

Pharmacotherapies and their influence on asymmetric dimethylargine (ADMA)

Renke Maas

▶ To cite this version:

Renke Maas. Pharmacotherapies and their influence on asymmetric dimethylargine (ADMA). Vascular Medicine, 2005, 10 (2), pp.S49-S57. 10.1191/1358863x05vm605oa . hal-00572121

HAL Id: hal-00572121 https://hal.science/hal-00572121

Submitted on 1 Mar 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Pharmacotherapies and their influence on asymmetric dimethylargine (ADMA)

Renke Maas

Abstract: Elevated plasma concentrations of the endogenous nitric oxide synthase inhibitor asymmetric dimethylarginine (ADMA) are found in various clinical settings, including renal failure, coronary heart disease, hypertension, diabetes and preeclampsia. In healthy people acute infusion of ADMA promotes vascular dysfunction, and in mice chronic infusion of ADMA promotes progression of atherosclerosis. Thus, ADMA may not only be a marker but also an active player in cardiovascular disease, which makes it a potential target for therapeutic interventions. This review provides a summary and critical discussion of the presently available data concerning the effects on plasma ADMA levels of cardiovascular drugs, hypoglycemic agents, hormone replacement therapy, antioxidants, and vitamin supplementation. We assess the evidence that the beneficial effects of drug therapies on vascular function can be attributed to modification of ADMA levels. To develop more specific ADMA-lowering therapies, mechanisms leading to elevation of plasma ADMA concentrations in cardiovascular disease need to be better understood. ADMA is formed endogenously by degradation of proteins containing arginine residues that have been methylated by S-adenosylmethionine-dependent methyltransferases (PRMTs). There are two major routes of elimination: renal excretion and enzymatic degradation by the dimethylarginine dimethylaminohydrolases (DDAH-1 and -2). Oxidative stress causing upregulation of PRMT expression and/or attenuation of DDAH activity has been suggested as a mechanism and possible drug target in clinical conditions associated with elevation of ADMA. As impairment of DDAH activity or capacity is associated with substantial increases in plasma ADMA concentrations, DDAH is likely to emerge as a prime target for specific therapeutic interventions.

Key words: asymmetric dimethylarginine (ADMA); cardiovascular disease; DDAH; drug therapy; nitric oxide synthase inhibitor

Introduction

The L-arginine–nitric oxide pathway is involved in the regulation of diverse physiological functions including endothelium-dependent vasodilation, platelet aggregation, neurotransmission, immune defense and apoptosis.¹ Accumulating evidence links asymmetric dimethylarginine (ADMA), an endogenous inhibitor of nitric oxide synthase (NOS), to human disease.² Elevated plasma ADMA concentrations are found in various clinical settings ranging from renal failure and liver failure to atherosclerosis, hypertension, diabetes and pre-eclampsia.^{3–11} Moreover, in patients with cardiovascular disease, end-stage renal disease, or critical illness, elevated plasma ADMA concentrations independently predict progression of atherosclerosis, future vascular events and/or overall mortality.^{12–15}

Several lines of evidence suggest that ADMA, being a NOS inhibitor, may not only be a marker of disease but a player. In human volunteers, infusion of ADMA has been shown impair to endothelium-dependent vasodilation, increase systemic and renal vascular resistance, and decrease cardiac output.^{3,16–18} These data suggest that chronic elevation of ADMA may directly promote vascular disease, which makes ADMA a promising new drug target. Studying the effects of commonly used drugs on plasma ADMA levels may be a first step towards gaining some insights into the mechanisms leading to elevated plasma ADMA concentrations. As ADMA may be both a marker and a player in cardiovascular disease it may therefore be difficult to discern indirect drug effects on ADMA levels related to improvement of

Institute of Experimental and Clinical Pharmacology, University Hospital Hamburg-Eppendorf, Hamburg, Germany

Address for correspondence: Renke Maas, Institute of Experimental and Clinical Pharmacology, University Hospital Hamburg-Eppendorf, Martinistrasse 32, 20246 Hamburg, Germany. E-mail: maas@uke.uni-hamburg.de

the underlying condition from direct drug effects (such as drug-induced augmentation of dimethylarginine dimethylaminohydrolase (DDAH) activity).

It is the purpose of this review to evaluate possible mechanisms leading to the elevation of ADMA plasma concentration and critically to review first in vitro and in vivo data concerning potential ADMAlowering therapies.

Formation and elimination of ADMA: mechanisms and targets for therapeutic interventions

Asymmetric dimethylation of L-arginine residues in proteins

The initial step in the endogenous formation of ADMA (Figure 1) is the asymmetric dimethylation of L-arginine residues of various cellular proteins

