

SPME-GC and sensors as rapid methods for monitoring the lipid oxidation in nuts

Sarah Pastorelli, Ana Rodriguez

► To cite this version:

Sarah Pastorelli, Ana Rodriguez. SPME-GC and sensors as rapid methods for monitoring the lipid oxidation in nuts. Food Additives and Contaminants, 2007, 24 (11), pp.1219-1225. 10.1080/02652030701426987. hal-00577466

HAL Id: hal-00577466 https://hal.science/hal-00577466

Submitted on 17 Mar 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Food Additives and Contaminants



SPME-GC and sensors as rapid methods for monitoring the lipid oxidation in nuts

Journal:	Food Additives and Contaminants
Manuscript ID: TFAC-2006-374.R1	
Manuscript Type:	Original Research Paper
Date Submitted by the Author:	20-Apr-2007
Complete List of Authors:	Pastorelli, Sarah; Joint Research Centre Rodriguez, Ana; Joint Research Centre
Methods/Techniques:	Chromatography - GC, Clean-up - SPME, Sensory
Additives/Contaminants:	Packaging active packaging
Food Types:	Dried fruit





http://mc.manuscriptcentral.com/tfac Email: fac@tandf.co.uk

1 2	Solid phase micro-extraction (SPME-GC) and sensors as rapid
3	methods for monitoring lipid oxidation in nuts
4	
5	S. Pastorelli*, L. Torri ¹ , A. Rodriguez, S. Valzacchi, S. Limbo ¹ and C.Simoneau
6	
7	European Commission DG-Joint Research Centre, Institute for Health and Consumer
8	Protection, Physical and Chemical Exposure Unit, TP 260, 21020 Ispra (VA) – Italy
9	¹ diSTAM, Department of Food Science and Microbiology, National University of
10	Milan, via Celoria, 2, 20133 Milan, Italy
11	
12	
13	*Corresponding author: Sarah Pastorelli
14	Joint Research Centre - IHCP
15	Physical and Chemical Exposure Unit
16	21020 Ispra (VA) Italy - T.P. 260
17	E-mail: sarah.pastorelli@jrc.it
18	
19	
20	
21	Abstract
22	Dry foods with high fat content are susceptible to lipid oxidation, which involves a
23	quality deterioration of the product, since this process is responsible for the generation
24	of off-flavours. Hexanal is considered to be a good shelf-life indicator of such oxidation

26 headspace analytical techniques. For this reason an electronic nose comprising 10 metal

products. In addition, due to its high volatility, hexanal can be easily determined by fast

2	
3	
Δ	
-	
3 4 5 6 7	
6	
7	
8	
0	
9	
10	
11	
12	
12	
13	
14	
15	
16	
17	
17	
9 10 11 12 13 14 15 16 17 18 9 20 21 22 24 25 26 27 28 9 30 132 33 45 67 89 30 31 32 33 45 36 7 89 30 31 32 33 34 35 67 89 30 30 31 32 33 34 35 36 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 38 37 37 38 37 37 38 37 37 38 37 37 37 37 37 37 37 37 37 37 37 37 37	
19	
20	
21	
21	
22	
23	
24	
25	
20	
26	
27	
28	
20	
23	
30	
31	
32	
33	
00	
34	
35	
36	
37	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

1

2

3

4

5

6

7

8

9

10

oxide semiconductors (MOS) and a solid-phase microextraction (SPME) coupled with gas chromatography and flame ionization detector (GC-FID) method were compared in order to determine hexanal formed in hazelnuts during storage under different conditions (room temperature, 40°C, UV light, with and without oxygen scavenger). The results obtained by the two methods showed a good correlation, confirming the possibility of using a multi-sensor system as a screening tool for the monitoring of shelf-life and oxidation state of nuts. **Keywords:** Multi-sensor system, electronic nose, SPME, hexanal, lipid oxidation, oxygen scavengers, shelf-life

- 11
- 12
- 13

 1 Introduction

Foods with high fat content like nuts are very good model products for shelf-life studies (Del Nobile 2001). These products are susceptible to rancidity because of the lipid oxidation (Brunton et al. 2000; Goodridge et al. 2003; Lee and Krochta 2002; Stashenko et al. 2002). Different storage variables such as light and temperature can influence the off-flavours development from the products generated by lipid oxidation (Jensen et al. 2005).

Hexanal is the main aldehyde formed during the oxidation of unsaturated fats (Shahidi2001). For this reason hexanal is a representative marker of the oxidative rancidity.

12 The purpose of the present paper was the development of a simple, efficient and fast 13 method to determine hexanal and to monitor its presence in hazelnuts samples stored 14 under controlled conditions.

