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#### **Food Additives and Contaminants**



### DIFFUSION OF HOMOLOGOUS MODEL MIGRANTS IN RUBBERY POLYSTYRENES: MOLAR MASS DEPENDENCE AND ACTIVATION ENERGY OF DIFFUSION

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# DIFFUSION OF HOMOLOGOUS MODEL MIGRANTS IN RUBBERY POLYSTYRENE: MOLAR MASS DEPENDENCE AND ACTIVATION ENERGY OF DIFFUSION

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#### **ABSTRACT:**

The published diffusion prediction models devoted to the diffusion of additives in food packaging simplify reality by having only a few parameters. Therefore, extrapolation of such models to barrier polymers, larger ranges of temperature and/or additive molecular weight (M<sub>W</sub>) are still questionable. Extra data is still required to generalize these existing prediction models. In this paper, diffusion of a specifically designed homologous set of model additives (from 236 to 1120 g.mol<sup>-1</sup>) was monitored in two polystyrenes at rubbery state (from 100 to 180°C): syndiotactic semi-crystalline polystyrene and its amorphous equivalent. Variations of D and Ea with migrant M<sub>W</sub> and temperature were found to be surprisingly low. Comparison of experimental behaviour with model predictions was performed. In their actual form, none of the models is clearly able to describe all experimental data, but clues can be given, showing converging of the different approaches.

Keywords: Diffusion coefficient; FRAP; molecular weight; activation energy; polystyrene;
 model probes

#### Introduction

To ensure consumer health, food packaging materials must be inert to packaged foodstuffs, in order to avoid contamination from harmful substances which can migrate from packaging materials. When dealing with plastic, regulations reinforce the concept of an Overall Migration Limit (OML) with a Specific Migration Limit (SML) for potentially toxic substances filed in a positive list. Petitioners for a new packaging material and control authorities have to test polymeric packaging to assess their compliance for food contact (e.g. US Food Law Act and EU regulation 2002/72/CE).

Migration is kinetically limited by the diffusion step within the polymer matrix, and is characterized by the associated diffusion coefficient D. The large amount of diffusion data in polyolefins has made the drawing of good prediction models possible. These are useful in predicting migration rate into foodstuffs or food simulants. Recently, food safety authorities have allowed their use in food packaging material assessments (e.g. art. 4 in EU regulation 2002/72/CE).

- - **Prediction of diffusion coefficients**

Different approaches have been developed to model diffusion in polymeric matrixes. From
Piringer's work, which set up a worst case model (eq. 1) to Limm and Hollifield's model (eq.
20 2), the diffusion processes can be simplified and implemented in ready-to-use models.

$$D^* = 10^4 \cdot \exp\left(A_P - \frac{C}{T} - 0.003M_W \left(45.M_W^{-1/3} - 1\right) - \frac{10454}{T}\right)$$
(1)

where  $D^*$  is the predicted overestimated diffusion coefficient (cm<sup>2</sup>.s<sup>-1</sup>), A'<sub>P</sub> and C are tabulated polymer parameters (dimensionless) T is the absolute temperature (K),  $M_W$  is the molecular weight of the diffusion species (g.mol<sup>-1</sup>).

3 4

6

$$D(T, M_{W}) = D_{0}^{'} \cdot \exp\left(\alpha \cdot M_{W}^{\frac{1}{2}} - K \frac{M_{W}^{\frac{1}{3}}}{T}\right)$$
(2)

where D is the predicted diffusion coefficient  $(cm^2.s^{-1})$ ,  $D'_{0}$  is the diffusion coefficient at infinite temperature (cm<sup>2</sup>.s<sup>-1</sup>),  $\alpha$  and K are tabulated polymer parameters (dimensionless), T is the absolute temperature (K),  $M_w$  is the molecular weight of the diffusion species (g.mol<sup>-1</sup>).

The reliability of such empirical models has been tested (Revnier et al. 1999; Helmroth et al. 2002; Rosca et Vergnaud 2006). Both models are able to describe the diffusion of additives in polyolefins, but prediction errors become significant when dealing with less mobile polymers, or when enlarging the extrapolation range to high temperatures or  $M_{WS}$ (Feigenbaum et al. 2005; Dole et al. 2006).

With the goal of creating a more general model, Dole et al. proposed a different approach to predict D with additive molecular weights that try to encompass both mobile and barrier polymers (Figure 1) (Dole et al. 2006). The  $D=f(M_W)$  correlations for a broad range of polymers were measured. They then showed that the negative slope of these  $[log_{10}D=f(M_W)]$ correlations decreased from high barrier polymers to lower barrier ones, while the Piringer model uses the same slope for all polymers and the Limm model integrates a weaker dependence. Consequently, as temperature directly affects polymer barrier properties (i.e. matrix mobility), Dole et al. suggested that this high slope variation leads to a variation of apparent Ea with temperature and M<sub>w</sub>. The empirical models above can still predict worst case values, but the safety margins may be drastically reduced from polyolefins to other extrapolated systems.

