



HAL
open science

BMP signalling permits population expansion by preventing premature myogenic differentiation in muscle satellite cells

Peter Zammit, Yusuke Ono, Frederico Calbaheu, Helge Amthor, Jennifer Morgan, Takenobu Katagiri

► **To cite this version:**

Peter Zammit, Yusuke Ono, Frederico Calbaheu, Helge Amthor, Jennifer Morgan, et al.. BMP signalling permits population expansion by preventing premature myogenic differentiation in muscle satellite cells. *Cell Death and Differentiation*, 2010, 10.1038/cdd.2010.95 . hal-00563514

HAL Id: hal-00563514

<https://hal.science/hal-00563514>

Submitted on 6 Feb 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

BMP signalling permits population expansion by preventing premature myogenic differentiation in muscle satellite cells

Yusuke Ono¹, Frederico Calbaheu¹, Jennifer E. Morgan², Takenobu Katagiri³, Helge Amthor⁴ and Peter S. Zammit^{1*}

1. King's College London, Randall Division of Cell and Molecular Biophysics, New Hunt's House, Guy's Campus, London, SE1 1UL, UK.
2. Dubowitz Neuromuscular Centre, UCL Institute of Child Health, 30 Guilford Street, London, WC1N 1EH
3. Division of Pathophysiology, Research Centre for Genomic Medicine, Saitama Medical University, 1397-1 Yamane, Hidaka-shi, Saitama 350-1241, Japan
4. UPMC INSERM UMR S 974 / CNRS UMR 7215, Institut de Myologie, 105 bd de l'Hôpital, 75013 Paris, France.

***Corresponding author:** Peter S. Zammit

King's College London, Randall Division of Cell and Molecular Biophysics, New Hunt's House, Guy's Campus, London, SE1 1UL, England.

E-mail: peter.zammit@kcl.ac.uk

Fax: +44 207 848 6435

Running title: BMP signalling regulates satellite cells

Key words: Satellite cell, BMP signalling, BMP4, Noggin, Pax7, Smad, Id1, MyoD, Skeletal muscle, Proliferation, Myogenic differentiation.

The number of words (except Abstract, Materials and Methods, References and Figure legends): **3498**

The number of figures: **9**

Abstract

Satellite cells are the resident stem cells of adult skeletal muscle, supplying myonuclei for homeostasis, hypertrophy and repair. Here, we have examined the role of Bone morphogenetic protein (BMP) signalling in regulating satellite cell function. Activated satellite cells expressed BMP receptor-1A (BMPR-1A) and contained phosphorylated Smad proteins, indicating that BMP signalling is operating during proliferation. Indeed, exogenous BMP4 stimulated satellite cell division and inhibited myogenic differentiation. Conversely, interfering with the interactions between BMPs and their receptors by the addition of either the BMP antagonist Noggin or soluble BMPR-1A fragments, induced precocious differentiation. Similarly, blockade of BMP signalling by siRNA-mediated knockdown of BMPR-1A, or disruption of the intracellular pathway by either Smad5 or Smad4 knockdown or inhibition of Smad1/5/8 phosphorylation with Dorsomorphin, also caused premature differentiation. BMP signalling acted to inhibit the up-regulation of genes associated with differentiation, in part, through regulating Id1. As satellite cells differentiated, Noggin levels increased to antagonise BMP signalling, since Noggin knockdown enhanced proliferation and impeded differentiation. Finally, interference of normal BMP signalling after muscle damage *in vivo* resulted in smaller regenerated myofibres. In conclusion, BMP signalling operates during routine satellite cell function to help coordinate the balance between proliferation and differentiation, before Noggin is activated to antagonise BMPs and facilitate terminal differentiation.

Introduction

Muscle satellite cells are the resident stem cells of skeletal muscle and supply myonuclei for postnatal muscle growth, and for maintenance and repair in adult (1). Satellite cells are located on the surface of myofibres and are mitotically quiescent in healthy adult muscle. In response to cues for routine myofibre homeostasis or hypertrophy, or the sporadic demands of muscle repair, satellite cells are activated to generate myoblasts that proliferate and eventually undergo myogenic differentiation to provide new myonuclei. Satellite cells also self-renew, thus maintaining a population of quiescent, undifferentiated precursors available to respond to repeated demand (2-4).