A multi-sensor system (electronic nose) and solid phase micro extraction (SPME) were
compared in order to evaluate the effect of storage conditions on packaged food.

According to the Gardner and Bartlett's definition (Gardner and Bartlett 1994), an electronic nose is "an instrument, which comprises an array of electronic chemical sensors with partial specificity and an appropriate pattern-recognition system, capable of recognising simple or complex odours". Different kinds of gas sensors have been developed and in function of their ability, the e-nose can be fit for various applications in food industry (Schaller et al. 1998). It was shown to be suitable both to evaluate the odour of packaging materials such as paper (Holmberg et al. 1995), paperboard (Willing

et al. 1998) and plastic films (Frank et al. 2001; Van Deventer and Mallikarjunan 2002) and to determine the sensory contamination of packaged foodstuffs (Heiniö et al. 2002). The aroma evaluation of foods can be performed in different steps of production from the raw material stage right to the final product in order to monitor the quality and freshness (Bartlett et al. 1997). In the last years, the use of an e-nose was tested both on different foods such as olive oil (Ulmer et al. 1997; Gonzáles Martín et al. 1999; García-Gonzáles et al. 2004), tomato (Berna et al. 2004), tea (Dutta et al. 2003), coffee (Ulmer et al. 1997; Singh et al. 1996), fruits (Di Natale et al. 2001; Saevels et al. 2003), bakery products (Needham et al. 2005) and on animal foods, e.g. meat (Berdagué and Talou 1993), poultry (Rajamäki et al. 2006), fish (Di Natale et al. 2001) and milk (Capone et al. 2001; Labreche et al. 2005). Moreover, the correlation of the sensor data with classical analytical techniques was investigated by various authors (Frank et al. 2001; Annor-Frempong et al. 1998; Bleibaum et al. 2002; Aishima 2004; Strathmann et al. 2005).

Solid phase micro extraction was selected as sampling technique to compare to e-nose data because it has been proven to be a simple, fast, solventless technique and it does not require sample preparation. Volatile profiles in different foodstuffs have been successfully characterised by SPME. Many SPME applications can be found in the literature for fruit (Augusto et al. 2000; De Lourdes-Cardeal et al. 2005), meat (Ramirez et al. 2004), vinegar (Natera-Marin et al. 2002), olive oil (García-Gonzáles et al. 2004), soybean (Boué et al. 2003), truffle (Diaz et al. 2002). The analysis of hexanal in several matrices has also been performed by SPME in turkey (Brunton et al. 2000), powder

infant milk formula (Fenaille et al. 2003) and freeze-dried chicken (Goodridge et al. 2003).

Due to the fact that both e-nose and SMPE has been successfully applied in food analysis, in the present work specific methods for hexanal determination in nuts have been carried out. The experimental design included also the use of oxygen scavengers. Commercial pouches for food active packaging were tested for their efficiency and the eventuality of an undesiderable absorption of flavours generated from the food spoilage was also investigated.

Materials and methods

Samples

Powdered hazelnuts purchased from a local supermarket were packed in transparent plastic pouches with high oxygen barrier properties. For each time-storage condition 3 different sachets were prepared: pouch with oxygen scavenger, pouch with 7 g of nuts, pouch with 7 g of nuts and an oxygen scavenger.

Samples were then stored at room temperature, at 40°C and under UV light. After each exposure period (0, 2, 5, 16, 31 days), pouches were opened and 6 head space vials (20 mL) were prepared for each package containing hazelnuts: 3 vials with 2 g of hazelnuts for e-nose evaluation and 3 vials with 0.1 g of hazelnuts for SPME analysis.

In the same way the oxygen scavenger powder was removed from the pouches, transferred in a 20 mL headspace vials and analysed by SPME/GC-FID technique.

1
2
3
4
5
6
7
8
à
10
10
10
12
13
14
15
16
17
18
19
20
21
22
23
13456789101123415167890212234256789011233456789333333333333333333333333333333333333
25
20
20
21
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
43 44
44 45
46 47
47 48
49
50
51
52
53
54
55
56
57
58
59
60

2 Reagents and Analytical Standards

Hexanal with purity of 98% and triacetin for standard solutions preparation were
purchased from Sigma-Aldrich (Steinheim, Germany).

