

Patterns of Antimicrobial Therapy in Severe Nosocomial Infections: Empiric Choices, Proportion of Appropriate Therapy, and Modification Rates–A Multicenter, Observational Survey in Critically Ill Patients.

Dirk Vogelaers, David de Bels, Frédéric Forêt, Sophie Cran, Eric Gilbert, Karen Schoonheydt, Stijn Blot

► To cite this version:

Dirk Vogelaers, David de Bels, Frédéric Forêt, Sophie Cran, Eric Gilbert, et al.. Patterns of Antimicrobial Therapy in Severe Nosocomial Infections: Empiric Choices, Proportion of Appropriate Therapy, and Modification Rates–A Multicenter, Observational Survey in Critically Ill Patients.. International Journal of Antimicrobial Agents, 2010, 35 (4), pp.375. 10.1016/j.ijantimicag.2009.11.015 . hal-00567278

HAL Id: hal-00567278 https://hal.science/hal-00567278

Submitted on 20 Feb 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Accepted Manuscript

Title: Patterns of Antimicrobial Therapy in Severe Nosocomial Infections: Empiric Choices, Proportion of Appropriate Therapy, and Modification Rates–A Multicenter, Observational Survey in Critically Ill Patients.

Authors: Dirk Vogelaers, David De Bels, Frédéric Forêt, Sophie Cran, Eric Gilbert, Karen Schoonheydt, Stijn Blot



PII: DOI: Reference:	S0924-8579(09) doi:10.1016/j.ija ANTAGE 3193		9.11.0	15	
To appear in:	International	Journal	of	Antimicrobial	Agents
Received date: Revised date: Accepted date:	17-9-2009 22-10-2009 23-11-2009				

Please cite this article as: Vogelaers D, De Bels D, Forêt F, Cran S, Gilbert E, Schoonheydt K, Blot S, Patterns of Antimicrobial Therapy in Severe Nosocomial Infections: Empiric Choices, Proportion of Appropriate Therapy, and Modification Rates–A Multicenter, Observational Survey in Critically III Patients., *International Journal of Antimicrobial Agents* (2008), doi:10.1016/j.ijantimicag.2009.11.015

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1	Patterns of Antimicrobial Therapy in Severe Nosocomial Infections:
2	Empiric Choices, Proportion of Appropriate Therapy, and Adaptation Rates –
3	A Multicenter, Observational Survey in Critically Ill Patients.
4	
5	Dirk Vogelaers ¹ , PhD, David De Bels ² , MD, Frédéric Forêt ³ , MD, Sophie Cran ⁴ , MD,
6	Eric Gilbert ⁵ , MD, Karen Schoonheydt ⁶ , MD, and Stijn Blot ¹ , PhD, for the
7	ANTHICUS Study Investigators*
8	
9	
10	Affiliations:
11	¹ General Internal Medicine & Infectious Diseases, Ghent University Hospital, Ghent,
12	Belgium
13	² Intensive Care Dept., Brugmann University Hospital, Brussels, Belgium
14 15	³ Dept. of Internal Medicine - Intensive Care Unit, CHR St. Joseph-Warquignies,
15	Boussu, Belgium
16	⁴ Intensive Care Dept., CHIREC, Brussels Belgium
17 18	 ⁵ Intensive Care Dept., CHR du Tournaisis, Tournai, Belgium ⁶ Intensive Care Dept., Ziekenhuisnetwerk Antwerpen, Antwerp, Belgium
19	^a Intensive Care Dept., Ziekennuisietwerk Antwerpen, Antwerp, Deigrunt
20	Word count: 3146
21	
22	Address of correspondence:
23	Prof. Dr. S. Blot
24	General Internal Medicine & Infectious Diseases
25	Ghent University Hospital
26	De Pintelaan 185
27	9000 Ghent, Belgium
28	T: +32 9 332 62 16
29	stijn.blot@UGent.be
30 31	
32	

33 Abstract

This prospective, observational multicenter (n=24) study investigated relationships 34 between antimicrobial choices and rates of empiric appropriate or adequate therapy, 35 and subsequent adaptation of therapy in 171 ICU patients with severe nosocomial 36 infections. Appropriate antibiotic therapy was defined as in-vitro susceptibility of the 37 causative pathogen and clinical response to the agent administered. In non-38 microbiologically documented infections, therapy was considered adequate in case of 39 favorable clinical response <5 days. Patients had pneumonia (n=127; 66 ventilator-40 associated), intra-abdominal infection (n=23), and bloodstream infection (n=21). 41 Predominant pathogens were Pseudomonas aeruginosa (n=29) Escherichia coli (n=26), 42 Staphylococcus aureus (n=22), and Enterobacter aerogenes (n=21). In 49.6% of infections 43 multidrug resistant (MDR) bacteria were involved, mostly extended-spectrum Beta-44 lactamase producing Enterobacteriaceae and MDR non-fermenting Gram-negative 45 bacteria. Prior antibiotic exposure and hospitalization in a general ward prior to ICU 46 admission were risk factors for MDR. Empiric therapy was appropriate/adequate in 47 63.7% of cases. Empiric schemes were classified according to coverage of (i) ESBL-48 Enterobacteriaceae + non-fermenting Gram-negative bacteria (meropenem-based), (ii) 49 non-fermenting Gram-negative bacteria (schemes with antipseudomonal agent), and 50 (iii) first-line agents not covering ESBL-Enterobacteriaceae nor non-fermenting Gram-51 negative bacteria. Meropenem-based schemes allowed for significantly higher rates 52 of appropriate/adequate therapy (p<0.001). This benefit remained when only 53 patients without risk factors for MDR were considered (p=0.021). In 106 patients 54 (61%) empiric therapy was adapted: in 60 cases following initial 55

- 56 inappropriate/inadequate therapy, in 46 patients in order to fine-tune empiric
- 57 therapy. In this study reflecting real-life practice first-line use of meropenem
- 58 provided significantly higher rates of appropriate/adequate therapy, irrespective of
- 59 presence of risk factors for MDR.
- 60
- 61 **Keywords:** antibiotic resistance multidrug resistance intensive care infection –

62 appropriate therapy

63 Introduction

To avoid nosocomial infection remains a daily challenge for healthcare workers 64 involved in the care of critically ill [1]. The prevalence of infection strongly depends 65 on the presence of particular iatrogenic risk factors such as the use of indwelling 66 devices and extensive surgery, as well as the severity of underlying disease and 67 critical illness. Despite several large scaled efforts to improve prevention of 68 healthcare-associated infection [2-7], in general about 20% to 50% of patients 69 70 hospitalized in intensive care units (ICUs) experience infection, either hospital or ICU-acquired [8, 9]. 71

72 Severe nosocomial infection carries a substantial economic burden due to extensive antimicrobial consumption and, even more importantly, increased length of 73 hospitalization [10-13]. In addition, severe infection seriously compromises the odds 74 of survival. Attributable mortality rates vary from as low as zero percent to a 75 dramatic 50% dependent of the type of infection, the causative pathogen, patient age, 76 associated co-morbidities, and overall quality of the anti-infective approach [13-21]. 77 Among all the modalities that are to be fulfilled in the aim to optimize clinical patient 78 outcomes, early initiation of the proper antimicrobial agent is a cornerstone [1]. 79 Failure to timely administer appropriate antimicrobial therapy results in a dramatic 80 increase in fatality rates [22-25]. Selecting an empiric regimen which covers the 81 causative pathogen, however, is hampered by the presence of multidrug resistant 82 83 (MDR) micro-organisms which is the utmost important cause of empiric inappropriate therapy. Since antimicrobial consumption in itself is a major trigger for 84

MDR development, physicians are frequently urged to thoughtfully use 'last-line
antibiotics'. In fact, the challenge is to achieve high rates of empiric appropriate
therapy in order to optimize patient survival, while avoiding unnecessary use of
antibiotics with the intention to minimize microbial selection pressure, and hence,
MDR development [1, 26].

In order to accomplish this goal, in the past decade two main strategies have been 90 91 proposed. In the surveillance-assisted strategy, the empiric regimen is selected mainly based on the presence or absence of MDR pathogens in routine surveillance 92 cultures. The strength of this approach is the high negative predictive value of 93 surveillance cultures to predict MDR involvement in subsequent infection. On the 94 condition that cultures are taken at least twice weekly, this strategy is capable to 95 combine high rates of empiric appropriate therapy with reduced consumption of 96 antibiotics in pneumonia and bacteremia [27-33]. The cost-effectiveness of this 97 98 approach, however, remains questionable. Another approach is the so-called 'deescalation strategy' which, on the condition of risk factors for MDR involvement, 99 recommends empiric start with a regimen which covers most potential MDR 100 pathogens such as extended-spectrum beta-lactamase producing (ESBL) 101 Enterobacteriaceae, non-fermenting Gram-negative bacteria, and methicillin-resistant 102 Staphylococcus aureus (MRSA) [34-36]. Once culture results are available, and if 103 104 feasible, narrowing the antimicrobial spectrum is advised. This concept, often referred to as 'de-escalation', is widely used and advocated [37-40]. This strategy has 105 106 been demonstrated to be successful and safe, and may also reduce antibiotic use and hence limiting the emergence of MDR [36, 41]. 107

The objective of the present study was to prospectively investigate patterns of antimicrobial therapy in critically ill patients with nosocomial infection. More precisely we formulated the following research questions: (i) What is the rate of empiric appropriate or adequate therapy achieved? (ii) Which antibiotic selections allow for the highest rates of empiric appropriate or adequate therapy, either in the presence or absence of risk factors for MDR pathogens? (iii) What is the rate by which empiric therapy (either appropriate/adequate or not) is adapted?

115

116

117 Methods

118 *Study design*. A prospective multicenter observational study was performed between

119 February 2006 and June 2007. In total 24 Belgian ICUs participated. In all centers the

- 120 study was approved by the local ethics committee and informed consent was
- 121 requested. In all centers antimicrobial prescribing was done or supervised by the
- 122 attending senior intensivist.

Inclusion criteria. Eligible patients were those who provided informed consent, were 123 at least 18 year of age, were hospitalized in the ICU, and experienced severe hospital-124 acquired infection, either pneumonia, intra-abdominal infection, primary 125 bloodstream infection, or secondary bloodstream infection originating from another 126 source than pneumonia or intra-abdominal infection (e.g. bacteremia secondary to 127 urinary tract infection or sinusitis). 128 Definitions. Infections were considered hospital- or ICU-acquired when diagnosed at 129 least 48 hours after respectively hospital or ICU admission. Severe bacterial 130 infections were defined following the International Sepsis Forum Consensus 131 Conference on Definitions of Infection in the ICU [42]. Definitions of invasive fungal 132 infections are described elsewhere [43, 44]. Pneumonia was considered ventilator-133 associated when occurring after at least 48 hours of mechanical ventilation. For the 134

135 purpose of the study, only the first episode per patient was considered. Pneumonia

- 136 or intra-abdominal infection with bacteremic breakthrough (i.e. positive blood
- 137 cultures) were classified according to the primary infection. Bloodstream infections

138 secondary to other sources than pneumonia or intra-abdominal infection were

139 classified as secondary bloodstream infection.

140 Definitions of MDR are described elsewhere [10, 32, 45]. *Candida* spp. were

141 considered MDR when resistant to fluconazole.

Following the epidemiological scope of the study aiming at insights into empiric 142 therapy in general, we also wanted to include microbiologically undocumented 143 144 infections with obvious signs of clinical sepsis. For this purpose we made a difference between appropriate and adequate empiric therapy. The term appropriate therapy 145 was valid for microbiologically documented infectious episodes and was defined as 146 *in vitro* susceptibility of the causative pathogen and clinical response to the agent 147 administered. The term adequate therapy was used in non-microbiologically 148 documented infections and was defined as favorable clinical response within five 149 days of therapy (resolution of signs of sepsis). 150

Severity of disease was assessed by means of the acute physiology and chronic health 151 evaluation (APACHE) II score [46]. The following co-morbid conditions were 152 registered: respiratory disease (chronic restrictive, obstructive or pulmonary vascular 153 disease resulting in severe exercise restriction), cardiac disease (New York Heart 154 Association Class IV), diabetes mellitus, hepatic disease (cirrhosis and portal 155 hypertension; episodes of past upper gastro-intestinal bleeding attributed to portal 156 hypertension or prior episodes of hepatic failure or encephalopathy), renal disease 157 158 (chronic glomerulonephritis, nephropathy, or chronic kidney disease), neurologic

159	disease (impairment of alertness or confusion) malignancy (hematologic cancer or
160	solid tumor), and neutropenia (absolute neutrophil count <1500 cells/mm³).
161	To assess relationships between empiric antibiotic selection and rates of appropriate
162	or adequate therapy, the empiric antimicrobial schemes were grouped according to
163	the spectrum of pathogens covered. Table 1 describes the classification of empiric
164	antimicrobial schemes. Because of the small numbers of empiric schemes covering
165	MRSA and the relative low occurrence rate of MRSA in the cohort, empiric antibiotic
166	schemes were classified in three major groups: (i) coverage of ESBL-Enterobacteriaceae
167	+ non-fermenting Gram-negative bacteria (meropenem [MER]-based schemes), (ii)
168	coverage of non-fermenting Gram-negative bacteria (schemes including an
169	antipseumononal agent), and (iii) schemes without coverage of either ESBL-
170	Enterobacteriaceae nor non-fermenting Gram-negative bacteria (first line agents). In
171	this analysis the added value of vancomycin as empiric therapy in order to cover
172	methicillin resistant Gram-positive pathogens was investigated separately.
173	Prior antibiotic exposure was defined as the administration of antimicrobial agents
174	within one month preceding the current infectious episode. Prior hospitalization was
175	defined as an hospital admission within the four months preceding the current ICU
176	admission.
177	Data analyses. Continuous variables are described as median (interquartile range
178	[IQR]) and discrete variables as n (%). For comparisons between groups the Mann
179	Whitney U test and Fisher Exact test or Chi-square test were used as appropriate.
180	Independent relationships with empiric appropriate or adequate therapy were

- assessed by means of a logistic regression analysis. Variables taken into account in
- the logistic regression analysis either showed a moderate relationship in univariate
- 183 analysis or a logic relationship with the dependent variable. Variables considered
- 184 were: age, APACHE II score, underlying diseases, hospitalization in another ward
- 185 prior to ICU admission, prior antibiotic exposure, and empiric antibiotic schemes.
- 186 Results of the regression analysis are reported as odds ratios (OR) and 95%

- 187 confidence intervals (CI).
- 188

190 Results

Demographics. During the study period 198 patients were included. Due to 191 incomplete patient files 27 patients were excluded, resulting in a final database 192 containing 171 patients. Patient characteristics are described in Table 2. Primary 193 infections were pneumonia (n=127, of which 66 ventilator-associated), intra-194 195 abdominal infection (n=23) bloodstream infection (n=21, of which 9 primary, 1 catheter-related, and 12 secondary to another source than pneumonia or intra-196 197 abdominal infection). One hundred and fifteen infections were ICU-acquired (66,7%). 198 About 75% of infections were microbiologically documented (Figure 1). This 199 represented 129 microbiologically documented infections in which 165 micro-200 organisms were isolated. Gram-negative bacteria were most common (n=122, 73.9%) with Pseudomonas aeruginosa (n=29), Escherichia coli (n=26), Enterobacter aerogenes 201 (n=21), *Klebsiella pneumonia* (n=7), and *Klebsiella oxytocca* (n=7) as predominant 202 203 pathogens. Among the 42 Gram-positive bacteria, *Staphylococcus aureus* (n=22), Streptococcus pneumoniae (n=11), and Enterococci (n=5) were most frequently isolated. 204 One fungal pathogen was isolated. In 64 of the 129 microbiologically documented 205 infections MDR pathogens were involved (49.6%). Figure 1 describes the breakdown 206 207 of the different types of resistance involved in these infections. Risk factors for multidrug resistance (MDR). Risk factors for MDR were assessed in 208 order to evaluate empiric antimicrobial regimens relative to their presence. No 209 210 specific underlying conditions appeared to predispose for MDR involvement. MDR

211 involvement was more frequent in patients with compared to without prior

antibiotic exposure (74.3% vs. 41.3%, p=0.001; relative risk 4.26, 95% CI: 1.80-10.09).
Hospitalization in another ward prior to ICU admission was also recognized as
significantly associated with MDR involvement (59.0% vs. 40.9%, p=0.041; relative
risk 2.08, 95% CI: 1.03-4.22). Figure 3 shows a breakdown of microbiologically
documented infections according to the presence of these two risk factors for MDR,
and subsequent MDR involvement.

218

Length of ICU stay prior to the development of infection, however, was negatively
associated with the risk of MDR involvement. Patients who experienced infections
without MDR pathogens developed this infection after a median of 3 days (IQR: 2-7
days), while infections caused by MDR pathogens occurred after a median of 2 days
(IQR: 0-4.25 days), (p=0.002).