Toward a general predictive model for additive diffusion in polymers? 1 The Dole approach provides a more complex view of diffusion: the  $D=f(M_W)$  and  $Ea=f(M_W)$ 2 slopes are functions of additive  $M_W$  but first strongly depend on polymer mobility (Table 1). 3 It is worth noting that both plasticization and T increase allow a barrier polymer to turn into a 4 more mobile one (plasticized vs. non-plasticized PVC, dry vs. 60%RH PA in Figure 1)

5 Piringer's model was built using diffusion coefficients of additives whose  $M_W$  was 6 smaller than 500 g.mol<sup>-1</sup>. Dole et al. studied monitored diffusion of additives whose  $M_W$  was 7 up to 807 g.mol<sup>-1</sup> but with very few data between 430 and 807 g.mol<sup>-1</sup>. Extrapolation for high 8  $M_W$  additives still needs to be evaluated. In contrast, tabulated parameters in Limm's model 9 were determined by fitting with D of Irganox 1076 (531 g.mol<sup>-1</sup>) and Irganox 1010 (1178 10 g.mol<sup>-1</sup>). Therefore, Limm's model may not accurately predict the diffusion of small 11 additives.

When dealing with additives of increasing M<sub>w</sub>, the overall size, shape, and chemical functionality vary. Therefore, diffusion data of commercial additives are quite scattered. However, the models mentioned above average the molecular size effect by considering only M<sub>w</sub>. Up to now, only the Piringer model was validated experimentally against a homologous series of alkanes and alcohols (Reynier et al. 2001; Reynier et al. 2001).

The same parallel can be drawn for the temperature range. The three models were all built for a specific temperature range. The Piringer model has to be used at a given temperature range depending on the studied polymer (175°C for PET, 70 °C maximum for PS). Limm and Hollifield fits α and K parameters between 40 and 70 °C. The Dole approach describes diffusion data at 40 °C but encompasses a great range of polymer mobility. This model is therefore able to describe diffusion over an equivalent range of temperature. Revnier et al. have shown that diffusion activation by a temperature increase or plasticization fits the general relationship drawn in Figure 1 (Reynier et al. 2001).

#### **Food Additives and Contaminants**

In order to determine the extent of D dependance on  $M_{W}$ -, T- and polymer mobility, it is still necessary to accumulate D data from homologous systems. This could then help to determine proper fitting model parameters for barrier polymers and could lead to a general model.

Therefore, this study was set up to follow the diffusion of a specifically designed homologous set of high M<sub>w</sub> probes (Pinte et al. 2008). These probes were chosen to be probably more accurately model commercial additives than alkanes used by Reynier et al. (Reynier et al. 2001). Moreover, they are dedicated to the use of the FRAP technique. The FRAP technique (Fluorescence Recovery After Photobleaching) is a microscopic technique used to monitor diffusion in both biological samples (Seiffert et Oppermann 2005), and material ones (Smith 1986; Tseng et al. 2000). It allows the observation of a micro-domain of the packaging material, without simulant (or food) interactions. This offers a quick determination of intrinsic apparent diffusion coefficients of model additives, especially when dealing with low mobility matrixes from. The chosen matrix is PS, for which little modelling validation has been obtained up to now for the following reasons:

- i PS can be either totally amorphous or semi crystalline, depending on its tacticity,
  ii It's relatively high glass transition temperature (Tg) facilitates measurements in
  the vicinity of Tg, even at a glassy state,
  - iii PS displays a large rubbery plateau without any important structural change (cold crystallization, melting). As a crystalline PS, syndiotactic PS (sPS) was preferred to isotactic PS because of its higher cold-crystallization and melting temperatures. When correctly annealed, sPS shows no post-crystallization or melting over the studied range of temperatures (100 to 180 °C).

To resume, the relationships  $D=f(M_W)$  and D=f(1/T) are established and discussed in this paper. Effects of crystallinity are also discussed. Comparison of experimental behavior with model predictions is made.

#### Material and methods

#### 6 Material

Amorphous atactic model polystyrene PS2M was obtained as a SEC (steric exclusion chromatography) standard from Sigma Aldrich (Saint Quentin Fallavier, France):  $\overline{M_{w}} = 1\,850\,000 \text{ g.mol}^{-1}, \ \overline{M_{n}} = 1\,760\,000 \text{ g.mo}^{1-1}, \ \overline{M_{w}}/\overline{M_{n}} = 1.05.$  Syndiotactic polystyrene was purchased from Scientific Polymer Products (Ontario, USA): approx.  $\overline{M_{w}} = 300\,000 \text{ g.mol}^{-1}, \text{ density}=1.05.$ 

12 The series of fluorescent probes was synthesized as previously described (Pinte et al. 13 2008). This set of five homologous molecules, as shown in table 2, was made by grafting a 14 fluorescent head (7-nitrobenzofurazan) onto an amino acid-like tail. This "tail" was 15 synthesized by oligomerization of the same repeating unit. Tail molecular weight was 16 increased by repeating the same 'monomer-like' unit. For more detailed synthetic procedures, 17 please refer to the above mentioned article and its Supporting Information.

#### 19 Methods

Both polymers were characterized by differential scanning calorimetry (MDSC 2940, TA Instruments). The heating rate was 10K.min<sup>-1</sup>. Glass transition temperatures were determined as the inflection point of heat flow from the second run, whereas melting temperature and melting enthalpy were determined from the first run. For sPS, crystallinity was found to be 70%, assuming a heat of fusion of 53 J.mol<sup>-1</sup> (Bruzaud et al. 2005). Both thermograms are shown in figure 2.