5

1

1

6 E-Nose

7 A commercial portable electronic nose (PEN 2 model) from WMA Airsense Analitics 8 Inc. was used. The sensor array system comprises 10 metal oxide semiconductors 9 (MOS) of different chemical composition and thickness to provide selectivity towards 10 volatile compound classes as indicated by the instrument supplier even if typically the 11 sensors react broadband-like on organic and inorganic gases (Table 1). These devices, 12 also called oxide or ceramic gas sensors, rely on changes of conductivity induced by the 13 adsorption of odour molecules. The elevated operative temperature (200-500°C) allows 14 avoiding interference from water and to aid rapid response and recovery times (Kohl 1992). The detection limit of hot sensors was in the range of 1 mg kg^{-1} . 15

- 16
- 17

[Table 1, please insert here]

18

19

20

21

For e-nose evaluations, 2 g of nut powder was placed in a 20 mL glass vial, sealed with a PTFE/silicone septum and a crimp top, stored at 25°C for 1 hour to equilibrate and analysed at the same temperature.

22

The measurement system permitted suction of the gaseous compounds from theheadspace of sample through the sensor array at 400 ml/min for 180 secs. Then a

second pump transported the filtered reference air to the sensor array (the flow rate was 600 ml/min) for 400 secs to rinse the system between two following samples. The dimensionless ratio between the conductivity change induced by the volatile compounds from the headspace sample and the conductivity variation due to the interaction between the reference air and the metal oxide semiconductor (analysing air sample the ratio for each gas sensor is equal to 1) has been considered as response furnished by the single gas sensors. SPME-GC-FID-MS Details concerning the development of the method and the analytical conditions are reported in a previous paper (Pastorelli et al. 2006).

- **Results and discussion**

E-nose and SPME have been used in this experimental work in order to monitor the shelf-life and oxidation state of hazelnuts during different storage conditions. Both techniques are rapid, convenient and solventless methods for the determination of volatiles compounds from sample materials.

The possibility of using an electronic nose with 10 metal oxide semiconductor sensors to evaluate the quality of nuts stored under different conditions was explored and compared with the SPME-GC-FID results. Figure 1 shows the principal component analysis (PCA) visualization of sensor array measurement performed on hazelnuts stored at room temperature, at 40°C and under UV light.

[Figure 1, please insert here] The two first principal components (PCs) accounted 89% of the total variability (71% and 18% respectively). The dominant effect was given by the first principal component (PC1) and a minor part by the second (PC2). The biplot (a scatter plot for scores and factor loadings) illustrated the mutual relationships between samples and sensors (Figure 1). In fact, the position of the samples in a part of the graph was determined by the influence of the response of the sensors collocated in the same area of the plot. In particular, in the left part of the graph the samples that have induced a higher interaction between volatile compounds and some metal oxide semiconductors (W5S, W6S, W1S, W1W, W2S, W2W, W3S) are shown. On the contrary, the position of the samples in the right part of the biplot was determined by a lower response of the above cited sensors. Moreover, the raw data used to perform the PCA (data not shown) indicated that the sensors coded as W1C, W3C and W5C provided the lowest electronic signals for all samples, similar to those obtained analysing the reference air. Therefore the samples resulted distributed along the PC1 (from positive to negative values) according to their increasing emission of volatile compounds.

An examination of the sensor correlation matrix and the overlapping of the W1C, W3C and W5C sensors revealed that those sensors were strongly correlated and so contributed little to the discrimination process. In order to evaluate the usefulness of these three sensors in data exploration, a PCA excluding them was performed.

24 [Figure 2, please insert here]

The new score plot (Figure 2) did not show significant differences in distribution of samples in comparison with previous results obtained considering all 10 sensors (Figure 1). On the basis of that, it is possible to affirm that the W1C, W3C and W5C sensors did not provide any essential information to discriminate nuts, because probably nut degradation did not produce aromatic compounds able to react with these metal oxide semiconductors (Table 1). In other words, satisfactory results could be achieved performing analysis with only the seven selected sensors (W5S, W6S, W1S, W1W, W2S, W2W, W3S).

PCA data could be used to differentiate sample degradation as function of different storage conditions (Figure 2). As can be seen, nuts stored under UV light and without oxygen scavenger in the pack were well separated from the others. The action of UV light associated with the presence of oxygen represented the worst storage condition that favoured the highest production and accumulation of volatile compounds in nuts, as demonstrated by the position of these samples in the part of score plot that reveals the highest quantity of volatile compounds detected from the electronic nose.

With regard to the storage temperature differences were not observed among samples stored at 40°C and at room temperature. In fact the response of the array of the 7 sensors did not permit separation of nuts into two groups as function of the storage temperature, on the contrary the samples were grouped in only one cluster.

In figure 2, it was also possible to appreciate a minor detection of volatile compounds in hazelnuts packed with oxygen scavenger (empty symbol) with respect to ones packed without the active device (full symbol) at the same storage time and condition. In fact, in all cases, the samples with oxygen scavenger were positioned in the plot more to the right than the nuts without oxygen scavenger.

