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Food Additives and Contaminants

Fatty acids esters of 3-chloropropane-1,2-diol in edible oils

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Fatty acids esters of 3-chloropropane-1,2-diol in edible oils

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Abstract

For EXECT THE TENDAM CONDUCT THE SURVEY THE SURVEY THE SURVEY THAN BY THE SURVEY THAN BY THE SURVEY THAN BY THE ONLY THE DUPLO THAN BY THE UPLO THE UPLO THE A series of 25 virgin and refined edible oils, obtained from the retail market was analyzed for their content of free 3-MCPD and 3-MCPD released from its esters with higher fatty acids (bound 3-MCPD). The oils contained free 3-MCPD ranging from \lt 3 μ g/kg (LOD) to µg/kg. Surprisingly, the bound 3-MCPD level was much higher and varied between < µg/kg (LOD) and 2462 µg/kg. On average, virgin oils had relatively low levels of bound 3-MCPD ranging from < 100 µg/kg (LOD) to < 300 µg/kg (LOQ). Higher levels of bound 3- MCPD were found in oils obtained from roasted oilseeds (337 µg/kg) and in the majority of refined oils (< 300 µg/kg to 2462 µg/kg), including refined olive oils. In general, it appears that the formation of bound 3-MCPD in oils is linked with the preliminary heat treatment of oilseeds and with the process of oil refining. The analysis of unrefined, de-gummed, bleached, and deodorized rapeseed oil showed that the level of bound MCPD decreased during the refining process. However, additional heating of seed oils for 30 min at temperatures ranging from 100^oC to 280^oC, and heating at 230^oC (260^oC) for up to 8 hours led to an increase of the bound 3-MCPD level. On the contrary, heating of olive oil resulted in a decrease of the bound 3-MCPD level. For comparison, the fat isolated from salami was analyzed for the intact fatty acid esters of 3-MCPD. This fat contained the bound 3-MCPD at the level of 1670 µg/kg and the fatty acid esters of 3-MCPD mainly consisted of 3-MCPD diesters, monoesters of 3- MCPD were present in smaller amounts. The major types of 3-MCPD diesters (about 85%) were mixed diesters of palmitic acid with C18 fatty acids (stearic, oleic, linoleic acids). These diesters were followed by 3-MCPD distearate (11%) and 3-MCPD dipalmitate (4%). Generally, very little 3-MCPD existed as the free compound (31 µg/kg).

Key words: *3-Chloropropane-1,2-diol, 3-MCPD, bound 3-MCPD, 3-MCPD fatty acid esters, edible oils, contaminants*

Introduction

3-Chloropropane-1,2-diol (also known as 3-MCPD) is a representative of the so called food borne or food processing contaminants. It was first identified in acid-hydrolyzed vegetable protein (acid-HVP) in 1981 (Davídek et al. 1982) as a reaction product of either acylglycerols or glycerol with hydrochloric acid and quite recently shown to be a racemic mixture of the corresponding enantiomers, (*R*)-3-MCPD and (*S*)-3-MCPD (Velíšek et al. 2002). 3-MCPD has been extensively monitored in acid-HVP because of its suspected carcinogenicity. In view of its toxicity, a provisional maximum tolerable daily intake of $2 \mu g$ per kg of body mass has been set (European Commission 2001) and a regulatory limit of 20 µg/kg, based on a 40% dry matter content, has been adopted for 3-MCPD in acid-HVP and soy sauce and came into force in the European Union in 2002 (European Commission 2001).

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a provisional maximum tolerable daily intake of 2 µg per kg of
ean Commission 2001) and a regulatory limit of 20 µg/kg, base
has been adopted for 3-MCPD in ac Recent studies (Crews 2002; Hamlet et al. 2002a; Hamlet et al. 2002b; Divinová 2004) have shown that elevated levels of 3-MCPD (\lt 5 to 134 μ g/kg) can occur not even in acid-HVP, soy sauces and related products but also in a wide range of retail processed foods such as meat, dairy and cereal products. Cereal products, particularly biscuits, crackers, doughnuts and bread, accounted for the highest levels of 3-MCPD reported (Hamlet et al. 2002b). Furthermore, 3-MCPD was found in malt products comprising brewing malts and malt extracts. The typical levels of 3-MCPD for specialist malts (having typical EBC colour units > 20) ranged from < 30 to < 500 µg/kg (Hamlet et al. 2002a), which is well over the maximum level for acid-HVP.