Satellite cell function is controlled by various regulatory pathways, chief amongst them being Notch/Delta and Wnt signalling (1). Many of these same regulatory networks also control embryonic myogenesis, in addition to many other processes both during development, and repair in adult. Another important network for organising embryonic and foetal myogenesis involves the Bone morphogenetic proteins (BMPs). BMPs belong to the transforming growth factor- β family and initiate signalling by binding to the transmembrane type 1 and type 2 BMP receptors. Upon BMP binding, type 1 and 2 receptors complex on the cell surface, allowing the constitutively active kinase of the type 2 receptor to transphosphorylate the type 1 receptor (5). This in turn, then phosphorylates the R-Smads - Smad1, Smad5 and Smad8 (pSmad1/5/8), which translocate to the nucleus to regulate transcription of target genes including *Ids* (6-8). Id proteins (comprising Id1, 2, 3, 4) bind ubiquitously expressed E-proteins to form inactive heterodimers, so preventing these same E-proteins from binding with tissue specific transcription factors such as MyoD and myogenin, a step necessary for their efficient function (9). BMP signalling can be modified in a number of ways, including via secreted antagonists such as Noggin, which bind BMPs with high affinity to interfere with interactions between BMPs and their receptors (5).

Although crucial for bone and cartilage formation and repair (5, 10), BMPs also inhibit myogenic differentiation to prevent ectopic myogenesis in the lateral plate mesoderm. In the dermomyotome though, BMP signalling itself must be inhibited, to permit the onset of myogenesis (11-14). BMPs not only impede myogenic differentiation in immortalised myogenic cell lines (e.g. C2) and primary myogenic cells *in vitro*, but also induce osteoblastic gene expression and differentiation towards the osteoblast lineage (15-18). Interestingly, although able to express early markers of osteogenic differentiation such as alkaline

phosphatase, many primary myogenic cells also retain expression of proteins associated with myogenesis (e.g. Pax7 and MyoD) after exposure to BMPs for several days (15, 17). Since intramuscular injection of certain BMPs (e.g. BMP2 and BMP4) can lead to ectopic bone formation *in vivo* (19, 20), these *in vitro* observations were thought to provide mechanistic insight into Fibrodysplasia ossificans progressiva (FOP): a rare disorder of skeletal malformations and progressive extra-skeletal ossification in muscle. FOP is caused by a mutation in *ALK-2*, which renders this BMP receptor type 1 constitutively active (19), and inhibition of this mutant *ALK-2* reduces ossification (21). However, Lounev et al. have recently shown that few (<5%) muscle progenitors actually contribute to BMP-induced heterotopic ossification *in vivo* (20).

Here, we investigated the role of BMP signalling in satellite cell function during adult myogenesis. During activation and proliferation, satellite cells robustly express BMP receptor type 1A (BMPR-1A), and pSmad1/5/8 and Smad4 are present in their nuclei, indicative of operational BMP signalling. Exogenous BMP4 can sustain satellite cell division and reduce differentiation, with BMP4 operating through BMPR-1A. Conversely, antagonising the interaction of BMP with its receptors, or perturbing intracellular BMP signalling by either inhibiting Smad1/5/8 phosphorylation or reducing Smad5 or Smad4 levels, all induced premature differentiation. Manipulation of BMP signalling affected Id1 levels: a known inhibitor of differentiation that regulates the function of the myogenic regulatory factors (that comprise of Myf5, MyoD, myogenin and MRF4) through sequestration of E-proteins. Satellite cell progeny then up-regulate the BMP antagonist Noggin as they differentiate, and knockdown of Noggin enhanced proliferation and impeded differentiation. Since blockade of BMP signalling after muscle injury hindered regeneration, our observations show that BMP signalling is a potent regulator of routine satellite cell function in adult.

Results

BMP pathway proteins are present in activated and proliferating satellite cells

Satellite cell function can be modelled *in vitro*. Upon stimulation by mitogen-rich medium (termed plating, or proliferation, medium here), isolated Pax7-expressing quiescent satellite cells are activated and up-regulate MyoD. Satellite cells then proliferate, before either down-regulating Pax7, maintaining MyoD, inducing myogenin and proceeding to differentiate, or down-regulating MyoD and maintaining Pax7 to return to a quiescent-like state, modelling self-renewal (2, 3).