2 3	1	
4	1	
5 6	2	Films of amorphous polystyrene for FRAP experiments (PS2M) were made as described in
7 8 9	3	(Pinte et al. 2008) by casting from an ethyl acetate solution onto a cover slide, followed by
9 10 11	4	solvent evaporation under dynamic vacuum at 90 $^{\circ}\text{C}$ , The cover slides were then mounted onto a
12 13	5	microscope slide by slight pressure and heating at 120_C for several seconds. ${ m sPS}$ is non soluble in
14 15 16	6	the same systems as aPS. Therefore, samples were made using the following method:
17 18	7	i dissolution in a refluxed toluene solution containing the proper amount of the
19 20	8	fluorescent probe,
21 22 23	9	ii casting on a hot plate (100 °C) to allow quick evaporation of toluene and to
24 25	10	avoid sPS precipitation,
26 27 28	11	iii removal of toluene traces at 90 °C in a dynamic vacuum oven for two days,
28 29 30	12	iv pressing a small piece of doped sPS between a glass microscope slide and a
31 32	13	glass cover slide at 270 °C.
33 34 35	14	In order to have fixed crystallinity sPS samples, sPS slides were annealed at 270 °C for
36 37	15	2 minutes and cooled at constant rate of 10 °C.min <sup>-1</sup> to allow crystallization to occur.
38 39	16	DSC showed no cold crystallization peak when heating these samples above Tg, up to
40 41 42	17	melting point.
43 44	18	
45 46 47	19	Fluorescent probe diffusion was monitored by confocal Fluorescence Recovery After
47 48 49	20	Photobleaching (FRAP) as described elsewhere (Pinte et al. 2008). This microscopic
50 51	21	technique has been previously used as a relevant method to collect D data in polymers (Smith
52 53 54	22	1986). FRAP creates an <i>in situ</i> concentration gradient in the bulk of small polymer samples.
55 56	23	The flux of the fluorescent species then induces an intensity fluctuation over the Region of
57 58	24	interest (ROI). From a practical point of view, FRAP experiments are conducted in three
59 60	25	steps: (i) reading the initial intensity with low laser power, (ii) bleaching a specified area by
	26	scanning a selected region of the sample (the ROI) with very high laser power, thus

photolysing the illuminated fluorescent probe, (iii) reading the recovery of the fluorescence intensity over time with low laser power. As intensity is assumed to be proportional to local probe concentration, D can be calculated from the fluorescence recovery kinetics. The bleached ROIs were ranging from 1.5 x 20 µm for low diffusing experiments to 67 x 61 µm for diffusion at high temperatures. The Shape ratio of the different probes used for FRAP experiment, denoting the ratio of a migrant's major dimension to its minor dimension was determined from NBDNEt2 to NBDNpip4. By 3D-drawing the molecules using MarvinSpace (software from ChemAxon), both dimension of the molecules then the shape ratio were easily determined. Experiments were set at a higher temperature than allowed on the microscope sample holder. Therefore, sample slides were mounted once for reading and bleaching, then demounted to be heated between two hot plates of a heating press, cooled down as quickly as possible and remounted to read the fluorescence profile evolution. The last steps were repeated until ROI intensity had varied enough to calculate D. Since the sample requires time to reach thermal equilibrium under the press, diffusion time (i.e. time spent in the heating press) was corrected to take this into account, assuming diffusion at lower temperatures to be

17 negligible.

To help re-localization in the sample when re-mounting under the confocal microscope, thin copper grids (Oxford Instruments, Saclay, France) were immersed in the PS in the casted drop of PS2M or in the molten piece of sPS. The hexagonal pattern (figure 3) provides valuable information when saved coordinates failed to precisely point to the region of interest. Due to the high temperatures required for the experiments, sandwiches of slides/PS sample/coverslide were sealed with epoxy glue instead of nail polish.

#### **Food Additives and Contaminants**

FRAP kinetics were processed as described elsewhere (Pinte et al. 2008). Due to quick diffusion (as high temperatures were explored), bleached ROIs were enlarged to extend recovery timescales. Unfortunately, bleaching was not efficient enough in the depth of the sample and apparent anomalous fluorescence recovery had to be corrected. As a consequence, the mathematical analysis implemented by Pinte et al. was slightly modified to eliminate this unwanted diffusion from planes below and above the focal plane and to analyze fluorescence recovery due to x-axis 1D diffusion.

Several authors using the FRAP method have reported immobile fractions when monitoring diffusion in complex systems (Miura 2004, Karboviak et al. 2006). Consequentially, equilibrium intensity in the bleached area may not be the same as in the initial one. The ratio of the final intensity to the initial one represents the percentage of mobile probes. The difference between this value and 100% then gives the amount of non mobile probes. Our diffusion experiments in syndiotactic polystyrene were therefore first conducted for a much longer time (6 months) than in the usual procedure in order to estimate the equilibrium intensity.

### **Results and discussion**

PS samples containing the probes  $(5 \times 10^{-10} \text{ mol.mg}^{-1})$  were obtained. All probes were correctly dispersed in the polymer matrixes, notwithstanding whether these were amorphous or crystalline. However, some patterns were observed in sPS, revealing the partial exclusion of fluorescent probes from more crystalline regions. This did not disturb FRAP experiments and diffusion of the different items of the homologous set was monitored in PS2M and in sPS for different temperatures ranging from 120 to 180 °C.

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Literature on the FRAP technique mentions that in the case of complex polymer systems, fluorescence recovery may not be total which is to be taken into account in the D calculation. For example, Equilibrium intensity is lower, denoting non mobile probes. This is supposed to be the case in semi-crystalline polymers where some of the probes could be entrapped and not available for diffusion. This was checked in this study but no immobile fraction was observed, even if some patterns could be observed in sPS samples (figure 3-sPS). Diffusion was found to be very similar in PS2M and in sPS. Equilibrium intensities were not different and close to the initial ones (less than a 10% decrease). Crystallinity did not induce a bigger change in mobile/non mobile probe ratios. Therefore, initial intensities were used as equilibrium intensities.