 For the development of a fast screening method the systems would need to calibrate more samples and a greater variety of conditions but the discrimination appeared to be feasible. The multi-sensor system provided a good differentiation among samples stored at different time-conditions. Anyway e-nose did not permit an identification of odour compounds responsible for the shelf-life state of the product. For this reason we used a SPME/GC-FID method for the identification of hexanal as indicator of hazelnuts oxidation state.

The results obtained with the SPME/GC-FID confirmed the findings of the investigation with the e-nose. The different hexanal content obtained in the SPME/GC-FID analysis was correlated to the results of the sensor array measurements. The hexanal content profile of the mean results out of triplicate measurements of different samples is described in the figure 3.

[Figures 3, please insert here]

- UV radiation seems to be the most critical factor for product oxidation. In fact, samples
- 24 stored under UV light showed a higher hexanal content compared with the ones stored

at room temperature and at 40°C. However no big differences have been detected
among samples stored at room temperature and at 40°C so that temperature seemed to
have a minor effect on oxidation process.

5 With samples stored under UV radiation hexanal content increased during storage time 6 while in all the other hazelnuts without oxygen scavenger the hexanal content remained 7 mostly constant or slightly decreased along exposure time. In the samples with oxygen 8 scavenger it was possible to observe a clearly reduction of hexanal content compared to 9 hazelnuts without active packaging under the same storage conditions. These results of 10 samples stored under UV showed the efficiency of oxygen scavenger in reducing 11 hazelnuts oxidation during storage.

In the case of samples stored at room temperature and at 40°C the hexanal content in the hazelnuts with an oxygen scavenger decreased progressively with time. Due to the fact that in hazelnuts without oxygen scavenger under these storage conditions there was no increase in hexanal content during exposure, the data obtained seemed to indicate that active packaging could have absorbed hexanal from package headspace. In order to check this phenomenon the oxygen scavenger content was analysed by SPME.

Hexanal was always found in the powder content in oxygen scavenger pouches and,
with the exception of 40°C, hexanal concentration increased along storage time (figure
4 shows the mean results out of triplicate measurements).

[Figures 4, please insert here]

1 Conclusion

An e-nose has been used as a screening tool to monitor the state of rancidity and therefore the shelf-life of the hazelnuts stored under controlled conditions. The portable electronic nose equipped with 10 metal oxide semiconductors provided a satisfactory differentiation among samples stored under diverse conditions. The identification of hexanal as hazelnut shelf-life marker was performed by solid phase micro extraction coupled to a GC-FID and confirmed by GC-MS.

9 E-nose measurement results and data obtained by SPME showed a good correlation
10 between off-flavours evaluation and hexanal content during storage. Both techniques
11 are complementary and could be used for monitoring the lipid oxidation in dry fat food.

References

Aishima TA. 2004. Correlating sensory attributes to gas chromatography-mass
spectrometry profiles and e-nose responses using partial least squares regression
analysis. Journal of chromatography A 1054:39-46.

Annor-Frempong IE, Nute GR, Wood JD, Whittington FW, West A. 1998.
Measurement of the responses to different odor intensities of "boar taint" using a
sensory panel and an electronic nose. Meat science 50:139-151.

Augusto F, Pires-Valente AL, dos Santos-Tada E, Rivellino SR. 2000. Screening of
Brazilian fruit aromas using solid-phase microextraction-gas chromatography-mass
spectrometry. Journal of chromatography A 873:117-127.

23 Bartlett PN, Elliott JM, Gardner JW. 1997. Electronic noses and their
24 application in the food industry. Food Technology 51:44-48.

Berdagué JL, Talou T. 1993. Examples of semiconductor gas sensors applied to meat
 products. Science des Aliments 13:141-148.
 Berna AZ, Lammertyn J, Saevels S, Di Natale C, Nicolaï BM. 2004. Electronic nose
 systems to study shelf life and cultivar effect on tomato aroma profile. Sensors and
 Actuators B 97:324-333.
 Bleibaum RN, Stone H, Tan T, Labreche S, Saint-Martin E, Isz S. 2002. Comparison of

7 sensory and consumer results with electronic nose and tongue sensors for apple juices.
8 Food quality and preference 13:409-422.

9 Boué SM, Shih BY, Carter-Wientjes CH, Cleveland TE. 2003. Identification of Volatile

10 Compounds in Soybean at Various Developmental Stages Using Solid Phase
11 Microextraction. Journal of Agricultural and Food Chemistry 51:4873-4876.