Rates of empiric appropriate or adequate therapy and subsequent adaptation. The presence 224 of MDR pathogens in infections was associated with a significant lower rate of 225 empiric appropriate therapy: 87.7% vs. 35.9% (p<0.001). Rates of empiric appropriate 226 and adequate therapy are illustrated in Figure 2. In 80 of the 129 microbiologically 227 documented infections empiric therapy was appropriate (62.0%). In non-228 microbiologically documented infections, empiric therapy was judged adequate in 29 229 of 42 infections (69.0%). In 106 patients (62.0%) empiric therapy was adapted: in 60 230 cases following initial inappropriate or inadequate therapy, and in 46 patients in 231 order to fine tune empiric therapy. 232

Empiric antimicrobial selection and rates of appropriate or adequate therapy. In 12 infections 233 the causative pathogen was methicillin-resistant (10 MRSA, 2 methicillin-resistant 234 *Staphylococcus epidermidis*). Vancomycin was 14 times administered as an empiric 235 agent, but in only three cases in which a methicillin-resistant pathogen proved to be 236 involved. As such the rate of inappropriate therapy in case of methicillin-resistance 237 involvement was 75%. Due to the relative low prevalence of methicillin-resistance as 238 well as the low added value of initiating a glycopeptide, the association (or not) of 239 such an agent did not alter the study results in this particular cohort. Therefore, we 240 analyzed rates of empiric appropriate or adequate therapy irrespective of the 241 coverage of MRSA. This is illustrated by Figure 4a. Coverage of ESBL-242 Enterobacteriaceae (by MER-based empiric schemes) allowed for significantly higher 243 rates of appropriate or adequate therapy. Because the advantage of an antibiotic 244 agent which covers ESBL-*Enterobacteriaceae* is obvious in settings with a high 245 prevalence of MDR, we hypothesized that the benefit of using a MER-based empiric 246 scheme would fade away when only patients without risk factors for MDR were 247 taken into account. However, in this analysis empiric schemes covering ESBL-248 Enterobacteriaceae remained superior (Figure 4b). 249

When MER-based empiric schemes were compared to other empiric regimens, a difference of approximately 30% in empiric appropriate or adequate therapy was observed when all patients were taken into account (89.2% vs. 56.0%; p<0.001), and this distinction remained when only patients without risk factors for MDR were considered (83.6% vs. 53.8%; p=0.040).

- 255 In a multivariable logistic regression analysis, the only factors independently
- associated with empiric appropriate or adequate therapy were a MER-based empiric
- scheme (OR 18.1, 95% CI 4.8 to 68.1; p<0.001) and the presence of MDR pathogens
- 258 (OR 0.04, 95% CI 0.01 to 0.10; p<0.001). No other variables reached the level of

259 significance.

In this prospective, observational study, which aims to reflect real life daily practice,

261 Discussion

262

rates of empiric appropriate or adequate therapy were only 63.7%. About half of the 263 infections were caused by MDR micro-organisms, mostly ESBL-Enterobacteriaceae and 264 *P. aeruginosa*. As such, only the inclusion of meropenem in the empiric scheme, 265 allowed for acceptable rates of appropriate therapy (approximately 90%). 266 Several reasons for the low rate of appropriate or adequate therapy can be proposed. 267 The overall prevalence of MDR in this particular cohort is high (~50%) and the rate of 268 appropriate or adequate therapy in infections caused by MDR pathogens was very 269 poor (35.9%). MDR is well recognized as a major determinant of inappropriate 270 therapy [47-52]. However, to a certain extent the involvement of MDR can be 271 predicted on basis of some typical risk factors of which the most essential are prior 272 antibiotic exposure and a length of ICU stay (or mechanical ventilation in case of 273 pneumonia) of >7 days [32, 53]. Other risk factors such as particular underlying 274 diseases (e.g. chronic obstructive pulmonary disease) and recent 275 surgery/hospitalization can also be taken into account but generally are not included 276 in the principal risk factors. Strangely enough, in this cohort of ICU patients with 277 nosocomial infection some of the classic patterns failed to predict the involvement of 278 MDR. For instance, prior hospitalization (<4 months of the current hospital 279 admission) was not associated with a higher likelihood of MDR, while length of ICU 280 281 stay was even inversely related to MDR involvement. On the other hand, prior

antibiotic exposure and hospitalization in a general ward prior to ICU admission

283 appeared to have a significant relationship with MDR involvement. Importantly, however, although these relationships were statistically significant, MDR pathogens 284 were isolated in about 40% of patients in the absence of these risk factors. As a 285 consequence, the predictive value of these risk factors was low. As such, the 286 generally accepted concepts of risk perception for MDR involvement failed in this 287 particular cohort. We have no plausible explanation for this observation. Given the 288 relationship between prior hospitalization at a general ward and MDR, the relative 289 high number of infections already present at time of ICU admission, and the short 290 length of stay in the ICU prior to infection, it appears that there exists a serious 291 problem of MDR in general wards in Belgian hospitals, thereby contributing to the 292 low rate of empiric appropriate therapy in ICUs. Indeed, already a decade ago a 293 higher incidence of Enterobacter aerogenes with an increasing resistance pattern has 294 been noticed in Belgian hospitals [54, 55]. 295

Following the high rate of ESBL-Enterobacteriaceae and P. aeruginosa, an MER-based 296 empiric scheme allowed for the highest rates of appropriate/adequate therapy. Due 297 298 to the failure of classic risk factors to predict MDR involvement performance rates of empiric schemes did not substantially alter when only patients without risk factors 299 were considered (Figure 3). Hence, based on the present study an approach 300 including coverage of both ESBL-Enterobacteriaceae and P. aeruginosa seems highly 301 302 warranted. Without firm supportive knowledge of local ecology in ICU and non-ICU settings indicating low MDR levels, an empiric strategy which does not cover ESBL-303 Enterobacteriaceae and P. aeruginosa is potentially dangerous, and as such supports the 304 de-escalation strategy. The present findings however, with the failure of MDR 305

prediction based on general characteristics in particular, provide perspectives for the
surveillance assisted approach in which individual colonization status is a major
element in steering empiric therapy.

309

310	In 62% of patients empiric therapy was adapted (n=106). In 60 cases adaptation
311	followed initial inappropriate or inadequate therapy, whereas in 46 patients empiric
312	therapy could be fine-tuned (true de-escalation). Noteworthy, when only
313	microbiologically documented infections are considered, 4 out of 129 patients
314	received inappropriate therapy even after the availability of culture results (Figure 1).
315	In MER-based empiric schemes (n=37) therapy was continued in 23 and adapted in
316	14 patients (in 4 cases because of initial inappropriate therapy and in 10 cases in
317	order to narrow down the spectrum). <mark>As this study supports the approach of empiric</mark>
318	coverage of ESBL-Enterobacteriaceae and P. aeruginosa, we stress the importance of de-
319	escalation whenever possible. Baran et al. found that previous exposition to
320	carbapenems was 44% in patients with infections caused by imipenem-resistant
321	Acinetobacter baumannii, whereas only 12% in patients infected by imipenem-
322	susceptible A. baumannii isolates [56]. Yet, in multivariate analysis, previous
323	antibiotic exposure and not carbapenem exposure in particular appeared to be an
	and block exposure and not careapenent exposure in particular appeared to be an
324	independent risk factor for imipenem-resistance. The link between exposure and
324 325	
	independent risk factor for imipenem-resistance. The link between exposure and

328	Surveillance of Antimicrobial Use and Antimicrobial Resistance in German ICUs
329	(SARI project), Meyer E et al. identified carbapenem use as an independent predictor
330	for a higher <i>Stenotrophomonas maltophilia</i> incidence [57]. Although this data do not
331	disclose a causal relationship, it is highly suggestive for the specific selection
332	pressure of this pathogen. Therefore, excessive use of carbapenems is to be avoided
333	by a strict de-escalation strategy and by avoidance of unnecessary long therapies.
334	
335	The most important advantage of this study is its non-interventional design,
336	reflecting real daily practice, thereby disclosing the true pain points in empiric
337	antibiotic therapy in critically ill patients. Due to the multicenter approach the data
338	might be valid for many tertiary care centers in Belgium and abroad.
339	This study has limitations. First, no outcome data are available as the primary aim of
340	the study was to describe antibiotic prescription patterns and how they perform in
341	terms of appropriate therapy. Yet, the relationship between initial empiric failure to
342	cover the causative pathogen and adverse outcomes has been demonstrated
343	repeatedly and is generally accepted as an important quality indicator [58, 59].
344	Secondly, the cohort does not represent a consecutive series of ICU patients with
345	nosocomial infections. Probably this might have lead to the selection bias towards
346	more severe infections, with a higher likelihood of MDR involvement. Yet, for what
347	concerns risk factors for MDR, including less severe infections would probably not
348	have changed the observations regarding the relationship between MDR infection
349	and risk factors.

350	In conclusion, in this prospective study reflecting real life practice in ICU patients
351	with nosocomial infections, the rate of appropriate or adequate empiric therapy was
352	63.7%. This study demonstrated that classic risk factors for MDR such as prior
353	antibiotic exposure and length of ICU stay, may be insufficient to predict MDR
354	involvement. As such, empiric first-line use of MER allowed for significantly higher
355	rates of appropriate or adequate therapy, irrespective of presence of these risk
356	factors, and may be recommended in settings with a high prevalence of MDR
357	pathogens. In addition, these data illustrate the necessity for strict infection
358	prevention and control.
359 360	
361	Declarations
362	Funding: This study was sponsored by an unrestricted grant by AstraZeneca,
363	Belgium. Dr. Blot is supported by a grant from the European Society of Intensive
364	Care Medicine and iMDsoft Patient Safety Research Award 2008.
365	Competing Interests: No conflicts of interest to declare
366	Ethical Approval: For this non-interventional, observational study, ethical approval
367	was given by the local ethics committees of the participating centres (n=24).
368	
369 370	

***The ANTHICUS Study Investigators are**: O. Abid (RHMS, Ath, Belgium), S. Cran

- 372 (CHIREC, Brussels, Belgium), D. De Bels (CHU Brugmann, Brussels, Belgium), F. De Leener
- 373 (CHR St. joseph-Warquignies, Boussu, Belgium), L. Finianos (Clinique St. Joseph, Liège,
- 374 Belgium), F. Forêt (CHR St. Joseph-Warquignies, Mons, Belgium), M. Genard (CIU Ambroise
- 375 Paré, Mons, Belgium), E. Gilbert (CHR du Tournaisis, Tournai, Belgium), E. Khodadadi (IRIS
- 376 Sud-Bracops, Brussels, Belgium), S. Machayekhi (CH Hornu Frameries, Hornu), D. Mircev
- 377 (IRIS Sud-Etterbeek-Ixelles, Brussels, Belgium), D. Neuberg (CH de L'Ardenne, Libramont,
- Belgium), J.-Y. Piette (CH Bois de l'Abbaye, Seraign, Belgium), P. Serpe (CH ND Bruyères,
- 379 Chenee, Belgium), F. Beernaert (H. Hart Ziekenhuis, Eeklo, Belgium), K. De Decker
- 380 (Antwerp University Hospital, Antwerp, Belgium), K. De Ridder (AZ Maria Middelares,
- 381 Sint-Niklaas, Belgium), C. Declercq (St. Josef Ziekenhuis, Izegem, Belgium), I. Demeyer (OLV
- 382 Aalst, Aalst, Belgium), B. Nonneman (ASZ, Aalst, Belgium), K. Schoonheydt (ZNA,

- 383 Antwerp, Belgium, W. Swinnen (AZ St. Blasius, Dendermonde, Belgium), L. Wosteyn (ASZ,
- 384 Roeselaere, Belgium).
- 385

386 **References**

Blot S. Limiting the attributable mortality of nosocomial infection and multidrug resistance in
 intensive care units. Clin Microbiol Infect 2008;14:5-13.

Lorente L, Blot S, Rello J. Evidence on measures for the prevention of ventilator-associated
 pneumonia. Eur Respir J 2007;30:1193-207.

391 3. Masterton RG, Mifsud AJ, Rao GG. Review of hospital isolation and infection control
 392 precautions. J Hosp Infect 2003;54:171-3.

Labeau SO, Vandijck DM, Rello J, Adam S, Rosa A, Wenisch C, et al. Centers for Disease
 Control and Prevention guidelines for preventing central venous catheter-related infection: results of
 a knowledge test among 3405 European intensive care nurses. Crit Care Med 2009;37:320-3.

Labeau S, Vandijck D, Rello J, Adam S, Rosa A, Wenisch C, et al. Evidence-based guidelines for
the prevention of ventilator-associated pneumonia: results of a knowledge test among European
intensive care nurses. J Hosp Infect 2008;70:180-5.

399 6. Yokoe DS, Classen D. Improving patient safety through infection control: a new healthcare
400 imperative. Infect Control Hosp Epidemiol 2008;29 Suppl 1:S3-11.

401 7. Berwick DM, Calkins DR, McCannon CJ, Hackbarth AD. The 100,000 lives campaign: setting a
402 goal and a deadline for improving health care quality. Jama 2006;295:324-7.

403 8. van der Kooi TI, de Boer AS, Mannien J, Wille JC, Beaumont MT, Mooi BW, et al. Incidence
404 and risk factors of device-associated infections and associated mortality at the intensive care in the
405 Dutch surveillance system. Intensive Care Med 2007;33:271-8.

Vincent JL, Bihari DJ, Suter PM, Bruining HA, White J, Nicolas-Chanoin MH, et al. The
prevalence of nosocomial infection in intensive care units in Europe. Results of the European
Prevalence of Infection in Intensive Care (EPIC) Study. EPIC International Advisory Committee. Jama
1995;274:639-44.

Blot SI, Depuydt P, Annemans L, Benoit D, Hoste E, De Waele JJ, et al. Clinical and economic
outcomes in critically ill patients with nosocomial catheter-related bloodstream infections. Clin Infect
Dis 2005;41:1591-8.

413 11. Vander Stichele RH, Elseviers MM, Ferech M, Blot S, Goossens H. Hospital consumption of
414 antibiotics in 15 European countries: results of the ESAC Retrospective Data Collection (1997-2002). J
415 Antimicrob Chemother 2006;58:159-67.

416 12. Vandijck DM, Depaemelaere M, Labeau SO, Depuydt PO, Annemans L, Buyle FM, et al. Daily
417 cost of antimicrobial therapy in patients with Intensive Care Unit-acquired, laboratory-confirmed
418 bloodstream infection. Int J Antimicrob Agents 2008;31:161-5.

419 13. Safdar N, Dezfulian C, Collard HR, Saint S. Clinical and economic consequences of ventilator420 associated pneumonia: a systematic review. Crit Care Med 2005;33:2184-93.

421 14. Blot S, Vandewoude K, Hoste E, Colardyn F. Reappraisal of attributable mortality in critically
422 ill patients with nosocomial bacteraemia involving Pseudomonas aeruginosa. J Hosp Infect

423 2003;53:18-24.

Blot S, Vandewoude K, Hoste E, J DW, Kint K, Rosiers F, et al. Absence of excess mortality in
critically ill patients with nosocomial Escherichia coli bacteremia. Infect Control Hosp Epidemiol
2003;24:912-5.

427 16. Garrouste-Orgeas M, Timsit JF, Tafflet M, Misset B, Zahar JR, Soufir L, et al. Excess risk of
428 death from intensive care unit-acquired nosocomial bloodstream infections: a reappraisal. Clin Infect
429 Dis 2006;42:1118-26.

430 17. Blot SI, Vandewoude KH, Colardyn FA. Clinical impact of nosocomial Klebsiella bacteremia in
431 critically ill patients. Eur J Clin Microbiol Infect Dis 2002;21:471-3.

43218.Blot SI, Vandewoude KH, Colardyn FA. Evaluation of outcome in critically ill patients with

433 nosocomial enterobacter bacteremia: results of a matched cohort study. Chest 2003;123:1208-13.

Blot S, Cankurtaran M, Petrovic M, Vandijck D, Lizy C, Decruyenaere J, et al. Epidemiology and
outcome of nosocomial bloodstream infection in elderly critically ill patients: a comparison between
middle-aged, old, and very old patients. Crit Care Med 2009;37:1634-41.

437 20. Agbaht K, Diaz E, Munoz E, Lisboa T, Gomez F, Depuydt PO, et al. Bacteremia in patients with
438 ventilator-associated pneumonia is associated with increased mortality: A study comparing

439 bacteremic vs. nonbacteremic ventilator-associated pneumonia. Crit Care Med 2007;35:2064-70.

440 21. Vandewoude KH, Blot SI, Benoit D, Colardyn F, Vogelaers D. Invasive aspergillosis in critically

441 ill patients: attributable mortality and excesses in length of ICU stay and ventilator dependence. J

442 Hosp Infect 2004;56:269-76.

443 22. Harbarth S, Garbino J, Pugin J, Romand JA, Lew D, Pittet D. Inappropriate initial antimicrobial
444 therapy and its effect on survival in a clinical trial of immunomodulating therapy for severe sepsis.
445 Am J Med 2003;115:529-35.

446 23. Ibrahim EH, Sherman G, Ward S, Fraser VJ, Kollef MH. The influence of inadequate

antimicrobial treatment of bloodstream infections on patient outcomes in the ICU setting. Chest2000;118:146-55.

Kollef MH, Sherman G, Ward S, Fraser VJ. Inadequate antimicrobial treatment of infections: a
risk factor for hospital mortality among critically ill patients. Chest 1999;115:462-74.

451 25. Kumar A, Roberts D, Wood KE, Light B, Parrillo JE, Sharma S, et al. Duration of hypotension
452 before initiation of effective antimicrobial therapy is the critical determinant of survival in human
453 septic shock. Crit Care Med 2006;34:1589-96.

454 26. Masterton R, Drusano G, Paterson DL, Park G. Appropriate antimicrobial treatment in
455 nosocomial infections-the clinical challenges. J Hosp Infect 2003;55 Suppl 1:1-12.

27. Depuydt P, Benoit D, Vogelaers D, Claeys G, Verschraegen G, Vandewoude K, et al. Outcome
in bacteremia associated with nosocomial pneumonia and the impact of pathogen prediction by

tracheal surveillance cultures. Intensive Care Med 2006;32:1773-81.

459 28. Jung B, Sebbane M, Chanques G, Courouble P, Verzilli D, Perrigault PF, et al. Previous
460 endotracheal aspirate allows guiding the initial treatment of ventilator-associated pneumonia.