Diffusion coefficients were measured for the five homologous probes, whose  $M_W$ ranged from 236 to 1120 g.mol<sup>-1</sup>, in both amorphous and semi-crystalline PS, with an experimental temperature increases from 100 °C to 180 °C. Those for PS2M are reported in table 3.

 $[log_{10}D=f(M_W)]$  correlations

18 Diffusion coefficients as a function of  $M_W$  for different temperatures are plotted on Figure 4. 19 The total decrease of  $log_{10}D$  with  $M_W$  for different temperatures is shown in table 3. 20 Increasing  $M_W$  from 236 to 1120 g.mol<sup>-1</sup> lead to a decrease of about two units of  $log_{10}D$  for 21 all the tested temperatures. Indeed, as shown in figure 4, an arbitrary line can fit all data at 22 rubbery state, by only vertical shifts as lines are parallel. As reported in Table 3, the slope can 23 be considered as almost constant, when not taking into account data at 100°C. The slope 24 seemed smaller at 100°C, when PS2M is at its DSC glass transition temperature.

#### **Food Additives and Contaminants**

The constancy of the  $\log_{10}D = f(M_W)$  slope correlations relates the PS2M data to the Piringer model, where influence of M<sub>W</sub> and T on D are independent. The D decrease is quite small (roughly from 1 to 2 orders of magnitude) for an 890 g.mol<sup>-1</sup> increase of the migrant molecular weight. For example, Reynier et al. reported a 4orderofmagnitude decrease for diffusion in polyolefins over the same range but for lower  $M_W$  (from 156 to 807 g.mol<sup>-1</sup>) (Reynier et al. 2001). But, the first molecule of the homologous series used in this study has a higher M<sub>w</sub> compared to Reynier et al.'s one which was made for a migrant molecular weight between Npip2 and Npip3. As reported by Dole et al., the sharp decrease observed forD mainly occurs for rather low M<sub>w</sub> that is to say between gases and small additives (M<sub>w</sub> less than 200 g.mol<sup>-1</sup>). For higher molecular weight additives, extrapolation of such D =f(Mw) curves could leads, in the present study, to a flatter slope of  $D=f(M_W)$  (Dole et al. 2006).

Migrant shape (steric hindrance) is known to be a limiting parameter for diffusion, whereas increasing migrant degrees of freedom counterbalances this by increasing diffusion (Al-Malaika et al. 1991). Reynier has reported a high decrease as predicted by the Piringer model when taking into account homologous series of both alkanes and alcohols as well as commercial additives for higher M<sub>W</sub> (Reynier et al. 2001). They observed that for high M<sub>W</sub> migrants, increasing the degrees of freedom of the arms lead to a relative increase of D (trilaurine and tripalmitine with long alkyl arms compared to Irgafos 168 and Irganox 1330, with highly hindered arms). In fact, in our study, D decrease with migrant Mw is rather low as reported above. This can be explained by the model additives series which are not shape-homologous over the whole range of  $M_W$  (as initially expected). Although they all have a worm-like shape (shape ratio <1 as shown in figure 5), and due to the repet unit (table 3), this ratio decreases along the series: NBDNet<sub>2</sub> has a more spherical shape than NBDNpip<sub>n</sub>. Moreover, when jumping from NBDNet<sub>2</sub> to NBDNpip<sub>1</sub>, no extra degree of freedom is added,

in contrast to the change from NBDNpip<sub>n</sub> to NBDNpip<sub>n+1</sub>. where the migrant linearity is improved. This provides extra degrees of freedom when increasing migrant  $M_W$  and thus, the influence of shape through  $M_W$  on D seems to be minimized.

The Piringer model implies a much higher dependence of D on  $M_W$  (between NBDNet<sub>2</sub> and NBDNpip<sub>4</sub>,  $\triangle log_{10}D=6.75$  instead of about 2 in this study). Extrapolation of the Limm model and the Dole approach to higher temperatures predicts variations closer to those observed in this study (respective predicted decrease  $\triangle log_{10}D$  of 2.75 and 3). D values have been reported to be less dependent on  $M_W$  in rubbery polymers than in more barrier ones (Dole et al. 2006). But the flatter slope, (figure 4), at 100 °C gives the opposite indication as predicted by Dole et al.

D=f(1/T) correlations

In consistency with any migration model, D was found to increase when increasing
temperature as shown in figure 6. The different D values between 120 °C and 180 °C ranged
from 3.2 to 3.9 log<sub>10</sub>D units (table 3).

Close to glass transition, D variations are emphasized. Although requiring further confirmation, the higher the M<sub>W</sub>, the stronger the dependence with T. For NBDNet<sub>2</sub> to NBDNpip<sub>3</sub>, the D jump temperature is between 100 and 120 °C whereas for NBDNpip<sub>4</sub>, it occurs at a higher temperature (the jump is related to the D data which leave the D = f(Mw)) line observed at high temperature). The Tg jump seems to be a function of M<sub>w</sub>, as it is of frequencies or heating speed in thermo-mechanical analysis (Sauerbrunn et al. 2003). Activation energies (Ea) are calculated from the Arrhenius law as usually describes by others authors (Hayashi et al.1994, Roe et al.1998). Ea is then proportional to the slope of the curves  $\log D = f(1/T)$  and determine from the least squares approach. For homogeneity, activation

energies were calculated between 120 and 180 °C, assuming they are constant over this range of temperature.