Brunton NP, Cronin DA, Monahan FJ, Durcan R. 2000. A comparison of solid-phase
microextraction (SPME) fibres for measurement of hexanal and pentanal in cooked
turkey. Food Chemistry 68:339-345.

Capone S, Epifani M, Quaranta F, Siciliano P, Taurino A, Vasanelli L. 2001.
Monitoring of rancidity of milk by means of an electronic nose and a dynamic PCA
analysis. Sensors and Actuators B 78:174-179.

De Lourdes-Cardeal Z, Moreira-Guimaraes E, Vilela-Parreira F. 2005. Analysis of
volatile compounds in some typical Brazilian fruits and juices by SPME-GC method.
Food Additives and Contaminants 22:508-513.

Del Nobile MA. 2001. Packaging design for potato chips. Journal of food engineering
47:211-215.

23 Diaz P, Señorans FJ, Reglero G, Ibañez E. 2002. Truffle Aroma Analysis by Headspace

24 Solid Phase Microextraction. Journal of Agricultural and Food Chemistry 50:468-6472.

Di Natale C, Magagnano A, Martinelli E, Paolesse R, Proietti E, D'Amico A. 2001. The
 evaluation of quality of post-harvest oranges and apples by means of an electronic nose.
 Sensors and Actuators B 78:26-31.
 Di Natale C, Olafsdottir G, Einarsson S, Martinelli E, Paolesse R, D'Amico A. 2001.
 Comparison and integration of different electronic noses for freshness evaluation of

6 cod-fish fillets. Sensors and Actuators B 77:572-578.

7 Dutta R, Kashwan KR, Gardner JW. 2003. Electronic nose based tea quality
8 standardization. Neural Networks 16:847-853.

9 Fenaille F, Visani P, Fumeaux R, Milo C, Guy PA. 2003. Comparison of Mass
10 Spectrometry-Based Electronic Nose and Solid Phase MicroExtraction Gas
11 Chromatography-Mass Spectrometry Technique to Assess Infant Formula Oxidation.
12 Journal of Agricultural and Food Chemistry 51:2790-2796.

Frank M, Ulmer H, Ruiz J, Visani P, Weimar U. 2001. Complementary analytical
measurements based upon gas chromatography-mass spectrometry, sensor system and
human sensory panel: a case study dealing with packaging materials. Analytica Chimica
Acta 431:11-29.

García-Gonzáles D, Aparicio R. 2004. Classification of different quality virgin olive
oils by metal-oxide sensors. European Food Research and Technology 218:484-487.

19 Gardner JW, Bartlett PN. 1994. A brief history of electronic noses. Sensors and20 Actuators B 18:211-220.

Gonzáles Martín Y, Pérez Pavón JL, Cordero BM, Pinto CG. 1999. Classification of
vegetable oils by linear discriminant analysis of Electronic Nose data. Analytica
Chimica 384:83-94.

Goodridge CF, Beaudry RM, Pestka JJ, Smith DM. 2003. Solid Phase Microextraction-Gas Chromatography for Quantifying Headspace Hexanal Above Freeze-Dried Chicken Myofibrils. Journal of Agricultural and Food Chemistry 51:4185-4190. Heiniö RL, Ahvenainen R. 2002. Monitoring of taints related to printed solid boards with an electronic nose. Food Additives and Contaminants 19:209-220. Holmberg M, Winquist F, Lundström I, Gardner JW, Hines EL. 1995. Identification of paper quality using a hybrid electronic nose. Sensors and Actuators B 27:246-249. Jensen PN, Danielsen B, Bertelsen G, Skibsted LH, Andersen ML. 2005. Storage stabilities of pork scratchings, peanuts, oatmeal and muesli: Comparison of ESR spectroscopy, headspace-GC and sensory evaluation for detection of oxidation in dry foods. Food Chemistry 91:25-38. Kohl D. 1992. Oxidic semiconductor gas sensors. In: Sberveglieri G, editor. Gas Sensors. Dordrecht: Kluwer Academic Publishers, p 43. Labreche S, Bazzo S, Cade S. 2005. Shelf life determination by electronic nose: application to milk. Sensors and Actuators B 106:199-206. Lee SY, Krochta JM. 2002. Accelerated Shelf Life Testing of Whey-Protein-Coated Peanuts Analyzed by Static Headspace Gas Chromatography. Journal of Agricultural and Food Chemistry 50:2022-2028. Natera-Marin R, Castro-Mejias R, Garcia-Moreno MV, Garcia-Rowe F, Garcia-Barroso C. 2002. Headspace solid-phase microextraction analysis of aroma compounds in vinegar: Validation study. Journal of chromatography A 967:261-267. Needham R, Williams J, Beales N. 2005. Early detection and differentiation of spoilage of bakery products. Sensors and Actuators B 106:20-23.