461 Intensive Care Med 2009;35:101-7.

462 29. Depuydt P, Benoit D, Vogelaers D, Decruyenaere J, Vandijck D, Claeys G, et al. Systematic
463 surveillance cultures as a toolto predict involvement of multidrug antibiotic resistant bacteria in
464 ventilator-associated pneumonia. Intensive Care Med 2008;34:675-82.

30. Blot S, Depuydt P, Vogelaers D, Decruyenaere J, De Waele J, Hoste E, et al. Colonization
status and appropriate antibiotic therapy for nosocomial bacteremia caused by antibiotic-resistant
gram-negative bacteria in an intensive care unit. Infect Control Hosp Epidemiol 2005;26:575-9.

468 31. Papadomichelakis E, Kontopidou F, Antoniadou A, Poulakou G, Koratzanis E, Kopterides P, et
469 al. Screening for resistant gram-negative microorganisms to guide empiric therapy of subsequent

470 infection. Intensive Care Med 2008;34:2169-75.

471 32. Depuydt PO, Blot SI, Benoit DD, Claeys GW, Verschraegen GL, Vandewoude KH, et al.

472 Antimicrobial resistance in nosocomial bloodstream infection associated with pneumonia and the

value of systematic surveillance cultures in an adult intensive care unit. Crit Care Med 2006;34:653-9.

474 33. Michel F, Franceschini B, Berger P, Arnal JM, Gainnier M, Sainty JM, et al. Early antibiotic

475 treatment for BAL-confirmed ventilator-associated pneumonia: a role for routine endotracheal476 aspirate cultures. Chest 2005;127:589-97.

477 34. Sandiumenge A, Diaz E, Bodí M, Rello J. Therapy of ventilator-associated pneumonia. A
478 patients-based approach based on the ten rules of "The Tarragona Strategy". Intensive Care Med
479 2003;29:876-83.

480 35. Kollef MH. Hospital-acquired pneumonia and de-escalation of antimicrobial treatment. Crit481 Care Med 2001;29:1473-5.

482 36. Leone M, Garcin F, Bouvenot J, Boyadjev I, Visintini P, Albanese J, et al. Ventilator-associated
483 pneumonia: breaking the vicious circle of antibiotic overuse. Crit Care Med 2007;35:379-85; quizz 86.

CCEPTED MANUSCR

484 37. Kollef M. Appropriate empirical antibacterial therapy for nosocomial infections: getting it 485 right the first time. Drugs 2003;63:2157-68. 486 38. Kollef MH. Treatment of ventilator-associated pneumonia: get it right from the start. Crit 487 Care Med 2003;31:969-70. 488 39. Rello J, Vidaur L, Sandiumenge A, Rodriguez A, Gualis B, Boque C, et al. De-escalation therapy 489 in ventilator-associated pneumonia. Crit Care Med 2004;32:2183-90. 490 Leone M, Martin C. How to break the vicious circle of antibiotic resistances? Curr Opin Crit 40. 491 Care 2008;14:587-92. 492 Masterton RG. The new treatment paradigm and the role of carbapenems. Int J Antimicrob 41. 493 Agents 2009;33:105-10. 494 42. Calandra T, Cohen J. The international sepsis forum consensus conference on definitions of 495 infection in the intensive care unit. Crit Care Med 2005;33:1538-48. 496 43. Blot S, Vandewoude K. Management of invasive candidiasis in critically ill patients. Drugs 497 2004;64:2159-75. 498 44. Vandewoude KH, Blot SI, Depuydt P, Benoit D, Temmerman W, Colardyn F, et al. Clinical 499 relevance of Aspergillus isolation from respiratory tract samples in critically ill patients. Crit Care 500 2006;10:R31. 501 45. Blot S, Vandewoude K, De Bacquer D, Colardyn F. Nosocomial bacteremia caused by 502 antibiotic-resistant gram-negative bacteria in critically ill patients: clinical outcome and length of 503 hospitalization. Clin Infect Dis 2002;34:1600-6. 504 46. Knaus WA, Draper EA, Wagner DP, Zimmerman JE. APACHE II: a severity of disease 505 classification system. Crit Care Med 1985;13:818-29. 506 47. Giamarellos-Bourboulis EJ, Papadimitriou E, Galanakis N, Antonopoulou A, Tsaganos T, 507 Kanellakopoulou K, et al. Multidrug resistance to antimicrobials as a predominant factor influencing 508 patient survival. Int J Antimicrob Agents 2006;27:476-81. 509 Leone M, Bourgoin A, Cambon S, Dubuc M, Albanese J, Martin C. Empirical antimicrobial 48. 510 therapy of septic shock patients: adequacy and impact on the outcome. Crit Care Med 2003;31:462-511 7. 512 49. Ortega M, Marco F, Soriano A, Almela M, Martinez JA, Munoz A, et al. Analysis of 4758 513 Escherichia coli bacteraemia episodes: predictive factors for isolation of an antibiotic-resistant strain 514 and their impact on the outcome. J Antimicrob Chemother 2009;63:568-74. 515 50. Kollef KE, Schramm GE, Wills AR, Reichley RM, Micek ST, Kollef MH. Predictors of 30-day 516 mortality and hospital costs in patients with ventilator-associated pneumonia attributed to 517 potentially antibiotic-resistant gram-negative bacteria. Chest 2008;134:281-7.

- 518 51. Albrecht SJ, Fishman NO, Kitchen J, Nachamkin I, Bilker WB, Hoegg C, et al. Reemergence of 519 gram-negative health care-associated bloodstream infections. Arch Intern Med 2006;166:1289-94.
- 520 52. Lautenbach E, Polk RE. Resistant gram-negative bacilli: A neglected healthcare crisis? Am J

521 Health Syst Pharm 2007;64:S3-21; quiz S2-4.

- 522 53. Trouillet JL, Chastre J, Vuagnat A, Joly-Guillou ML, Combaux D, Dombret MC, et al. Ventilator523 associated pneumonia caused by potentially drug-resistant bacteria. Am J Respir Crit Care Med
 524 1998;157:531-9.
- 525 54. De Gheldre Y, Glupczynski Y, Struelens M, De Mol P. Emergence of Enterobacter aerogenes
 526 as a major antibiotic-resistant nosocomial pathogen in Belgian hospitals. Clin Microbiol Infect
 527 1999;5:622-7.

528 55. De Gheldre Y, Struelens MJ, Glupczynski Y, De Mol P, Maes N, Nonhoff C, et al. National

- epidemiologic surveys of Enterobacter aerogenes in Belgian hospitals from 1996 to 1998. J Clin
 Microbiol 2001;39:889-96.
- 531 56. Baran G, Erbay A, Bodur H, Onguru P, Akinci E, Balaban N, et al. Risk factors for nosocomial 532 imipenem-resistant Acinetobacter baumannii infections. Int J Infect Dis 2008;12:16-21.
- 533 57. Meyer E, Schwab F, Gastmeier P, Rueden H, Daschner FD, Jonas D. Stenotrophomonas
- maltophilia and antibiotic use in German intensive care units: data from Project SARI (Surveillance of
- 535 Antimicrobial Use and Antimicrobial Resistance in German Intensive Care Units). J Hosp Infect

536 2006;64:238-43.

537 58. Degoricija V, Sharma M, Legac A, Gradiser M, Sefer S, Vucicevic Z. Survival analysis of 314 538 episodes of sepsis in medical intensive care unit in university hospital: impact of intensive care unit 539 performance and antimicrobial therapy. Croat Med J 2006;47:385-97.

- 540 59. Dellinger RP, Carlet JM, Masur H, Gerlach H, Calandra T, Cohen J, et al. Surviving Sepsis
- 541 Campaign guidelines for management of severe sepsis and septic shock. Intensive Care Med

542 2004;30:536-55.

- 543
- 544
- 545

- 546 Figure Legends
- 547 Figure 1 Review of 171 nosocomial infections distributed for microbiological
- 548 documentation and multidrug resistance involvement.
- 549 MDR, multidrug resistance or multidrug resistant
- 550 ESBL, extended spectrum beta-lactamase producing Enterobacteriaceae
- 551 MRSA, methicillin- resistant *Staphylococcus aureus*
- 552 MRSE, methicillin-resistant *Staphylococcus epidermidis*

553

- Figure 2 Rates of empiric appropriate or adequate therapy in ICU patients with
 nosocomial infections.
- 556 Cont., continuation of empiric therapy
- 557 Adapt., adaptation of empiric therapy
- 558

559 Figure 3 – Breakdown of microbiologically documented infections according to the

- 560 presence of risk factors for multidrug resistance, and multidrug resistance
- 561 involvement.
- 562 MDR, multidrug resistance
- 563

Figure 4 - Rates of empiric appropriate or adequate therapy according to three
major groups of empiric antibiotic regimens (coverage of methicillin-resistant
pathogens not considered).

- 567 Figure 4a, all patients considered (n=170; in one patient no empiric therapy was initiated)
- 568 Figure 4b, only patients considered without risk factors for multidrug resistance involvement
- 569 (hospitalization at a general ward prior to ICU admission and prior antibiotic exposure),
- 570 (n=78).
- 571 Group 1: coverage of ESBL-*Enterobacteriaceae* + *P. aeruginosa* (meropenem-based schemes)
- 572 Group 2: coverage of *P. aeruginosa* (schemes containing an antipseudomonal agent)

573 Group 3: no coverage of ESBL nor non-fermenting Gram-negative bacteria (first line agents)

574

575

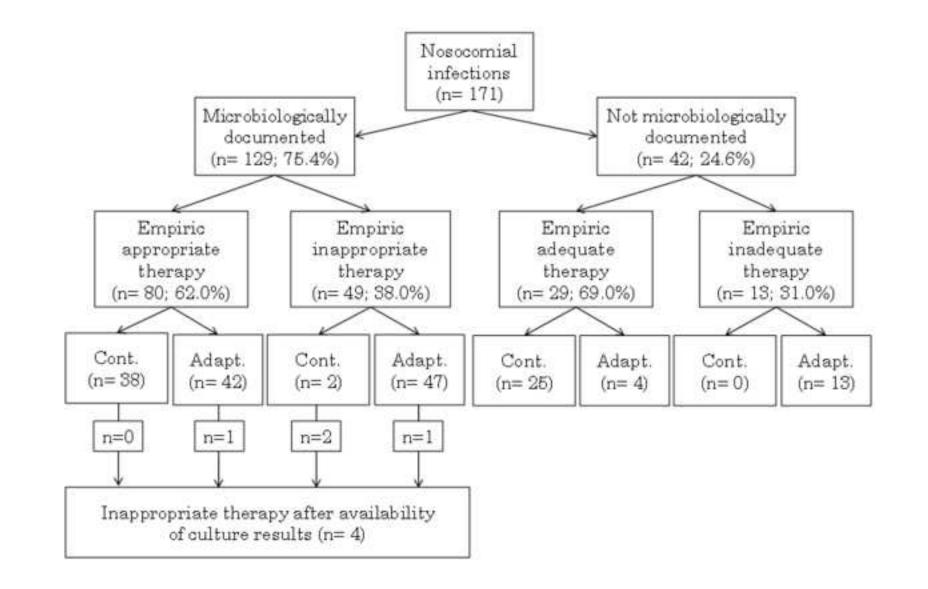
A cooled Manus

Table 1 – Empiric antibiotic regimens clustered in six major groups according to the pathogens covered.

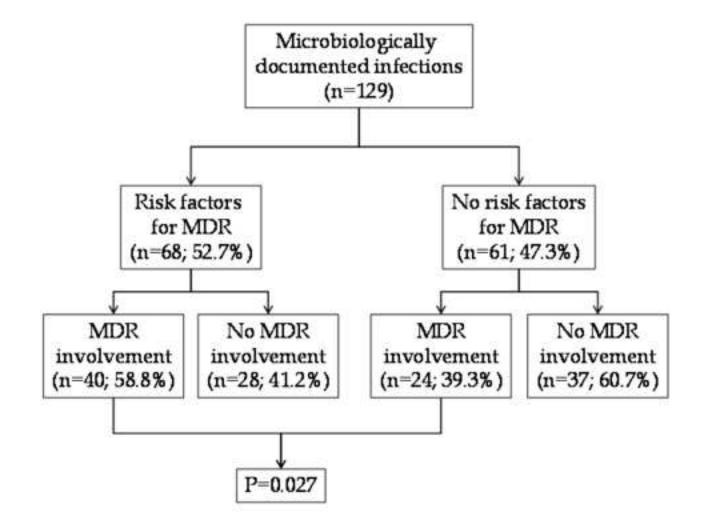
Coverage	Empiric regimens	n
ESBL- Enterobacteriaceae + P. aeruginosa + MRSA (n=6)	MER+VAN	5
	MER+VAN+AMI	1
		X
ESBL- Enterobacteriaceae + P. aeruginosa (n=31)	MER	18
	MER+AMI	9
	MER+OXA	1
	MER+CIP+AMI	1
	MER+PIPtaz+AMI	2
<i>P. aeruginosa</i> + MRSA (n=7)	PIPtaz+VAN+LEVO	1
	PIPtaz+VAN	1
	CAZ+VAN	1
A	CEFE+VAN	2
	CEFE+VAN+AMI	2
P. aeruginosa (n=74)	PIPtaz	12
	PIPtaz+AMI	12
	PIPtaz+FLU	2
	PIPtaz+AMOXIclav	1
N X	AMOXIclav+AMI	4
	CAZ	2
	CEFE	8
	CIP	2
6	LEVO	3
	CEFE+AMI	6
	CAZ+AMI	7
	CEFE+CIP	3
	CAZ+MOXI	2
	CEFE+FLU	1
	CIP+AMOXIclav	2
	LEVO+AMOXIclav	3
	LEVO+AMOXIclav+CLA	1
	LEVO+AMI	1
	CIP+OXA	1

	OXA+AZT	1
MRSA (n=1)	VAN	1
Non-MDR pathogens or <i>P. aeruginosa</i> (n=51)	AMOXIclav	32
	AMOXIclav+metronidazol	1
	CEF	7
	MOXI	1
	CTX	1
	CTX+metronidazol	3
	CEF+CLA	1
	temocillin	3
	OXA	2
No empiric therapy (n=1)	6	1

Legend: MER, meropenem AMI, aminoglycoside VAN, vancomycin OXA, oxacillin CIP, ciprofloxacin PIPtaz, piperacillin/tazobactam LEVO, levofloxacin FLU, fluconazole AMOXIclav, amoxicillin/clavulanate CEF, cefuroxim CAZ, ceftazidime CTX, ceftriaxone CEFE, cefepim MOXI, moxifloxacin CLA, clarithromycin







1	Patterns of Antimicrobial Therapy in Severe Nosocomial Infections:
2	Empiric Choices, Proportion of Appropriate Therapy, and Modification Rates –
3	A Multicenter, Observational Survey in Critically Ill Patients.
4	
5	Dirk Vogelaers ¹ , PhD, David De Bels ² , MD, Frédéric Forêt ³ , MD, Sophie Cran ⁴ , MD,
6	Eric Gilbert ⁵ , MD, Karen Schoonheydt ⁶ , MD, and Stijn Blot ¹ , PhD, for the
7	ANTHICUS Study Investigators*
8	
9	
10	Affiliations:
11	¹ General Internal Medicine & Infectious Diseases, Ghent University Hospital, Ghent,
12	Belgium
13	² Intensive Care Dept., Brugmann University Hospital, Brussels, Belgium
14	³ Dept. of Internal Medicine - Intensive Care Unit, CHR St. Joseph-Warquignies,
15	Boussu, Belgium
16	 ⁴ Intensive Care Dept., CHIREC, Brussels Belgium ⁵ Intensive Care Dept., CHR du Tournaisis, Tournai, Belgium
17 18	⁶ Intensive Care Dept., Ziekenhuisnetwerk Antwerpen, Antwerp, Belgium
19	intensive Care Dept., Ziekennuisietwerk Antwerpen, Antwerp, Deigrun
20	Word count: 3146
21	
22	Address of correspondence:
23	Prof. Dr. S. Blot
24	General Internal Medicine & Infectious Diseases
25	Ghent University Hospital
26	De Pintelaan 185
27	9000 Ghent, Belgium
28	T: +32 9 332 62 16
29 30 31 32	<u>stijn.blot@UGent.be</u>

33 Abstract

This prospective, observational multicenter (n=24) study investigated relationships 34 between antimicrobial choices and rates of empiric appropriate or adequate therapy, 35 and subsequent modification of therapy in 171 ICU patients with severe nosocomial 36 infections. Appropriate antibiotic therapy was defined as in-vitro susceptibility of the 37 causative pathogen and clinical response to the agent administered. In non-38 microbiologically documented infections, therapy was considered adequate in the 39 case of favorable clinical response <5 days. Patients had pneumonia (n=127; 66 40 ventilator-associated), intra-abdominal infection (n=23), and bloodstream infection 41 42 (n=21). Predominant pathogens were *Pseudomonas aeruginosa* (n=29) *Escherichia coli* (n=26), Staphylococcus aureus (n=22), and Enterobacter aerogenes (n=21). In 49.6% of 43 infections multidrug resistant (MDR) bacteria were involved, mostly extended-44 spectrum Beta-lactamase producing Enterobacteriaceae and MDR non-fermenting 45 Gram-negative bacteria. Prior antibiotic exposure and hospitalization in a general 46 ward prior to ICU admission were risk factors for MDR. Empiric therapy was 47 appropriate/adequate in 63.7% of cases. Empiric schemes were classified according 48 to coverage of (i) ESBL-producing Enterobacteriaceae and non-fermenting Gram-49 negative bacteria ("meropenem-based"), (ii) non-fermenting Gram-negative bacteria 50 (schemes with an antipseudomonal agent), and (iii) first-line agents not covering 51 ESBL-Enterobacteriaceae nor non-fermenting Gram-negative bacteria. Meropenem-52 based schemes allowed for significantly higher rates of appropriate/adequate 53 therapy (p<0.001). This benefit remained when only patients without risk factors for 54 MDR were considered (p=0.021). In 106 patients (61%) empiric therapy was 55

56	modified: in 60 cases following initial inappropriate/inadequate therapy, in 46
57	patients in order to refine empiric therapy. In this study which reflects real-life
58	practice, first-line use of meropenem provided significantly higher rates of the
59	appropriate/adequate therapy, irrespective of presence of risk factors for MDR.
60	

61 **Keywords:** antibiotic resistance – multidrug resistance – intensive care – infection –

62 appropriate therapy

63 Introduction

Avoiding nosocomial infection remains a daily challenge for healthcare workers
involved in the care of critically ill [1]. Despite several large scaled efforts to improve
prevention of healthcare-associated infection [2-7], in general about 20% to 50% of
patients hospitalized in intensive care units (ICUs) experience infection, either
hospital or ICU-acquired [8, 9].