Ea are between 140 and 180 °C slightly increased with molecular weight (table 4), whereas between 120 and 180 °C, it could be considered as constant, considering the data scattering. Results on a reduced range limit comparison with the Piringer model, which takes a constant Ea into account, and make the PS2M data closer to the Limm and Hollifield model (respective increase of Ea from Net<sub>2</sub> to Npip<sub>4</sub> is 1.5 vs 1.7). The effect of M<sub>w</sub> seems to prevail over the shape factor ratio, in contrast to diffusion coefficients. This odd behavior, however, makes one relationship clear: the weaker the dependence of the  $D=f(M_W)$  slope with temperature, the smaller Ea variations.

11 Increasing linearity for higher  $M_W$  leads to a weaker shape factor influence and 12 therefore decreasing dependence of Ea with  $M_W$ .

Ea were then compared to data in rubbery PS and LDPE. Hall et al. studied the diffusion of lophine in rubbery PS between 100 and 145 °C. (Hall et al. 1999). From their data, the calculated Ea (272 kJ.mol<sup>-1</sup>) is one and a half times larger than Hall et al.'s equivalent Ea  $(M_W \text{ of lophine is between that of NBDNet}_2 \text{ and NBDNpip}_1)$ . This difference may lie in the different shape and degree of freedom. As previously mentioned, the homologous set is worm shaped with a propyl spacer between rigid cores providing degrees of freedom, whereas lophine is more spherical with three short and rigid phenyl arms. This difference in Eas is logical because the Ea represents the energy required to open a sufficient hole between polymer chains for the migrant to diffuse: thus, a greater probe diameter means a higher Ea (Al-Malaika et al. 1991; Reynier et al. 2001). However, these reported Eas are very high compared to those calculated by Al-Malaika et al. (Al-Malaika et al. 1991) for homologous linear 2-hydroxy benzophenones in LDPE (between 69 and 86 kJ.mol<sup>-1</sup> from 5 to 100 °C). 

This could be explain by the mobility of the matrix: the temperature gap to the Tg is far bigger in Al-Malaika's experiment (for LDPE: T-Tg ~ 150 °C) than in ours (0 < T-Tg < 80 °C) or in Hall's (0 < T-Tg < 45 °C). This corroborates the suggestion of Dole et al. (Dole et al. 2006) that Ea may decrease with increasing temperature, but over a larger range and farther from the Tg than studied here.

#### **D=f(crystallinity)**

8 Crystallinity is reported to decrease D by hampering migrant diffusion as described by 9 Hedenqvist et al. (Hedenqvist et al. 1996). Some authors even describe an exponential 10 relationship between D and cristallinity (Klopffer and Flaconneche 2001).

Hedenqvist et al. report a five-fold reduction of D in linear PE and a two-fold one in branched PE when increasing crystallinity by 50% (Hedenqvist et al. 1996). The same observation was made by Chiang et al. when comparing quenched pseudo amorphous sPS with isothermally crystallized sPS (Chiang et al. 2002). Such reductions are of the same magnitude of what was found in our system (Figure 7). Nevertheless, larger Ds in sPS than in PS2M at low temperature are odd. (from Figure 7 and other results not shown). In addition, no increasing difference between D<sub>amorphous</sub> and D<sub>crystalline</sub> with the additive M<sub>W</sub> was found, in contrast to what Hedenqvist et al. reported. The torturous diffusion path in sPS might be counterbalanced by the increase of free volume due to increased tacticity and Tg heights. Highly tactic polymers have been reported to have higher free volume than atactic ones due to special chain conformation (Dammert et al. 1999; Soldera et Grohens 2002). When getting close enough to Tg, tortuosity effect may become less important than free volume size and chain mobility.

D 25

### 1 Conclusions

Diffusion in two different PS, one amorphous and one semi-crystalline, was monitored at the rubbery state for five homologous probes, offering a M<sub>W</sub> range of almost 900 g.mol<sup>-1</sup>. Diffusion was found to be less dependent on temperature or migrant molecular weight than calculated from prediction models.. However, D decrease for M<sub>w</sub> range was found close to the Limm model. This low dependence might be explained by the higher M<sub>w</sub> of the first molecule of the model migrant series. Another hypothesis may be offered to explain the low correlation between D and M<sub>w</sub>. Even if the probes are chemically homologous, they are unfortunately not ideally homologous; their shape factor varies along the series from relatively square to worm-like. This variation may counteract the decrease of D as M<sub>w</sub> gets higher. Bearing this fact in mind, none of the models can be excluded, based on the study's data. Moreover, all the three models can partially explain the observed effects of M<sub>w</sub> and T on D.

Whatever hypothesis is taken into account, the following general relationship is accurate: when D is not strongly affected by Mw or T, neither is Ea. Further experiments are required to develop a general model for: (i) a more shape-homologous series of designed migrants, and (ii) diffusion in glassy matrices, where very little data are still available because of very long diffusion time The Frap technique could help to acquire information about that issue.