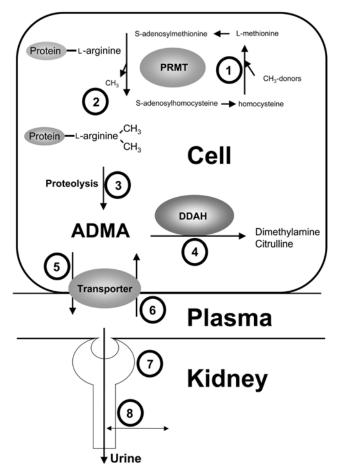


Figure 1 Formation and elimination of ADMA: (1) *S*adenosylmethionine is required as a source of methyl groups; (2) asymmetric methylation of L-arginine residues in proteins by type I protein-arginine methyltransferases (PRMTs); (3) proteolysis leads to liberation of ADMA; (4) degradation of ADMA by dimethylarginine dimethlyaminohydrolase (DDAH); (5) carrier-mediated excretion of ADMA; (6) carrier-mediated uptake of ADMA; (7) renal filtration of ADMA; (8) carrier-mediated secretion and re-uptake of ADMA. by specific protein L-arginine methyltransferases (PRMTs).^{19,20} Type 1 PRMTs are considered the predominant enzymes responsible for asymmetrical dimethylation,²⁰ and S-adenosylmethionine (SAM) derived from L-methionine appears to be the principal methyl group donor for this reaction.²¹ Moreover, elevation of SAM promotes PRMT activity and subsequent formation of ADMA. The product of this methylation reaction, S-adenosylhomocysteine (which inhibits PRMT activity) is hydrolysed to L-homocysteine, which, in turn, can be remethylated by methionine synthase to L-methionine.²² This reaction, which closes the methylation cycle, depends on vitamin B_{12} and 5-methyltetrahydrofolic acid as methyl group donors. In principle, each step of this methylationremethylation cycle constitutes a potential target for therapeutic approaches to lower SAM levels and thus reduce substrate availability for asymmetric dimethylation of L-arginine residues.

Practical and clinical considerations, however, make formation of SAM and PRMTs unlikely targets. First, post-translational methylation of L-arginine residues is considered a universal biological process affecting hundreds of proteins.¹⁹ In most cases the effects of this methylation are still unknown. Thus, interference with this mechanism may have unforseen consequences. Secondly, based upon their role in the homocysteinemethionine cycle, one would expect folate and vitamin B_{12} supplementation to increase synthesis of SAM and subsequent ADMA formation. However, it was shown in patients with increased plasma homocysteine levels that folate lowered elevated plasma ADMA concentrations.²³ In this study, which unfortunately lacked a placebo group, the beneficial effect of folate on ADMA levels was attributed to the reduction of homocysteine by folate. In contrast, in another study, supplementation with folate and B vitamins had no impact on ADMA levels.24

Proteolysis and formation of ADMA

When proteins are degraded, asymmetrically dimethylated L-arginine residues are released as ADMA. Considering the wide variety of asymmetrically dimethylated proteins it is very unlikely that sufficiently specific inhibitors of the proteasome involved in degrading these proteins can be found. Moreover, specific or nonspecific inhibition of protein degradation could lead to accumulation of these proteins or peptide fragments, which potentially could cause a form of storage disease. Hence, proteolysis is unlikely to present a suitable drug target for reducing ADMA levels.

Specific metabolism of ADMA by DDAH

ADMA is selectively degraded by DDAH to citrulline and dimethylamine.²⁵ In humans there are two major isoforms of DDAH (DDAH-1 and DDAH-2) with

	Kidney	Liver	Endothelium
Predominant DDAH isoform	DDAH-1>2	DDAH-1>>2	DDAH-2>>>1
Organ weight (kg)	0.3 (0.15 × 2)	1.5	~1.5
Increase of ADMA in organ failure	Yes 1.5–10×	Yes 6–12×	Likely but causality not established
Net organ extraction (%)	16 (arterial-venous)	~30ª	?

 Table 1
 Major sites of ADMA elimination.

^aEstimate based on rat data.

partly overlapping tissue distributions²⁶ in kidney, liver and small intestine (Table 1). Presently, it is not known what percentage of the endogenously formed ADMA is directly metabolized in the cells where it is formed and how much is released from the cells into other tissues to be metabolized or excreted. It is also not known which DDAH isoform or which organ dominates ADMA degradation in humans in vivo. Rising plasma concentrations in renal³ or liver failure⁴ strongly suggest, however, that at least some tissues release a substantial fraction of the endogenously formed ADMA to be metabolized or excreted elsewhere. With an estimated 'organ weight' similar to that of the liver, the endothelium could be another major site of ADMA metabolism (Table 1).²⁷ Current estimates of the relative expression of the two isoforms are mainly based on mRNA data. DDAH-1 expression tends to dominate in tissues with a high expression of the neuronal isoform of NOS (nNOS), whereas DDAH-2 expression seems to follow the expression pattern of endothelial NOS isoform (eNOS). Analysis of the DDAH-2 gene and its promotor regions revealed potential binding sites for regulators of transcription (STAT, PPAR/RXR, NF-KB and IRF-1) as well as features consistent with the function of a housekeeping gene (absence of canonical TATA boxes and a CpG island around the transcription start sites). Specific activators of DDAH expression are not yet known, but activation of DDAH expression is of course an interesting drug target.

In cell culture all-*trans*-retinoic acid increased DDAH-2 expression.²⁸ Regulation of DDAH-1 expression is even less well understood. In smooth muscle cells interleukin-1 beta upregulates DDAH-1 expression at the protein level.²⁹ DDAH contains four highly redox sensitive cysteine groups, which appear to be crucial for its catalytic function.^{25,30} DDAH activity may also be regulated by *S*-nitrosylation of a reactive cysteine residue (Cys-249),³⁰ providing some form of an indirect negative feedback, especially for inducible NOS (iNOS)-derived nitric oxide (NO).

Therapeutic augmentation of DDAH expression or activity: a word of caution

Although increased NO synthesis due to upregulation of DDAH expression or activity is usually desired in cardiovascular disease, it may have catastrophic effects when occurring in tumors (by promoting NOmediated angiogenesis). Indeed, recent studies suggest that DDAH activity is increased in human tumors.³¹ Moreover, overexpression of DDAH-1 or -2 enhances tumor growth and tumor angiogenesis.³²

Transport and exchange of ADMA

When assessing drug effects on ADMA levels, a major drawback of all available clinical data has to be considered. We currently measure the plasma concentration, which may represent only some form of 'cellular spillover'. Little is known regarding the correlation of plasma ADMA and intracellular ADMA concentrations in vivo. Thus, even a normal or normalized plasma ADMA concentration may not completely exclude severe derangement of intracellular ADMA concentrations by disease or drugs.