While levels in retail foods have not been found to exceed 134 μ g/kg (Crews 2002; Hamlet et al. 2002a; Hamlet et al. 2002b; Divinová et al. 2004a), the potential for additional generation of 3-MCPD from < 5 to 275 µg/kg, resulting from domestic cooking procedures, has been also reported (Crews 2001; Breitling-Utzmann 2003; Breitling-Utzmann 2004). Significant levels of 3-MCPD were found notably in the high temperature crust region of grilled cheeses (Crews 2001) and toasted breads (Crews 2001; Breitling-Utzmann 2003; Breitling-Utzmann 2004). More than 500 μ g/kg 3-MCPD can be found in well-toasted breads (Breitling-Utzmann 2003; Breitling-Utzmann 2004). However, at the moment there are no limits for any food product or food ingredient other than acid-HVP and soy sauce.

Model experiments simulating processed foods (Velíšek et al. 2003; Calta et al. 2004) showed that 3-MCPD evidently arised from either acylglycerols or glycerol in a reaction with naturally present or intentionally added sodium chloride. The 3-MCPD level strongly depends on temperature and the content of lipids, glycerol, salt and water. It is believed that the precursors (acylglycerols and glycerol) react with negatively charged nucleophiles, hydroxyl and chloride anions, which leads first to 3-MCPD esters and finally to free 3-MCPD. Generation of 3-MCPD in model dough systems showed that free glycerol is the key precursor of 3-MCPD in leavened doughs (Hamlet et al. 2004a). Furthermore, monoacylglycerols, lysophospholipids, and phosphatidylglycerols present in flour improvers were important precursors of 3-MCPD in leavened doughs (Hamlet et al. 2004b).

Recent findings (Rétho and Blanchard 2005) indicate that also bromine anions react with acylglycerols and glycerol yielding 3-bromopropane-1,2-diol and 2-bromo-1,3-propanediol that were found in vegetable oils at the level ranging from 7 to 45 μ g.kg⁻¹.

The mechanisms of 3-MCPD formation in processed foods are still very little understood. However, there have been recent studies of mechanism of its formation in cereal products e.g. from glycerol (Hamlet et al. 2004a), mono- and diacylglycerols and glycerol phosphate (Hamlet et al. 2004b).

3-MCPD in leavened doughs (Hamlet et al. 2004a).

rols, lysophospholipids, and phosphatidylglycerols present in f

precursors of 3-MCPD in leavened doughs (Hamlet et al. 2004

s (Rétho and Blanchard 2005) indicate that als Fatty acid esters of 3-MCPD were the known precursors in the formation of 3-MCPD in model mixtures consisting of hydrochloric acid and triacylglycerols, phospholipids, soybean oil, soybean meal, wheat and maize lipids (Velíšek et al. 1980; Davídek et al. 1982). They were also found as the constituents of raw acid-HVP (Velíšek et al. 1980) and reported in the neutral fraction of goat's milk lipids (Cerbulis et al. 1984), where their occurrence was tentatively ascribed to the use of chlorine-based sanitizers.

Our recent findings indicate that the formation of 3-MCPD esters (monoesters and diesters with higher fatty acids) is characteristic of a variety of processed foods (Divinová et al. 2004b; Svejkovská et al. 2004; Doležal et al. 2005). These compounds represent a new class of food contaminants, the bound form of 3-MCPD, from which free 3-MCPD could be released by a lipase-catalyzed hydrolysis reaction. In many cases the amount of bound 3- MCPD exceeded that of the free 3-MCPD.