To first determine the expression profile of BMPR-1A, we immunostained satellite cells retained in their niche on myofibres isolated from the extensor digitorum longus (EDL) muscle. BMPR-1A was undetectable in the vast majority of quiescent satellite cells on freshly isolated (T0) myofibres (**Fig. 1A**). However, after culture in plating medium for 48 (T48) or 72 hours (T72), BMPR-1A became robustly expressed in activated and proliferating satellite cells (**Fig. 1A**). *BMPR-1B* was not detectable by Q-PCR (data not shown), consistent with its absence in C2C12 myoblasts (22). Intracellular BMP signalling through BMPR-1A operates via phosphorylation of the carboxyl terminal of R-Smad proteins. Immunostaining for either pSmad1/5/8 or Smad5, revealed a strong nuclear signal in activated and proliferating (T48 and T72), but not quiescent (T0), satellite cells (**Fig. 1B and 1C**). The co-activator (Co-Smad) Smad4, which facilitates translocation of Smad1/5/8 to the nucleus and promotes their transcriptional activity (5), was also present in the nuclei of activated satellite cells (**Fig. 1C**). Thus, the expression dynamics of BMPR-1A, pSmad1/5/8, Smad4 and Smad5 mirror each other, and indicate that BMP signalling is operating in activated and proliferating satellite cells.

Since Noggin-mediated BMP antagonism contributes to initiation of the myogenic program during embryogenesis (13), we hypothesised that Noggin may also modify BMP signalling in satellite cells. Noggin was undetectable in quiescent (T0), and at low levels in activated Pax7⁺, satellite cells (**Fig. 2A and B**). By contrast, Noggin was highly expressed by differentiating myoblasts at T72, as shown by co-immunostaining for Noggin and myogenin (**Fig. 2B**). Culturing satellite cells attached to a myofibre provides a useful model for studying satellite cell activation, proliferation and the initial stages of differentiation. Later events, such as myoblast fusion into large multinucleated myotubes however, are better studied in satellite cells removed

from their associated myofibre and plated onto Matrigel-coated culture dishes. Such plated satellite cell-derived myoblasts cultured in differentiation medium for 2 days, clearly exhibited high Noggin levels in myotubes and other Pax7⁻ cells (**Fig. 2C and D**).

BMP regulates the balance between proliferation and differentiation in satellite cells

We next examined the effects of stimulating BMP signalling, and used administration of recombinant BMP4 to achieve this, since it is present in serum (23). Addition of recombinant BMP4 protein (100 ng/ml) to EDL satellite cells retained in their niche on the myofibre, significantly increased the number of Pax7⁺MyoD⁺ satellite cells, but decreased the number of cells with the differentiating Pax7⁻MyoD⁺ phenotype (**Fig. 3A**). Inhibition of differentiation by BMP4 was confirmed by co-immunostaining for myogenin and Pax7 (**Fig. 3B**). Plating medium contains chick embryo extract and horse serum, so is rich in growth factors, and contains BMP4 (23). Under these culture conditions, satellite cell proliferation is likely to be near maximal, hence addition of BMP4 did not further increase cell division (**Fig. 3A and B**). Since satellite cells retained expression of genes associated with myogenesis, these short exposures to BMP4 did not induce any obvious loss of myogenic identity.

To interfere with BMP binding to its receptors, we added recombinant Noggin protein (50 ng/ml) or soluble BMPR-1A fragments (sBMPR-1Af) (200 ng/ml) to cultures of satellite cells associated with a myofibre for 72 hours. Exposure to either Noggin or sBMPR-1Af resulted in a significant reduction in the mean total satellite cell number per myofibre, particularly of cells with the Pax7⁺MyoD⁺ phenotype (**Fig. 3A**). Numbers of differentiating Pax7⁻MyoD⁺ cells were already increased at T48, and became the predominant phenotype at T72, signifying a precocious onset of differentiation (**Fig. 3A**). There were also significantly fewer self-renewing Pax7⁺MyoD⁻ cells present at T72 (**Fig. 3A**). Co-immunostaining for Pax7 and myogenin confirmed that both Noggin and sBMPR-1Af enhanced differentiation, revealing a significant increase in cells with the Pax7⁻myogenin⁺ phenotype (**Fig. 3B**). These effects were not restricted to satellite cells from somite-derived muscles, since those from the branchiomeric masseter muscle, which has a different embryonic origin (24), also exhibited fewer differentiating Pax7⁻MyoD⁺ cells in the presence of BMP4, but increased differentiation when exposed to Noggin (**Fig. 3C**).

Culture in proliferation medium does not readily support differentiation of plated satellite cells. Addition of recombinant BMP4 resulted in a higher proportion of cells co-immunostained for Pax7 and

MyoD (**Fig. 3D**). The differentiation index under these culture conditions however, is already so low that any inhibitory effect of BMP4 on formation of myosin heavy chain (MyHC) containing myocytes was not significant (**Fig. 3E, quantified in F**). Addition of either Noggin or sBMPR-1Af however, triggered a robust differentiation, as shown by both the reduced proportion of plated cells with Pax7 (**Fig. 3D**), and the marked increase in myotubes containing MyHC (**Fig. 3E and F**). The proportion of satellite cell progeny with the self-renewing Pax7⁺MyoD⁻ phenotype was also significantly increased, from effectively zero in control cultures, to ~0.7% after exposure to either Noggin or sBMPR-1Af (**Fig. 3D**).