Given its structural similarities to L-arginine, the cationic amino acid ADMA most likely shares carrier systems with L-arginine to leave or enter cells.³³ Indeed, early in vitro data suggest that transport of ADMA may at least in part be mediated by y+ carriers.³⁴ Clinical data regarding the precise transport mechanisms for ADMA and their regulation in vivo are not currently available.

Renal filtration and excretion of ADMA

Elevated plasma levels of ADMA were first reported in patients with chronic renal failure by Vallance and co-workers.³ We now know that the combination of renal filtration, tubular secretion, tubular reabsorption and degradation by renal DDAH result in a net renal extraction of about 16% of circulating ADMA.³⁵ Presently it is difficult, however, to estimate the relative contribution of each mechanism. It is interesting that symmetric dimethylarginine, which is chemically very similar to ADMA but not metabolized by DDAH, has a renal extraction rate of only 10%. This difference may be interpreted as evidence for a substantial contribution of renal DDAH activity to the renal extraction of ADMA.

Exogenous sources of ADMA

Protein-L-arginine methylation is a universal biological principle found in bacteria as well as in plants and animals.³⁶ It is therefore very likely that free and protein-incorporated ADMA³⁷ is also taken in with the diet, modulated by dietary components, or formed by resident colonic bacteria. This offers several further targets for ADMA-lowering therapies that have not yet been investigated.

Major disease mechanisms leading to elevation of ADMA in human disease

Shear stress

In vitro, shear stress may have divergent effects on ADMA.³⁸ Moderate shear stress was found to upregulate PRMT1 expression while DDAH activity was unchanged, resulting in enhanced cellular ADMA release. In contrast, when cells were exposed to a higher shear stress, PRMT1 and DDAH activity both increased, resulting in normalization of ADMA levels.

High blood glucose and impaired glucose tolerance

Elevation of plasma ADMA levels have been observed in a rat model of diabetes, and DDAH activity was found to be significantly impaired in human cells exposed to high concentrations of glucose.³⁹ Since it was shown that impairment of DDAH activity could be alleviated by polyethylene glycol-conjugated superoxide dismutase, impairment of DDAH activity by glucose-induced oxidative stress was suggested as the major underlying mechanism. In humans the situation appears to be more complex. In volunteers without diabetes, plasma ADMA levels were found to be correlated with individual insulin sensitivity but not with blood glucose level.¹⁰ Clinical data concerning plasma ADMA levels in patients with diabetes are somewhat conflicting: some studies have shown an elevation of ADMA⁴⁰ while others demonstrated a reduction.⁴¹

Oxidative stress

Several independent lines of evidence indicate that impairment of DDAH activity by oxidative stress is the final mechanism leading to elevation of ADMA in diverse clinical conditions.⁴² DDAH contains four highly redox sensitive cysteine groups, which appear to be crucial for its catalytic function.^{25,30}

Oxidized low-density lipoprotein (oxLDL) but not native LDL was found to decrease DDAH activity⁴³ and increase cellular ADMA liberation. In cell culture, oxLDL and high concentrations of native LDL lead to upregulation of PRMTs and cellular ADMA synthesis that could be prevented by antioxidants.⁴⁴

Unfortunately, impairment of DDAH activity by oxidative stress or upregulation of PRMTs may not be easy to overcome in humans. A Finnish study found no association between plasma ADMA levels and dietary vitamin E or C intake.⁴⁵ In an open label study, 800 IU vitamin E over 6 weeks had no significant effect on plasma ADMA.⁴⁶ Moreover, in a very recent

clinical trial *N*-acetyl-cysteine had no effect on plasma ADMA levels.⁴⁷

Hyperhomocysteinemia

In animals⁴⁸ and humans^{49,50} experimental hyperhomocysteinemia is associated with elevation of plasma ADMA. Two (not necessarily exclusive) mechanisms have been proposed to explain this finding:

- 1. Homocysteine may be remethylated to form Lmethionine, which, in turn, serves as a methyl group donator for the methylation of L-arginine.²² Increased ADMA release by endothelial cells under high concentrations of methionine or homocysteine was reduced by the methylation inhibitor *S*-adenosylmethionine.⁴⁴ In monkeys, dietary supplementation with B vitamins normalized homocysteine but did not alter ADMA levels or improve endothelial function.⁴⁸
- 2. Homocysteine may also directly interact with the sulfhydryl groups of DDAH. In an in vitro binding assay, biotinylated homocysteine was found to bind directly to DDAH-1.⁵¹ This binding could be prevented by an antioxidant.

Drug therapy and ADMA: early clinical data

Statin therapy

Statins are among the most effective drugs for lowering cardiovascular risk associated with elevation of LDL cholesterol. Several lines of evidence suggest that the beneficial effects of statins are at least in part mediated by LDL-independent (pleiotropic) mechanisms such as improvement of endothelial function by increasing the bioavailability of NO. In animal experiments as well as in patients a high LDL cholesterol level was found to be associated with elevated plasma ADMA concentrations.^{48,52} Even though this finding does not seem to be found consistently in all patient populations,⁵³ several investigators have tested the hypothesis that statin treatment may improve vascular function by reducing plasma ADMA (Table 2). In the majority of studies, statin treatment had no impact on plasma ADMA levels.^{54–56} This would clearly argue against a direct connection between ADMA and LDL cholesterol in humans. Only rosuvastatin was shown to lower ADMA⁵⁷ but, given the negative results for the other statins, this finding remains to be replicated in other populations.