2
~
3
1
4
5
0
6
-
1
0
0
9
10
4.4
11
12
12
13
14
15
15
16
17
2 3 4 5 6 7 8 9 101 12 3 4 15 6 7 8 9 101 12 3 4 15 6 7 8 9 101 12 3 4 15 6 7 8 9 21 22 3 4 25 6 7 8 9 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3
10
10
13
20
~
21
າງ
<u>_</u> _
23
24
0E
25
26
20
27
28
20
29
30
00
31
00
32
22
33
34
04
35
00
36
37
51
38
00
39
40
40
41
- T I
42
42
43
43
43 44
43 44
43 44 45
43 44 45 46
43 44 45 46
43 44 45 46 47
43 44 45 46 47
43 44 45 46 47 48
43 44 45 46 47 48 49
43 44 45 46 47 48 49
43 44 45 46 47 48 49 50
43 44 45 46 47 48 49 50
43 44 45 46 47 48 49 50 51
43 44 45 46 47 48 49 50 51
43 44 45 46 47 48 49 50 51 52
43 44 45 46 47 48 49 50 51
43 44 45 46 47 48 49 50 51 52 53
43 44 45 46 47 48 49 50 51 52 53 54
43 44 45 46 47 48 49 50 51 52 53 54
43 44 45 46 47 48 49 50 51 52 53 53 55
43 44 45 46 47 48 49 50 51 52 53 54
43 44 45 46 47 48 49 50 51 52 53 54 55 56
43 44 45 46 47 48 49 50 51 52 53 55 56 57
43 44 45 46 47 48 49 50 51 52 53 55 56 57
43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58
43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58
43 44 45 46 47 48 49 50 51 52 53 55 56 57

1 Pastorelli S, Valzacchi S, Rodriguez A, Simoneau C. 2006. Solid-phase microextraction 2 method for the determination of hexanal in hazelnuts as an indicator of the interaction of 3 active packaging materials with food aroma compounds. Food Additives and 4 Contaminants 23:1236-1241. 5 Rajamäki T, Alakomi H, Ritvanen T. 2006. Application of an electronic nose for quality 6 assessment of modified atmosphere packaged poultry meat. Food Control 17:5-13. 7 Ramirez MR, Estevez M, Morcuende D, Cava R. 2004. Effect of the Type of Frying 8 Culinary Fat on Volatile Compounds Isolated in Fried Pork Loin Chops by Using 9 SPME-GC-MS. Journal of Agricultural and Food Chemistry 52:7637-7643. 10 Saevels S, Lammertyn J, Berna AZ, Veraberbeke EA, Di Natale C, Nicolaï BM. 2003. 11 An electronic nose and a mass spectrometry-based electronic nose for assessing apple 12 quality during shelf life. Postharvest biology and technology 30:3-14. 13 Schaller E, Bosset JO, Escher F. 1998. Electronic noses' and their application to food. 14 Lebensmittel Wissenschaft und Technology 31:305-316. Shahidi F. 2001. Headspace volatile aldehydes as indicators of lipid oxidation in foods. 15 16 2001. Advances in Experimental Medicine and Biology 488: 13-123. 17 Singh S, Hines EL, Gardner JW. 1996. Fuzzy neural computing of coffee and tainted-18 water data from an electronic nose. Sensors and Actuators B 30:185-190. 19 Stashenko EE, Puertas MA, Martinez JR. 2002. SPME determination of volatile 20 aldehydes for evaluation of in-vitro antioxidant activity. Analytical and Bioanaytical 21 Chemistry 373:70-74. 22 Strathmann S, Pastorelli S, Simoneau C. 2005. Investigation of the interaction of active 23 packaging material with food aroma compounds. Sensors and Actuators B 106:83-87.

Ulmer H, Mitrovics J, Noetzel G, Weima,r U, Göpel W. 1997. Odours and flavours identified with hybrid modular sensor systems. Sensors and Actuators B 43:24-33. Van Deventer D, Mallikarjunan P. 2002. Optimizing an electronic nose for analysis of volatiles from printing inks on assorted plastic films. Innovative Food Science and Emerging Technologies 3:93-99. Vas G, Vékey K J. 2004. Solid-phase microextraction: a powerful sample preparation tool prior to mass spectrometric analysis. Journal of mass spectrometry 39:233-254. Willing BIL, Brundin A., Lundström I. 1998. Odour analysis of paperboard, the correlation between human senses and electronic sensors using multivariate analysis. Packaging Technology and Science 11:59-67. en..