Severe nosocomial infection carries a substantial economic burden due to 69 antimicrobial consumption and increased length of hospitalization [10-13]. In 70 addition, severe infection seriously compromises survival. Attributable mortality 71 72 varies from zero, to as much as 50% depending on a variety of factors [13-21]. Early initiation of the appropriate antimicrobial agents is a cornerstone in optimising 73 clinical patient outcomes, [1]. Failure of timely administration of appropriate 74 antimicrobial therapy results in a dramatic increase in mortality [22-25]. Selecting an 75 empiric regimen which covers causative pathogens, however, is hampered by the 76 presence of multidrug resistant (MDR) micro-organisms which is the utmost 77 important cause of empiric inappropriate therapy. Since antimicrobial consumption 78 itself is a major trigger for MDR development, physicians are frequently urged to use 79 'last-resort antibiotics' thoughtfully. The challenge is to achieve high rates of empiric 80 appropriate therapy, while avoiding unnecessary use of antibiotics, and hence, MDR 81 development [1, 26]. 82

In order to achieve this goal, two main strategies have been proposed. In the
surveillance-assisted strategy, the empiric regimen is selected mainly based on the

presence or absence of MDR pathogens in routine surveillance cultures. The strength 85 of this approach is the high negative predictive value of surveillance cultures to 86 predict MDR involvement in subsequent infection. If cultures are taken at least twice 87 weekly, this strategy is capable to combine high rates of empiric appropriate therapy 88 with reduced consumption of antibiotics in pneumonia and bacteremia [27-33]. The 89 cost-effectiveness of this approach, however, remains unclear. Another approach is 90 the 'de-escalation strategy' which, if there are risk factors for MDR, recommends an 91 empiric start with a regimen which covers most potential MDR pathogens such as 92 extended-spectrum beta-lactamase producing (ESBL) Enterobacteriaceae, non-93 fermenting Gram-negative bacteria, and meticillin-resistant Staphylococcus aureus 94 (MRSA) [34-36]. Once culture results are available, and if feasible, narrowing the 95 antimicrobial spectrum is advised. This concept, often referred to as 'de-escalation', is 96 widely used and advocated [37-40]. This strategy has been shown to be successful 97 and safe, and may also reduce antibiotic use and hence limit the emergence of MDR 98 [36, 41]. 99

The objective of the present study was prospectively to investigate patterns of 100 antimicrobial therapy in critically ill patients with nosocomial infection. More 101 precisely we formulated the following research questions: (i) What is the rate of 102 empiric appropriate or adequate therapy achieved? (ii) Which antibiotic selections 103 allow for the highest rates of empiric appropriate or adequate therapy, either in the 104 presence or absence of risk factors for MDR pathogens? (iii) What is the rate by 105 106 which empiric therapy (either appropriate/adequate or not) is subsequently modified? 107

108

109

S.

110 Methods

Study design. A prospective multicenter observational study was performed between February 2006 and June 2007. 24 Belgian ICUs participated. In all centres the study was approved by the local ethics committee and informed consent was requested. In all centers antimicrobial prescribing was by, or supervised by, the attending senior intensivist.

116	Inclusion criteria. Eligible patients were those who gave informed consent, were at
117	least 18 years of age, were hospitalized in the ICU, and experienced severe hospital-
118	acquired infection; either pneumonia, intra-abdominal infection, primary or
119	secondary bloodstream infection originating from a source other than pneumonia or
120	intra-abdominal infection (e.g. bacteremia secondary to urinary tract infection or
121	sinusitis).

Definitions. Infections were considered hospital- or ICU-acquired when diagnosed at 122 least 48 hours after hospital or ICU admission respectively. Severe bacterial 123 infections were defined according to the International Sepsis Forum Consensus 124 Conference on Definitions of Infection in the ICU [42]. Definitions of invasive fungal 125 infections are described elsewhere [43, 44]. Pneumonia was considered ventilator-126 127 associated when occurring after at least 48 hours of mechanical ventilation. For the purpose of the study, only the first episode per patient was considered. Pneumonia 128 or intra-abdominal infection with bacteraemic breakthrough (i.e. positive blood 129 130 cultures) were classified according to the primary infection. Bloodstream infections

131 secondary to other sources than pneumonia or intra-abdominal infection were

132 classified as secondary bloodstream infection.

133 Definitions of MDR are described elsewhere [10, 32, 45]. *Candida* spp. were

134 considered MDR when resistant to fluconazole.

Following the epidemiological scope of the study, aiming at insights into empiric 135 therapy in general, we also wanted to include microbiologically unconfirmed 136 137 infections with obvious signs of clinical sepsis. For this purpose we made a difference between appropriate and adequate empiric therapy. The term appropriate therapy 138 was used for microbiologically documented infectious episodes and was defined as 139 *in vitro* susceptibility of the causative pathogen with clinical response to the agent 140 administered. The term adequate therapy was used in non-microbiologically 141 confirmed infections, and was defined as favourable clinical response within five 142 days of treatment (resolution of signs of sepsis). 143

Severity of disease was assessed by means of the acute physiology and chronic health 144 evaluation (APACHE) II score [46]. The following co-morbid conditions were 145 recorded: respiratory disease (chronic restrictive, obstructive or pulmonary vascular 146 disease resulting in severe exercise restriction), cardiac disease (New York Heart 147 Association Class IV), diabetes mellitus, hepatic disease (cirrhosis and portal 148 hypertension; episodes of past upper gastro-intestinal bleeding attributed to portal 149 hypertension or prior episodes of hepatic failure or encephalopathy), renal disease 150 151 (chronic glomerulonephritis, nephropathy, or chronic kidney disease), neurologic

152	disease (impairment of alertness or confusion) malignancy (haematologic or solid
153	tumor), and neutropenia (absolute neutrophil count <1500 cells/mm ³).

To assess relationships between empiric antibiotic selection and rates of appropriate 154 and adequate therapy, the empiric antimicrobial schemes were grouped according to 155 the spectrum of pathogens covered. Table 1 gives the classification of empiric 156 antimicrobial schemes. Because of the small numbers of empiric schemes covering 157 158 MRSA and the relative low occurrence rate of MRSA in the cohort, empiric antibiotic schemes were classified in three major groups: (i) coverage of ESBL-producing 159 Enterobacteriaceae and non-fermenting Gram-negative bacteria ("meropenem 160 [MER]-based" schemes), (ii) coverage of non-fermenting Gram-negative bacteria 161 162 (schemes including an antipseumononal agent), and (iii) schemes without coverage of either of ESBL-producing Enterobacteriaceae nor non-fermenting Gram-negative 163 bacteria (first line agents). In this analysis the added value of vancomycin as empiric 164 165 therapy in order to cover meticillin resistant Gram-positive pathogens was investigated separately. 166

Prior antibiotic exposure was defined as the administration of antimicrobial agents
within one month preceding the current infectious episode. Prior hospitalization was
defined as an hospital admission within the four months preceding the current ICU
admission.

Data analyses. For comparisons between groups of continuous variables the Mann
Whitney U test and Fisher Exact test or Chi-square test were used as appropriate.
Independent relationships with empiric appropriate or adequate therapy were

- assessed by means of a logistic regression analysis. Variables taken into account in
- the logistic regression analysis either showed a moderate relationship in univariate
- analysis or a logic relationship with the dependent variable. Variables considered
- 177 were: age, APACHE II score, underlying diseases, hospitalization in another ward
- 178 prior to ICU admission, prior antibiotic exposure, and empiric antibiotic schemes.
- 179 Results of the regression analysis are reported as odds ratios (OR) and 95%

- 180 confidence intervals (CI).
- 181

183 **Results**

Demographics. During the study period 198 patients were included. Due to 184 incomplete patient files 27 patients were excluded, resulting in a final database 185 containing 171 patients. Patient characteristics are given in Table 2. Primary 186 infections were pneumonia (n=127, of which 66 ventilator-associated), intra-187 abdominal infection (n=23) bloodstream infection (n=21, of which 9 primary, 1 188 catheter-related, and 12 secondary to a source other than pneumonia or intra-189 190 abdominal infection). 115 infections were ICU-acquired (66,7%). Approximately 75% 191 of infections were microbiologically documented (Figure 1). This represented 129 192 microbiologically documented with 165 micro-organisms. Gram-negative bacteria 193 were most common (n=122, 73.9%) with Pseudomonas aeruginosa (n=29), Escherichia coli (n=26), Enterobacter aerogenes (n=21), Klebsiella pneumonia (n=7), and Klebsiella 194 oxytocca (n=7) as predominant pathogens. Among the 42 Gram-positive bacteria, 195 *Staphylococcus aureus* (n=22), *Streptococcus pneumoniae* (n=11), and enterococci (n=5) 196 were most frequently isolated. One fungal pathogen was isolated. In 64 of the 129 197 microbiologically documented infections MDR pathogens were involved (49.6%). 198 Risk factors for multidrug resistance (MDR). Risk factors for MDR were assessed in 199 order to evaluate empiric antimicrobial regimens relative to their presence. No 200 201 specific underlying conditions appeared to predispose for MDR involvement. MDR involvement was more frequent in patients with prior antibiotic exposure (74.3% vs. 202 203 41.3%, p=0.001; relative risk 4.26, 95% CI: 1.80-10.09). Hospitalization in another ward prior to ICU admission was also recognized as significantly associated with 204

MDR involvement (59.0% vs. 40.9%, p=0.041; relative risk 2.08, 95% CI: 1.03-4.22).
Figure 2 shows a breakdown of microbiologically documented infections according
to the presence of these two risk factors for MDR, and subsequent MDR involvement.

Length of ICU stay prior to the development of infection, however, was negatively
associated with the risk of MDR involvement. Patients who experienced infections
without MDR pathogens developed this infection after a median of 3 days (IQR: 2-7
days), while infections caused by MDR pathogens occurred after a median of 2 days
(IQR: 0-4.25 days), (p=0.002).

Rates of empiric appropriate or adequate therapy and subsequent modification. The presence 214 of MDR pathogens in infections was associated with a significant lower rate of 215 empiric appropriate therapy: 87.7% vs. 35.9% (p<0.001). Rates of empiric appropriate 216 and adequate therapy are illustrated in Figure 3. In 80 of the 129 microbiologically 217 documented infections, empiric therapy was appropriate (62.0%). In non-218 microbiologically documented infections, empiric therapy was judged adequate in 29 219 of 42 infections (69.0%). In 106 patients (62.0%) empiric therapy was modified: in 60 220 cases following initial inappropriate or inadequate therapy, and in 46 patients in 221 order to refine empiric therapy. 222

Empiric antimicrobial selection and rates of appropriate or adequate therapy. In 12 infections
the causative pathogen was meticillin-resistant (10 MRSA, 2 meticillin-resistant *Staphylococcus epidermidis*). Vancomycin administered empirically in 14 cases, but in
only was a meticillin-resistant pathogen involved. The rate of inappropriate therapy

227 in case of meticillin-resistance involvement was thus 75%. Due to the relative low prevalence of meticillin-resistance and the low added value of initiating a 228 glycopeptide, the association of such an agent did not alter the study results in this 229 particular cohort. Therefore, we analyzed rates of empiric appropriate or adequate 230 therapy irrespective of the coverage of MRSA (Figure 4a). Coverage of of ESBL-231 producing Enterobacteriaceae (by MER-based empiric schemes) allowed for 232 significantly higher rates of appropriate or adequate therapy. We hypothesized that 233 the benefit of using a MER-based empiric scheme would disappear when only 234 patients without risk factors for MDR were taken into account. However, in this 235 analysis empiric schemes covering of ESBL-producing Enterobacteriaceae remained 236 superior (Figure 4b). 237

When MER-based empiric schemes were compared with others, a difference of
approximately 30% in empiric appropriate or adequate therapy was observed when
all patients were included (89.2% vs. 56.0%; p<0.001), and this distinction remained
when only patients without risk factors for MDR were considered (83.6% vs. 53.8%;
p=0.040).

In a multivariable logistic regression analysis, the only factors independently
associated with empiric appropriate or adequate therapy were a MER-based empiric
scheme (OR 18.1, 95% CI 4.8 to 68.1; p<0.001) and the presence of MDR pathogens
(OR 0.04, 95% CI 0.01 to 0.10; p<0.001). No other variables reached statistical
significance.

248

249 Discussion

In this prospective, observational study, rates of empiric appropriate or adequate therapy were only 63.7%. About half of the infections were caused by MDR microorganisms, mostly ESBL-*Enterobacteriaceae* and *P. aeruginosa*. As such, only the inclusion of meropenem in the empiric scheme, allowed for acceptable rates of appropriate therapy (approximately 90%).

Several reasons for the low rate of appropriate or adequate therapy are possible. The 255 overall prevalence of MDR in this particular cohort is high (~50%) and the rate of 256 appropriate or adequate therapy in infections caused by MDR pathogens was low 257 (35.9%). MDR is recognized as a determinant of inappropriate therapy [47-52]. The 258 involvement of MDR can be partly predicted by some typical risk factors such as 259 prior antibiotic exposure and length of ICU stay (or mechanical ventilation in case of 260 pneumonia) of >7 days [32, 53]. Other risk factors such as underlying diseases and 261 recent surgery/hospitalization can also be taken into account. However, in our 262 cohort of ICU patients with nosocomial infection some of the recognized risk factors 263 failed to predict the involvement of MDR. Also, MDR pathogens were isolated in 264 about 40% of patients in the absence of these risk factors. As a consequence, the 265 predictive value of these risk factors was low. As such, the generally accepted 266 concepts of risk for MDR involvement were not supported by this particular cohort. 267 It appears that there exists a serious problem of MDR in general wards in Belgian 268 269 hospitals, thereby contributing to the low rate of appropriate empiric therapy.

270	Indeed, a decade ago a higher incidence of <i>Enterobacter aerogenes</i> with an increasing
271	resistance pattern was noticed in Belgian hospitals [54, 55].

Following the high rate of of ESBL-producing Enterobacteriaceae and *P. aeruginosa*, a 272 MER-based empiric scheme gave the highest rates of appropriate/adequate therapy. 273 Due to the failure of classic risk factors to predict MDR involvement, performance of 274 empiric schemes did not substantially alter when only patients without risk factors 275 276 were examined (Figure 2). Hence, the present study suggests coverage of both of ESBL-producing Enterobacteriaceae and *P. aeruginosa* is warranted and supports the 277 de-escalation strategy. The present findings however, with the failure of MDR 278 prediction based on general characteristics in particular, provide perspectives for the 279 280 surveillance assisted approach in which individual colonization status is a major element in steering empiric therapy. 281

282

In 62% (n=106) of patients empiric therapy was modified. In 60 cases modification 283 followed initial inappropriate or inadequate therapy, in the other 46 patients empiric 284 therapy could be refined (true de-escalation). It is noteworthy that when only 285 microbiologically documented infections are considered, four of 129 patients 286 received inappropriate therapy even after culture results were available (Figure 1). In 287 MER-based empiric schemes (n=37) therapy was continued in 23 and modofied in 14 288 patients (in four cases because of initial inappropriate therapy and in 10 to narrow 289 290 the spectrum). This supports the approach of empiric antibiotic coverage followed by de-escalation. Likweise Baran et al. found that previous exposure to carbapenems 291

2.