Irrespective of the potential effects of statins on ADMA, recent studies suggest that ADMA may modulate the effect of statins on vascular function.⁵⁸

Angiotensin converting enzyme inhibitors and angiotensin receptor blockers

In various conditions such as hypertension, diabetes and coronary artery disease, angiotensin converting

Drug (ref.)	Duration (weeks)	n	% change	n	
	Duration (weeks)		70 change	μ	
Pravastatin 40 mg/d (Eid et al 2003 ⁵⁴)	8	32	-9	ns	
Pravastatin 40 mg/d (Janatuinen et al 2001 ⁵⁵)	26	21	No change	ns	
Simvastatin 80 mg/d (Päivä et al 2003 ⁵³)	8	24	No change	ns	
Atorvastatin 40 mg/d (Päivä et al 2003 ⁵³)	8	24	No change	ns	
Simvastatin 20 mg/d (Pereira et al 2003 ⁵⁶)	8	25	No change	ns	
Rosuvastatin 10 mg/d (Lu et al 2004 ⁵⁷)	6	23	-18	<0.001	

 Table 2
 Effect of statin therapy on plasma ADMA levels.

ns, not significant.

enzyme (ACE) inhibitors and angiotensin receptor blockers (ARBs) were found to improve endothelial function and NO availability.^{59–61} Several studies have assessed the effect of ACE inhibitors or ARBs on plasma ADMA levels in these patient groups (Table 3).^{62–66} In patients with hypertension, diabetes or syndrome X, the reduction in plasma ADMA achieved by ACE inhibition was in the range of 12–20%, while plasma L-arginine levels did not seem to be affected.⁶³ ARBs were found to reduce plasma ADMA by 10–16%.^{62,66} All of these studies were small and had rather short observation periods of 1–12 weeks. Thus, confirmation of these promising results by larger trials will be required.

The results of a recent study in eNOS knockout mice suggests that ADMA may also exert NO-independent effects via upregulation of ACE and augmentation of oxidative stress through AT₁-dependent pathways.⁶⁷ Thus, ACE inhibitors and ARBs may help to prevent ADMA-induced pathology independently of their effects on plasma ADMA levels.

Hypoglycemic agents

In patients with type 2 diabetes, 3 months of treatment with 0.5–1.0 g/d metformin alone (n = 16) or in combination with sulfonylurea (n = 15) reduced plasma ADMA concentrations by 28.5% and 32.0% respectively.⁶⁸ Some caution is warranted, however, as no placebo data are available from that study. Furthermore, in contrast to what one would expect

from the in vitro data, no relationship was observed between the decrease in ADMA concentration and glycemic control. Thus, it is possible that the reduction of ADMA levels by metformin is mediated via glucose-independent mechanisms. In this respect it may be of interest to note that metformin has structural similarities to ADMA.⁶⁹

In an uncontrolled study the PPAR-gamma agonist rosiglitazone (12 weeks of treatment with a final dose of up to 8 mg/d) improved insulin sensitivity and reduced plasma ADMA concentrations by 30% in seven insulin-resistant patients.¹⁰ These data certainly await further confirmation by larger controlled trials because, in vitro, neither PPAR-gamma agonists (ciglitazone) nor PPAR-gamma antagonists (T007) significantly altered cellular ADMA liberation.⁷⁰ Apart from the effects on blood glucose there is, however, at least support from theoretical considerations because a PPAR/PXR binding site has been identified in the promotor region of DDAH-2.²⁸

Hormone replacement therapy

Hormone replacement therapy (HRT) has frequently been shown to improve endothelial function and NO synthesis.^{71,72} Moreover, ADMA levels were found to decrease in normal pregnancy.¹¹ The precise mechanism by which estrogen reduces ADMA has remained obscure so far. The effect of various forms of HRT on ADMA plasma concentration was explored in some small randomized controlled trials (Table 4).^{73–75} In all

Drug (ref.)	Duration (weeks)	n	(% change)	р
Enalapril 20 mg/d (Delles et al 2002 ⁶²)	1	20	-17%	< 0.05
Enalapril 10 g/d (Chen et al 2002 ⁶³)	8	10	-20%	<0.01
Enalapril 20 mg/d (Napoli et al 2004 ⁶⁴)	12	38	−16 %	<0.05
Zofenopril15–30 mg/d (Napoli et al 2004 ⁶⁴)	12	38	−24%	<0.01
Perindopril 4 mg/d (Ito et al 2001 ⁶⁶)	4	7	−12%ª	<0.05
Perindopril 4 mg/d (Ito et al 2002 ⁶⁵)	1	11	−16 %ª	0.02
Eprosartan 600 mg/d (Delles et al 2002 ⁶²)	1	20	16	<0.05
Eprosartan 300 mg/d + enalapril 20 mg/d (Delles et al 2002 ⁶²)	1	20	18	<0.05
Losartan 50 mg/d (Ito et al 2001 ⁶⁶)	4	7	10	<0.05

^aData extracted from graphs in publication.

