sites, temperature and pH optima. For recombinant HhoA, and to a lesser extent for HtrA, we observed an interesting shift in the pH optimum from slightly acidic to alkaline in the presence of Mg^{2+} and Ca^{2+} ions. All three proteases formed different homo-oligomeric complexes with and without substrate, implying mechanistic differences in comparison to each other and to the well-studied *Escherichia coli* orthologues DegP and DegS. Deletion of the PDZ domain decreased, but not abolished proteolytic activity of all three proteases, and prevented substrate-induced formation of complexes higher than trimers by HtrA and HhoA. In summary, biochemical characterisation of HtrA, HhoA and HhoB lays the foundation for a better understanding of their overlapping, but not completely redundant stress resistance functions in *Synechocystis* sp. PCC 6803.

Key words: biochemical characterisation, oligomerisation, PDZ domain, serine proteases, substrate specificity, *Synechocystis* sp. PCC 6803

Abbreviations used: BSA, bovine serum albumin; Deg, degradation of periplasmic proteins; HtrA, high temperature requirement A; PDZ domain, domains originally identified in the post-synaptic density protein, the *Drosophila* tumour suppressor disc-protein and the tight-junction protein Zo-1; rHtrA, rHhoA, rHhoB, recombinant HtrA, HhoA, HhoB, respectively; ROS, reactive oxygen species.

INTRODUCTION

The comparatively self-sufficient lifestyle of photosynthetic organisms comes at a price, a high inherent risk of damage to proteins and pigments by reactive oxygen species (ROS) that are formed as byproducts of oxygenic photosynthesis, particularly under unfavourable environmental conditions [1]. In the natural environment cyanobacteria, such as *Synechocystis* sp. PCC 6803 (hereafter *Synechocystis* 6803), which was isolated from a fresh-water lake, are exposed to tremendous changes in light intensities and temperatures. Under these conditions formation of ROS and subsequent damage to proteins is increased [2]. To cope with this problem, *Synechocystis* 6803 contains a variety of chaperones and proteases that monitor proper fold and function of proteins [3]. Particularly interesting candidates are enzymes of the multifunctional Deg/HtrA family of ATP-independent serine endopeptidases.

Deg/HtrA proteases are defined by the combination of a catalytic domain of the S1B serine proteases, containing histidine-aspartate-serine as a catalytic triad, with one or more C-terminal PDZ domains [4]. The PDZ domains mediate specific protein-protein interactions by preferentially binding to the C-terminal 3-4 amino acid residues of the target proteins [5-7]. In *Escherichia coli*, DegP/HtrA is a major heat shock protein that acts both as chaperone and as unspecific housekeeping protease on denatured proteins [8], whereas its membrane-anchored paralogue DegS/HhoB acts as a sensor for folding stress in the periplasm and initiates the σ^E stress response [9-11]. *Synechocystis* 6803 has three genes encoding Deg/HtrA proteases, named *htrA/degP* (slr1204), *hhoA/degQ* (sll1679) and *hhoB/degS* (sll1427) after their *E. coli* orthologues [12]. However, phylogenetic analysis showed that the three Deg/HtrA paralogues of *Synechocystis* 6803 are more closely related to each other than to the *E. coli* orthologues carrying the same name, and *vice versa* [13]. In order to avoid confusion and prevent assumptions purely based on the name, we will hereafter use the designations HtrA, HhoA and HhoB for the *Synechocystis* 6803 Deg/HtrA proteases, and DegP, DegQ and DegS for the *E. coli* paralogues.

Crystallographic studies showed that the basic functional unit of Deg/HtrA proteases is a homo-trimer that is stabilised by residues of the protease domains [11,14,15]. The *E. coli* chaperone/protease DegP further stacks two of the trimers with the catalytic centres facing each other and thus forms a very compact, proteolytically inactive, hexameric complex [14]. DegP is activated through reversible conformational changes triggered by binding of peptides to the first of its two PDZ domains [6,16-19] or by a shift to higher temperature [8]. In solution, activated DegP forms large spherical 12- or 24-mers, composed of 4 or 8 homo-trimers, enclosing the substrates [20,21]. On lipid membranes, *E. coli* DegP/HtrA forms bowl-shaped structures with 4-, 5-, or 6-fold symmetry with a trimer as the basic structural unit [22]. In contrast, the *E. coli* stress sensor DegS is a trimer that is anchored in the plasma membrane by its N-terminal transmembrane domain, exposing the protease and PDZ domains to the periplasm in a proteolytically inactive conformation [11]. Binding of activating peptides, such as the C-termini of unfolded outer membrane porins, to the PDZ domains triggers conformational changes that activate DegS [5,7,10].

In comparison to the detailed studies on *E. coli* Deg/HtrA proteases, little is known about the physiological function and biochemical mechanism of *Synechocystis* 6803 orthologues. Bioinformatic analysis of the primary sequences of HtrA, HhoA and HhoB showed that all three proteins contain only a single C-terminal PDZ domain [13,23,24], predicted secretory signal peptides of 34 and 25 residues for HhoA and HhoB, respectively, and a N-terminal

transmembrane-spanning domain of HtrA [23]. Biochemical analysis of the recombinant HhoA (rHhoA) suggested that similar to *E. coli* DegP [25] also this protease functions as a general housekeeping enzyme in the removal of misfolded proteins.

Earlier studies investigating the physiological function of cyanobacterial Deg/HtrA proteases focused mainly on their proposed role in the selective degradation of the D1 protein from photosystem II, which is particularly susceptible to photooxidative damage and quickly turned over under high intensity light stress (reviewed in [26-28]). Analyses of a *Synechocystis* 6803 triple $\Delta htrA/hhoA/hhoB$ mutant showed that these proteases are not essential for the degradation of the D1 protein when exposed to high intensity light [29] or UV-B radiation [30]. Nevertheless, Deg/HtrA proteases play vitally important roles in the resistance to heat and high intensity light stresses [3,29,31]. The triple $\Delta htrA/hhoA/hhoB$ mutant accumulated more oxidatively damaged proteins than the wild type, did not grow at high intensity light or high temperature, and showed severe deficits in phototaxis [29]. This study also suggested an astonishing overlap in function of Deg/HtrA proteases because these severe phenotypes were only observed for the triple $\Delta htrA/hhoA/hhoB$ mutant, but not for any of the double deletion mutants. This is particularly puzzling because proteomic analyses found the *Synechocystis* 6803 Deg/HtrA proteases in different subcellular compartments. HtrA was found in the outer membrane [32], HhoA in the periplasm [33] and associated with the plasma membrane [34], while no experimental data for the predicted periplasmic localisation of HhoB have been reported.

In this study, we have expressed all three proteases as soluble recombinant enzymes (rHtrA, rHhoA and rHhoB, respectively) in active and inactive variants in order to explore similarities and differences in their enzymatic properties. We show that rHtrA, rHhoA and rHhoB can degrade unfolded model substrates, but with different preferences regarding cleavage site, pH and temperature. Furthermore, accumulation of distinct degradation intermediates and formation of different oligomeric complexes indicate diverging molecular mechanisms, both between the three cyanobacterial Deg/HtrA proteases, as well as in comparison to the well-studied *E. coli* orthologues.

EXPERIMENTAL

Strains and culture conditions

A glucose-tolerant wild type strain of *Synechocystis* sp. PCC 6803, was grown at a light intensity of $50 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ at 30°C in liquid BG-11 medium [35] buffered with 10 mM TES, pH 8.2.