We have analyzed 20 samples of selected retail food products for their bound 3-MCPD content. The levels of bound 3-MCPD found in the analyzed foods varied between traces (140 µg/kg) and 6100 µg/kg and were 10 to 396 times higher than the free 3-MCPD levels. Five foods of plant origin that were processed at high temperatures (particularly crisp bread, salty crackers, doughnuts, French fries and dark malt) contained elevated levels of bound 3-MCPD. The highest level of bound 3-MCPD (6100 μ g/kg) was found in a sample of French fries (Svejkovská et al. 2004). The level of bound 3-MCPD in roast coffee was relatively low and varied between 6 µg/kg (soluble coffees) and 390 µg/kg (decaffeinated coffee) and exceeded the free 3-MCPD level 8 to 33 times (Doležal et al. 2005). The presence of bound 3-MCPD in pickled olives and herrings suggests that these compounds can also form at relatively low temperatures and even in acid media (Svejkovská et al. 2004).

For Peer Assume 20 and 390 µg/kg (decafreinated coffee D level 8 to 33 times (Doležal et al. 2005). The presence of bou and herrings suggests that these compounds can also form at d even in acid media (Svejkovská et al. Analysis of white bread showed (Hamlet and Sadd 2004) that the highest levels of bound 3- MCPD were found in the crust (547 μ g/kg) and toast (160 μ g/kg) while in crumb they were found in much lower amount (26 µg/kg). The corresponding levels of the free 3-MCPD were 477 μ g/kg, 93 μ g/kg and < 0.5 μ g/kg, respectively, i.e. 6 to 52 times lower. It was also showed that free 3-MCPD can be released from its esters by a commercial lipase (from *Aspergillus oryzae*) hydrolysis.

The vegetable oils recently analyzed by Rétho and Blanchard (2005) contained very low levels of 3-MCPD, between 3 and 7 µg/kg. The possible formation of 3-MCPD in oils by a lipase-catalyzed transesterification of triacylglycerols has recently been reported (Robert et al. 2004). It is not yet known whether this phenomenon is due to hydrolysis of 3-MCPD esters or lipase catalyzed formation from triacylglycerols and chlorides.

Due to the occurrence of 3-MCPD esters in a variety of retail and homemade foods, we assumed that these lipophilic compounds could also arise in edible oils during the refining process. Described herein are our investigations into the determination of 3-MCPD esters in a variety of oils and fats. Our attention was mainly focused on the role of various factors that can influence the level of 3-MCPD esters during industrial processing of oils and their culinary use.

Materials and methods

Chemicals and samples

3-Chloropropane-1,2-diol (> 98%, 3-MCPD) was purchased from E. Merck (Darmstadt, Germany), 3-MCPD-*d5* (99.4%) from Dr. Ehrenstorfer (Augsburg, Germany), phenylboronic acid (PBA, ≥ 97%) from Fluka Chemie (Buchs, Switzerland). 1,2-Diacyl-3-chloro-1,2 propanediols were synthesised according to KRAFT et al. 1979 and purified on a silica gel column using light petroleum ether/diethyl ether mixtures. All other reagents and solvents were of analytical purity.

Vegetable oils as well as roasted almonds and peanuts were obtained from retail outlets in Prague. The sample of salami was provided by Central Science Laboratory, York, UK. Unrefined (mixture of pressed and extracted oil), degummed, bleached, deodorized rapeseed oils were supplied by Setuza a.s. (Ústí n. L., Czechia).

Instrumentation

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cal purity.
as well as roasted almonds and peanuts were obtained from n
ample of salami was provided by Central Science Laborato
ture of pressed and extracte GC/MS analysis of 4-chloromethyl-2-phenyl 1,3,2-dioxaborolane, obtained by derivatization of 3-MCPD using phenylboronic acid, was carried out on an Agilent Technologies 6890N gas chromatograph (Agilent Technologies, Palo Alto, CA, USA) equipped with a Series 5973 quadrupole mass selective detector Agilent 5973 MSD (70 eV) and data processing system (MSD ChemStation, G1701CA version C.00.00). Gas chromatography was performed on a capillary column Equity^{™-1} (30 m x 0.25 mm i.d., thickness of 1 µm, Supelco, PA, USA). The injector was held at 250° C (splittles), the column temperature was programmed from 80° C (1 min) to 300^oC (37 min) at a rate of 10^oC/min. Helium at a flow rate of 0.8 ml/min was used as the carrier gas, 1 µl sample was injected. For quantification purposes, single ion monitoring was used to monitor ions at *m/z* 147 (3-MCPD) and at *m/z* 150 (3-MCPD-*d5*). Ions at m/z 91 and 196 (3-MCPD) and at m/z 93 and 201 (3-MCPD- d_5) were used as qualifiers.