Drug and dose (ref.)	Duration (weeks)	ADMA (% change; µmol/l)	p	L-arginine (% change; µmol/l)	p	∟-arginine/ ADMA ratio (% change)	p	Placebo
17-beta-estradiol 2 mg/d (Post et al 2003 ⁷⁴)	12	-5.2	0.03	-7.3	0.02	-1.6	0.75	Yes
17-beta-estradiol 2 mg/d + dydrogesterone 10 mg/d (Post et al 2003 ⁷⁴)	12	-8.0	0.02	-7.9	0.14	0.3	0.83	Yes
17-beta-estradiol 2 mg/d + trimegestone 10 mg/d (Post et al 2003 ⁷⁴)	12	-21	<0.01	-36	<0.001	-19	<0.01	Yes
Conjugated equine estrogen 0.625 mg/d (Teerlink et al 2003 ⁷⁵)	104	-9.7	<0.01	No change; data not reported	-	Not reported	-	Yes
17-beta-estradiol 100 mg implant (Holden et al 2003 ⁷³)	2	-18	<0.05	Not reported	-	Not reported	-	No

 Table 4
 Effect of hormone therapy on ADMA plasma concentration.

-, data not cited.

of these trials ADMA levels were modestly (5–21%) reduced by 17-beta-estradiol alone or when sequentially combined with cyclogesterone or trimegestone. Unfortunately, L-arginine levels also declined, resulting in no change or even a decrease in the L-arginine/ ADMA ratio. Thus, unless it can be proven that the decrease in L-arginine was the result of increased consumption in NO synthesis (and not upregulation of arginase activity), it remains questionable how much benefit can be conferred by HRT on patients with an elevated ADMA.

Conclusion and outlook

Specific ADMA-lowering therapies are not yet available. Promising results from first pilot studies with widely used drugs such as ACE-inhibitors, ARBs, hypoglycemic agents and HRT await confirmation in large placebo-controlled trials.

To develop more specific therapies, further insights into the mechanisms leading to an elevation of ADMA in vivo are needed. Tools such as high-throughput methods for the quantification of ADMA and isoformspecific DDAH inhibitors (and activators) will play a key role. Important insights may also come from the DDAH-transgenic mice⁷⁶ and DDAH1/2 knock out models currently under development. So far, most conditions associated with an elevation of ADMA are chronic, so ADMA-lowering therapies will have to be long term. Future studies will therefore have to address carefully the long-term effects of ADMAlowering therapies on angiogenesis and tumor growth.

Until specific ADMA-lowering therapies become available, treatment of modifiable risk factors leading to elevation of ADMA or depletion of L-arginine, and supplementation with L-arginine, may be the best available options for patients with high plasma ADMA concentrations because NO synthesis is correlated with the L-arginine/ADMA ratio.^{77,78}

References

- 1 Moncada S, Higgs A. The L-arginine-nitric oxide pathway. *N* Engl J Med 1993; **329**: 2002–12.
- 2 Cooke JP. Asymmetrical dimethylarginine: the Über marker? *Circulation* 2004; **109**: 1813–18.
- 3 Vallance P, Leone A, Calver A, Collier J, Moncada S. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. *Lancet* 1992; **339**: 572–75.
- 4 Tsikas D, Rode I, Becker T, Nashan B, Klempnauer J, Frölich JC. Elevated plasma and urine levels of ADMA and 15(S)-8-iso-PGF2alpha in end-stage liver disease. *Hepatology* 2003; 38: 1063–64.
- 5 Miyazaki H, Matsuoka H, Cooke JP et al. Endogenous nitric oxide synthase inhibitor: a novel marker of atherosclerosis. *Circulation* 1999; **99**: 1141–46.
- 6 Kielstein JT, Böger RH, Bode-Böger SM et al. Marked increase of asymmetric dimethylarginine in patients with incipient primary chronic renal disease. *J Am Soc Nephrol* 2002; **13**: 170–76.
- 7 Yoo JH, Lee SC. Elevated levels of plasma homocyst(e)ine and asymmetric dimethylarginine in elderly patients with stroke. *Atherosclerosis* 2001; **158**: 425–30.
- 8 Taddei S, Virdis A, Mattei P et al. Aging and endothelial function in normotensive subjects and patients with essential hypertension. *Circulation* 1995; **91**: 1981–87.
- 9 Tarnow L, Hovind P, Teerlink T, Stehouwer CD, Parving HH. Elevated plasma asymmetric dimethylarginine as a marker of cardiovascular morbidity in early diabetic nephropathy in type 1 diabetes. *Diabetes Care* 2004; **27**: 765–69.
- 10 Stühlinger MC, Abbasi F, Chu JW et al. Relationship between insulin resistance and an endogenous nitric oxide synthase inhibitor. *JAMA* 2002; 287: 1420–26.