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#### Table 1. Factors influencing diffusion coefficients and activation energies according to the three models

Model D influencing factors Ea influencing factors Piringer's model  $M_W$ Т polymer polymer Limm and Т polymer  $M_{W}$ Mw polymer Hollifield's model Dole's model Т polymer  $M_W$  $M_W$ polymer 

Т

strong dependence (bold), average dependence (normal), weak dependance (italic)

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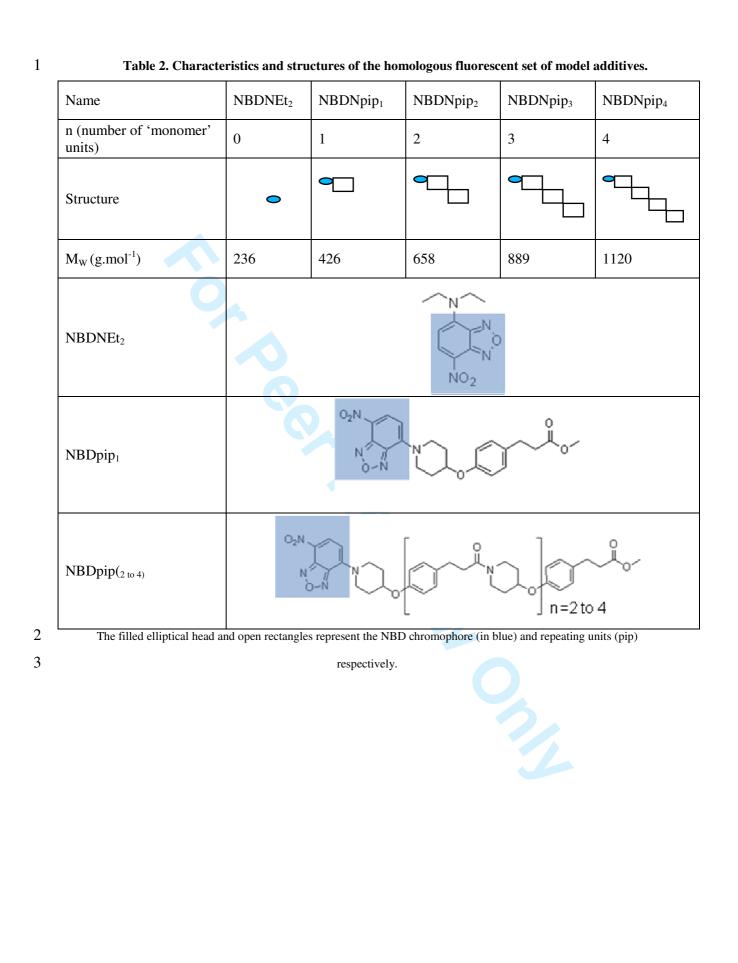


Table 3. Diffusion coefficients measured in amorphous polystyrene (PS2M) at various temperatures (D
are in 10 <sup>-12</sup> cm <sup>2</sup> .s <sup>-1</sup> ). Probes legend are given in table 2.

Probes			Tempera	ature (°C)		
$M_W (g.mol^{-1})$	180	160	140	120	100	$\Delta log_{10}D^b$
236	17000	1400	48	9,5	0,026	3.2
426	12000	1000	60	7,0	0,0081	3.2
658	5100	480	4	0,64	nd	3.9
889	600	140	10	0, 29	0,0035	3.3
1120	500	84	0,13	0.07	0,0008	3.8
$\Delta log_{10}D^{a}$	-1.5	-1.2	-2.6	-2.1	-1.2	
	$236$ $426$ $658$ $889$ $1120$ $\Delta \log_{10} D^{a}$ Slog_{10} D were calculated fr	236       17000         426       12000         658       5100         889       600         1120       500 $\Delta \log_{10} D^a$ -1.5 $\Delta \log_{10} D$ were calculated from NBDNEt <sub>2</sub> Alog <sub>10</sub> D were calculated between 120°C and alog <sub>10</sub> D were calculated	236       17000       1400         426       12000       1000         658       5100       480         889       600       140         1120       500       84 $\Delta \log_{10} D^a$ -1.5       -1.2 $Mog_{10} D$ were calculated from NBDNEt <sub>2</sub> to NBDNpip <sub>4</sub> NBDNpip <sub>4</sub> $Mog_{10} D$ were calculated between 120°C and 180°C	236       17000       1400       48         426       12000       1000       60         658       5100       480       4         889       600       140       10         1120       500       84       0,13 $\Delta log_{10} D^a$ -1.5       -1.2       -2.6 $log_{10} D$ were calculated from NBDNEt <sub>2</sub> to NBDNpip <sub>4</sub> NBDNpip <sub>4</sub>	236         17000         1400         48         9,5           426         12000         1000         60         7,0           658         5100         480         4         0,64           889         600         140         10         0,29           1120         500         84         0,13         0.07           Δlog <sub>10</sub> D <sup>a</sup> -1.5         -1.2         -2.6         -2.1	236         17000         1400         48         9,5         0,026           426         12000         1000         60         7,0         0,0081           658         5100         480         4         0,64         nd           889         600         140         10         0,29         0,0035           1120         500         84         0,13         0.07         0,0008 $\Delta \log_{10} D^a$ -1.5         -1.2         -2.6         -2.1         -1.2

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Table 4. Calculated activation energies (Ea) for the model additives in actactic polystyrene (PS2M).
Arrhenius plot of PS2M diffusion coefficients.