292 was 44% in patients with infections caused by imipenem-resistant *Acinetobacter* baumannii, but only 12% in patients infected by imipenem-susceptible strains [56]. 293 However multivariate analysis showed it was previous antibiotic exposure, and not 294 carbapenem exposure specifically, that was an independent risk factor for imipenem-295 resistance. The link between exposure and specific resistance development is 296 intuitive, but such findings suggest the aim of reducing microbial selection pressure 297 is valid for all antimicrobial agents. Further, based on the Surveillance of 298 Antimicrobial Use and Antimicrobial Resistance in German ICUs (SARI project), 299 Meyer E et al. identified carbapenem use as an independent predictor for increased 300 incidence of Stenotrophomonas maltophilia [57]. 301 302 This study has its limitations. Firstly, no outcome data are available since the primary 303 aim of the study was to describe antibiotic prescription patterns and how they 304 perform in terms of appropriate therapy. However, the relationship between initial 305 empiric failure to cover the causative pathogen and adverse outcomes was 306 demonstrated repeatedly, and is generally accepted as an important quality indicator 307 [58, 59]. Secondly, the cohort does not represent a consecutive series of ICU patients 308 with nosocomial infections This may have led to selection bias for more severe 309 infections, with a higher likelihood of MDR involvement, although there is no reason 310 311 to believe this would have changed the relationship between MDR infection and risk factors. 312

In conclusion, in this prospective study, which reflects real life practice in ICUpatients with nosocomial infections, the rate of appropriate or adequate empiric

315	therapy was 63.7%. This study demonstrated that classic risk factors for MDR such as
316	prior antibiotic exposure and length of ICU stay, may be insufficient to predict MDR
317	involvement. As such, empiric first-line use of MER allowed for significantly higher
318	rates of appropriate or adequate therapy, irrespective of presence of these risk
319	factors, and may be recommended in settings with a high prevalence of MDR
320	pathogens. In addition, these data illustrate the necessity for strict infection
321	prevention and control.
322	
323	
324	Declarations
325	Funding: This study was sponsored by an unrestricted grant by AstraZeneca,
326	Belgium. Dr. Blot is supported by a grant from the European Society of Intensive
327	Care Medicine and iMDsoft Patient Safety Research Award 2008.
328	Competing Interests: No conflicts of interest to declare
329	Ethical Approval: For this non-interventional, observational study, ethical approval
330	was given by the local ethics committees of the participating centres ($n=24$).
331	

333

***The ANTHICUS Study Investigators are**: O. Abid (RHMS, Ath, Belgium), S. Cran

- 335 (CHIREC, Brussels, Belgium), D. De Bels (CHU Brugmann, Brussels, Belgium), F. De Leener
- 336 (CHR St. joseph-Warquignies, Boussu, Belgium), L. Finianos (Clinique St. Joseph, Liège,
- 337 Belgium), F. Forêt (CHR St. Joseph-Warquignies, Mons, Belgium), M. Genard (CIU Ambroise
- 338 Paré, Mons, Belgium), E. Gilbert (CHR du Tournaisis, Tournai, Belgium), E. Khodadadi (IRIS
- 339 Sud-Bracops, Brussels, Belgium), S. Machayekhi (CH Hornu Frameries, Hornu), D. Mircev
- 340 (IRIS Sud-Etterbeek-Ixelles, Brussels, Belgium), D. Neuberg (CH de L'Ardenne, Libramont,
- Belgium), J.-Y. Piette (CH Bois de l'Abbaye, Seraign, Belgium), P. Serpe (CH ND Bruyères,
- 342 Chenee, Belgium), F. Beernaert (H. Hart Ziekenhuis, Eeklo, Belgium), K. De Decker
- 343 (Antwerp University Hospital, Antwerp, Belgium), K. De Ridder (AZ Maria Middelares,
- 344 Sint-Niklaas, Belgium), C. Declercq (St. Josef Ziekenhuis, Izegem, Belgium), I. Demeyer (OLV
- 345 Aalst, Aalst, Belgium), B. Nonneman (ASZ, Aalst, Belgium), K. Schoonheydt (ZNA,

- 346 Antwerp, Belgium, W. Swinnen (AZ St. Blasius, Dendermonde, Belgium), L. Wosteyn (ASZ,
- 347 Roeselaere, Belgium).
- 348

349 **References**

Blot S. Limiting the attributable mortality of nosocomial infection and multidrug resistance in
 intensive care units. Clin Microbiol Infect 2008;14:5-13.

Lorente L, Blot S, Rello J. Evidence on measures for the prevention of ventilator-associated
 pneumonia. Eur Respir J 2007;30:1193-207.

Masterton RG, Mifsud AJ, Rao GG. Review of hospital isolation and infection control
 precautions. J Hosp Infect 2003;54:171-3.

Labeau SO, Vandijck DM, Rello J, Adam S, Rosa A, Wenisch C, et al. Centers for Disease
 Control and Prevention guidelines for preventing central venous catheter-related infection: results of

a knowledge test among 3405 European intensive care nurses. Crit Care Med 2009;37:320-3.

Labeau S, Vandijck D, Rello J, Adam S, Rosa A, Wenisch C, et al. Evidence-based guidelines for
the prevention of ventilator-associated pneumonia: results of a knowledge test among European
intensive care nurses. J Hosp Infect 2008;70:180-5.

362 6. Yokoe DS, Classen D. Improving patient safety through infection control: a new healthcare
363 imperative. Infect Control Hosp Epidemiol 2008;29 Suppl 1:S3-11.

364 7. Berwick DM, Calkins DR, McCannon CJ, Hackbarth AD. The 100,000 lives campaign: setting a
365 goal and a deadline for improving health care quality. Jama 2006;295:324-7.

8. van der Kooi TI, de Boer AS, Mannien J, Wille JC, Beaumont MT, Mooi BW, et al. Incidence
and risk factors of device-associated infections and associated mortality at the intensive care in the
Dutch surveillance system. Intensive Care Med 2007;33:271-8.

369 9. Vincent JL, Bihari DJ, Suter PM, Bruining HA, White J, Nicolas-Chanoin MH, et al. The

370 prevalence of nosocomial infection in intensive care units in Europe. Results of the European

Prevalence of Infection in Intensive Care (EPIC) Study. EPIC International Advisory Committee. Jama
1995;274:639-44.

Blot SI, Depuydt P, Annemans L, Benoit D, Hoste E, De Waele JJ, et al. Clinical and economic
outcomes in critically ill patients with nosocomial catheter-related bloodstream infections. Clin Infect
Dis 2005;41:1591-8.

376 11. Vander Stichele RH, Elseviers MM, Ferech M, Blot S, Goossens H. Hospital consumption of
377 antibiotics in 15 European countries: results of the ESAC Retrospective Data Collection (1997-2002). J
378 Antimicrob Chemother 2006;58:159-67.

379 12. Vandijck DM, Depaemelaere M, Labeau SO, Depuydt PO, Annemans L, Buyle FM, et al. Daily
380 cost of antimicrobial therapy in patients with Intensive Care Unit-acquired, laboratory-confirmed
381 bloodstream infection. Int J Antimicrob Agents 2008;31:161-5.

382 13. Safdar N, Dezfulian C, Collard HR, Saint S. Clinical and economic consequences of ventilator 383 associated pneumonia: a systematic review. Crit Care Med 2005;33:2184-93.

Blot S, Vandewoude K, Hoste E, Colardyn F. Reappraisal of attributable mortality in critically
ill patients with nosocomial bacteraemia involving Pseudomonas aeruginosa. J Hosp Infect
2003;53:18-24.

Blot S, Vandewoude K, Hoste E, J DW, Kint K, Rosiers F, et al. Absence of excess mortality in
critically ill patients with nosocomial Escherichia coli bacteremia. Infect Control Hosp Epidemiol
2003;24:912-5.

Garrouste-Orgeas M, Timsit JF, Tafflet M, Misset B, Zahar JR, Soufir L, et al. Excess risk of
death from intensive care unit-acquired nosocomial bloodstream infections: a reappraisal. Clin Infect
Dis 2006;42:1118-26.

393 17. Blot SI, Vandewoude KH, Colardyn FA. Clinical impact of nosocomial Klebsiella bacteremia in
394 critically ill patients. Eur J Clin Microbiol Infect Dis 2002;21:471-3.

Blot SI, Vandewoude KH, Colardyn FA. Evaluation of outcome in critically ill patients with
 nosocomial enterobacter bacteremia: results of a matched cohort study. Chest 2003;123:1208-13.

Blot S, Cankurtaran M, Petrovic M, Vandijck D, Lizy C, Decruyenaere J, et al. Epidemiology and
outcome of nosocomial bloodstream infection in elderly critically ill patients: a comparison between
middle-aged, old, and very old patients. Crit Care Med 2009;37:1634-41.

Agbaht K, Diaz E, Munoz E, Lisboa T, Gomez F, Depuydt PO, et al. Bacteremia in patients with
 ventilator-associated pneumonia is associated with increased mortality: A study comparing

402 bacteremic vs. nonbacteremic ventilator-associated pneumonia. Crit Care Med 2007;35:2064-70.

403 21. Vandewoude KH, Blot SI, Benoit D, Colardyn F, Vogelaers D. Invasive aspergillosis in critically

404 ill patients: attributable mortality and excesses in length of ICU stay and ventilator dependence. J
405 Hosp Infect 2004;56:269-76.

406 22. Harbarth S, Garbino J, Pugin J, Romand JA, Lew D, Pittet D. Inappropriate initial antimicrobial
407 therapy and its effect on survival in a clinical trial of immunomodulating therapy for severe sepsis.
408 Am J Med 2003;115:529-35.

409 23. Ibrahim EH, Sherman G, Ward S, Fraser VJ, Kollef MH. The influence of inadequate

410 antimicrobial treatment of bloodstream infections on patient outcomes in the ICU setting. Chest

411 2000;118:146-55.

412 24. Kollef MH, Sherman G, Ward S, Fraser VJ. Inadequate antimicrobial treatment of infections: a

risk factor for hospital mortality among critically ill patients. Chest 1999;115:462-74.

Kumar A, Roberts D, Wood KE, Light B, Parrillo JE, Sharma S, et al. Duration of hypotension
before initiation of effective antimicrobial therapy is the critical determinant of survival in human
septic shock. Crit Care Med 2006;34:1589-96.

417 26. Masterton R, Drusano G, Paterson DL, Park G. Appropriate antimicrobial treatment in
418 nosocomial infections-the clinical challenges. J Hosp Infect 2003;55 Suppl 1:1-12.

27. Depuydt P, Benoit D, Vogelaers D, Claeys G, Verschraegen G, Vandewoude K, et al. Outcome
in bacteremia associated with nosocomial pneumonia and the impact of pathogen prediction by

421 tracheal surveillance cultures. Intensive Care Med 2006;32:1773-81.

422 28. Jung B, Sebbane M, Chanques G, Courouble P, Verzilli D, Perrigault PF, et al. Previous

423 endotracheal aspirate allows guiding the initial treatment of ventilator-associated pneumonia.

424 Intensive Care Med 2009;35:101-7.

29. Depuydt P, Benoit D, Vogelaers D, Decruyenaere J, Vandijck D, Claeys G, et al. Systematic
surveillance cultures as a toolto predict involvement of multidrug antibiotic resistant bacteria in
ventilator-associated pneumonia. Intensive Care Med 2008;34:675-82.

30. Blot S, Depuydt P, Vogelaers D, Decruyenaere J, De Waele J, Hoste E, et al. Colonization
status and appropriate antibiotic therapy for nosocomial bacteremia caused by antibiotic-resistant
gram-negative bacteria in an intensive care unit. Infect Control Hosp Epidemiol 2005;26:575-9.

431 31. Papadomichelakis E, Kontopidou F, Antoniadou A, Poulakou G, Koratzanis E, Kopterides P, et

al. Screening for resistant gram-negative microorganisms to guide empiric therapy of subsequentinfection. Intensive Care Med 2008;34:2169-75.

434 32. Depuydt PO, Blot SI, Benoit DD, Claeys GW, Verschraegen GL, Vandewoude KH, et al.

435 Antimicrobial resistance in nosocomial bloodstream infection associated with pneumonia and the

436 value of systematic surveillance cultures in an adult intensive care unit. Crit Care Med 2006;34:653-9.

437 33. Michel F, Franceschini B, Berger P, Arnal JM, Gainnier M, Sainty JM, et al. Early antibiotic

438 treatment for BAL-confirmed ventilator-associated pneumonia: a role for routine endotracheal439 aspirate cultures. Chest 2005;127:589-97.

34. Sandiumenge A, Diaz E, Bodí M, Rello J. Therapy of ventilator-associated pneumonia. A
patients-based approach based on the ten rules of "The Tarragona Strategy". Intensive Care Med
2003;29:876-83.

443 35. Kollef MH. Hospital-acquired pneumonia and de-escalation of antimicrobial treatment. Crit444 Care Med 2001;29:1473-5.

445 36. Leone M, Garcin F, Bouvenot J, Boyadjev I, Visintini P, Albanese J, et al. Ventilator-associated
446 pneumonia: breaking the vicious circle of antibiotic overuse. Crit Care Med 2007;35:379-85; quizz 86.

CCEPTED MANUSCR

447 37. Kollef M. Appropriate empirical antibacterial therapy for nosocomial infections: getting it 448 right the first time. Drugs 2003;63:2157-68. 449 38. Kollef MH. Treatment of ventilator-associated pneumonia: get it right from the start. Crit 450 Care Med 2003;31:969-70. 451 39. Rello J, Vidaur L, Sandiumenge A, Rodriguez A, Gualis B, Boque C, et al. De-escalation therapy 452 in ventilator-associated pneumonia. Crit Care Med 2004;32:2183-90. 453 Leone M, Martin C. How to break the vicious circle of antibiotic resistances? Curr Opin Crit 40. 454 Care 2008;14:587-92. 455 Masterton RG. The new treatment paradigm and the role of carbapenems. Int J Antimicrob 41. 456 Agents 2009;33:105-10. Calandra T, Cohen J. The international sepsis forum consensus conference on definitions of 457 42. 458 infection in the intensive care unit. Crit Care Med 2005;33:1538-48. 459 43. Blot S, Vandewoude K. Management of invasive candidiasis in critically ill patients. Drugs 460 2004;64:2159-75. 461 44. Vandewoude KH, Blot SI, Depuydt P, Benoit D, Temmerman W, Colardyn F, et al. Clinical 462 relevance of Aspergillus isolation from respiratory tract samples in critically ill patients. Crit Care 463 2006;10:R31. 464 45. Blot S, Vandewoude K, De Bacquer D, Colardyn F. Nosocomial bacteremia caused by 465 antibiotic-resistant gram-negative bacteria in critically ill patients: clinical outcome and length of 466 hospitalization. Clin Infect Dis 2002;34:1600-6. 467 46. Knaus WA, Draper EA, Wagner DP, Zimmerman JE. APACHE II: a severity of disease 468 classification system. Crit Care Med 1985;13:818-29. 469 47. Giamarellos-Bourboulis EJ, Papadimitriou E, Galanakis N, Antonopoulou A, Tsaganos T, 470 Kanellakopoulou K, et al. Multidrug resistance to antimicrobials as a predominant factor influencing 471 patient survival. Int J Antimicrob Agents 2006;27:476-81. 472 48. Leone M, Bourgoin A, Cambon S, Dubuc M, Albanese J, Martin C. Empirical antimicrobial 473 therapy of septic shock patients: adequacy and impact on the outcome. Crit Care Med 2003;31:462-474 7. 475 49. Ortega M, Marco F, Soriano A, Almela M, Martinez JA, Munoz A, et al. Analysis of 4758 476 Escherichia coli bacteraemia episodes: predictive factors for isolation of an antibiotic-resistant strain 477 and their impact on the outcome. J Antimicrob Chemother 2009;63:568-74. 478 50. Kollef KE, Schramm GE, Wills AR, Reichley RM, Micek ST, Kollef MH. Predictors of 30-day 479 mortality and hospital costs in patients with ventilator-associated pneumonia attributed to 480 potentially antibiotic-resistant gram-negative bacteria. Chest 2008;134:281-7. 22

- 481 51. Albrecht SJ, Fishman NO, Kitchen J, Nachamkin I, Bilker WB, Hoegg C, et al. Reemergence of 482 gram-negative health care-associated bloodstream infections. Arch Intern Med 2006;166:1289-94.
- 483 52. Lautenbach E, Polk RE. Resistant gram-negative bacilli: A neglected healthcare crisis? Am J
 484 Health Syst Pharm 2007;64:S3-21; guiz S2-4.
- 485 53. Trouillet JL, Chastre J, Vuagnat A, Joly-Guillou ML, Combaux D, Dombret MC, et al. Ventilator486 associated pneumonia caused by potentially drug-resistant bacteria. Am J Respir Crit Care Med
 487 1998;157:531-9.
- 488 54. De Gheldre Y, Glupczynski Y, Struelens M, De Mol P. Emergence of Enterobacter aerogenes
 489 as a major antibiotic-resistant nosocomial pathogen in Belgian hospitals. Clin Microbiol Infect
 490 1999;5:622-7.
- 491 55. De Gheldre Y, Struelens MJ, Glupczynski Y, De Mol P, Maes N, Nonhoff C, et al. National
- 492 epidemiologic surveys of Enterobacter aerogenes in Belgian hospitals from 1996 to 1998. J Clin
 493 Microbiol 2001;39:889-96.
- 494 56. Baran G, Erbay A, Bodur H, Onguru P, Akinci E, Balaban N, et al. Risk factors for nosocomial
 495 imipenem-resistant Acinetobacter baumannii infections. Int J Infect Dis 2008;12:16-21.
- 496 57. Meyer E, Schwab F, Gastmeier P, Rueden H, Daschner FD, Jonas D. Stenotrophomonas
- 497 maltophilia and antibiotic use in German intensive care units: data from Project SARI (Surveillance of
- 498 Antimicrobial Use and Antimicrobial Resistance in German Intensive Care Units). J Hosp Infect
- 499 2006;64:238-43.
- 500 58. Degoricija V, Sharma M, Legac A, Gradiser M, Sefer S, Vucicevic Z. Survival analysis of 314 501 episodes of sepsis in medical intensive care unit in university hospital: impact of intensive care unit 502 performance and antimicrobial therapy. Croat Med J 2006;47:385-97.
- 503 59. Dellinger RP, Carlet JM, Masur H, Gerlach H, Calandra T, Cohen J, et al. Surviving Sepsis
 504 Campaign guidelines for management of severe sepsis and septic shock. Intensive Care Med
- 505 2004;30:536-55.
- 506
- 507
- 508

- 509 Figure Legends
- 510 Figure 1 Review of 171 nosocomial infections distributed for microbiological
- 511 documentation and multidrug resistance involvement.
- 512 MDR, multidrug resistance or multidrug resistant
- 513 ESBL, extended spectrum beta-lactamase producing Enterobacteriaceae
- 514 MRSA, meticillin- resistant *Staphylococcus aureus*
- 515 MRSE, meticillin-resistant *Staphylococcus epidermidis*

516

- 517 Figure 3 Rates of empiric appropriate or adequate therapy in ICU patients with
- 518 nosocomial infections.
- 519 Cont., continuation of empiric therapy
- 520 Modif., modification of empiric therapy
- 521
- 522 Figure 2 Breakdown of microbiologically documented infections according to the
- 523 presence of risk factors for multidrug resistance, and multidrug resistance
- 524 involvement.
- 525 MDR, multidrug resistance
- 526
- 527 Figure 4 Rates of empiric appropriate or adequate therapy according to three

528 **major groups of empiric antibiotic regimens** (coverage of meticillin-resistant

- 529 pathogens not considered).
- 530 Figure 4a, all patients considered (n=170; in one patient no empiric therapy was initiated)
- 531 Figure 4b, only patients considered without risk factors for multidrug resistance involvement
- 532 (hospitalization at a general ward prior to ICU admission and prior antibiotic exposure),
- 533 (n=78).
- Group 1: coverage of of ESBL-producing and *Pseudomonas aeruginosa* (meropenem-basedschemes)

536 Group 2: coverage of *P. aeruginosa* (schemes containing an antipseudomonal agent)

Cox Cox

- 537 Group 3: no coverage of of ESBL-producing, or non-fermenting, Gram-negative bacteria
- 538 (first line agents)
- 539
- 540

Table 1 – Empiric antibiotic regimens clustered in six major groups according to the pathogens covered.