Plasmid construction

Genomic DNA was isolated from *Synechocystis* 6803 using a kit (Roche Diagnostics). Cloning of recombinant HhoA constructs has been described previously [25]. The sequences encoding N-terminally truncated active HtrA and HhoB protease constructs (Figure 1A) were amplified by PCR from genomic DNA with gene specific primers (Operon Biotechnologies) and ligated into the pET151-D/TOPO expression vector using the Champion directional cloning kit (Invitrogen) according to manufacturer's instructions. The following primers were used: For rHtrA 5'-CAC CTC CAT TGC TCC TGC CAA TGA G -3' and 5'-GCA TTA TTG GGG AGG GGC GCT GG -3' and for rHhoB 5'-CAC CAG TCT GAC TCC TGC CCC GGT -3' and 5'-GCA TTA TTG ACC CAA ATC TTC GGG A -3'. Plasmids encoding proteolytically inactive variant were

obtained by changing the codons for the active site Ser residues to Ala with the Quikchange II point mutagenesis kit (Stratagene). The codon for Ser₂₉₆ of rHtrA was mutated to Ala (rHtrA_{SA}) using the primer pair 5'- CAT TAA CCC TGG TAA CTC TGG TGG TC -3' and 5'- CAC CAG CGT TAC CCG GGT TAA TGG CC - 3', which also introduce a silent point mutation that generates an additional *Sma*I restriction site. The codon for Ser₂₅₈ of rHhoB was mutated to Ala (rHhoB_{SA}) using the primers 5'- GGC AAT GCC GGT GGG CCC CTG CTC - 3' and 5'- CAG GGG CCC ACC GGC ATT GCC TGG -3', which also introduces an *Apa*I restriction site through silent point mutation. Active and inactive PDZ domain deletion constructs were obtained by the same PCR-based cloning strategy, using the plasmids encoding active or inactive HtrA and HhoB as templates. The following primers were used: For rHtrA_{SA}ΔPDZ, 5'- CAC CTC CAT TGC TCC TGC CAA TGA G -3' and 5'- GC TCA TTT GCC CGT AGC AAT TAA CTG -3', for rHhoBΔPDZ and rHhoB_{SA}ΔPDZ, 5'- CAC CAG TCT GAC TCC TGC CCC GGT -3' and 5'- GC TCA CAT TTT GCC CTT GGT AAA CAG -3'. All inserted sequences and orientation in the plasmids were confirmed by DNA sequencing (GATC Biotech AG).

Expression and purification of recombinant protein

Chemically competent *E. coli* BL21(DE3)Star cells (Invitrogen) transformed with expression plasmids were grown in 50 to 100 ml Lysogeny Broth (LB) minimal medium containing 100 μg mg⁻¹ ampicillin (LB/Amp100) at 37°C over night. These pre-cultures were harvested by centrifugation (3 000 g, 8 min, 4°C), resuspended and further grown in 1 to 2 l LB/Amp100 at 19°C to OD₆₀₀ = 0.4 to 0.6. Expression of recombinant protease constructs was induced by the addition of 0.1 mM isopropyl-1-thio-D-galactoside (IPTG). Cultures were further grown over night at 19°C and then harvested by centrifugation (5 000 g, 10 min, 4°C). Cell pellets were resuspended in 10 to 20 ml binding buffer (50 mM HEPES-NaOH, 300 mM NaCl, pH 8.0) and stored at -20°C.

E. coli cells were lysed on ice by 10 to 15 repeats of 10 s sonication with 20 s cooling interval and centrifuged (26 000 g, 1 h, 4°C). For protease activity assays with β-casein and complex formation assays, recombinant protease constructs were purified from the soluble cell lysate by nickel affinity chromatography with HisTrap column connected to an Äkta purifier FPLC system (GE Healthcare) as described [25]. For all protease activity and reconstitution assays, recombinant protease constructs were purified at 5°C from filtered soluble cell lysates (45 μm filter) using His GraviTrap affinity columns (GE Healthcare) according to manufacturer's instructions. Application in binding buffer was followed by a washing step with 10 ml binding buffer containing 60 mM imidazole and an elution step with 5 ml elution buffer (50 mM HEPES-NaOH, 300 mM NaCl, 500 mM imidazole, pH 8.0). Eluate was collected in 1 ml-fractions, then diluted 10-fold with distilled water and concentrated using Amicon Ultra centrifugal filter devices with a molecular weight cut-off of 10 kDa (Amicon) in order to decrease the imidazole concentration and the buffer strength. Protein concentrations were determined by Bradford protein assay (Biorad) with bovine serum albumin (BSA) solutions as standard.

Analysis of protease complex formation

Elution fractions containing approximately 200 μg purified Deg/HtrA protease constructs were incubated either alone or in a 1:3, 1:1 or 3:1 molar ratio with β-casein (Sigma-Aldrich) as model substrate for 10 min at room temperature. Complex sizes were estimated by size-exclusion chromatography using a pre-packed Superdex 200 column (GE Healthcare). The column was

repeatedly calibrated with high and mid-weight molecular mass markers (GE Healthcare) according to manufacturer's instructions.

Protease activity assays

For protease activity assays, purified protease constructs were incubated with an excess of β -casein, bovine serum albumin (BSA), or lysozyme in 250 mM Tris, pH 7.0, supplemented with 20 mM CaCl_2 for 2 h at 40°C. Unfolding of lysozyme was induced by incubation with 20 mM DTT for 3 h at 40°C prior to protease activity assays.

Fluorogenic protease activity assays were performed using the Enzchek Protease Assay (Invitrogen). For each reaction, 3 μg purified recombinant protease were incubated with 0.5 μg BODIPY TR-X casein in a total volume of 200 μl in the presence of 0.1 mM NaN_3 . To measure the pH-optima, 10 mM MES (up to pH 6.5) or HEPES (from pH 7.0) were used to buffer the solutions and the measurements were performed at 30°C. Fluorescence signals measured at a wavelength of 617 nm were followed with an Infinite 200 microplate reader (Tecan) using 589 nm as excitation wavelength. For kinetic measurements at different pH, a fluorescence signal increase was followed over time, recording 6 replicates every 10 min, while incubating the sample plate in darkness at 30°C inside the microplate reader. The activity was calculated from the initial rate of fluorescence emission. For determination of the temperature dependence, assays were incubated in 10 mM MES and at the optimal pH of the three Deg/HtrA proteases (pH 5.5 for rHtrA, pH 6.5 for rHhoA and rHhoB) at a temperature range from 10°C to 65°C in a PCR cyclor before measuring total fluorescence at room temperature after 40 min incubation.

Sequencing of proteolytic β -casein fragments

To identify β -casein cleavage sites 200 pmol purified recombinant proteases (active, or inactive variants as negative controls) were incubated in the presence of 10 mM CaCl_2 with 75 μg β -casein at 40°C for several hours. Samples were taken after 0, 30, 90 min and 10 h and analysed by SDS-PAGE (see below). Proteolytic fragments of cleaved β -casein were excised from the gel and sent for N- and C-terminal sequencing to the Protein Analysis Centre (PAC, Karolinska Institute, Stockholm, Sweden).

Protein analysis

Proteins were solubilised in lithium dodecyl sulphate (LDS) buffer and separated by SDS-PAGE as described [36]. Gels were stained with Coomassie brilliant blue R250 [37].

Bioinformatics

Sequence analysis was performed using SMART [38], TargetP [39] and SignalP [40] programs. For engineering of mature, soluble protease constructs, the secondary structures of HhoA, HhoB and HtrA were analysed with PsiPred v2.5 [41] and compared to published structures of *E. coli* DegP [14], DegS [11], and human HtrA2/Omi [15] and sequence alignments [4,23].

RESULTS AND DISCUSSION

Engineering and heterologous expression of Deg/HtrA protease constructs of *Synechocystis* 6803

To investigate the enzymatic properties of three cyanobacterial Deg/HtrA proteases in detail, we expressed rHtrA, rHhoA and rHhoB in their active form as recombinant fusion proteins with C-terminal His-tag using the *E. coli* expression system. For controls, inactive versions of the proteases with mutations in the catalytic site were generated by site-directed point mutagenesis, expressed and purified under similar conditions. Based on earlier studies of membrane-anchored human HtrA2 [15] and *E. coli* DegS [10,11], the putative transmembrane-domain of HtrA from *Synechocystis* 6803 was truncated after the residue Gln₉₃ (Figure 1A and Supplementary Figure S1). Both active rHtrA, as well as inactive rHtrA_{SA}, with Ser₂₉₆ replaced by Ala, were obtained in sufficient quantity and purity after purification by Ni²⁺-affinity chromatography (Figure 1B). rHhoA and its proteolytically inactive version rHhoA_{SA}, both lacking the predicted signal peptide, were expressed and purified as described (Figure 1A and B; [25]). HhoB contains a predicted N-terminal signal sequence of 25 residues and a long additional N-terminal domain of 56 residues (compared to only 23 residues in HhoA) between signal peptide and protease domain. Based on our experience with rHhoA, we first tried to express rHhoB lacking only the predicted signal sequence. However, this construct could not be expressed as soluble protein and its yield was extremely low (data not shown). Therefore, we further truncated HhoB after Gln₅₀, placing the introduced His-tag in approximately the same distance from the protease domain as in rHhoA. A third rHhoB construct was truncated after Gln₇₄, close to the predicted first helix of the protease domain (Supplementary Figure S1). Of these, only the rHhoB construct truncated after Gln₅₀ yielded low amounts of active enzyme (Figure 1B, hereafter rHhoB). Also the inactive version rHhoB_{SA}, with Ser₂₅₈ replaced by Ala, was obtained in similar yield and purity (Figure 1B).