Model additive	$M_W(g.mol^{-1})$	Ea (kJ	Ea (kJ.mol <sup>-1</sup> )	
		between 120 and 180 °C	between 140 and 180 °C	
Net <sub>2</sub>	236	187	170	
Npip <sub>1</sub>	436	189	179	
Npip <sub>2</sub>	658	219	237	
Npip <sub>3</sub>	889	188	212	
Npip <sub>4</sub>	1120	239 behavior and a constant Ea be	249	

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5 6	2	Figure captions
7 8	3	
9 10	4	Figure 1. Diffusive behaviour of different migrants in polymers at 40°C, redrawn from
11 12	5	Dole et al.'s data (Dole et al. 2006).
13 14	6	Circles refer to diffusion in mobile polymers, such as LLDPE, LDPE, plasticized PVC,
15 16	7	plasticized PP), triangles show D for barrier polymers (dry PA, dry EVOH, unplasticized
17	8	PVC, PET, PMMA, PS), and squares refer to intermediate polymers (PA at 60%RH, PP).
18 19	9	Diffusion was carried out at 40°C. The arrow indicates an increasing matrix mobility,
20 21	10	embodied by the system glass transition temperature.
22 23 24	11	
25 26	12	Figure 2. Thermal characterization of virgin PS2M (acatic polystyrene) and sPS
27 28	13	(semicrystalline syndiotactic polystyrene sPS)
29 30	14	
31 32	15	Figure 3. syndioctactic polystyrene (sPS sample), showing non homogeneously dispersed
33 34	16	Npip2 probes (left), and homogeneous dispersion in amorphous polystyrene PS2M
35 36	17	(right). Zoom: 1.9, picture size: 125x125 µm. The black pattern of the copper hexagonal
37 38	18	grid can be seen immersed in the sPS sample.
39 40	19	
41 42	20	Figure 4. Diffusion coefficients in PS2M as a function of $M_W$ for different temperatures.
43 44	21	Plain lines have the same slope, they are vertically shifted to best describe $D=f(M_W)$ for each
45 46	22	temperature. The dotted line (T=100°C) has a different slope.
47 48 49	23	
50	24	Figure 5. Shape ratio of the different probes used for FRAP experiment, from NBDNEt <sub>2</sub>
51 52 53	25	to NBDNpip <sub>4</sub> .
54	26	The dimension ratio, which is equal to the largest cross section from head to tail, was
55 56	27	estimated by drawing the molecules in MarvinSpace (ChemAxon).
57 58	28	
59 60	29	Figure 6. Arrhenius plot of PS2M diffusion coefficients.

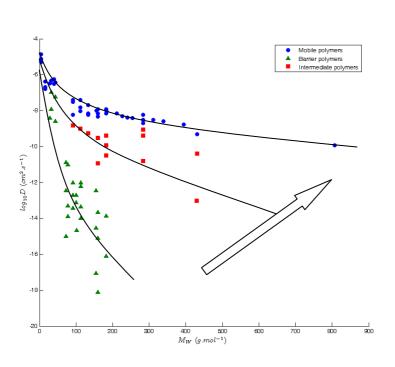
Lines are drawn only as guides. The dotted line (Npip4), however, has a lower slope than

other plain lines.

#### Figure 7. Comparison of the diffusion of two probes in amorphous or semi crystalline

polystyrenes.

<text>



### Figure 1. Diffusive behaviour of different migrants in polymers at 40°C, redrawn from Dole et al.'s data (Dole et al. 2006).

Circles refer to diffusion in mobile polymers, such as LLDPE, LDPE, plasticized PVC, plasticized PP), triangles show D for barrier polymers (dry PA, dry EVOH, unplasticized PVC, PET, PMMA, PS), and squares refer to intermediate polymers (PA at 60%RH, PP). Diffusion was carried out at 40°C. The arrow indicates an increasing matrix mobility, embodied by the system glass transition temperature.

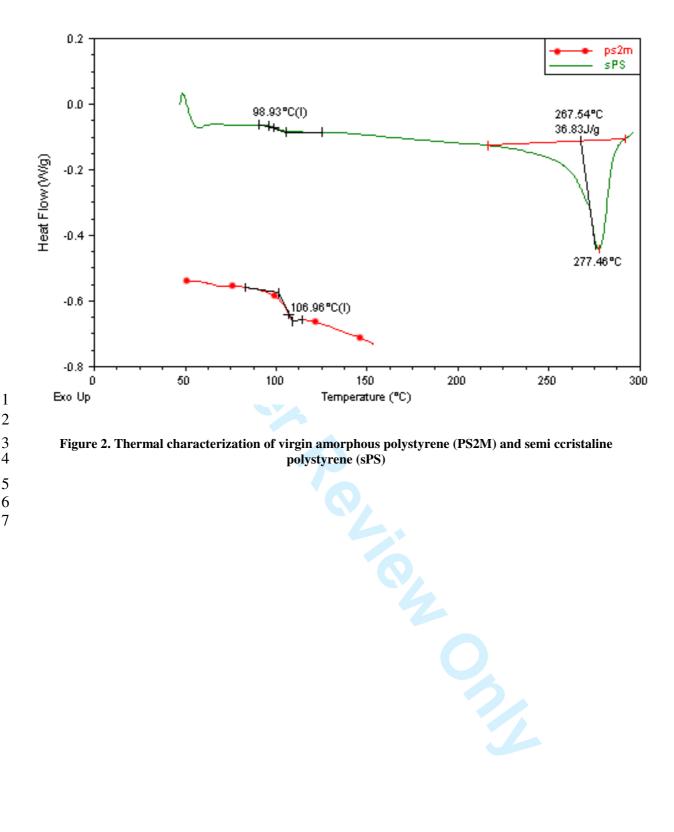
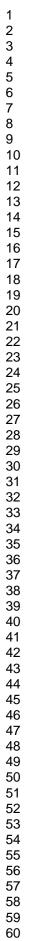


Figure 2. Thermal characterization of virgin amorphous polystyrene (PS2M) and semi ccristaline