The fat isolated from salami was separated by preparative TLC (Davídek et al. 1980) into the individual fraction and the fractions containing diesters and monoesters of 3-MCPD were analyzed by GC using the same instrument as above and a DB-1HT fused silica capillary column (15 m x 0.25 mm i.d., thickness of 1 µm, J&W Scientific, CA, USA). One 1 µl sample was injected into the split injection port $(1:4)$ held at 300^oC with a solvent delay of 2 min. The oven was initially set to 100^oC, then programmed to 340^oC at a rate of 10^oC/min and kept at 340° C for 26 min. The helium carrier gas was supplied at a constant rate of 0.7 ml/min. The analytes were ionized by EI at 70 eV and the obtained mass spectra of 3-MCPD esters with fatty acids were consistent with the spectra of the synthesized standards (KRAFT et al. 1979, Velíšek et al. 1980). The individual esters were quantified using 1,2-dipalmitoyl-3 chloro-1,2-propanediol (purity of 96.9) as the external standard.

Isolation of fat from nuts and salami

from nuts and salami

from the roasted almonds, roasted peanuts, and salami the mate

diethyl ether (80 ml) using Ultra Turrax T25 (Janke and

Switzerland). The obtained extract was filtered off and the

two portions of di To obtain fat from the roasted almonds, roasted peanuts, and salami the material $(20 g)$ was extracted with diethyl ether (80 ml) using Ultra Turrax T25 (Janke and Kunkel, IKA-Labortechnik, Switzerland). The obtained extract was filtered off and the residue again extracted with two portions of diethyl ether (20 ml each). The combined extracts were washed with water (20 ml), dried with anhydrous sodium sulfate and the solvent evaporated to dryness. The oil content was determined using the Soxhlet method (extraction with either diethyl ether - nuts or hexane - salami).

Heating of oils

The oil (20 g) was placed to a beaker and heated in an oven at the desiderative temperature (100^oC, 150^oC, 200^oC, 230^oC, 260, and 280^oC) for 30 minutes. Other portions of the oil were heated at either 260° C or 280° C for 0.5, 1, 2, 4, and 8 hours. The oils were let to cool to room temperature and analyzed for the content of free and bound 3-MCPD.

Free 3-MCPD

The oil (5 g) placed in a 100 ml Erlenmeyer flask was dissolved in a hexane : acetone mixture (1:1, v/v, 30 ml) and 50 µl of an aqueous solution of $3-MCPD-d₅$ (internal standard, 0.5 µg) were added. The solution was transferred to a 100 ml separatory funnel and the Erlenmeyer flask was washed again with two 10 ml portions of the hexane/acetone mixture. The combined extracts was transferred to a separatory funnel and extracted with water (10 ml). The aqueous phase was evaporated to dryness using a vacuum rotary evaporator at 55° C. The residue was dissolved in 20% sodium chloride (2 ml), derivatised with phenylboronic acid (0.4 ml) and used for the determination of free 3-MCPD according to the procedure of Divinová et al. (2004b). Three parallel determinations of each sample were made.

Bound 3-MCPD

The oil (100 mg) placed in a 10 ml distillation flask was dissolved in tetrahydrofurane (1 ml). To this solution 1.8 ml sulphuric acid solution (98%, 1.8 ml in 100 ml methanol) was added and the mixture was heated at 40° C for 16 hours. The cooled mixture was then neutralised

with saturated NaHCO₃ solution (0.5 ml), 1 ml of 3-MCPD- d_5 (internal standard, 1 μ g) was added and the solution was evaporated to dryness using a vacuum rotary evaporator at 55° C. The residue dissolved in 20% sodium chloride (2 ml), extracted with two portions of hexane (2 ml), derivatised with phenylboronic acid (0.2 ml) and used for the determination of bound 3-MCPD according to the above procedure (Divinová et al. 2004b). Three parallel determinations of each sample were made.