Coverage	Empiric regimens	n
ESBL- Enterobacteriaceae + P. aeruginosa + MRSA (n=6)	MER+VAN	5
	MER+VAN+AMI	1
		X
ESBL- Enterobacteriaceae + P. aeruginosa (n=31)	MER	18
	MER+AMI	9
	MER+OXA	1
	MER+CIP+AMI	1
	MER+PIPtaz+AMI	2
<i>P. aeruginosa</i> + MRSA (n=7)	PIPtaz+VAN+LEVO	1
	PIPtaz+VAN	1
	CAZ+VAN	1
	CEFE+VAN	2
	CEFE+VAN+AMI	2
P. aeruginosa (n=74)	PIPtaz	12
	PIPtaz+AMI	12
	PIPtaz+FLU	2
	PIPtaz+AMOXIclav	1
<u>v</u>	AMOXIclav+AMI	4
	CAZ	2
	CEFE	8
	CIP	2
	LEVO	3
	CEFE+AMI	6
	CAZ+AMI	7
	CEFE+CIP	3
	CAZ+MOXI	2
	CEFE+FLU	1
	CIP+AMOXIclav	2
	LEVO+AMOXIclav	3
	LEVO+AMOXIclav+CLA	1
	LEVO+AMI	1
	CIP+OXA	1

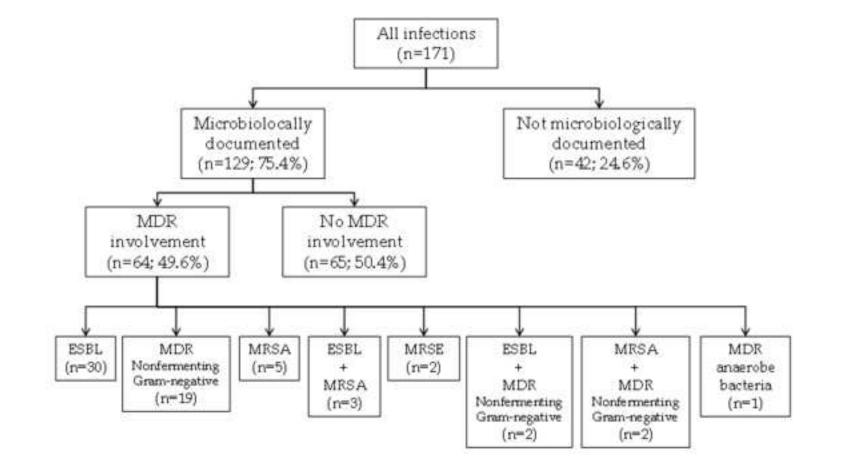
	OXA+AZT	1
MRSA (n=1)	VAN	1
Non-MDR pathogens or <i>P. aeruginosa</i> (n=51)	AMOXIclav	32
	AMOXIclav+metronidazole	1
	CEF	7
	MOXI	1
	CTX	1
	CTX+metronidazole	3
	CEF+CLA	1
	temocillin	3
	OXA	2
No empiric therapy (n=1)	5	1

Legend: MER, meropenem AMI, aminoglycoside VAN, vancomycin OXA, oxacillin CIP, ciprofloxacin PIPtaz, piperacillin/tazobactam LEVO, levofloxacin FLU, fluconazole AMOXIclav, amoxicillin/clavulanate CEF, cefuroxim CAZ, ceftazidime CTX, ceftriaxone CEFE, cefepim MOXI, moxifloxacin CLA, clarithromycin

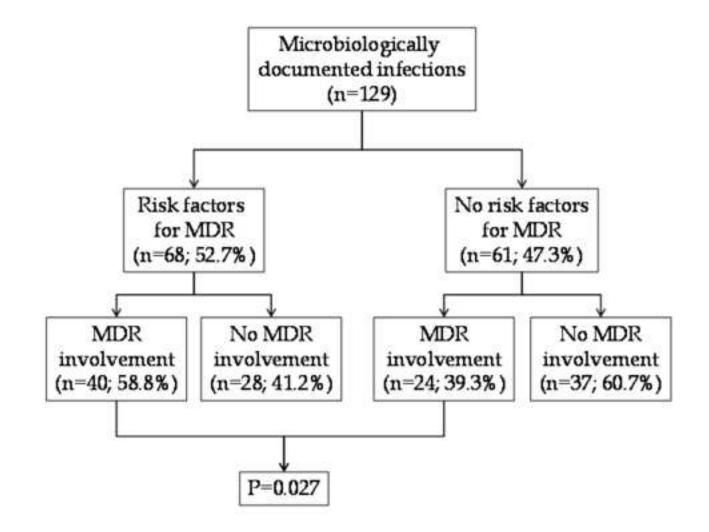
Characteristic			
Gender (male)	107 (62.0)		
Age (years)	67 (55 – 76)		
APACHE II score	20 (15 – 24)		
Recent trauma / surgery	39 (22.8)		
Co-morbidities	139 (81.3)		
Respiratory disease	66 (38.6)		
Cardiac disease	59 (34.5)		
Neurologic disease	28 (16.4)		
Diabetes mellitus	36 (21.1)		
Hepatic disease	11 (6.4)		
Renal disease	20 (11.7)		
Malignancy	28 (16.4)		
Neutropenia	1 (0.6)		
Corticosteroid therapy	20 (11.7)		
Prior hospitalization	51 (30.4)		
Hospitalization at another ward prior to ICU admission	84 (50.0)		
Prior antibiotic use	49 (28.7)		
ICU-acquired infection	115 (66.7)		
ICU stay before onset of infection (days)	3.8 (0 - 6)		

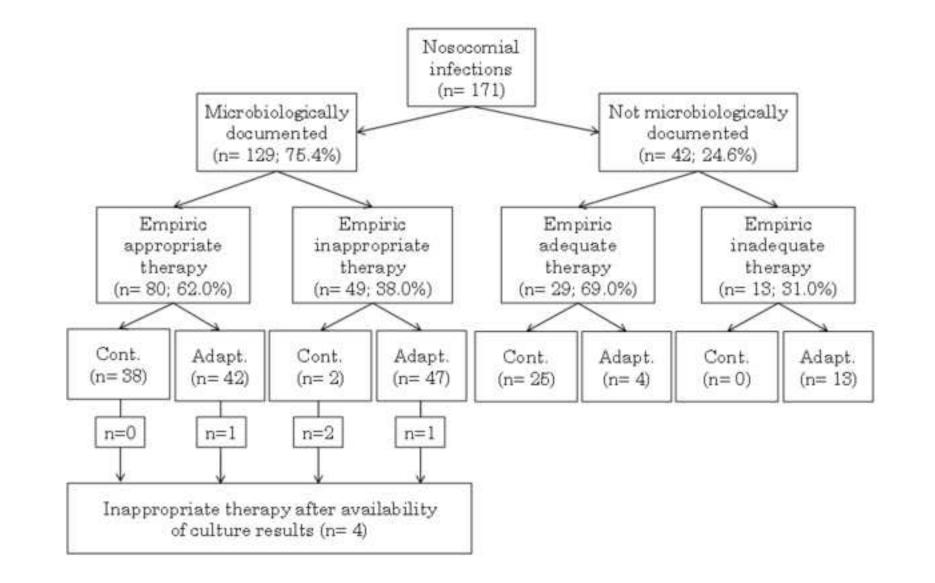
Table 2 – Characteristics of 171 ICU patients with a nosocomial infection.

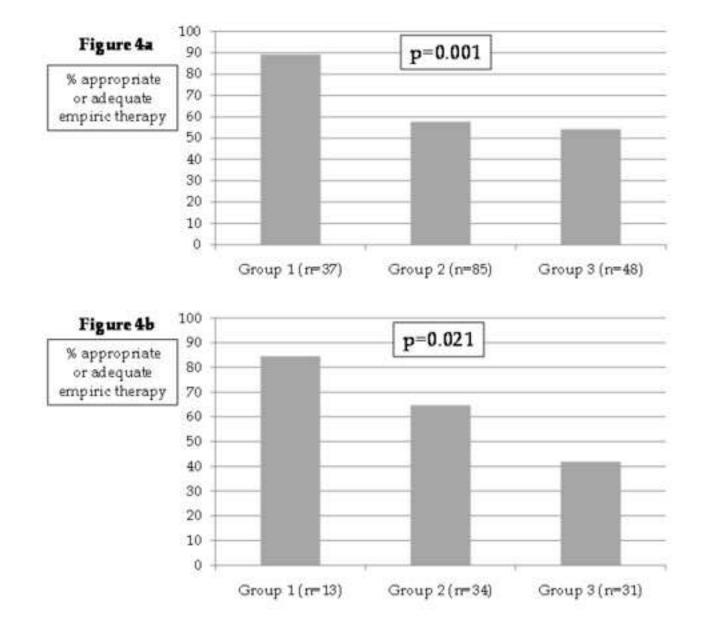
Data are described as n (%) or median (interquartile range)











Ċ

Patterns of antimicrobial therapy in severe nosocomial infections: empirical choices, proportion of appropriate therapy and adaptation rates—a multicentre observational survey in critically ill patients

Dirk Vogelaers ^a, David De Bels ^b, Frédéric Forêt ^c, Sophie Cran ^d, Eric Gilbert ^e, Karen Schoonheydt ^f, Stijn Blot ^{a,*}; for the ANTHICUS Study Investigators ¹

 ^a General Internal Medicine & Infectious Diseases, Ghent University Hospital, De
 Pintelaan 185, 9000 Ghent, Belgium
 ^b Intensive Care Department, Brugmann University Hospital, Brussels, Belgium
 ^c Department of Internal Medicine–Intensive Care Unit, CHR St-Joseph Warquignies, Boussu, Belgium
 ^d Intensive Care Department, CHIREC, Brussels, Belgium

- ^e Intensive Care Department, CHR du Tournaisis, Tournai, Belgium
- [†] Intensive Care Department, Ziekenhuisnetwerk Antwerpen, Antwerp, Belgium

ARTICLE INFO

Article history:

Received 17 September 2009

Accepted 23 November 2009

Keywords:

Antibiotic resistance

Multidrug resistance

Intensive care

Infection

Appropriate therapy

* Corresponding author. Tel.: +32 9 332 62 16.

E-mail address: stijn.blot@UGent.be (S. Blot).

¹ The ANTHICUS Study Investigators are: O. Abid (RHMS, Ath, Belgium); S. Cran (CHIREC, Brussels, Belgium); D. De Bels (CHU Brugmann, Brussels, Belgium); F. De Leener (CHR St-Joseph Warquignies, Boussu, Belgium); L. Finianos (Clinique St-Joseph, Liège, Belgium); F. Forêt (CHR St-Joseph Warquignies, Mons, Belgium); M. Genard (CIU Ambroise Paré, Mons, Belgium); E. Gilbert (CHR du Tournaisis, Tournai, Belgium); E. Khodadadi (IRIS Sud–Bracops, Brussels, Belgium); S. Machayekhi (CH Hornu-Frameries, Hornu, Belgium); D. Mircev (IRIS Sud–Etterbeek-Ixelles, Brussels, Belgium); D. Neuberg (CH de l'Ardenne, Libramont, Belgium); J.-Y. Piette (CH Bois de l'Abbaye, Seraign, Belgium); P. Serpe (CH Notre Dame Bruyères, Chenée, Belgium); F. Beernaert (H. Hart Ziekenhuis, Eeklo, Belgium); K. De Decker (Antwerp University Hospital, Antwerp, Belgium); K. De Ridder (AZ Maria Middelares, Sint-Niklaas, Belgium); C. Declercq (St-Josef Ziekenhuis, Izegem, Belgium); I. Demeyer (OLV Aalst, Aalst, Belgium); B. Nonneman (ASZ, Aalst, Belgium); K. Schoonheydt (ZNA, Antwerp, Belgium); W. Swinnen (AZ St-Blasius, Dendermonde, Belgium); and L. Wosteyn (ASZ, Roeselare, Belgium).

ABSTRACT

This prospective, multicentre, observational study investigated relationships between antimicrobial choices and rates of empirical appropriate or adequate therapy as well as subsequent adaptation of therapy in 171 Intensive Care Unit (ICU) patients with severe nosocomial infections. Appropriate antibiotic therapy was defined as in vitro susceptibility of the causative pathogen and clinical response to the agent administered. In non-microbiologically documented infections, therapy was considered adequate in the case of a favourable clinical response within 5 days. Patients had pneumonia (n = 127; 66 ventilator-associated), intra-abdominal infection (n = 23) and bloodstream infection (n = 21). Predominant pathogens were Pseudomonas aeruginosa (n = 29), Escherichia coli (n = 26), Staphylococcus aureus (n = 22) and Enterobacter aerogenes (n = 21). Multidrug-resistant (MDR) bacteria were involved in 49.6% of infections, mostly extended-spectrum β -lactamase (ESBL)producing Enterobacteriaceae and MDR non-fermenting Gram-negative bacteria. Prior antibiotic exposure and hospitalisation in a general ward prior to ICU admission were risk factors for multidrug resistance. Empirical therapy was appropriate/adequate in 63.7% of cases. Empirical schemes were classified according to coverage of (i) ESBL-producing Enterobacteriaceae + non-fermenting Gram-negative bacteria (meropenem-based), (ii) non-fermenting Gram-negative bacteria (schemes with an antipseudomonal agent) and (iii) first-line agents not covering ESBL-producing Enterobacteriaceae or non-fermenting Gram-negative bacteria. Meropenem-based schemes allowed for significantly higher rates of appropriate/adequate therapy (P < 0.001). This benefit remained when only patients without risk factors for MDR infection were considered (P = 0.021). Empirical therapy was adapted in 106 patients (62%), in 60 cases following initial

inappropriate/inadequate therapy and in 46 patients to fine-tune empirical therapy. In this study reflecting real-life practice, first-line use of meropenem provided significantly higher rates of appropriate/adequate therapy, irrespective of presence of risk factors for multidrug resistance.

1. Introduction

Avoiding nosocomial infection remains a daily challenge for healthcare workers involved in care of the critically ill [1]. The prevalence of infection strongly depends on the presence of particular iatrogenic risk factors, such as the use of indwelling devices and extensive surgery, as well as the severity of underlying disease and critical illness. Despite several large-scale efforts to improve the prevention of healthcare-associated infection [2–7], in general ca. 20–50% of patients hospitalised in the Intensive Care Unit (ICU) experience infection, either hospital- or ICU-acquired [8,9].

Severe nosocomial infection carries a substantial economic burden owing to extensive antimicrobial consumption and, even more importantly, increased length of hospitalisation [10–13]. In addition, severe infection seriously compromises the likelihood of survival. Attributable mortality rates vary from as low as 0% to a dramatic 50% depending of the type of infection, the causative pathogen, patient age, associated co-morbidities and overall quality of the anti-infective approach [13–21]. Among all the processes that should be fulfilled with the aim of optimising clinical patient outcome, early initiation of the proper antimicrobial agent is a cornerstone [1]. Failure to administer appropriate antimicrobial therapy in a timely manner results in a dramatic increase in the fatality rate [22–25]. However, selecting an empirical regimen that covers the causative pathogen is hampered by the presence of multidrug-resistant (MDR) microorganisms, which is the most important cause of inappropriate empirical therapy. Since antimicrobial consumption in itself is a major trigger for the development of multidrug resistance, physicians are frequently urged

to use 'last-line antibiotics' carefully. In fact, the challenge is to achieve high rates of appropriate empirical therapy in order to optimise patient survival whilst avoiding unnecessary use of antibiotics with the intention to minimise microbial selection pressure and hence development of multidrug resistance [1,26].

To accomplish this goal, in the past decade two main strategies have been proposed. In the surveillance-assisted strategy, the empirical regimen is selected mainly based on the presence or absence of MDR pathogens in routine surveillance cultures. The strength of this approach is the high negative predictive value of surveillance cultures in predicting the involvement of MDR microorganisms in subsequent infection. On the condition that cultures are taken at least twice weekly, this strategy is able to combine high rates of appropriate empirical therapy with reduced consumption of antibiotics in pneumonia and bacteraemia [27-33]. However, the cost effectiveness of this approach remains questionable. Another approach is the so-called 'deescalation strategy' that, on the condition of risk factors for MDR involvement, recommends initiation of empirical therapy with a regimen that covers most potential MDR pathogens such as extended-spectrum β -lactamase (ESBL)-producing Enterobacteriaceae, non-fermenting Gram-negative bacteria and meticillin-resistant Staphylococcus aureus (MRSA) [34-36]. Once culture results are available, and if feasible, narrowing the antimicrobial spectrum is advised. This concept, often referred to as 'de-escalation', is widely used and advocated [37–40]. This strategy has been demonstrated to be successful and safe and may also reduce antibiotic use and hence limit the emergence of multidrug resistance [36,41].