1 Table 1. Codes and selectivity of e-nose gas sensors.

W1Caromatic compoundsW5Sbroad range compounds, polar compounds, nitrogen oxides and ozoneW3Cammonia, aromatic compounds, aldeides, chetonsW6ShydrogenW5Calkanes, aromatic compounds, less polar compoundsW1Smethane, broad range compoundsW1Wsulphur compounds, terpenes and sulphur organic compoundsW2Salcohols, partially aromatic compounds, chetonsW3Smethane	Sensor code	General description
W3Cammonia, aromatic compounds, aldeides, chetonsW6ShydrogenW5Calkanes, aromatic compounds, less polar compoundsW1Smethane, broad range compoundsW1Wsulphur compounds, terpenes and sulphur organic compoundsW2Salcohols, partially aromatic compounds, chetonsW2Waromatic compounds, sulphur organic compoundsW3Smethane	W1C	aromatic compounds
w6S hydrogen w5C alkanes, aromatic compounds, less polar compounds w1S methane, broad range compounds w1W sulphur compounds, terpenes and sulphur organic compounds w2S alcohols, partially aromatic compounds, chetons w2W aromatic compounds, sulphur organic compounds w3S methane	W5S	broad range compounds, polar compounds, nitrogen oxides and ozone
W5Calkanes, aromatic compounds, less polar compoundsW1Smethane, broad range compoundsW1Wsulphur compounds, terpenes and sulphur organic compoundsW2Salcohols, partially aromatic compounds, chetonsW2Waromatic compounds, sulphur organic compoundsW3Smethane	W3C	ammonia, aromatic compounds, aldeides, chetons
W1S methane, broad range compounds W1W sulphur compounds, terpenes and sulphur organic compounds W2S alcohols, partially aromatic compounds, chetons W2W aromatic compounds, sulphur organic compounds W3S methane	W6S	hydrogen
W1Wsulphur compounds, terpenes and sulphur organic compoundsW2Salcohols, partially aromatic compounds, chetonsW2Waromatic compounds, sulphur organic compoundsW3Smethane	W5C	alkanes, aromatic compounds, less polar compounds
W2Salcohols, partially aromatic compounds, chetonsW2Waromatic compounds, sulphur organic compoundsW3Smethane	W1S	methane, broad range compounds
W2W aromatic compounds, sulphur organic compounds W3S methane	W1W	sulphur compounds, terpenes and sulphur organic compounds
W3S methane	W2S	alcohols, partially aromatic compounds, chetons
	W2W	aromatic compounds, sulphur organic compounds
	W3S	methane
		P

2

3

4

5

6

7

8

9

10

11

12

13

14

15

UV light

1

Figure 1. PCA of the response of 10 sensors to the headspace of samples (N=Nuts, NS=

Nuts+Scavenger) stored under different conditions (RT=Room Temperature, 40=40°C,

Figure 2. Score plot from the PCA analysis of samples (N=Nuts, NS= Nuts+Scavenger)

stored under different conditions (RT=Room Temperature, 40=40°C, UV= UV light) at

Figure 3. Hexanal content of samples stored with and without oxygen scavenger at

Figure 4. Hexanal content in scavenger powder at room temperature, 40°C and under

UV= UV light) at different storage times (t=0, 2, 5, 16, 31 days).

different storage times (t=0, 2, 5, 16, 31 days).

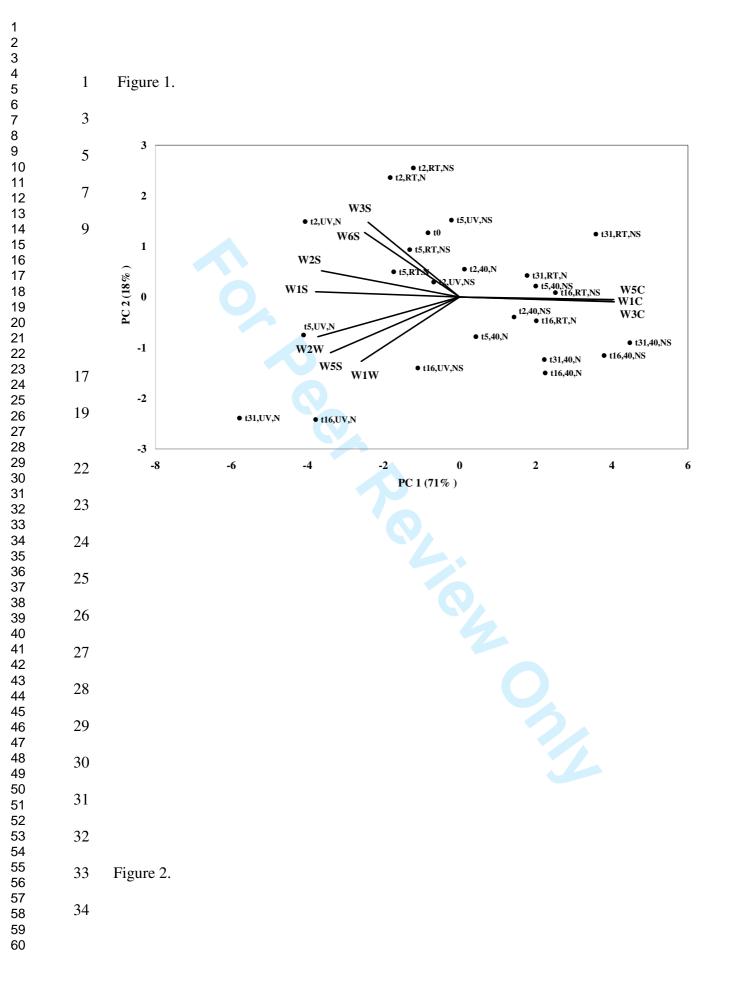
room temperature, 40°C and under UV light