Results and discussion

Our recent experiments simulating processed foods have revealed that the models consisting of the recognized 3-MCPD precursors (i.e. glycerol, triolein, lecithin) and heated (230°C, 30 min) with sodium chloride at a level typically present in foods yielded both enantiomers of 3- MCPD, i.e. (*R*)-3-MCPD and (*S*)-3-MCPD, in equimolar amounts (Velíšek et al. 2002). Stereospecific analysis of diesters of 3-MCPD found in goat's milk also showed that they were a racemic mixture (Myher et al. 1986). In total, six different compounds, four monoesters and two diesters, can be derived from racemic 3-MCPD and a fatty acid

(**Figure 1**).

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Follow Example 12 and the method shave revealed that the most dead 3-MCPD precursors (i.e. glycerol, triolein, lecithin) and head an enloride at a level typically present in foods yielded both ench-
 For PEP The method used throughout our research and focused on the occurrence of 3-MCPD esters in foods (Divinová et al. 2004b) was based on the determination of 3-MCPD released from the esters by methanolysis and thus did not have the potential to differentiate between the monoesters and diesters of 3-MCPD or the esters derived from its individual enantiomers. As a part of the present work, the above method has been optimized to increase its sensitivity so that the limit of detection (LOD) and limit of quantification (LOQ) of 3-MCPD released from the esters, when expressed as 3-MCPD (so called bound 3-MCPD), was 100 µg/kg oil and µg/kg oil, respectively. The LOD and LOQ for the method used to determine the free 3- MCPD was 3 μ g/kg oil and 9 μ g/kg oil, respectively. Using these two methods, a series of retail market oils, oils isolated from nuts, and oils obtained from our industrial partner was analyses for the levels of free and bound 3-MCPD. Groups of retail market oils were selected to focus on virgin oils produced by different producers and from different raw materials, as well as on various grades of oils that might contain 3-MCPD esters resulting from heat processing during the refining process (Table I).

It can be seen that the analyzed oils, with only three exceptions, contained free 3-MCPD at the levels of either LOD or LOQ (3 replications). The levels of 3-MCPD higher than the LOQ were only found in virgin hazel nut oil (10 μ g/kg), virgin soybean oil (12 μ g/kg), and virgin wheat germ oil (24 μ g/kg) and they did not exceed the level of 50 μ g/kg dry matter adopted for 3-MCPD in acid-HVP and soy sauce (European Commission 2001).

The bound 3-MCPD was found in oils at levels ranging from \lt 100 μ g/kg (LOD) to µg/kg. As it can be seen (Figure 2), the average bound 3-MCPD level of oils (half of the value of either LOD or LOQ was used to calculate the average values) is low in virgin seed oils obtained from unroasted seeds (63 µg/kg), and increases in the following sequence: virgin olive oils (75 µg/kg), virgin germ oils (100 µg/kg), virgin seed oils from gained roasted seeds (337 μ g/kg), refined seed oils (524 μ g/kg), and refined olive oils (1464 μ g/kg).

For Propagation In one of a Force Emagning From Trevelor in the term of the term of the term of the Herbert Computer of the Herbert of the computation of parti Within the sets of virgin oils, levels exceeding 300 μ g/kg (LOQ) were only found in sesame oil obtained from roasted seeds (337 μ g/kg). Refined seed oils (< 300-1234 μ g/kg) and refined olive oils $($300-2462 \text{ µg/kg})$ had higher levels of bound 3-MCPD. The amount of$ 2462 ug of bound 3-MCPD/kg found in refined olive oil then corresponds e.g. to 7780 ug/kg of 3-MCPD monooleate or to 13110 µg/kg of 3-MCPD dioleate and exceeds by 500% the level adopted for 3-MCPD in acid-HVP and soy sauce (European Commission 2001).