The objective of the present study was prospectively to investigate patterns of antimicrobial therapy in critically ill patients with nosocomial infection. More precisely, the following research questions were formulated. (i) What is the rate of empirical appropriate or adequate therapy achieved? (ii) Which antibiotic selections allow for the highest rates of empirical appropriate or adequate therapy, either in the presence or absence of risk factors for MDR pathogens? And (iii) what is the rate by which empirical therapy (either appropriate/adequate or not) is adapted?

2. Methods

2.1. Study design

A prospective, multicentre observational study was performed between February 2006 and June 2007. In total, 24 Belgian ICUs participated. Informed consent was requested from all patients. In all centres, antimicrobial prescribing was done or supervised by the attending senior intensive care physician.

2.2. Inclusion criteria

Eligible patients were those who provided informed consent, were at least 18 year of age, hospitalised in the ICU and experienced severe hospital-acquired infection, either pneumonia, intra-abdominal infection, primary bloodstream infection or secondary bloodstream infection originating from a source other than pneumonia or intra-abdominal infection (e.g. bacteraemia secondary to urinary tract infection or sinusitis).

2.3. Definitions

Infections were considered hospital- or ICU-acquired when they were diagnosed >48 h after hospital or ICU admission, respectively. Severe bacterial infections were defined following the International Sepsis Forum Consensus Conference on Definitions of Infection in the ICU [42]. Definitions of invasive fungal infections are described elsewhere [43,44]. Pneumonia was considered ventilator-associated when occurring after >48 h of mechanical ventilation. For the purpose of the study, only the first episode per patient was considered. Pneumonia or intra-abdominal infections with bacteraemic breakthrough (i.e. positive blood cultures) were classified according to the primary infection. Bloodstream infection secondary to sources other than pneumonia or intra-abdominal infection were classified as secondary bloodstream infection.

Definitions of multidrug resistance are described elsewhere [10,32,45]. *Candida* spp. were considered MDR when resistant to fluconazole.

Following the epidemiological scope of the study aiming at insights into empirical therapy in general, we also wanted to include microbiologically undocumented infections with obvious signs of clinical sepsis. For this purpose, a difference was made between appropriate and adequate empirical therapy. The term appropriate therapy was valid for microbiologically documented infectious episodes and was defined as in vitro susceptibility of the causative pathogen and clinical response to the agent administered. The term adequate therapy was used in non-microbiologically documented infections and was defined as favourable clinical response within 5 days of therapy (resolution of signs of sepsis).

Severity of disease was assessed by means of the Acute Physiology and Chronic Health Evaluation (APACHE) II score [46]. The following co-morbid conditions were registered: respiratory disease (chronic restrictive, obstructive or pulmonary vascular disease resulting in severe exercise restriction); cardiac disease (New York Heart Association Class IV); diabetes mellitus; hepatic disease (cirrhosis and portal hypertension, episodes of past upper gastrointestinal bleeding attributed to portal hypertension or prior episodes of hepatic failure or encephalopathy); renal disease (chronic glomerulonephritis, nephropathy or chronic kidney disease); neurological disease (impairment of alertness or confusion); malignancy (haematological cancer or solid tumour); and neutropenia (absolute neutrophil count <1500 cells/mm³).

To assess relationships between empirical antibiotic selection and rates of appropriate or adequate therapy, the empirical antimicrobial schemes were grouped according to the spectrum of pathogens covered. Table 1 describes the classification of empirical antimicrobial schemes. Because of the small number of empirical schemes covering MRSA and the relative low occurrence rate of MRSA in the cohort, empirical antibiotic schemes were classified in three major groups: (i) coverage of ESBL-producing Enterobacteriaceae + non-fermenting Gram-negative bacteria (meropenem-based schemes); (ii) coverage of non-fermenting Gram-negative bacteria (schemes including an antipseudomonal agent); and (iii) schemes without coverage of either ESBL-producing Enterobacteriaceae or non-fermenting Gramnegative bacteria (first-line agents). In this analysis, the added value of vancomycin as empirical therapy to cover meticillin-resistant Gram-positive pathogens was investigated separately.

Prior antibiotic exposure was defined as administration of antimicrobial agents within 1 month preceding the current infectious episode. Prior hospitalisation was defined as a hospital admission within the 4 months preceding the current ICU admission.

2.4. Data analysis

Continuous variables are described as median [interquartile range (IQR)] and discrete variables as number (%). For comparison between groups, the Mann–Whitney *U*-test and Fisher's exact test or χ^2 test were used as appropriate. Independent relationships with empirical appropriate or adequate therapy were assessed by means of logistic regression analysis. Variables taken into account in the logistic regression analysis either showed a moderate relationship in the univariate analysis or a logic relationship with the dependent variable. Variables considered were age, APACHE II score, underlying diseases, hospitalisation in another ward prior to ICU admission, prior antibiotic exposure and empirical antibiotic schemes. Results of the regression analysis are reported as odds ratios (OR) and 95% confidence interval (CI).

3. Results

3.1. Demographics

During the study period, 198 patients were included. Owing to incomplete patient files, 27 patients were excluded, resulting in a final database containing 171 patients. Patient characteristics are described in Table 2. Primary infections were pneumonia

(n = 127, of which 66 were ventilator-associated), intra-abdominal infection (n = 23) and bloodstream infection (n = 21, of which 9 were primary, 1 was catheter-related and 11 secondary to a source other than pneumonia or intra-abdominal infection). One hundred and fifteen infections (67.3%) were ICU-acquired. Approximately 75% of infections were microbiologically documented (Fig. 1). This represented 129 microbiologically documented infections in which 165 microorganisms were isolated. Gram-negative bacteria were most common (n = 122; 73.9%), with *Pseudomonas aeruginosa* (n = 29), *Escherichia coli* (n = 26), *Enterobacter aerogenes* (n = 21), *Klebsiella pneumonia* (n = 7) and *Klebsiella oxytoca* (n = 7) being the predominant pathogens. Among the 42 Gram-positive bacteria, *S. aureus* (n = 22), *Streptococcus pneumoniae* (n = 11) and enterococci (n = 5) were most frequently isolated. One fungal pathogen was isolated. MDR pathogens were involved in 64 (49.6%) of the 129 microbiologically documented infections. Fig. 1 shows the breakdown of the different types of resistance involved in these infections.

3.2. Risk factors for multidrug resistance

Risk factors for multidrug resistance were assessed in order to evaluate empirical antimicrobial regimens relative to their presence. No specific underlying conditions appeared to predispose for involvement of MDR organisms. MDR involvement was more frequent in patients with prior antibiotic exposure compared with patients without prior antibiotic exposure [74.3% vs. 41.3%; P = 0.001; relative risk (RR) = 4.26, 95% Cl 1.80–10.09]. Hospitalisation in another ward prior to ICU admission was also recognised as significantly associated with MDR involvement (59.0% vs. 40.9%; P = 0.041; RR = 2.08, 95% Cl 1.03–4.22). Fig. 2 shows a breakdown of

microbiologically documented infections according to the presence of these two risk factors for multidrug resistance and subsequent MDR involvement.

Length of ICU stay prior to the development of infection was negatively associated with the risk of MDR involvement. Patients who experienced infections without MDR pathogens developed this infection after a median of 3 days (IQR 2–7 days), whilst infections caused by MDR pathogens occurred after a median of 2 days (IQR 0–4.25 days) (P = 0.002).

3.3. Rates of empirical appropriate or adequate therapy and subsequent adaptation The presence of MDR pathogens in infections was associated with a significant lower rate of empirical appropriate therapy (87.7% vs. 35.9%; P < 0.001). Rates of empirical appropriate and adequate therapy are illustrated in Fig. 3. Empirical therapy was appropriate in 80 (62.0%) of the 129 microbiologically documented infections. In the 42 non-microbiologically documented infections, empirical therapy was judged adequate in 29 (69.0%). Empirical therapy was adapted in 106 patients (62.0%), in 60 cases following initial inappropriate or inadequate therapy and in 46 patients to fine-tune empirical therapy.

3.4. Empirical antimicrobial selection and rates of appropriate or adequate therapy In 12 infections the causative pathogen was meticillin-resistant (10 MRSA and 2 meticillin-resistant *Staphylococcus epidermidis*). Vancomycin was administered 14 times as an empirical agent, but in only 3 cases in which a meticillin-resistant pathogen proved to be involved. As such, the rate of inappropriate therapy in the

case of meticillin resistance involvement was 75%. Owing to the relative low prevalence of meticillin resistance as well as the low added value of initiating a glycopeptide, the association (or not) of such an agent did not alter the study results in this particular cohort. Therefore, rates of empirical appropriate or adequate therapy were analysed irrespective of the coverage of MRSA. This is illustrated in Fig. 4a. Coverage of ESBL-producing Enterobacteriaceae (by meropenem-based empirical schemes) allowed for significantly higher rates of appropriate or adequate therapy. Because the advantage of an antibiotic agent that covers ESBL-producing Enterobacteriaceae is obvious in settings with a high prevalence of multidrug resistance, it was hypothesised that the benefit of using a meropenem-based empirical scheme would fade away when only patients without risk factors for MDR were taken into account. However, in this analysis empirical schemes covering ESBL-producing Enterobacteriaceae remained superior (Fig. 4b).

When meropenem-based empirical schemes were compared with other empirical regimens, a difference of ca. 30% in empirical appropriate or adequate therapy was observed when all patients were taken into account (89.2% vs. 56.0%; P < 0.001) and this distinction remained when only patients without risk factors for MDR infection were considered (83.6% vs. 53.8%; P = 0.040).

In a multivariate logistic regression analysis, the only factors independently associated with empirical appropriate or adequate therapy were a meropenem-based empirical scheme (OR = 18.1, 95% CI 4.8–68.1; P < 0.001) and the presence of MDR pathogens (OR 0.04, 95% CI 0.01–0.10; P < 0.001). No other variables reached the level of significance.

4. Discussion

In this prospective, observational study that aimed to reflect real-life daily practice, rates of empirical appropriate or adequate therapy were only 63.7%. Approximately one-half of the infections were caused by MDR microorganisms, mostly ESBL-producing Enterobacteriaceae and *P. aeruginosa*. As such, only the inclusion of meropenem in the empirical scheme allowed for acceptable rates of appropriate therapy (ca. 90%).

Several reasons for the low rate of appropriate or adequate therapy can be proposed. The overall prevalence of multidrug resistance in this particular cohort is high (ca. 50%) and the rate of appropriate or adequate therapy in infections caused by MDR pathogens was very poor (35.9%). Multidrug resistance is well recognised as a major determinant of inappropriate therapy [47–52]. However, to a certain extent the involvement of multidrug resistance can be predicted on the basis of some typical risk factors, of which the most essential are prior antibiotic exposure and length of ICU stay (or mechanical ventilation in the case of pneumonia) of >7 days [32,53]. Other risk factors, such as a particular underlying diseases (e.g. chronic obstructive pulmonary disease) and recent surgery/hospitalisation, can also be taken into account but generally are not included in the principal risk factors. Strangely enough, in this cohort of ICU patients with nosocomial infection some of the classic patterns failed to predict the involvement of multidrug resistance. For instance, prior hospitalisation (within 4 months of the current hospital admission) was not associated with a higher likelihood of multidrug resistance, whilst length of ICU stay was even

inversely related to MDR involvement. On the other hand, prior antibiotic exposure and hospitalisation in a general ward prior to ICU admission appeared to have a significant relationship with MDR involvement. Importantly, however, although these relationships were statistically significant, MDR pathogens were isolated in ca. 40% of patients in the absence of these risk factors. As a consequence, the predictive value of these risk factors was low. As such, the generally accepted concepts of risk perception for MDR involvement failed in this particular cohort. We have no plausible explanation for this observation. Given the relationship between prior hospitalisation in a general ward and multidrug resistance, the relatively high number of infections already present at the time of ICU admission and the short length of stay in the ICU prior to infection, it appears that there exists a serious problem of multidrug resistance in general wards in Belgian hospitals, thereby contributing to the low rate of empirical appropriate therapy in ICUs. Indeed, already a decade ago a higher incidence of *E. aerogenes* with an increasing resistance pattern had been noticed in Belgian hospitals [54,55].

Following the high prevalence rate of ESBL-producing Enterobacteriaceae and *P. aeruginosa*, a meropenem-based empirical scheme allowed for the highest rates of appropriate/adequate therapy. Owing to the failure of classic risk factors to predict MDR involvement (Fig. 2), performance rates of empirical schemes did not substantially alter when only patients without risk factors were considered (Fig. 4b). Hence, based on the present study, an approach including coverage both of ESBL-producing Enterobacteriaceae and *P. aeruginosa* seems highly warranted. Without firm supportive knowledge of local ecology in ICU and non-ICU settings indicating low MDR levels, an empirical strategy that does not cover ESBL-producing

Enterobacteriaceae and *P. aeruginosa* is potentially dangerous and as such supports the de-escalation strategy. However, the present findings, with the failure of MDR prediction based on general characteristics in particular, provide perspectives for the surveillance-assisted approach in which individual colonisation status is a major element in steering empirical therapy.

Empirical therapy was adapted in 62% of patients (n = 106). In 60 cases adaptation followed initial inappropriate or inadequate therapy, whereas in 46 patients empirical therapy could be fine tuned (true de-escalation). Of note is that when only microbiologically documented infections are considered, 4 of 129 patients received inappropriate therapy even after the availability of culture results (Fig. 3). In meropenem-based empirical schemes (n = 37), therapy was continued in 23 patients and adapted in 14 patients (in 4 cases because of initial inappropriate therapy and in 10 cases to narrow down the spectrum). As this study supports the approach of empirical coverage of ESBL-producing Enterobacteriaceae and P. aeruginosa, we stress the importance of de-escalation whenever possible. Baran et al. [56] reported that the rate of previous exposure to carbapenems was 44% in patients with infections caused by imipenem-resistant Acinetobacter baumannii whereas it was only 12% in patients infected by imipenem-susceptible A. baumannii isolates. Yet in multivariate analysis, previous antibiotic exposure and not carbapenem exposure in particular appeared to be an independent risk factor for imipenem resistance. The link between exposure and specific resistance development is obvious and should not be disregarded, but this suggests that the mission to reduce microbial selection pressure is valid for all antimicrobial agents and not only for carbapenems. Furthermore, based on the Surveillance of Antimicrobial Use and Antimicrobial

Resistance in German ICUs (SARI project), Meyer et al. [57] identified carbapenem use as an independent predictor for a higher *Stenotrophomonas maltophilia* incidence. Although these data do not disclose a causal relationship, they are highly suggestive for the specific selection pressure of this pathogen. Therefore, excessive use of carbapenems is to be avoided by a strict de-escalation strategy and by avoidance of unnecessary long therapies.

The most important advantage of this study is its non-interventional design, reflecting real daily practice, thereby disclosing the true problem points in empirical antibiotic therapy in critically ill patients. Owing to the multicentre approach, the data might be valid for many tertiary care centres in Belgium and abroad.

However, the study has limitations. First, no outcome data are available as the primary aim of the study was to describe antibiotic prescription patterns and how they perform in terms of appropriate therapy. However, the relationship between failure of initial empirical therapy to cover the causative pathogen and adverse outcomes has been demonstrated repeatedly and is generally accepted as an important quality indicator [58,59]. Second, the cohort does not represent a consecutive series of ICU patients with nosocomial infections. It is possible that this might have led to a selection bias towards more severe infections, with a higher likelihood of MDR involvement.

In conclusion, in this prospective study reflecting real-life practice in ICU patients with nosocomial infections, the rate of appropriate or adequate empirical therapy was 63.7%. This study demonstrated that classic risk factors for multidrug resistance such

as prior antibiotic exposure and length of ICU stay may be insufficient to predict MDR involvement. As such, empirical first-line use of meropenem allowed for significantly higher rates of appropriate or adequate therapy, irrespective of the presence of these risk factors, and may be recommended in settings with a high prevalence of MDR pathogens. In addition, these data illustrate the necessity for strict infection prevention and control.

Funding

This study was sponsored by an unrestricted grant from AstraZeneca (Belgium). SB is supported by a grant from the European Society of Intensive Care Medicine and iMDsoft Patient Safety Research Award 2008.

Competing interests

None declared.

Ethical approval

Ethical approval was given by the local ethics committees of the participating centres

$$(n = 24).$$

References

- [1] Blot S. Limiting the attributable mortality of nosocomial infection and multidrug resistance in intensive care units. Clin Microbiol Infect 2008;14:5–13.
- [2] Lorente L, Blot S, Rello J. Evidence on measures for the prevention of ventilatorassociated pneumonia. Eur Respir J 2007;30:1193–207.
- [3] Masterton RG, Mifsud AJ, Rao GG. Review of hospital isolation and infection control precautions. J Hosp Infect 2003;54:171–3.
- [4] Labeau SO, Vandijck DM, Rello J, Adam S, Rosa A, Wenisch C, et al. Centers for Disease Control and Prevention guidelines for preventing central venous catheterrelated infection: results of a knowledge test among 3405 European intensive care nurses. Crit Care Med 2009;37:320–3.
- [5] Labeau S, Vandijck D, Rello J, Adam S, Rosa A, Wenisch C, et al. Evidencebased guidelines for the prevention of ventilator-associated pneumonia: results of a knowledge test among European intensive care nurses. J Hosp Infect 2008;70:180–5.
- [6] Yokoe DS, Classen D. Improving patient safety through infection control: a new healthcare imperative. Infect Control Hosp Epidemiol 2008;29(Suppl 1):S3–11.
- [7] Berwick DM, Calkins DR, McCannon CJ, Hackbarth AD. The 100,000 lives campaign: setting a goal and a deadline for improving health care quality. JAMA 2006;295:324–7.
- [8] van der Kooi TI, de Boer AS, Mannien J, Wille JC, Beaumont MT, Mooi BW, et al. Incidence and risk factors of device-associated infections and associated mortality at the intensive care in the Dutch surveillance system. Intensive Care Med 2007;33:271–8.