To investigate the role of the single PDZ domain in each of the three *Synechocystis* 6803 Deg/HtrA proteases, we further created PDZ domain deletion constructs by truncation within the predicted loops connecting the protease and PDZ domains (Figure 1A and Supplementary Figure S1). rHtrA and rHtrA_{SA} were truncated after Lys₃₄₂ to yield rHtrA Δ PDZ and rHtrA_{SA} Δ PDZ, respectively. Cloning and expression of rHhoA Δ PDZ and its proteolytically inactive variant rHhoA_{SA} Δ PDZ, both truncated after Gly₂₈₂, were expressed and purified as described before (Figure 1A and C; [25]). Similarly, rHhoB and rHhoB_{SA} were truncated after Met₃₀₅, resulting in rHhoB Δ PDZ and rHhoB_{SA} Δ PDZ, respectively. All deletion constructs were obtained in good quantity and reasonable purity after Ni²⁺-affinity chromatography (Figure 1C).

All three Deg/HtrA proteases in *Synechocystis* 6803 are protein quality control factors

We assayed the activity and substrate specificity of the recombinant Deg/HtrA constructs using model substrates that differed with regard to their folding states, such as bovine β -casein, BSA, and lysozyme. All three protease constructs with intact PDZ domain, rHtrA, rHhoA and rHhoB, were able to cleave the naturally unfolded β -casein, but not correctly folded globular BSA or lysozyme (Figure 2A). Reduction of lysozyme with DTT, which induces partial unfolding by breaking disulfide bonds, allowed its cleavage by HhoA and to a lower extent by rHtrA, but not by HhoB (Figure 2A, lower panel). Under the chosen conditions, equal amounts of rHtrA, rHhoA and rHhoB showed very different activity: Little uncleaved β -casein remained in assays with rHtrA, and a prominent degradation fragment appeared. rHhoA completely degraded the excess of β -casein, whereas rHhoB showed only little activity and generated a prominent degradation fragment with a slightly lower molecular mass than intact β -casein (Figure 2A). All substrates tested were not degraded in the presence of inactive protease variants, thus excluding

that the substrate degradation by active variants was mediated by impurities in the preparation (Figure 2A).

We consistently assayed a lower activity for rHhoB than for rHtrA and rHhoA. We can not exclude that this is merely caused by unfortunate engineering of the rHhoB truncation constructs, but the chosen truncation of HhoB at Gln₅₀ should have affected neither the fold of the protease domain, nor any other important domain [24]. Instead, this truncation made rHhoB more similar to rHhoA, which is the most active recombinant protease in our hands. Thus, the low activity of rHhoB might also reflect, similarly to *E. coli* DegS [9], a more stringent substrate specificity, a function as chaperone rather than protease, a requirement for yet unidentified cofactors, such as allosteric activators [17-19], or a need for the formation of a hetero-oligomeric complex. A formation of hetero-hexameric complex was reported for chloroplast-located Deg5 and Deg8 in *Arabidopsis thaliana* [42].

Taken together, this provides evidence that all three proteases, HtrA, HhoA and HhoB, might act as protein quality control factors degrading denatured and damaged proteins [29]. This further suggests remarkable mechanistic differences to Deg/HtrA proteases in *E. coli*, where DegP and DegQ with two PDZ domains [17,43] act as quality control factors, whereas DegS, containing a transmembrane domain and a single PDZ domain, acts as a stress sensor [11,17].

Deletion of the PDZ domain severely impaired proteolytic activity

rHtrA Δ PDZ showed only residual activity against β -casein and reduced lysozyme (Figure 2B). Also rHhoA Δ PDZ degraded both unfolded substrates less efficiently than its variant with the intact PDZ domain, and larger degradation fragments accumulated after proteolysis of reduced lysozyme (Figure 2B). rHhoB Δ PDZ showed only minute activity against β -casein and no activity was detected against reduced lysozyme (Figure 2B). As expected, no degradation of substrates tested was assayed in the presence of inactive protease variants lacking PDZ domains (Figure 2B). Taken together, this data suggests that the PDZ domains are not absolute requirements for proteolytic activity of all three proteases but they are important for the regulation of proteolytic activity and/or for presenting the substrate to the protease domain.

rHtrA, rHhoA and rHhoB generate distinct β -casein degradation products

Proteolysis of the model substrate β -casein by recombinant Deg/HtrA proteases with intact PDZ domains was further investigated in more detail, and different enzymatic kinetics and distinct degradation intermediates were observed (Figure 3A). In the presence of rHtrA only traces of β -casein could be detected after 30 min of incubation, but proteolytic fragments 1 and 2 remained stable and were not further degraded during 90 min of incubation (Figure 3A, left panel). Similarly, most of the β -casein was cleaved by rHhoA within 30 min and the generated proteolytic fragment 3, but not fragments 4 and 5, disappeared within 90 min (Figure 3A, middle panel). Longer incubation times lead to a complete degradation of all fragments (Figure 2A, [25]). In contrast, a very low proteolytic activity was assayed for rHhoB. Significant amounts of β -casein were detected even after 10 h of incubation with this protease and the proteolytic fragment 6 accumulated progressively with the time of incubation (Figure 3A, right panel). This suggests that *Synechocystis* 6803 Deg/HtrA proteases release their substrates after defined cleavage sites, similar to *E. coli* DegS [10], *A. thaliana* Deg2 [44] and Deg15 [45,46]. This is a remarkable contrast to *E. coli* DegP, which digests unfolded substrate proteins progressively without accumulation of intermediates [6,21].

Cleavage site specificity

The cleavage specificity of proteases is defined by the amino acid composition of the substrate around the cleavage site, with non-prime site residues P3, P2, P1 preceding the cleavage site, and prime-site residues P1', P2', P3' following the cleavage site [47]. To gain insight into the cleavage site specificity of rHtrA, rHhoA and rHhoB, we analysed the most abundant proteolytic β -casein fragments generated by each protease by N- and C-terminal sequencing. The majority of the sequenced fragments contained the N-terminus of mature β -casein, while cleavage occurred at the C-terminal part of this substrate (Figure 3B and Table 1). Three C-terminal cleavage sites, Val₁₆₂, Gln₁₄₁ and Val₁₃₀, were identified for rHtrA (Figure 3B and Table 1). A common cleavage site at Leu₁₆₅ was found for rHhoA and rHhoB, however, rHhoA additionally cleaved β -casein at Ala₁₀₁ generating two proteolytic fragments, the N-terminal 1-101 as well as the C-terminal 102-199 amino acid fragments (Figure 3B and Table 1).

E. coli DegP showed a strong preference for small hydrophobic residues (valine, alanine, threonine and isoleucine) in the P1 position, whereas P2 and P3 positions are less defined [6]. Our data show a similar trend for rHtrA and rHhoA (Figure 2B). It was further demonstrated that PDZ1, the first of the two PDZ domains of *E. coli* DegP, recognised the hydrophobic C-terminus of substrates and presented the substrate for cleavage [6]. Since DegP cleavage occurred at a hydrophobic site approximately 12 to 16 amino acids further towards the N-terminus, the resulting cleavage fragment could again be bound by a PDZ1 domain of the trimer, resulting in processive cleavage of the substrate into peptides of 10 to 20 residues [6]. Similarly, the PDZ domains of *Synechocystis* 6803 Deg/HtrA proteases might preferentially bind the hydrophobic residues at the C-terminus of β -casein (-PFPIIV) and present the substrate to the catalytic site for cleavage, explaining the absence of larger C-terminal fragments. However, all three *Synechocystis* Deg/HtrA proteases are not as processive as *E. coli* DegP, possibly due to the lack of a second PDZ domain.