Finally, we determined the effects of exogenous BMP4 on satellite cells following serum deprivation, which normally promotes myogenin expression, cell cycle exit, and fusion into myotubes. However, addition of recombinant BMP4 (200 ng/ml) to differentiation medium led to a significant drop in the proportion of cells containing myogenin (**Fig. 3G, quantified in H**) and fusing into myotubes (**Fig. 3G, quantified in I**). However, MyoD remained readily detectable in the nuclei of these cells after exposure to BMP4 for 2 days (**Fig. 3G**). Importantly, BMP4 caused a 1.46±0.2-fold increase in total cell number, indicating that BMP4 not only inhibits differentiation, but also stimulates satellite cell proliferation, despite serum depletion (**Fig. 3J**).

BMP operates through BMPR-1A to prevent precocious differentiation

To determine whether BMP signalling was transduced through BMPR-1A in activated satellite cells, we used siRNA-mediated knockdown (25), which reduced the mean total number of satellite cells per myofibre when assayed at T72 (**Fig. 4A, quantified in B**). In particular, BMPR-1A knockdown significantly decreased the number of both Pax7⁺MyoD⁺ cells and Pax7⁺MyoD⁻ self-renewed cells, but not the number of Pax7⁺MyoD⁺ differentiating cells, which became the predominant phenotype (**Fig. 4B**). The efficient reduction in BMPR-1A levels by siRNA was confirmed by Western blot analysis of plated satellite cells, which also revealed decreased Pax7 levels, but an increase in the amount of MyHC (**Fig. 4C**). Immunostaining (**Fig. 4D, quantified E-G**) confirmed that reducing BMPR-1A led to a reduction in the proportion of Pax7⁺ cells (**Fig. 4E**), which was accompanied by a significant reduction in Ki67⁺ proliferating cells (**Fig. 4D and F**). In contrast, there was an increase in the proportion of MyHC⁺ differentiated cells after BMPR-1A knockdown (**Fig. 4D and G**).

Blockade of intracellular BMP signalling promotes myogenic differentiation

We next analysed the pSmad1/5/8 signalling cascade (5). The protein kinase inhibitor Dorsomorphin is a specific inhibitor of Smad1/5/8 phosphorylation by BMP receptors (26). Dorsomorphin (1-5 μM) was not toxic to plated satellite cells, as shown using the TUNEL assay (data not shown). Exposure of cultured myofibres to Dorsomorphin significantly decreased the mean number of associated Pax7⁺MyoD⁺ satellite cells, and so Pax7⁺MyoD⁺ differentiating cells became the main phenotype (**Fig. 5A**). Dorsomorphin also induced myogenic differentiation in a dose dependent (0.1-1.0 μM) manner in plated satellite cells maintained in proliferation medium (**Fig. 5B**). Additionally, we also targeted Smad signalling using siRNA-mediated knockdown of Smad5 or Smad4, which effectively reduced the levels of the translated protein (**Fig. 5C and D**). Co-immunostaining for Ki67 and MyHC revealed that Smad5 or Smad4 knockdown significantly reduced cell proliferation, and induced precocious differentiation (**Fig. 5E, quantified in F/G**).

BMP signalling regulates the ability of MyoD to activate its transcriptional targets

To understand how interference with BMP signalling promotes myogenic differentiation, we investigated Id1, a downstream target of BMPs (6-8) and a negative controller of MyoD and myogenin (9, 27). We recently showed that Id1 is highly expressed in proliferating satellite cells, before being down-regulated during myogenic differentiation (25). Immunostaining showed that exposure to Dorsomorphin to inhibit Smad1/5/8 phosphorylation, reduced Id1 levels in both satellite cells retained in their niche on the myofibre (**Fig. 6A**), and in plated satellite cells (**Fig. 6B**).

Muscle creatine kinase (CKM) is a well-characterised gene and provides a useful tool to quantify both the ability of MyoD to transactivate its target genes, and myogenic differentiation (27). Therefore, we measured the activity of a *CKM-luciferase* construct (*CKM-LUCpCS2*) in response to both exogenous Id1, and perturbation of BMP signalling (**Fig. 6C**). As expected, *CKM-LUCpCS2* activity was significantly decreased after transfection with *Id1* (**Fig. 6C**). Importantly, transfection of a plasmid encoding a constitutively active BMPR-1A (caBMPR-1A) that phosphorylates Smad1/5/8 in the absence of BMP ligands (28), also significantly reduced *CKM-LUCpCS2* activity (**Fig. 6C**). In contrast, inhibition of Smad1/5/8 phosphorylation with Dorsomorphin significantly increased the activity of *CKM-LUCpCS2*, but Id1 was able to completely reverse this Dorsomorphin-mediated increase (**Fig. 6C**). Finally, caBMPR-1A also reduced *CKM-LUCpCS2* activity back to control levels, despite the presence of Dorsomorphin (**Fig. 6C**).