- 11 Holden DP, Fickling SA, Whitley GS, Nussey SS. Plasma concentrations of asymmetric dimethylarginine, a natural inhibitor of nitric oxide synthase, in normal pregnancy and preeclampsia. *Am J Obstet Gynecol* 1998; **178**: 551–56.
- 12 Valkonen VP, Päivä H, Salonen JT et al. Risk of acute coronary events and serum concentration of asymmetrical dimethylarginine. *Lancet* 2001; **358**: 2127–28.
- 13 Zoccali C, Bode-Böger S, Mallamaci F et al. Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: a prospective study. *Lancet* 2001; **358**: 2113–17.
- 14 Lu TM, Ding YA, Lin SJ, Lee WS, Tai HC. Plasma levels of asymmetrical dimethylarginine and adverse cardiovascular events after percutaneous coronary intervention. *Eur Heart* J 2003; 24: 1912–19.
- 15 Zoccali C, Benedetto FA, Maas R et al. CREED Investigators. Asymmetric dimethylarginine, C-reactive protein, and carotid intima-media thickness in end-stage renal disease. *J Am Soc Nephrol* 2002; **13**: 490–96.
- 16 Calver A, Collier J, Leone A, Moncada S, Vallance P. Effect of local intra-arterial asymmetric dimethylarginine (ADMA) on the forearm arteriolar bed of healthy volunteers. *J Hum Hypertens* 1993; 7: 193–94.
- 17 Achan V, Broadhead M et al. Asymmetric dimethylarginine causes hypertension and cardiac dysfunction in humans and is actively metabolized by dimethylarginine dimethylaminohydrolase. *Arterioscler Thromb Vasc Biol* 2003; **23**: 1455–59.
- 18 Kielstein JT, Impraim B, Simmel S et al. Cardiovascular effects of systemic nitric oxide synthase inhibition with asymmetrical dimethylarginine in humans. *Circulation* 2004; **109**: 172–77.
- 19 Boisvert FM, Cote J, Boulanger MC, Richard S. A proteomic analysis of arginine-methylated protein complexes. *Mol Cell Proteomics* 2003; 2: 1319–30.
- 20 Tang J, Frankel A, Cook RJ et al. PRMT1 is the predominant type 1 protein arginine methyltransferase in mammalian cells. *J Biol Chem* 2000; **275**: 7723–30.
- 21 Böger RH, Sydow K, Borlak J et al. LDL cholesterol upregulates synthesis of asymmetrical dimethylarginine in human endothelial cells: involvement of S-adenosylmethioninedependent methyltransferases. *Circ Res* 2000; 87: 99–105.
- 22 Lentz SR, Rodionov RN, Dayal S. Hyperhomocysteinemia, endothelial dysfunction, and cardiovascular risk: the potential role of ADMA. *Atheroscler Suppl* 2003; **4**(4): 61–65.
- 23 Holven KB, Haugstad TS, Holm T, Aukrust P, Ose L, Nenseter MS. Folic acid treatment reduces elevated plasma levels of asymmetric dimethylarginine in hyperhomocysteinaemic subjects. *Br J Nutr* 2003; **89**: 359–63.
- 24 Sydow K, Schwedhelm E, Arakawa N et al. ADMA and oxidative stress are responsible for endothelial dysfunction in hyperhomocyst(e)inemia: effects of L-arginine and B vitamins. *Cardiovasc Res* 2003; **57**: 244–52.
- 25 Ogawa T, Kimoto M, Sasaoka K. Purification and properties of a new enzyme, NG,NG-dimethylarginine dimethylaminohydrolase, from rat kidney. *J Biol Chem* 1989; 264: 10205–209.
- 26 Tran CT, Fox MF, Vallance P, Leiper JM. Chromosomal localization, gene structure, and expression pattern of DDAH1: comparison with DDAH2 and implications for evolutionary origins. *Genomics* 2000; **68**: 101–105.
- 27 McCarty MF. Vascular endothelium is the organ chiefly responsible for the catabolism of plasma asymmetric dimethylarginine – an explanation for the elevation of plasma ADMA

in disorders characterized by endothelial dysfunction. *Med Hypotheses* 2004; **63**: 699–708.

- 28 Achan V, Tran CT, Arrigoni F, Whitley GS, Leiper JM, Vallance P. all-*trans*-Retinoic acid increases nitric oxide synthesis by endothelial cells: a role for the induction of dimethylarginine dimethylaminohydrolase. *Circ Res* 2002; **90**: 764–69.
- 29 Ueda S, Kato S, Matsuoka H et al. Regulation of cytokineinduced nitric oxide synthesis by asymmetric dimethylarginine: role of dimethylarginine dimethylaminohydrolase. *Circ Res* 2003; **92**: 226–33.
- 30 Leiper J, Murray-Rust J, McDonald N, Vallance P. S-nitrosylation of dimethylarginine dimethylaminohydrolase regulates enzyme activity: further interactions between nitric oxide synthase and dimethylarginine dimethylaminohydrolase. *Proc Natl Acad Sci U S A* 2002; **99**: 13527–32.
- 31 Kostourou V, Robinson SP, Whitley GS, Griffiths JR. Effects of overexpression of dimethylarginine dimethylaminohydrolase on tumor angiogenesis assessed by susceptibility magnetic resonance imaging. *Cancer Res* 2003; 63: 4960–66.
- 32 Kostourou V, Robinson SP, Cartwright JE, Whitley GS. Dimethylarginine dimethylaminohydrolase 1 enhances tumour growth and angiogenesis. *Br J Cancer* 2002; **87** 673–80.
- 33 Brosnan ME, Brosnan JT. Renal arginine metabolism. J Nutr 2004; 134(10 Suppl): 2796S–97S.
- 34 Closs EI, Basha FZ, Habermeier A, Forstermann U. Interference of L-arginine analogues with L-arginine transport mediated by the y+ carrier hCAT-2B. *Nitric Oxide* 1997; 1: 65–73.
- 35 Nijveldt RJ, Van Leeuwen PA, Van Guldener C, Stehouwer CD, Rauwerda JA, Teerlink T. Net renal extraction of asymmetrical (ADMA) and symmetrical (SDMA) dimethylarginine in fasting humans. *Nephrol Dial Transplant* 2002; 17: 1999–2002.
- 36 McBride AE, Silver PA. State of the arg:protein methylation at arginine comes of age. *Cell* 2001; **106**: 5–8.
- 37 Al Banchaabouchi M, Marescau B, Possemiers I, D'Hooge R, Levillain O, De Deyn PP. NG, NG-dimethylarginine and NG, NG-dimethylarginine in renal insufficiency. *Pflugers Arch* 2000; **439**: 524–31.
- 38 Osanai T, Saitoh M, Sasaki S, Tomita H, Matsunaga T, Okumura K. Effect of shear stress on asymmetric dimethylarginine release from vascular endothelial cells. *Hypertension* 2003; 42: 985–90.
- 39 Lin KY, Ito A, Asagami T et al. Impaired nitric oxide synthase pathway in diabetes mellitus: role of asymmetric dimethylarginine and dimethylarginine dimethylaminohydrolase. *Circulation* 2002; **106**: 987–92.
- 40 Abbasi F, Asagmi T, Cooke JP et al. Plasma concentrations of asymmetric dimethylarginine are increased in patients with type 2 diabetes mellitus. *Am J Cardiol* 2001; **88**: 1201–203.
- 41 Päivä H, Lehtimaki T, Laakso J et al. Plasma concentrations of asymmetric-dimethyl-arginine in type 2 diabetes associate with glycemic control and glomerular filtration rate but not with risk factors of vasculopathy. *Metabolism* 2003; **52**: 303–307.
- 42 Sydow K, Münzel T. ADMA and oxidative stress. *Atheroscler Suppl* 2003; **4**(4): 41–51.
- 43 Ito A, Tsao PS, Adimoolam S, Kimoto M, Ogawa T, Cooke JP. Novel mechanism for endothelial dysfunction: dysregulation of dimethylarginine dimethylaminohydrolase. *Circulation* 1999; **99**: 3092–95.