Inactive *Synechocystis* Deg/HtrA proteases form different oligomers

Earlier structural studies have shown that the basic unit of Deg/HtrA proteases is a homo-trimer [11,14,15]. Two trimers of *E. coli* DegP are stacked to a cage-like hexamer by both PDZ domains of the subunits [14,48,49]. Homology modelling predicted all three *Synechocystis* 6803 Deg/HtrA proteases to form trimers [23]. Surprisingly, we found earlier that also rHhoA assembles into a hexamer in solution [25], despite the fact that it only contains one PDZ domain. Here we assayed the oligomerisation state of rHtrA_{SA} and rHhoB_{SA} by size-exclusion chromatography. rHtrA_{SA} eluted with a volume of 13.2 ml, corresponding to an apparent molecular mass of 140 kDa (Figure 4A), which is consistent with a trimer formation. Published crystal structures show that Deg/HtrA protease trimers from *Homo sapiens* and *E. coli* are disk-shaped and wider than tall if viewed from the side [11,15]. Therefore, such trimers have a larger hydrodynamic radius than globular proteins of the same molecular mass, resulting in a larger-than-expected apparent molecular mass when estimated by the elution volume in a size-exclusion chromatography calibrated with globular standard marker proteins. However, the rHtrA_{SA} complexes were unstable, and we frequently observed monomers eluting at approximately 14.6 ml (apparent molecular mass of ca. 60 kDa). In agreement with our earlier experiments [25], rHhoA_{SA} formed a hexamer eluting with a volume of 11.5 ml (apparent molecular mass 320 kDa) (Figure 4B, [22]). rHhoB_{SA} eluted at 13.1 ml, indicating a trimeric complex with an apparent molecular mass of approximately 160 kDa (Figure 4C). For rHtrA_{SA} and rHhoB_{SA} additional peaks were observed at the void volume of the Superdex 200 size-exclusion columns, but very little or no

protein was detected in these fractions after SDS-PAGE and Coomassie-blue staining (Figs 4A and C, lower panels), indicating that these peaks might represent SDS-insoluble aggregates.

Substrate binding to rHtrA_{SA} and rHhoA_{SA} changes their oligomerisation states

In the presence of substrates *E. coli* DegP massively re-arranges and assembles several trimers into large oligomeric complexes, mediated by interactions of both PDZ domains [16,20,21]. To test the effect of substrate binding on the oligomerisation state of cyanobacterial Deg/HtrA proteases, we incubated rHtrA_{SA}, rHhoA_{SA} and rHhoB_{SA} with β -casein as model substrate prior to size-exclusion chromatography. In the presence of β -casein, the elution diagram of rHtrA_{SA} displayed an additional peak at ca. 10.5 ml, indicating the formation of a complex with an apparent molecular mass of approximately 580 kDa (Figure 4A). β -casein alone elutes at 13.5 ml (data not shown, compare with Figure 4C), but co-elutes with rHtrA_{SA} in the 10.5 ml-peak, clearly demonstrating that this complex is the result of β -casein binding by rHtrA_{SA} (Figure 4A). As for rHtrA_{SA} alone, an additional prominent peak is detected at the void volume of the column, but only little protein is observed in the sample aliquot of this fraction after SDS-PAGE analysis (Figure 4A). Due to steric constraints, the formation of such higher-order oligomeric HtrA-substrate complexes appears unlikely under physiological conditions since HtrA is anchored in the outer membrane [30]. This suggests that HtrA may function similar to human HtrA2/Omi, which is anchored as an inactive zymogen to the inner mitochondrial membrane [48]. A soluble, active form of HtrA2/Omi is released by cleavage of the N-terminal 133 amino acids that contain the transmembrane anchor (Figure 1A). This soluble form translocates from the mitochondria to the cytosol and contributes to apoptosis [50]. A similar release of the cyanobacterial HtrA protease from the outer membrane would also help to explain the overlap in its function with HhoA and HhoB under certain conditions.

Similar to rHtrA_{SA}, also rHhoA_{SA} changed its complex size dramatically in the presence of β -casein, featuring prominent additional peaks at 8.3 ml and at 9.5 ml (Figure 4B). Both rHhoA_{SA} and β -casein co-eluted in the peak at 9.5 ml, indicating the formation of a large rHhoA_{SA}/ β -casein complex of approximately 670 kDa. Hardly any protein was observed in the fraction containing the early peak at 8.3 ml, which is close to the column void volume and might therefore contain insoluble aggregates. In contrast, the elution profile of rHhoB_{SA} in the presence of β -casein showed no additional peaks and the elution profiles of rHhoB_{SA} and β -casein alone were very similar, indicating that these two proteins did not form a stable complex under conditions tested (Figure 4C). This observation is consistent with the low activity of rHhoB observed in proteolytic assays (Figure 2C) and may indicate either a problem of the rHhoB constructs that impairs formation of higher-oligomeric complexes or a lack of required cofactors, such as specific peptides [16-19]. It was reported that binding of specific effector-peptides to the PDZ domain-1 of *E. coli* DegP in a resting conformation causes a local rearrangement that is allosterically transmitted to the substrate-binding pocket of the protease domain and such activated protease state assembles into larger oligomeric particles [18-21]. Also *E. coli* DegS is activated by the binding of allosteric effectors to the PDZ domain, but in contrast to DegP, protease activation and substrate proteolysis are separated events [10,11].

Our studies on cyanobacterial Deg/HtrA proteases indicate that the formation of large oligomeric complexes in the presence of a substrate might be a common feature for this enzyme family (Figure 4). Unfortunately, size-exclusion chromatography is not suitable for a reliable determination of the exact oligomerisation state of rHtrA_{SA}/ β -casein and rHhoA_{SA}/ β -casein complexes due to the unknown number of bound β -casein molecules and its unknown effect on

the hydrodynamic radius of the complexes. However, in comparison to published data by Krojer and colleagues [21] and Jiang and colleagues [20], the formation of 12- or 24-meric complexes for rHhoA_{SA}, and 9- to 12-mers for rHtrA_{SA} is estimated.

Deletion of PDZ domains abolishes formation of oligomers beyond a trimer

We had shown earlier [25] that the deletion the PDZ domain of rHhoA prevents formation of a hexamer. Instead, rHhoAΔPDZ assembled into a trimer, which is consistent with the homology model that attributed trimerisation to the N-terminus of the protease domain [23]. Similarly, rHtrAΔPDZ and rHhoBΔPDZ were detected as trimers (data not shown). However, the trimers of all three constructs were not very stable and monomers were frequently observed. The equilibrium of trimers and monomers appeared concentration-dependent, with monomers prevailing at low protein concentrations. All three PDZ-domain deletion constructs, rHtrAΔPDZ, rHhoAΔPDZ and rHhoBΔPDZ, were not able to assemble to larger complexes in the presence of β-casein (summarised in Figure 5). This is in agreement with reports showing that peptide binding to the PDZ domain of *E. coli* DegP [16-18] promotes formation of higher-order-particles.

Mg²⁺ and Ca²⁺ increase activity of rHtrA, rHhoA and rHhoB

We characterised the proteolytic activity of rHtrA, rHhoA and rHhoB in biochemical terms using BODIPY TR-X-labelled casein as a fluorogenic substrate. The effect of increasing concentrations of divalent ions on the proteolytic activity of three Deg/HtrA proteases was tested. The values were normalised to the protease activity without the addition of divalent cations to display the actual increase of activity in the presence of divalent ions. While the activity of rHtrA and rHhoA increased almost 4- to 6-times in the presence of 5 mM CaCl₂ and remained constant at higher CaCl₂ concentrations, the activity of rHhoB increased more gradually up to 14-times reaching the maximal level at 10 mM CaCl₂ (Figure 6A). Similar results were obtained after the addition of MgCl₂ (Supplementary Figure S2A). The reason for this activation by Mg²⁺ or Ca²⁺ is not clear at the moment, but either: (i) divalent cations could interact with the substrate and make it more susceptible for degradation, and/or (ii) binding of divalent cations could affect the structure and thus activates the Deg/HtrA protease. The much stronger effect of Mg²⁺ or Ca²⁺ ions on rHhoB activity (Figure 6A) can be explained by the almost negligible activity in the absence of divalent ions (Figure 6B).