- [9] Vincent JL, Bihari DJ, Suter PM, Bruining HA, White J, Nicolas-Chanoin MH, et al. The prevalence of nosocomial infection in intensive care units in Europe. Results of the European Prevalence of Infection in Intensive Care (EPIC) Study. EPIC International Advisory Committee. JAMA 1995;274:639–44.
- [10] Blot SI, Depuydt P, Annemans L, Benoit D, Hoste E, De Waele JJ, et al. Clinical and economic outcomes in critically ill patients with nosocomial catheterrelated bloodstream infections. Clin Infect Dis 2005;41:1591–8.
- [11] Vander Stichele RH, Elseviers MM, Ferech M, Blot S, Goossens H. Hospital consumption of antibiotics in 15 European countries: results of the ESAC Retrospective Data Collection (1997–2002). J Antimicrob Chemother 2006;58:159–67.
- [12] Vandijck DM, Depaemelaere M, Labeau SO, Depuydt PO, Annemans L, Buyle FM, et al. Daily cost of antimicrobial therapy in patients with Intensive Care Unitacquired, laboratory-confirmed bloodstream infection. Int J Antimicrob Agents 2008;31:161–5.
- [13] Safdar N, Dezfulian C, Collard HR, Saint S. Clinical and economic consequences of ventilator-associated pneumonia: a systematic review. Crit Care Med 2005;33:2184–93.
- [14] Blot S, Vandewoude K, Hoste E, Colardyn F. Reappraisal of attributable mortality in critically ill patients with nosocomial bacteraemia involving *Pseudomonas aeruginosa*. J Hosp Infect 2003;53:18–24.
- [15] Blot S, Vandewoude K, Hoste E, De Waele J, Kint K, Rosiers F, et al. Absence of excess mortality in critically ill patients with nosocomial *Escherichia coli* bacteremia. Infect Control Hosp Epidemiol 2003;24:912–5.

- [16] Garrouste-Orgeas M, Timsit JF, Tafflet M, Misset B, Zahar JR, Soufir L, et al. Excess risk of death from intensive care unit-acquired nosocomial bloodstream infections: a reappraisal. Clin Infect Dis 2006;42:1118–26.
- [17] Blot SI, Vandewoude KH, Colardyn FA. Clinical impact of nosocomial *Klebsiella* bacteremia in critically ill patients. Eur J Clin Microbiol Infect Dis 2002;21:471–3.
- [18] Blot SI, Vandewoude KH, Colardyn FA. Evaluation of outcome in critically ill patients with nosocomial *Enterobacter* bacteremia: results of a matched cohort study. Chest 2003;123:1208–13.
- [19] Blot S, Cankurtaran M, Petrovic M, Vandijck D, Lizy C, Decruyenaere J, et al. Epidemiology and outcome of nosocomial bloodstream infection in elderly critically ill patients: a comparison between middle-aged, old, and very old patients. Crit Care Med 2009;37:1634–41.
- [20] Agbaht K, Diaz E, Munoz E, Lisboa T, Gomez F, Depuydt PO, et al. Bacteremia in patients with ventilator-associated pneumonia is associated with increased mortality: a study comparing bacteremic vs. nonbacteremic ventilatorassociated pneumonia. Crit Care Med 2007;35:2064–70.
- [21] Vandewoude KH, Blot SI, Benoit D, Colardyn F, Vogelaers D. Invasive aspergillosis in critically ill patients: attributable mortality and excesses in length of ICU stay and ventilator dependence. J Hosp Infect 2004;56:269–76.
- [22] Harbarth S, Garbino J, Pugin J, Romand JA, Lew D, Pittet D. Inappropriate initial antimicrobial therapy and its effect on survival in a clinical trial of immunomodulating therapy for severe sepsis. Am J Med 2003;115:529–35.

- [23] Ibrahim EH, Sherman G, Ward S, Fraser VJ, Kollef MH. The influence of inadequate antimicrobial treatment of bloodstream infections on patient outcomes in the ICU setting. Chest 2000;118:146–55.
- [24] Kollef MH, Sherman G, Ward S, Fraser VJ. Inadequate antimicrobial treatment of infections: a risk factor for hospital mortality among critically ill patients. Chest 1999;115:462–74.
- [25] Kumar A, Roberts D, Wood KE, Light B, Parrillo JE, Sharma S, et al. Duration of hypotension before initiation of effective antimicrobial therapy is the critical determinant of survival in human septic shock. Crit Care Med 2006;34:1589–96.
- [26] Masterton R, Drusano G, Paterson DL, Park G. Appropriate antimicrobial treatment in nosocomial infections—the clinical challenges. J Hosp Infect 2003;55(Suppl 1):1–12.
- [27] Depuydt P, Benoit D, Vogelaers D, Claeys G, Verschraegen G, Vandewoude K, et al. Outcome in bacteremia associated with nosocomial pneumonia and the impact of pathogen prediction by tracheal surveillance cultures. Intensive Care Med 2006;32:1773–81.
- [28] Jung B, Sebbane M, Chanques G, Courouble P, Verzilli D, Perrigault PF, et al. Previous endotracheal aspirate allows guiding the initial treatment of ventilatorassociated pneumonia. Intensive Care Med 2009;35:101–7.
- [29] Depuydt P, Benoit D, Vogelaers D, Decruyenaere J, Vandijck D, Claeys G, et al. Systematic surveillance cultures as a tool to predict involvement of multidrug antibiotic resistant bacteria in ventilator-associated pneumonia. Intensive Care Med 2008;34:675–82.
- [30] Blot S, Depuydt P, Vogelaers D, Decruyenaere J, De Waele J, Hoste E, et al. Colonization status and appropriate antibiotic therapy for nosocomial bacteremia

caused by antibiotic-resistant Gram-negative bacteria in an intensive care unit. Infect Control Hosp Epidemiol 2005;26:575–9.

- [31] Papadomichelakis E, Kontopidou F, Antoniadou A, Poulakou G, Koratzanis E, Kopterides P, et al. Screening for resistant Gram-negative microorganisms to guide empiric therapy of subsequent infection. Intensive Care Med 2008;34:2169– 75.
- [32] Depuydt PO, Blot SI, Benoit DD, Claeys GW, Verschraegen GL, Vandewoude KH, et al. Antimicrobial resistance in nosocomial bloodstream infection associated with pneumonia and the value of systematic surveillance cultures in an adult intensive care unit. Crit Care Med 2006;34:653–9.
- [33] Michel F, Franceschini B, Berger P, Arnal JM, Gainnier M, Sainty JM, et al. Early antibiotic treatment for BAL-confirmed ventilator-associated pneumonia: a role for routine endotracheal aspirate cultures. Chest 2005;127:589–97.
- [34] Sandiumenge A, Diaz E, Bodí M, Rello J. Therapy of ventilator-associated pneumonia. A patients-based approach based on the ten rules of "The Tarragona Strategy". Intensive Care Med 2003;29:876–83.
- [35] Kollef MH. Hospital-acquired pneumonia and de-escalation of antimicrobial treatment. Crit Care Med 2001;29:1473–5.
- [36] Leone M, Garcin F, Bouvenot J, Boyadjev I, Visintini P, Albanese J, et al.
 Ventilator-associated pneumonia: breaking the vicious circle of antibiotic overuse.
 Crit Care Med 2007;35:379–85; quiz 386.
- [37] Kollef M. Appropriate empirical antibacterial therapy for nosocomial infections: getting it right the first time. Drugs 2003;63:2157–68.
- [38] Kollef MH. Treatment of ventilator-associated pneumonia: get it right from the start. Crit Care Med 2003;31:969–70.

- [39] Rello J, Vidaur L, Sandiumenge A, Rodriguez A, Gualis B, Boque C, et al. Deescalation therapy in ventilator-associated pneumonia. Crit Care Med 2004;32:2183–90.
- [40] Leone M, Martin C. How to break the vicious circle of antibiotic resistances?Curr Opin Crit Care 2008;14:587–92.
- [41] Masterton RG. The new treatment paradigm and the role of carbapenems. Int J Antimicrob Agents 2009;33:105–10.
- [42] Calandra T, Cohen J. The International Sepsis Forum Consensus Conference on Definitions I infection in the Intensive Care Unit. Crit Care Med 2005;33:1538– 48.
- [43] Blot S, Vandewoude K. Management of invasive candidiasis in critically ill patients. Drugs 2004;64:2159–75.
- [44] Vandewoude KH, Blot SI, Depuydt P, Benoit D, Temmerman W, Colardyn F, et al. Clinical relevance of *Aspergillus* isolation from respiratory tract samples in critically ill patients. Crit Care 2006;10:R31.
- [45] Blot S, Vandewoude K, De Bacquer D, Colardyn F. Nosocomial bacteremia caused by antibiotic-resistant Gram-negative bacteria in critically ill patients: clinical outcome and length of hospitalization. Clin Infect Dis 2002;34:1600–6.
- [46] Knaus WA, Draper EA, Wagner DP, Zimmerman JE. APACHE II: a severity of disease classification system. Crit Care Med 1985;13:818–29.
- [47] Giamarellos-Bourboulis EJ, Papadimitriou E, Galanakis N, Antonopoulou A, Tsaganos T, Kanellakopoulou K, et al. Multidrug resistance to antimicrobials as a predominant factor influencing patient survival. Int J Antimicrob Agents 2006;27:476–81.

- [48] Leone M, Bourgoin A, Cambon S, Dubuc M, Albanese J, Martin C. Empirical antimicrobial therapy of septic shock patients: adequacy and impact on the outcome. Crit Care Med 2003;31:462–7.
- [49] Ortega M, Marco F, Soriano A, Almela M, Martinez JA, Munoz A, et al. Analysis of 4758 *Escherichia coli* bacteraemia episodes: predictive factors for isolation of an antibiotic-resistant strain and their impact on the outcome. J Antimicrob Chemother 2009;63:568–74.
- [50] Kollef KE, Schramm GE, Wills AR, Reichley RM, Micek ST, Kollef MH. Predictors of 30-day mortality and hospital costs in patients with ventilatorassociated pneumonia attributed to potentially antibiotic-resistant Gram-negative bacteria. Chest 2008;134:281–7.
- [51] Albrecht SJ, Fishman NO, Kitchen J, Nachamkin I, Bilker WB, Hoegg C, et al. Reemergence of Gram-negative health care-associated bloodstream infections. Arch Intern Med 2006;166:1289–94.
- [52] Lautenbach E, Polk RE. Resistant Gram-negative bacilli: a neglected healthcare crisis? Am J Health Syst Pharm 2007;64(23 Suppl 14):S3–21; quiz S22–4.
- [53] Trouillet JL, Chastre J, Vuagnat A, Joly-Guillou ML, Combaux D, Dombret MC, et al. Ventilator-associated pneumonia caused by potentially drug-resistant bacteria. Am J Respir Crit Care Med 1998;157:531–9.
- [54] De Gheldre Y, Glupczynski Y, Struelens M, De Mol P. Emergence of Enterobacter aerogenes as a major antibiotic-resistant nosocomial pathogen in Belgian hospitals. Clin Microbiol Infect 1999;5:622–7.

- [55] De Gheldre Y, Struelens MJ, Glupczynski Y, De Mol P, Maes N, Nonhoff C, et al. National epidemiologic surveys of *Enterobacter aerogenes* in Belgian hospitals from 1996 to 1998. J Clin Microbiol 2001;39:889–96.
- [56] Baran G, Erbay A, Bodur H, Onguru P, Akinci E, Balaban N, et al. Risk factors for nosocomial imipenem-resistant *Acinetobacter baumannii* infections. Int J Infect Dis 2008;12:16–21.
- [57] Meyer E, Schwab F, Gastmeier P, Rueden H, Daschner FD, Jonas D. Stenotrophomonas maltophilia and antibiotic use in German intensive care units: data from Project SARI (Surveillance of Antimicrobial Use and Antimicrobial Resistance in German Intensive Care Units). J Hosp Infect 2006;64:238–43.
- [58] Degoricija V, Sharma M, Legac A, Gradiser M, Sefer S, Vucicevic Z. Survival analysis of 314 episodes of sepsis in medical intensive care unit in university hospital: impact of intensive care unit performance and antimicrobial therapy. Croat Med J 2006;47:385–97.
- [59] Dellinger RP, Carlet JM, Masur H, Gerlach H, Calandra T, Cohen J, et al. Surviving Sepsis Campaign guidelines for management of severe sepsis and septic shock. Intensive Care Med 2004;30:536–55.

Fig. 1. Review of 171 nosocomial infections by microbiological documentation and multidrug resistance involvement. MDR, multidrug resistance or multidrug resistant; ESBL, extended-spectrum β -lactamase-producing Enterobacteriaceae; MRSA, meticillin- resistant *Staphylococcus aureus*; MRSE, meticillin-resistant *Staphylococcus epidermidis*.

Fig. 2. Breakdown of microbiologically documented infections according to the presence of risk factors for multidrug resistance (MDR) and MDR involvement.

Fig. 3. Rates of empirical appropriate or adequate therapy in Intensive Care Unit patients with nosocomial infections. Cont., continuation of empirical therapy; Adapt., adaptation of empirical therapy.

Fig. 4. Rates of empirical appropriate or adequate therapy according to three major groups of empirical antibiotic regimens (coverage of meticillin-resistant pathogens not considered): (a) all patients considered (n = 170; in 1 patient no empirical therapy was initiated); and (b) only patients without risk factors for multidrug resistance involvement considered (hospitalisation in a general ward prior to Intensive Care Unit admission and prior antibiotic exposure) (n = 78). Group 1, coverage of extended-spectrum β -lactamase (ESBL)-producing Enterobacteriaceae + *Pseudomonas aeruginosa* (meropenem-based schemes); Group 2, coverage of *P. aeruginosa* (schemes containing an antipseudomonal agent); and Group 3, no coverage of ESBL-producing or non-fermenting Gram-negative bacteria (first-line agents).

Table 1

Empirical antibiotic regimens clustered in six major groups according to the

pathogens covered

Coverage	Empirical regimens	n
ESBL-producing Enterobacteriaceae + Pseudomonas	MER+VAN	5
aeruginosa + MRSA ($n = 6$)		
	MER+VAN+AMI	1
ESBL-producing Enterobacteriaceae + <i>P. aeruginosa</i> (<i>n</i> =	MER	18
31)		
	MER+AMI	9
	MER+OXA	1
	MER+CIP+AMI	1
	MER+PIP/TAZ+AMI	2
P. aeruginosa + MRSA (n = 7)	PIP/TAZ+VAN+LEV	1
	PIP/TAZ+VAN	1
	CAZ+VAN	1
	CEP+VAN	2
	CEP+VAN+AMI	2
P. aeruginosa (n = 74)	PIP/TAZ	1
	PIP/TAZ+AMI	1
	PIP/TAZ+FLU	2
	PIP/TAZ+AMC	1
	AMC+AMI	4
	CAZ	2
	CEP	8
	CIP	2
	LEV	3
	CEP+AMI	6
	CAZ+AMI	7
	CEP+CIP	3
	CAZ+MOX	2

	CEP+FLU	1
	CIP+AMC	2
	LEV+AMC	3
	LEV+AMC+CLA	1
	LEV+AMI	1
	CIP+OXA	1
	OXA+AZT	1
$MRSA\;(n=1)$	VAN	1
Non-MDR pathogens or <i>P. aeruginosa</i> (<i>n</i> = 51)	AMC	32
	AMC+MTR	1
	CEF	7
	MOX	1
	CFX	1
	CFX+MTR	3
	CEF+CLA	1
	ТМО	3
	OXA	2
No empirical therapy $(n = 1)$		1

ESBL, extended-spectrum β-lactamase; MRSA, meticillin-resistant *Staphylococcus aureus*; MDR, multidrug-resistant; MER, meropenem; VAN, vancomycin; AMI, aminoglycoside; OXA, oxacillin; CIP, ciprofloxacin; PIP/TAZ, piperacillin/tazobactam; LEV, levofloxacin; CAZ, ceftazidime; CEP, cefepime; FLU, fluconazole; AMC, amoxicillin/clavulanic acid; MOX, moxifloxacin; CLA, clarithromycin; AZT, aztreonam; MTR, metronidazole; CEF, cefuroxime; CFX, ceftriaxone; TMO, temocillin.

Table 2

Characteristics of 171 Intensive Care Unit (ICU) patients with nosocomial infections

Characteristic	n (%) ^a
Gender (male)	107 (62.6)
Age (years) [median (IQR)]	67 (55–76)
APACHE II score [median (IQR)]	20 (15–24)
Recent trauma/surgery	39 (22.8)
Co-morbidities	139 (81.3)
Respiratory disease	66 (38.6)
Cardiac disease	59 (34.5)
Neurological disease	28 (16.4)
Diabetes mellitus	36 (21.1)
Hepatic disease	11 (6.4)
Renal disease	20 (11.7)
Malignancy	28 (16.4)
Neutropenia	1 (0.6)
Corticosteroid therapy	20 (11.7)
Prior hospitalisation	51 (29.8)
Hospitalisation at another ward prior to ICU admission	84 (49.1)
Prior antibiotic use	49 (28.7)
ICU-acquired infection	115 (67.3)
ICU stay before onset of infection (days) [median (IQR)]	3.8 (0–6)

IQR, interquartile range; APACHE, Acute Physiology and Chronic Health Evaluation.

^a Data are n (%) unless otherwise stated.