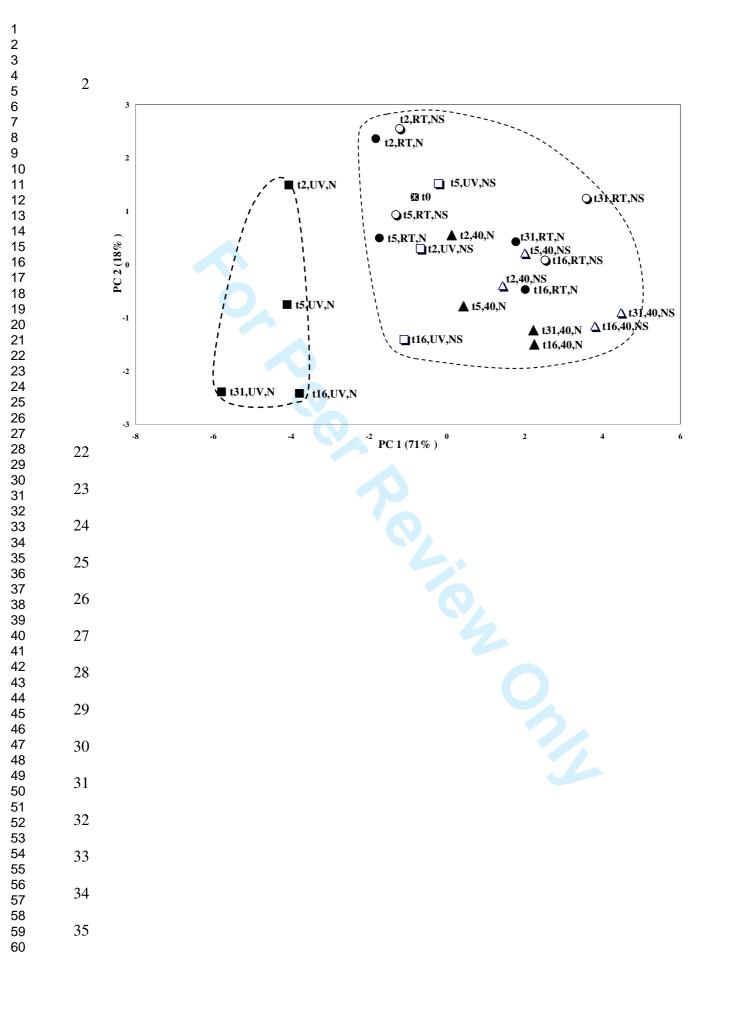
2
3
4
5
4 5 6 7
7
1
8
9
10
11
10
12
13
14
15
16
17
10
10
19
20
21
22
22
9 9 10 11 12 13 14 15 16 7 18 9 21 22 32 25 26 27 28 9 0 12 33 34 35 67 89 20 31 23 34 35 37 89 20 30 31 23 34 35 37 89 30 30 31 32 33 34 35 36 37 38 38 37 37 38 39 30 31 37 37 37 37 38 37 37 37 37 37 37 37 37 37 37 37 37 37
24
25
26
27
28
20
29
30
31
32
33
24
34
35
36
37
38
30
40
40
41
42 43
43
44
45
40
46
47
48
49
50
50 51
52
53
54
55
56
57
58
59

60

Figure l	legends
----------	---------

http://mc.manuscriptcentral.com/tfac Email: fac@tandf.co.uk





1 Figure 3.