Overlapping, but distinct pH and temperature profiles of rHtrA, rHhoA and rHhoB

The pH preference profiles were measured for all three Deg/HtrA proteases in the absence (Figure 6B) and the presence (Figure 6C) of CaCl₂ at a range from pH 4.5 to 8.0. In the absence of CaCl₂ rHtrA showed a maximal proteolytic activity between pH 5.5 and 6.0, whereas rHhoA had its optimum at pH 6.5 (Figure 6B). Almost no proteolytic activity was detected for rHhoB (Figure 6B). However, the addition of 10 mM CaCl₂, resulted in strong changes in the pH optima leading to their broadening and a shift towards a basic pH (Figure 6C). For rHtrA the highest activity was detected at pH 7.0, whereas the activity of rHhoA progressively increased with higher pH (Figure 6C, please note that the fluorescence detector gain was decreased after the addition of divalent cations to avoid saturation). This striking shift in the pH optimum of HhoA in the presence of Ca²⁺ is in agreement with earlier observations [25]. Similar pH preference profiles were also observed after the addition of MgCl₂, ruling out a Ca²⁺-specific effect (data not shown). The activity of rHhoB in the presence of Ca²⁺ was constantly high at pH between 5.5 and 6.5 and decreased slightly at pH 6.5 to 8.0 (Figure 6C).

In the absence of Ca^{2+} , rHtrA, rHhoA and rHhoB were all most active at slightly acidic conditions. In the presence of 10 mM Ca^{2+} , both rHtrA and rHhoB showed a broader pH optimum with a maximum between pH 5.5 and 7.0. In contrast, rHhoA showed a striking shift of its pH optimum to basic conditions. To our knowledge, such a strong shift in the pH preference has not been reported for any other protease.

The effect of temperature on the proteolytic activity of Deg/HtrA proteases was assessed by measuring total fluorescence at one fixed time point after 40 min incubation at the indicated temperature (Figure 6D). To ensure that neither the substrate was limiting at the chosen time point, nor saturation of the detector occurred at/or near the fluorescence maxima, control experiments were performed, in which the fluorescence emission generated by degradation of BODIPY TR-X-labelled casein was measured over time at 30°C (Supplementary Figure S2B). Only low proteolytic activity was recorded for the three active Deg/HtrA proteases at a temperature ranging between 10 to 25°C (Figure 6D). While rHtrA had its temperature optimum between 30°C and 40°C the activity of rHhoA increased with rising temperature reaching its maximum at around 50 to 55°C and decreased rapidly at higher temperatures, probably due to denaturation of the enzyme (Figure 6D). A similar temperature curve has also been observed previously for rHhoA using Resorufin-labelled casein as substrate [25]. The temperature dependency of rHhoB was comparable to rHhoA, however, its optimum was shifted to slightly lower temperatures and the maximal activity was reached already at 45 to 50°C (Figure 6D).

Implications for the physiological function of HtrA, HhoA and HhoB in *Synechocystis* 6803

All three *Synechocystis* 6803 Deg/HtrA proteases were able to act on unfolded protein substrates, suggesting functions as general protein quality control proteases consistent with the stress-sensitive phenotype of the triple mutant $\Delta htrA/hhoA/hhoB$ [29]. The overlapping biochemical properties of HtrA, HhoA and HhoB may allow any two of the enzymes to compensate for the lack of the third, which could explain the striking lack of stress-sensitive phenotypes of the double deletion mutants in comparison to the triple mutant $\Delta htrA/hhoA/hhoB$ [29]. However, our data also suggests that each of the three enzymes could contribute differently to the overall proteolytic potential with varying environmental conditions. For instance, *Synechocystis* 6803 cells prefer to grow at alkaline conditions [51] and are commonly grown in BG11-media, which contain approximately 0.5 mM of combined Mg^{2+} and Ca^{2+} ions. Thus, HhoA would be the most active of the three *Synechocystis* 6803 Deg/HtrA proteases under optimal physiological conditions.

The observed activation of all three proteases by Ca^{2+} and Mg^{2+} ions and the slightly acidic pH optimum of HtrA and HhoB may be particularly important for an effective stress response to high intensity light. Sufficiently high intensity light saturates photosynthesis rates, causing acidification of the thylakoid lumen and increased formation of ROS [1]. This further leads to oxidative damage of photosynthetic protein complexes and release of Ca^{2+} from the thylakoid membranes to the lumen [26]. These conditions then would activate any of the three Deg/HtrA proteases. Since no reliable localisation studies exist for Deg/HtrA proteases in cyanobacteria it is not clear whether these enzymes are present in the periplasmic space, in the thylakoid lumen or in both compartments. Unfortunately, purification of the cyanobacterial thylakoid lumen proteome poses massive technical challenges [52] and thus neither a comprehensive proteomic analysis has been reported, nor is it possible to probe this compartment for the presence of Deg/HtrA proteases with antibodies. However, three Deg/HtrA proteases have been detected in the

thylakoid lumen of higher plants, where no other general proteases were reported [53,54]. Furthermore, out of the reported 16 paralogues of *A. thaliana*, the three cyanobacterial proteases show the highest similarity to the three thylakoid lumen-located Deg/HtrA enzymes [24]. Hence it is very likely that at least one of the *Synechocystis* 6803 Deg/HtrA proteases, such as e.g. HhoB, is located in the thylakoid lumen.

The temperature profiles of rHtrA, rHhoA and rHhoB give an indication how these enzymes may contribute to high temperature resistance. While rHtrA activity decreased at temperatures higher than 30°C, which is the preferred growth temperature of *Synechocystis* 6803, the activity of rHhoA and rHhoB increased to well above lethal temperatures. The soluble enzymes HhoA and HhoB should, therefore, play a more important role in protein quality control at high temperature than HtrA. Interestingly, *htrA* is the only gene regulated by the two-component system Hik10-Rre3 under salt and hyperosmotic stresses, which suggests an additional, more specific function of HtrA [55].

Conclusions

In summary, this family-wide biochemical characterisation of the *Synechocystis* 6803 Deg/HtrA proteases suggests complementary physiological functions of the three enzymes and highlights considerable mechanistic differences of HtrA, HhoA and HhoB, as compared to their well-studied *E. coli* orthologues. We have shown that the three Deg/HtrA proteases of *Synechocystis* 6803, expressed as soluble recombinant His-tagged fusion proteins, can act as protein quality control proteases. Distinct pH- and temperature-optima of the three enzymes, different cleavage sites and oligomerisation status speak in favour of partially overlapping but not completely redundant physiological functions.

AUTHOR CONTRIBUTION

Pitter Huesgen engineered Deg/HtrA constructs, performed oligomerisation studies together with Helder Miranda, Manuela Perthold and Holger Schuhmann, analysed the data and wrote draft of the paper. Helder Miranda and Lam Xuan Tam expressed and purified Deg/HtrA constructs and performed proteolytic assays. Holger Schuhmann performed bioinformatic analysis. Christiane Funk and Iwona Adamska co-conceived the research and edited the manuscript.