- 44 Böger RH, Sydow K, Borlak J et al. LDL cholesterol upregulates synthesis of asymmetrical dimethylarginine in human endothelial cells: involvement of S-adenosylmethioninedependent methyltransferases. *Circ Res* 2000; **87**: 99–105.
- 45 Päivä H, Lehtimaki T, Laakso J et al. Dietary composition as a determinant of plasma asymmetric dimethylarginine in subjects with mild hypercholesterolemia. *Metabolism* 2004; 53: 1072–75.
- 46 Saran R, Novak JE, Desai A et al. Impact of vitamin E on plasma asymmetric dimethylarginine (ADMA) in chronic kidney disease (CKD): a pilot study. *Nephrol Dial Transplant* 2003; **18**: 2415–20.
- 47 Doshi S, McDowell IF, Goodfellow J et al. Relationship between *S*-adenosylmethionine, *S*-adenosylhomocysteine, asymmetric dimethylarginine and endothelial function in healthy human subjects during experimental hyper- and hypohomocysteinemia. *Metabolism* 2005; **54**: 351–60.
- 48 Böger RH, Bode-Böger SM, Sydow K, Heistad DD, Lentz SR. Plasma concentration of asymmetric dimethylarginine, an endogenous inhibitor of nitric oxide synthase, is elevated in monkeys with hyperhomocyst(e)inemia or hypercholesterolemia. Arterioscler Thromb Vasc Biol 2000; 20: 1557–64.
- 49 Stühlinger MC, Oka RK, Graf EE et al. Endothelial dysfunction induced by hyperhomocyst(e)inemia: role of asymmetric dimethylarginine. *Circulation* 2003; **108**: 933–38.
- 50 Böger RH, Lentz SR, Bode-Böger SM, Knapp HR, Haynes WG. Elevation of asymmetrical dimethylarginine may mediate endothelial dysfunction during experimental hyperhomocyst(e)inaemia in humans. *Clin Sci (Lond)* 2001; **100**: 161–67.
- 51 Stühlinger MC, Oka RK, Graf EE et al. Endothelial dysfunction induced by hyperhomocyst(e)inemia: role of asymmetric dimethylarginine. *Circulation* 2003; **108**: 933–38.
- 52 Böger RH, Bode-Böger SM, Szuba A et al. Asymmetric dimethylarginine (ADMA): a novel risk factor for endothelial dysfunction: its role in hypercholesterolemia. *Circulation* 1998; **98**: 1842–47.
- 53 Päivä H, Laakso J, Lehtimaki T, Isomustajarvi M, Ruokonen I, Laaksonen R. Effect of high-dose statin treatment on plasma concentrations of endogenous nitric oxide synthase inhibitors. *J Cardiovasc Pharmacol* 2003; **41**: 219–22.
- 54 Eid HM, Eritsland J, Larsen J, Arnesen H, Seljeflot I. Increased levels of asymmetric dimethylarginine in populations at risk for atherosclerotic disease. Effects of pravastatin. *Atherosclerosis* 2003; **166**: 279–84.
- 55 Janatuinen T, Laaksonen R, Vesalainen R et al. Effect of lipidlowering therapy with pravastatin on myocardial blood flow in young mildly hypercholesterolemic adults. *J Cardiovasc Pharmacol* 2001; 38: 561–68.
- 56 Pereira EC, Bertolami MC, Faludi AA, Salem M, Bersch D, Abdalla DS. Effects of simvastatin and L-arginine on vasodilation, nitric oxide metabolites and endogenous NOS inhibitors in hypercholesterolemic subjects. *Free Radic Res* 2003; **37**: 529–36.
- 57 Lu TM, Ding YA, Leu HB, Yin WH, Sheu WH, Chu KM. Effect of rosuvastatin on plasma levels of asymmetric dimethylarginine in patients with hypercholesterolemia. *Am J Cardiol* 2004; 94: 157–61.
- 58 Janatuinen T, Laakso J, Laaksonen R et al. Plasma asymmetric dimethylarginine modifies the effect of pravastatin on myocardial blood flow in young adults. *Vasc Med* 2003; 8: 185–89.
- 59 Joannides R, Bizet-Nafeh C, Costentin A et al. Chronic ACE inhibition enhances the endothelial control of arterial mechanics and flow-dependent vasodilatation in heart failure. *Hypertension* 2001; **38**: 1446–50.