The average ratio of free 3-MCPD to bound 3-MCPD was 19 within the set of virgin seed oils and even 12 if the roasted sesame seed oil was not included. Virgin olive oils (extra virgin olive oils have a free acidity, expressed as oleic acid, of not more than 1 gram per 100 grams, virgin olive oils have a free acidity of less than 2 grams per 100 grams) had their average free 3-MCPD/bound 3-MCPD ratio 22, while this ratio increased to 150 within the set of refined seed oils and even to 325 within the set of refined olive oils.

It appears that the formation of 3-MCPD esters in oils is linked either with the preliminary heat treatment of seeds (e.g. roasting as in the case of roasted sesame seed oil) or with the process of oil refining (refined seed and olive oils). Moreover, there is certain dissimilarity between seed oils and olive oils as the latter have been obtained from the pulp. The pulp oils have high water content and active lipases yielding by hydrolysis partial acylglycerols during handling of fruits and their subsequent storage prior processing. Partial acylglycerols may

then become the precursors of 3-MCPD esters (Calta et al. 2004). Virgin olive oils are produced by thrifty procedures where temperature does not exceed 40° C while refined olive oils are commonly obtained by physical refination (degumming and bleaching). Moreover, the diatomaceous earth used for bleaching is sometimes purified by hydrochloric acid, which results in the formation of metal chlorides that may become the precursors of MCPD esters. Olive oil is a blend of virgin oil with refined oil and olive pomace oil is a blend of refined pomace olive oil and possibly some virgin oil (Firestone 2005).

filuence of heat treatment on oilseeds, we have analyzed the le D in retail roasted almonds and roasted peanuts, both unsalted after are summarized in Table II. For example, the amount of bout of four monasted salted almon To check the influence of heat treatment on oilseeds, we have analyzed the level of free and bound 3-MCPD in retail roasted almonds and roasted peanuts, both unsalted and salted. The obtained results are summarized in Table II. For example, the amount of bound 3-MCPD in the oil extracted from roasted salted almonds was 1035 µg/kg while the level of bound 3- MCPD in virgin almond oil was < 100 µg/kg. Similarly, roasted salted peanuts contained bound 3-MCPD at the level of 883 μ g/kg, which is much higher than the level (< 100 μ g/kg) found in virgin peanut oil (Table I). The presence of salt did not markedly influence the level of 3-MCPD esters as it is usually added to the cooled roasted product.

We have also analyzed unrefined, degummed, bleached and deodorized rapeseed oil obtained from our industrial partner and, surprisingly, the level of bound MCPD esters decreased during the refining process (Table III). Bleaching was done at about $90-100^{\circ}$ C for 30 min and deodorization at 230–250°C for approximately 30 min. Therefore, it seems that the chemical composition of the oil given by its origin (pulp *versus* seed) plays a crucial role in the formation of MCPD esters.

We were further interested how different batches of oil from the same producer differed in their bound 3-MCPD level. Therefore, 6 batches of rapeseed oil produced in 2005 and 2006 were analyzed and it was found that the bound 3-MCPD levels ranged from 381 to 484 µg/kg (Table I).

We were also interested whether the thermal treatment of commercial oils exerts influence upon the level of bound 3-MCPD. Therefore, samples of refined rapeseed oil (with the original amount of 3-MCPD esters of 381 µg/kg), virgin wheat germ oil (<100 µg/kg), and refined olive oil (2462 μ g/kg) were heated for 2 hrs at temperatures ranging from 100^oC to

280^oC, then at 230^oC and 260^oC for 0.5, 1, 2, 4 and 8 hrs and analyzed for the content of bound 3-MCPD. The extreme temperatures of 260° C and 280° C and very long times of heating were done for comparison. The achieved results can be seen in Figure 3 and Figure 4, respectively.