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REFERENCES

1. Barber, J. and Andersson, B. (1992) Too much of a good thing: light can be bad for photosynthesis. *Trends Biochem. Sci.* **17**, 61-66
2. Melis, A. (1999) Photosystem-II damage and repair cycle in chloroplasts: what modulates the rate of photodamage? *Trends Plant Sci.* **4**, 130-135
3. Sokolenko, A., Pojidaeva, E., Zinchenko, V., Panichkin, V., Glaser, V. M., Herrmann, R. G. and Shestakov, S. V. (2002) The gene complement for proteolysis in the cyanobacterium *Synechocystis* sp. PCC 6803 and *Arabidopsis thaliana* chloroplasts. *Curr. Genet.* **41**, 291-310
4. Clausen, T., Southan, C. and Ehrmann, M. (2002) The HtrA family of proteases: implications for protein composition and cell fate. *Mol. Cell* **10**, 443-455
5. Hasselblatt, H., Kurzbauer, R., Wilken, C., Krojer, T., Sawa, J., Kurt, J., Kirk, R., Hasenbein, S., Ehrmann, M. and Clausen, T. (2007) Regulation of the σ^E stress response by DegS: how the PDZ domain keeps the protease inactive in the resting state and allows integration of different OMP-derived stress signals upon folding stress. *Genes Dev.* **21**, 2659-1670
6. Krojer, T., Pangerl, K., Kurt, J., Sawa, J., Stingl, C., Mechtler, K., Huber, R., Ehrmann, M. and Clausen, T. (2008) Interplay of PDZ and protease domain of DegP ensures efficient elimination of misfolded proteins. *Proc. Natl. Acad. Sci. USA* **105**, 7702-7707
7. Sohn, J., Grant, R. A. and Sauer, R. T. (2007) Allosteric activation of DegS, a stress sensor PDZ protease. *Cell* **131**, 572-583
8. Spiess, C., Beil, A. and Ehrmann, M. (1999) A temperature-dependent switch from chaperone to protease in a widely conserved heat shock protein. *Cell* **97**, 339-347
9. Alba, B. M., Zhong, H. J., Pelayo, J. C. and Gross, C. A. (2001) *degS* (*hhoB*) is an essential *Escherichia coli* gene whose indispensable function is to provide σ^E activity. *Mol. Microbiol.* **40**, 1323-1333
10. Walsh, N. P., Alba, B. M., Bose, B., Gross, C. A. and Sauer, R. T. (2003) OMP peptide signals initiate the envelope-stress response by activating DegS protease via relief of inhibition mediated by its PDZ domain. *Cell* **113**, 61-71
11. Wilken, C., Kitzing, K., Kurzbauer, R., Ehrmann, M. and Clausen, T. (2004) Crystal structure of the DegS stress sensor: How a PDZ domain recognizes misfolded protein and activates a protease. *Cell* **117**, 483-494
12. Kaneko, T., Sato, S., Kotani, H., Tanaka, A., Asamizu, E., Nakamura, Y., Miyajima, N., Hirose, M., Sugiura, M., Sasamoto, S. et al. (1996) Sequence analysis of the genome of the unicellular cyanobacterium *Synechocystis* sp. strain PCC6803. II. Sequence determination of the entire genome and assignment of potential protein-coding regions. *DNA Res.* **3**, 109-136
13. Huesgen, P. F., Schuhmann H. and Adamska, I. (2005) The family of Deg proteases in cyanobacteria and chloroplasts of higher plants. *Physiol. Plant.* **123**, 413-420
14. Krojer, T., Garrido-Franco, M., Huber, R., Ehrmann, M. and Clausen, T. (2002) Crystal structure of DegP (HtrA) reveals a new protease-chaperone machine. *Nature* **416**, 455-459
15. Li, W., Srinivasula, S. M., Chai, J., Li, P., Wu, J. W., Zhang, Z., Alnemri, E. S. and Shi, Y. (2002) Structural insights into the pro-apoptotic function of mitochondrial serine protease HtrA2/Omi. *Nat. Struct. Biol.* **9**, 436-441

16. Krojer, T., Sawa, J., Huber, R. and Clausen, T. (2010) HtrA proteases have a conserved activation mechanism that can be triggered by distinct molecular cues. *Nat. Struct. Mol. Biol.* **17**, 844-852
17. Meltzer, M., Hasenbein, S., Mamant, N., Merdanovic, M., Poepsel, S., Hauske, P., Kaiser, M., Huber, R., Krojer, T., Clausen, T. et al. (2009) Structure, function and regulation of the conserved serine proteases DegP and DegS of *Escherichia coli*. *Res. Microbiol.* **160**, 660-666
18. Merdanovic, M., Mamant, N., Meltzer, M., Poepsel, S., Auckenthaler, A., Melgaard, R., Hauske, P., Nagel-Steger, L., Clarke, A. R., Kaiser, M. et al. (2010) Determinants of structural and functional plasticity of a widely conserved protease chaperone complex. *Nat. Struct. Mol. Biol.* **17**, 837-844
19. Meltzer, M., Hasenbein, S., Hauske, P., Kucz, N., Merdanovic, M., Grau, S., Beil, A., Jones, D., Krojer, T., Clausen, T. et al. (2008) Allosteric activation of HtrA protease DegP by stress signals during bacterial protein quality control. *Angew. Chem. Int. Ed. Engl.* **47**, 1332-1334
20. Jiang, J., Zhang, X., Chen, Y., Wu, Y., Zhou, Z. H., Chang, Z. and Su, S. F. (2008) Activation of DegP chaperone-protease via formation of large cage-like oligomers upon binding to substrate proteins. *Proc. Natl. Acad. Sci. USA* **105**, 11939-11944
21. Krojer, T., Sawa, J., Schäfer, E., Saibil, H. R., Ehrmann, M. and Clausen, T. (2008) Structural basis for the regulated protease and chaperone function of DegP. *Nature* **453**, 885-890
22. Shen, Q. T., Bai, X. C., Chang, L. F., Wu, Y., Wang, H. W. and Sui, S. F. (2009) Bowl-shaped oligomeric structures on membranes as DegP's new functional forms in protein quality control. *Proc. Natl. Acad. Sci. USA* **106**, 4858-4863
23. Jansen, T., Kidron, H., Taipaleenmäki, H., Salminen, T. and Mäenpää, P. (2005) Transcriptional profiles and structural models of the *Synechocystis* sp. PCC 6803 Deg proteases. *Photosynth. Res.* **84**, 57-63
24. Kieselbach, T. and Funk, C. (2003) The family of Deg/HtrA proteases: from *Escherichia coli* to *Arabidopsis*. *Physiol. Plant.* **119**, 337-346
25. Huesgen, P. F., Scholz, P. and Adamska, I. (2007) The serine protease HhoA from *Synechocystis* sp. strain PCC 6803: substrate specificity and formation of a hexameric complex are regulated by the PDZ domain. *J. Bacteriol.* **189**, 6611-6618
26. Adir, N., Zer, H., Shochat, S. and Ohad, I. (2003) Photoinhibition—a historical perspective. *Photosynth. Res.* **76**, 343-370
27. Edelman, M. and Mattoo, A. K. (2008) D1-protein dynamics in photosystem II: the lingering enigma. *Photosynth. Res.* **98**, 609-620
28. Huesgen, P. F., Schuhmann, H. and Adamska, I. (2009) Deg/HtrA proteases as components of a network for photosystem II quality control in chloroplasts and cyanobacteria. *Res. Microbiol.* **160**, 726-732
29. Barker, M., de Vries, R., Nield, J., Komenda, J. and Nixon, P. J. (2006) The Deg proteases protect *Synechocystis* sp. PCC 6803 during heat and light stresses but are not essential for removal of damaged D1 protein during the photosystem two repair cycle. *J. Biol. Chem.* **281**, 30347-30355
30. Cheregi, O., Sicora, C., Kos, P. B., Barker, M., Nixon, P. J. and Vass, I. (2006) The role of the FtsH and Deg proteases in the repair of UV-B radiation-damaged photosystem II in the cyanobacterium *Synechocystis* PCC 6803. *Biochim. Biophys. Acta.* **1767**, 820-828

31. Silva, P., Choi, Y. J., Hassan, H. A. and P. Nixon, P. J. (2002) Involvement of the HtrA family of proteases in the protection of the cyanobacterium *Synechocystis* PCC 6803 from light stress and in the repair of photosystem II. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **357**, 1461-1470
32. Huang, F., Hedman, E., Funk, C., Kieselbach, T., Schröder, W. P. and Norling, B. (2004) Isolation of outer membrane of *Synechocystis* sp. PCC 6803 and its proteomic characterization. *Mol. Cell. Proteomics* **3**, 586-595
33. Fulda, S., Huang, F., Nilsson, F., Hagemann, M. and Norling, B. (2000) Proteomics of *Synechocystis* sp. strain PCC 6803. Identification of periplasmic proteins in cells grown at low and high salt concentrations. *Eur. J. Biochem.* **267**, 5900-5907
34. Huang, F., Fulda, S., Hagemann, M. and Norling, B. (2006) Proteomic screening of salt-stress-induced changes in plasma membranes of *Synechocystis* sp. strain PCC 6803. *Proteomics* **6**, 910-920
35. Rippka, R., Deruelles, J., Waterbury, J. B., Herdman, M. and Stanier, R. Y. (1979) Generic assignments, strain histories and properties of pure cultures of cyanobacteria. *J. Gen. Microbiol.* **111**, 1-61
36. Laemmli, U. K. (1970) Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* **227**, 680-685
37. Sambrook, J., Fritsch, E. F. and Maniatis, T. (1989) *Molecular Cloning: A Laboratory Manual*, 2nd edn, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY
38. Letunic, I., Copley, R. R., Schmidt, S., Ciccarelli, F. D., Doerks, T., Schultz, J., Ponting, C. P. and Bork, P. (2004) SMART 4.0: towards genomic data integration. *Nucleic Acids Res.* **32**, D142-144
39. Emanuelsson, O., Nielsen, H., Brunak, S. and von Heijne, G. (2000) Predicting subcellular localization of proteins based on their N-terminal amino acid sequence. *J. Mol. Biol.* **300**, 1005-1016
40. Nielsen, H., Engelbrecht, J., Brunak, S. and von Heijne, G. (1997) Identification of prokaryotic and eukaryotic signal peptides and prediction of their cleavage sites. *Protein Eng.* **10**, 1-6
41. Bryson, K., McGuffin, L. J., Marsden, R. L., Ward, J. J., Sodhi, J. S. and Jones, D. T. (2005) Protein structure prediction servers at University College London. *Nucleic Acids Res.* **33**, W36-38
42. Sun, X., Peng, L., Guo, J., Chi, W., Ma, J., Lu, C. and Zhang, L. (2007) Formation of DEG5 and DEG8 complexes and their involvement in the degradation of photodamaged photosystem II reaction center D1 protein in *Arabidopsis*. *Plant Cell.* **19**, 1347-1361
43. Kolmar, H., Waller, P. R. and Sauer, R. T. (1996) The DegP and DegQ periplasmic endoproteases of *Escherichia coli*: specificity for cleavage sites and substrate conformation. *J. Bacteriol.* **178**, 5925-5929
44. Haußühl, K., Andersson, B. and Adamska, I. (2001) A chloroplast DegP2 protease performs the primary cleavage of the photodamaged D1 protein in plant photosystem II. *EMBO J.* **20**, 713-722
45. Helm, M., Luck, C., Prestele, J., Hierl, G., Huesgen, P. F., Fröhlich, F., Arnold, G. J., Adamska, I., Gorg, A., Lottspeich, F. et al. (2007) Dual specificities of the glyoxysomal/peroxisomal processing protease Deg15 in higher plants. *Proc. Natl. Acad. Sci. USA* **104**, 11501-11506