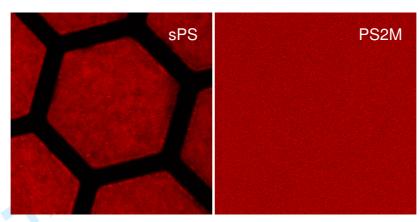
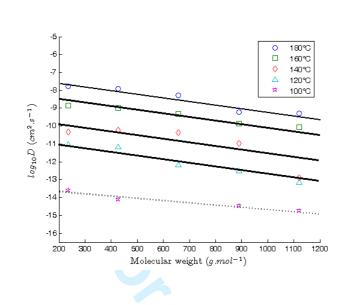
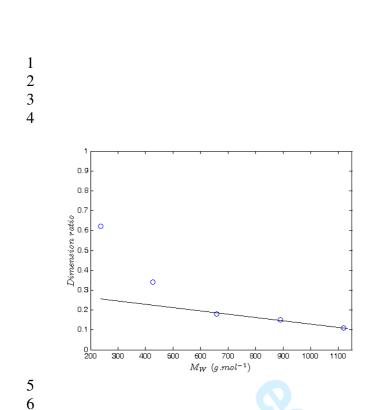


Figure 3. Semi-cristallilne polystyrene (sPS) sample, showing non homogeneously dispersed Npip2 probes
 (left), and homogeneous dispersion in amorphous polystyrene (PS2M) (right). Zoom: 1.9, picture size:
 125x125 μm. The black pattern of the copper hexagonal grid can be seen immersed in the sPS sample.



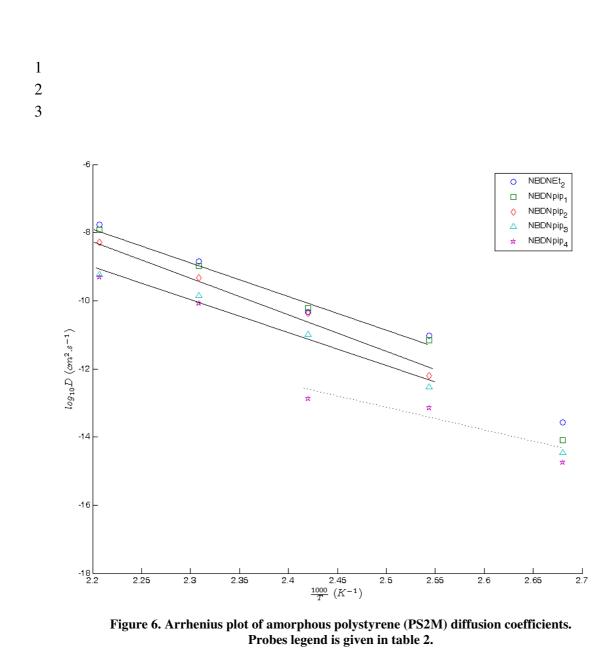
## Figure 4. Diffusion coefficients in amorphous polystyrene (PS2M) as a function of probes M<sub>W</sub> for different temperatures.

Plain lines have the same slope, they are vertically shifted to best describe D=f(M<sub>w</sub>) for each temperature. The dotted line (T=100°C) has a different slope.



## Figure 5. Shape ratio of the different probes used for FRAP experiment, from NBDNEt<sub>2</sub> to NBDNpip<sub>4</sub> 'see tables 2 for probe chemical structures)

The dimension ratio, which is equal to the largest cross section from head to tail, was estimated by drawing the molecules in MarvinSpace (ChemAxon).



Lines are drawn only as guides. The dotted line (Npip4), however, has a lower slope than other

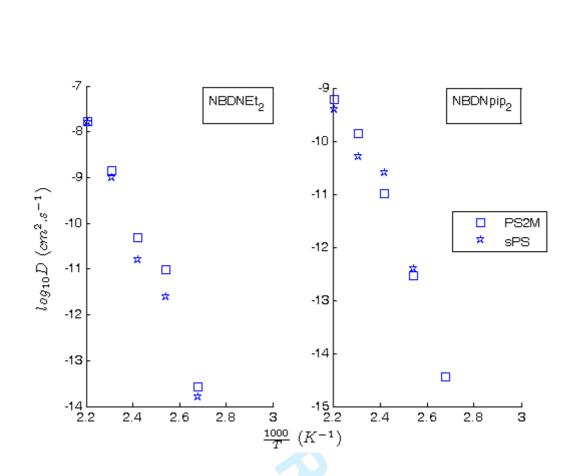


Figure 7. Comparison of the diffusion of two probes in amorphous (PS2M) or semi crystalline polystyrenes (sPS). Probes structures are given in table 2.

#### List of abbreviations

- $M_W$ Molecular weight
  - D diffusion coefficient (
- Ea Activation energy
- FRAP Fluorescence recovery after photo bleaching
- A'<sub>P</sub> tabulated polymer parameters (from the Piringer diffusion model)
- C tabulated polymer parameters (from the Piringer diffusion model)
- T temperature

- D\* overestimated diffusion coefficient (obtained from the Piringer model)
- D'0 diffusion coefficient at infinite temperature (from the Limm model)
- $\alpha$  tabulated polymer parameters (from the Limm model)
- K tabulated polymer parameters (from the Limm model)
- PVC, polyvinylchloride
- PA polyamide
- RH relative humidity
- PET polyethylene terephtalate
- PS2M Polystyrene with High molecular weight (amorphous PS)
- sPS syndiotactique polystyrene (semi-crystalline)
- ROI region of interest
- l of the prove. NBD : NBD is the fluorescent kernel of the probes (nitrobenzoxadiazol)