Our findings indicate that the level of bound 3-MCPD in rapeseed and wheat germ oils increased with temperature and time of heating while the level of bound 3-MCPD in olive oil decreased(Figure 3 and Figure 4). The increase in bound 3-MCPD level is probably due to the formation of 3-MCPD esters *de novo* while its decrease in olive oil is mainly caused by rapid decomposition, oxidation, and polymerization of 3-MCPD esters (Svejkovská et al. 2006). Heating of oils at 260° C (Figure 5 and Figure 6) first leads to an increase of the bound 3-MCPD level in olive and rapeseed oils (achieved in about one hour), which is followed by a decrease with prolonged heating. Again, the bound forms of 3-MCPD in olive oil are more susceptible to decomposition, oxidation, and polymerization. The amount of free 3-MCPD did not change under the experimental conditions and remained at the level of $\langle 9 \mu g/gg \rangle$.

re 3 and Figure 4). The increase in bound 3-MCPD level is prob
MCPD esters *de novo* while its decrease in olive oil is mainly oxidation, and polymerization of 3-MCPD esters (Svejkovsk
at 260°C (Figure 5 and Figure 6) firs Since the 3-MCPD fatty acid esters derived from fats and oils seem to be universally distributed in processed foods (Divinová et al. 2004, Hamlet and Sadd 2004, Doležal et al. 2005), the individual naturally occurring esters of 3-MCPD were investigated in the last stage of this study. We have analyzed the fat isolated from salami (fermented for 3 days at 62°C , water 28.0%, protein 19.3%, fat 45.2%, salt 4.31%, carbohydrates 1.9%) that contained free and bound 3-MCPD at the level of 31 µg/kg and 1670 µg/kg, respectively. The bound 3- MCPD mainly consisted of 3-MCPD diesters, monoesters of 3-MCPD were present in smaller amounts. The major types of 3-MCPD diesters (about 85%) were mixed diesters of palmitic acid with C18 fatty acids (stearic, oleic, linoleic acids). These diesters were followed by 3- MCPD distearate (11%) and 3-MCPD dipalmitate (4%). Analogous results were obtained analyzing bakery products.

Further research aimed at the reaction pathways leading to the formation of 3-MCPD esters (that seem to be universally distributed in processed foods) and conditions under which these compounds form is ongoing.

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Only Figure 5. Changes in bound 3-MCPD level in olive oils with time of heating at 230° C and 260° C.
- Figure 6. Changes in bound 3-MCPD level in rapeseed oils with time of heating at 230° C and 260° C.

Table I. The content of bound 3-MCPD in retail edible oils.

* Half of the value of LOQ (or LOD) was used. ** Different batches from the same producer. *** Retail grades according to The International Olive Oil Council (IOOC) standards.

Olive Oil Council (IOOC) standards.

Table II. The content of oil, free and bound 3-MCPD in roasted almonds and peanuts.

Table III. Changes in the content of free and bound 3-MCPD in rapeseed oil during physical refining

* Half of the value of LOQ was used.

Nue of LOQ was used.
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Figure 1. Fatty acid monoesters and diesters of 3-MCPD.

$$
\begin{array}{ccc}\nC_{1} & C_{2} & C_{3} & C_{4} & C_{5} & C_{5} & C_{6} & C_{7} & C_{8} & C_{8} & C_{9} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{11} & C_{12} & C_{13} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{10} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{10} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{10} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{19} & C_{10} & C_{10} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{19} & C_{10} & C_{10} & C_{10} & C_{11} & C_{12} & C_{13} & C_{14} & C_{15} & C_{16} & C_{17} & C_{18} & C_{19} & C_{19} & C_{10} & C
$$

(*R*)-1-acyl-3-chloropropane-1,2-diol (*S*)-1-acyl-3-chloropropane-1,2-diol (*R*)-1,2-diacyl-3-chloropropane-1,2-diol

R-CO-O-C-H CH₂-OH $CH₂-Cl$ OH Cl

$$
\begin{array}{c}\nCH_2-OH \\
H=C=O-CO-F \\
\vdots \\
CH_2-CI\n\end{array}
$$

(*S*)-2-acyl-3-chloropropane-1,2-diol

H
\n
$$
CD-R
$$

\n $HP - C - 0 - C0 - R$
\n $CH_2 - O - CO - R$
\n $CH_2 - Cl$

(*R*)-2-acyl-3-chloropropane-1,2-diol (*S*)-1,2-diacyl-3-chloropropane-1,2-diol

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Figure 3. Changes in bound 3-MCPD level in rapeseed, wheat germ, and olive oils heated to various temperatures for 30 min.