46. Schuhmann, H., Huesgen, P. F., Gietl, C. and Adamska, I. (2008) The DEG15 serine protease cleaves peroxisomal targeting signal 2-containing proteins in *Arabidopsis*. *Plant Physiol.* **148**, 1847-1856
47. Schechter, I. and Berger, A. (1967) On the size of the active site in proteases. I. Papain. *Biochem. Biophys. Res. Commun.* **27**, 157-162
48. Iwanczyk, J., Damjanovic, D., Kooistra, J., Leong, V., Jomaa, A., Ghirlando, R. and Ortega, J. (2007) Role of the PDZ domains in *Escherichia coli* DegP protein. *J. Bacteriol.* **189**, 3176-3186
49. Jomaa, A., Damjanovic, D., Leong, V., Ghirlando, R., Iwanczyk, J. and Ortega, J. (2007) The inner cavity of *Escherichia coli* DegP protein is not essential for molecular chaperone and proteolytic activity. *J. Bacteriol.* **189**, 706-716
50. Vande Walle, L., Lamkanfi, M. and Vandenberghe, P. (2008) The mitochondrial serine protease HtrA2/Omi: an overview. *Cell Death Differ.* **15**, 453-460
51. Kurian, D., Phadwal, K. and Mäenpää, P. (2006) Proteomic characterization of acid stress response in *Synechocystis* sp. PCC 6803. *Proteomics* **6**, 3614-3624
52. Spence, E., Sarcina, M., Ray, N., Møller, S. G., Mullineaux, C. W. and Robinson, C. (2003) Membrane-specific targeting of green fluorescent protein by the Tat pathway in the cyanobacterium *Synechocystis* PCC6803. *Mol. Microbiol.* **48**, 1481-1489
53. Peltier, J. B., Emanuelsson, O., Kalume, D. E., Ytterberg, J., Friso, G., Rudella, A., Liberles, D. A., Soderberg, L., Roepstorff, P., von Heijne, G. et al. (2002) Central functions of the lumenal and peripheral thylakoid proteome of *Arabidopsis* determined by experimentation and genome-wide prediction. *Plant Cell* **14**, 211-236
54. Schubert, M., Petersson, U. A., Haas, B. J., Funk, C., Schröder, W. P. and Kieselbach, T. (2002) Proteome map of the chloroplast lumen of *Arabidopsis thaliana*. *J. Biol. Chem.* **277**, 8354-8365
55. Murata, N. and Suzuki, I. (2006) Exploitation of genomic sequences in a systematic analysis to access how cyanobacteria sense environmental stress. *J. Exp. Bot.* **57**, 235-247

Table 1 Cleavage sites within β -casein by rHtrA, rHhoA and rHhoB assayed by N- and C-terminal sequencing.

^a The mature secreted version of β -casein (processed at the N-terminal Ala₁₅) was used for these studies (N1-199 amino acids).

Protease	Generated fragments (amino acids) ^a	Cleavage site	Number in the gel
rHtrA	N1-162	FPPQSV↓LSL	1
	N1-130	LTLTDV↓ENL	2a
	N1-141	PLPLLQ↓SWM	2b
rHhoA	N1-165	QSVLSL↓SQS	3
	N1-101	SKVKEA↓MAP	4
	N102-199	KEA↓MAPKHK	5
rHhoB	N1-165	QSVLSL↓SQS	6

FIGURE LEGENDS

Figure 1 Purification of recombinant active and inactive variants of *Synechocystis* 6803 Deg/HtrA proteases

(A) Schematic representations of rHtrA, rHhoA and rHhoB constructs generated for this study, *E. coli* DegP and DegS and human HtrA2/Omi. Hatched grey boxes mark putative transmembrane domains, white boxes show predicted signal peptides, light grey boxes indicate protease domains with positions of catalytic His and Asp marked in white and the position of a catalytic Ser mutated to Ala in inactive variants indicated in black, and dark grey boxes mark PDZ domains. Proteins are drawn to scale, with length of 100 amino acid residues indicated. (B) Coomassie-stained SDS-PAGE gels showing the elution fractions after Ni²⁺-affinity chromatography, containing recombinant active rHtrA, rHhoA and rHhoB, and inactive rHtrA_{SA}, rHhoA_{SA} and rHhoB_{SA} variants of proteases. (C) Coomassie-stained SDS-PAGE gels showing the elution fractions after Ni²⁺-affinity chromatography, containing recombinant active rHtrAΔPDZ, rHhoAΔPDZ and rHhoBΔPDZ, and inactive rHtrA_{SA}ΔPDZ, rHhoA_{SA}ΔPDZ and rHhoB_{SA}ΔPDZ variants of proteases lacking PDZ domains.

Figure 2 Proteolytic activity of *Synechocystis* 6803 Deg/HtrA proteases against substrates with different folding states

Naturally unfolded β-casein, globular, folded BSA, folded lysozyme and partially denatured, reduced lysozyme were used as model substrates. (A) Protease constructs with intact PDZ domains. (B) Protease constructs without PDZ domains, e.g. C-terminally truncated after the protease domain.

Figure 3 Cleavages site specificity of *Synechocystis* 6803 Deg/HtrA-proteases

(A) Degradation of β-casein by recombinant rHtrA, rHhoA and rHhoB, analysed by Coomassie-stained SDS-PAGE. M indicates the molecular mass protein marker. Arrows with numbers highlight bands that were excised from the gel for N- and C-terminal protein sequencing. Proteolytic assays using rHtrA were performed in 50 mM MES, 20 mM CaCl₂, pH 5.5, assays with rHhoA and rHhoB were carried out in 50 mM Tris, 20 mM CaCl₂, pH 7.0. Representatives of at least three replicates are shown. (B) Amino acid sequence of mature β-casein. Black bars show residues identified by N-terminal sequencing of fragments excised from gels in (A), dashed bars indicate residues identified by C-terminal sequencing and arrows indicate deduced cleavage sites.