- 60 Higashi Y, Sasaki S, Nakagawa K, Matsuura H, Kajiyama G, Oshima T. Effect of the angiotensin-converting enzyme inhibitor imidapril on reactive hyperemia in patients with essential hypertension: relationship between treatment periods and resistance artery endothelial function. *J Am Coll Cardiol* 2001; **37**: 863–70.
- 61 Hornig B, Landmesser U, Kohler C et al. Comparative effect of ace inhibition and angiotensin II type 1 receptor antagonism on bioavailability of nitric oxide in patients with coronary artery disease: role of superoxide dismutase. *Circulation* 2001; **103**: 799–805.
- 62 Delles C, Schneider MP, John S, Gekle M, Schmieder RE. Angiotensin converting enzyme inhibition and angiotensin II AT1-receptor blockade reduce the levels of asymmetrical N(G), N(G)-dimethylarginine in human essential hypertension. Am J Hypertens 2002; **15**: 590–93.
- 63 Chen JW, Hsu NW, Wu TC, Lin SJ, Chang MS. Long-term angiotensin-converting enzyme inhibition reduces plasma asymmetric dimethylarginine and improves endothelial nitric oxide bioavailability and coronary microvascular function in patients with syndrome X. *Am J Cardiol* 2002; **90**: 974–82.
- 64 Napoli C, Sica V, de Nigris F et al. Sulfhydryl angiotensinconverting enzyme inhibition induces sustained reduction of systemic oxidative stress and improves the nitric oxide pathway in patients with essential hypertension. *Am Heart J* 2004; **148**: e5.
- 65 Ito A, Egashira K, Narishige T, Muramatsu K, Takeshita A. Angiotensin-converting enzyme activity is involved in the mechanism of increased endogenous nitric oxide synthase inhibitor in patients with type 2 diabetes mellitus. *Circ J* 2002; 66: 811–15.
- 66 Ito A, Egashira K, Narishige T, Muramatsu K, Takeshita A. Renin-angiotensin system is involved in the mechanism of increased serum asymmetric dimethylarginine in essential hypertension. *Jpn Circ J* 2001; 65: 775–78.
- 67 Suda O, Tsutsui M, Morishita T et al. Asymmetric dimethylarginine produces vascular lesions in endothelial nitric oxide synthase-deficient mice: involvement of renin-angiotensin system and oxidative stress. *Arterioscler Thromb Vasc Biol* 2004; **24**: 1682–88.
- 68 Asagami T, Abbasi F, Stühlinger M et al. Metformin treatment lowers asymmetric dimethylarginine concentrations in patients with type 2 diabetes. *Metabolism* 2002; **51**: 843–46.
- 69 Vallance P, Leiper J. Cardiovascular biology of the asymmetric dimethylarginine:dimethylarginine dimethylaminohydrolase pathway. *Arterioscler Thromb Vasc Biol* 2004; 24: 1023–30.
- 70 Albsmeier J. Bedeutang endogener Inhibitoren der NO-Synthase für kardiovaskuläre Erkrankungen und ihre pharmakologische Therapie (Relevance of endogenous NOS inhibitors for cardiovascular disease and its therapy) [Thesis FBCh 2550]. Hamburg: University of Hamburg, 2004. Retrieved 7 May, 2005, from: http://www.chemie.unihamburg.de/bibliothek/2004/Dissertation Albsneier.pdf
- 71 Vehkavaara S, Hakala-Ala-Pietila T, Virkamaki A et al. Differential effects of oral and transdermal estrogen replacement therapy on endothelial function in postmenopausal women. *Circulation* 2000; **102**: 2687–93.
- 72 Imthurn B, Rosselli M, Jaeger AW, Keller PJ, Dubey RK. Differential effects of hormone-replacement therapy on endogenous nitric oxide (nitrite/nitrate) levels in postmenopausal women substituted with 17 beta-estradiol valerate and cyproterone acetate or medroxyprogesterone acetate. *J Clin Endocrinol Metab* 1997; **82**: 388–94.

- 73 Holden DP, Cartwright JE, Nussey SS, Whitley GS. Estrogen stimulates dimethylarginine dimethylaminohydrolase activity and the metabolism of asymmetric dimethylarginine. *Circulation* 2003; **108**: 1575–80.
- 74 Post MS, Verhoeven MO, van der Mooren MJ, Kenemans P, Stehouwer CD, Teerlink T. Effect of hormone replacement therapy on plasma levels of the cardiovascular risk factor asymmetric dimethylarginine: a randomized, placebo-controlled 12-week study in healthy early postmenopausal women. *J Clin Endocrinol Metab* 2003; **88**: 4221–26.
- 75 Teerlink T, Neele SJ, de Jong S, Netelenbos JC, Stehouwer CD. Oestrogen replacement therapy lowers plasma levels of

asymmetrical dimethylarginine in healthy postmenopausal women. *Clin Sci (Lond)* 2003; **105**: 67–71.

- 76 Dayoub H, Achan V, Adimoolam S et al. Dimethylarginine dimethylaminohydrolase regulates nitric oxide synthesis: genetic and physiological evidence. *Circulation* 2003; 108: 3042–47.
- 77 Tsikas D, Böger RH, Sandmann J, Bode-Böger SM, Frölich JC. Endogenous nitric oxide synthase inhibitors are responsible for the L-arginine paradox. *FEBS Lett* 2000; 478: 1–3.
- 78 Böger RH, Bode-Böger SM. The clinical pharmacology of L-arginine. *Annu Rev Pharmacol Toxicol* 2001; **41**: 79–99.