Figure 4 Complex formation by inactive variants of *Synechocystis* 6803 Deg/HtrA proteases in the absence and the presence of substrate

Size-exclusion chromatography of (A) rHtrA_{SA}, (B) rHhoA_{SA}, and (C) rHhoB_{SA}. Black lines show elution profiles obtained in the absence of substrate, and grey lines show profiles after pre-

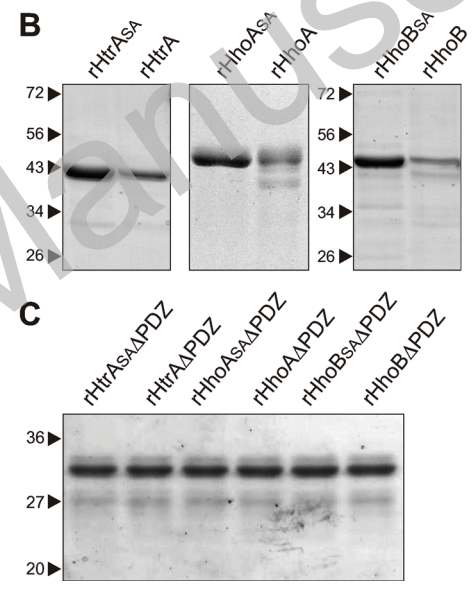
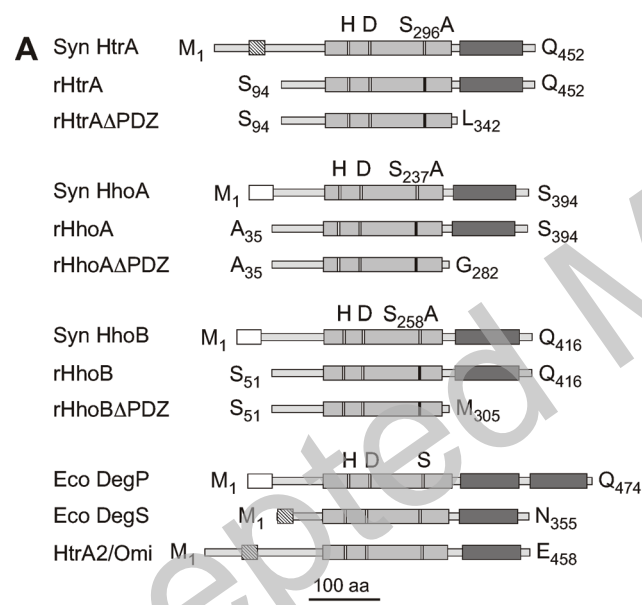
incubation with β -casein in a 3:1 molar ratio of protease:substrate (A, B) or 1:3 ratio (C). Lower panels show aliquots of the size-exclusion chromatography fractions separated by SDS-PAGE and visualised by Coomassie blue staining. The exclusion volume of the columns and the elution volumes of selected marker proteins are indicated with black triangles.

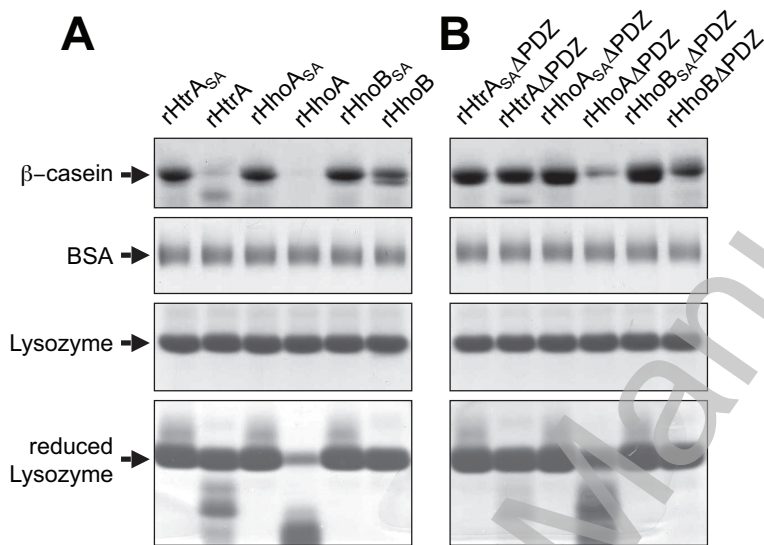
Figure 5 Schematic overview of the oligomerisation data

Our results demonstrated that in contrast to trimeric rHhoB, trimeric rHtrA and hexameric rHhoA form higher-order oligomers in the presence of β -casein. Deletion of the PDZ domain locks all three proteases in the trimer state.

Figure 6 Biochemical properties of rHtrA, rHhoA and rHhoB assayed with a fluorogenic casein substrate

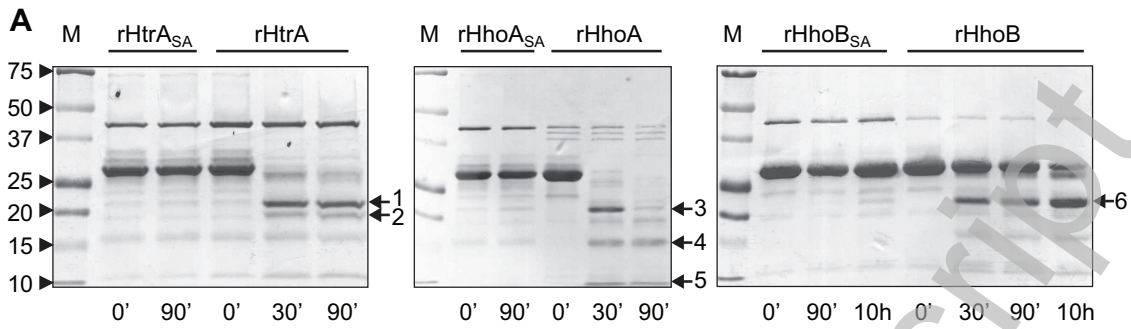
In all panels data points obtained for rHtrA are represented by diamonds, rHhoA by triangles, rHhoB by circles. Open symbols connected by dashed lines indicate absence of divalent ions, filled symbols connected by solid lines indicate presence of divalent ions. Please note that detector gain settings may differ between panels. Error bars indicated standard deviation ($n=3$). (A) Effect of Ca^{2+} ion concentration on proteolytic activity. Activity was measured by the initial rate of fluorescence generation in 10 mM MOPS buffer, pH 6.5, and normalised to the signal observed without CaCl_2 . (B) Effect of pH on protease activity in the absence of divalent ions. Activity as measured by the initial rate of fluorescence generation. (C) Effect of pH on protease activity in the presence of 10 mM CaCl_2 . (D) Effect of temperature on protease activity as measured by the total fluorescence after 40 min, normalised to the fluorescence observed at 10°C.





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1,2,3,4,6

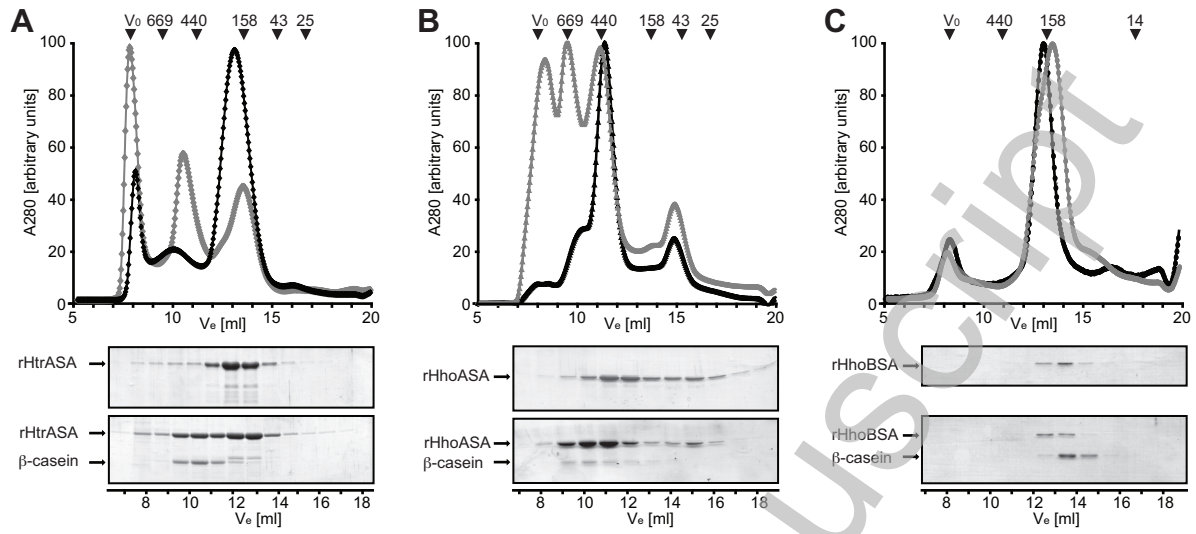
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PVVVPPFLQPEVMGVSKVKEA↓MAPKHKEMPFPKYPVEPFETESQS⁴LTLDV↓ENLHL⁵PLPLLO↓SWMHQPHQPLPPTVM

FPPQSV↓LSL↓SQSKVLPVPQKAVPYQQRDMP¹IQAFLLYQEPVLGPV^{3,6}RGFF⁵PIIV

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