Thus, BMP signalling in satellite cells acts to limit the transcriptional activation of genes controlled by MyoD and enhanced during differentiation.

Noggin antagonises BMPs to facilitate myogenic differentiation

Noggin is up-regulated as satellite cells differentiate (**Fig. 2**). To examine the role of Noggin, we first performed siRNA-mediated knockdown in plated satellite cells, and immunostaining showed that it worked efficiently (**Fig. 7A**). Plated satellite cells transfected with Noggin siRNA and co-immunostained for Ki67 and MyHC (**Fig. 7B**, quantified **C-E**) had a significantly increased proportion of proliferating Ki67⁺ satellite cells (**Fig. 7C**). However, the proportion of differentiating MyHC⁺ cells and the fusion index were both markedly reduced, with only myocytes or small myotubes present (**Fig. 7B, D and E**). Consistent with these observations, Western blot analysis revealed a slight decrease in myogenin levels (**Fig. 7F**). We also found that the fusion defect induced by Noggin knockdown could be rescued by Dorsomorphin-mediated blockade of intracellular BMP signalling (**Fig. 7G**, quantified **H**), indicating that endogenous Noggin from differentiating cells is required for proper myotube formation, through suppression of BMP signalling.

BMP signalling is operative during muscle regeneration *in vivo*

Finally, we evaluated whether BMP signalling was also operative in satellite cells in regenerating muscle *in vivo*. Cardiotoxin was used to induce damage in the gastrocnemius muscle of adult mice. Three days later, the regenerating muscles were removed, cryosectioned and immunostained for pSmad1/5/8, together with MyoD to identify satellite cell-derived myoblasts: 97.4 ± 2.9% (n= 50 cells from each of 3 mice) of MyoD⁺ cells contained pSmad1/5/8 (**Fig. 8A**). Undamaged areas of the muscle periphery provided the control, and no pSmad1/5/8 immunostaining was associated with myofibres in these non-regenerating regions (data not shown).

Regenerating gastrocnemius muscle was also injected with either 50 µl of 10 µM Dorsomorphin (**Fig. 8B-D**) or 50 µl of 100 µg/ml sBMPR-1Af (**Fig. 8E-G**) 1 and 3 days after cardiotoxin-induced injury. The contralateral regenerating muscle served as the control, and was injected with 50 µl of vehicle. Regenerating muscles were removed on day 8 and immunostained for MyHC and collagen type 1 (**Fig. 8B and E**). Administration of Dorsomorphin resulted in a significant ~20% decrease in the mean perimeter of centrally nucleated regenerating myofibres compared to controls, while the reduction was nearer ~40% with sBMPR-1Af

(**Fig. 8C and F**). A significant ~1.5-fold increase in collagen type 1 deposition following either Dorsomorphin or sBMPR-1Af administration was also observed (**Fig. 8D and G**).

Discussion

Here we investigated the role of BMP signalling in normal adult myogenesis. We found that exogenous BMP4 was able to sustain proliferation in satellite cells and inhibit differentiation. Conversely, blocking interaction of BMP with its receptors, down-regulating BMPR-1A or perturbing intracellular BMP signal-mediation, all induced rapid differentiation. Thus, BMP signalling initially serves to allow expansion of the satellite cell pool by stimulating proliferation and preventing precocious myogenic differentiation (**Fig. 9**).

The source of BMPs that act on satellite cells *in vivo* remains to be determined, but myogenic cells can express and secrete BMP4 (29, 30), with higher levels produced by myoblasts from dystrophic muscle (31). Additionally, muscle damage may also expose satellite cells to BMP4 in serum (23), and/or from cells such as macrophages, since microglia/macrophages produce BMPs after CNS damage (32). While prolonged exposure of myogenic cells to high levels of BMPs can induce expression of various osteogenic genes (16), we found that satellite cells exposed to BMP4 for the shorter periods likely encountered during muscle repair/regeneration *in vivo*, did not obviously compromise their myogenic identity.

Satellite cells respond to BMPs because they express BMPR-1A, which was undetectable on most quiescent satellite cells, but up-regulated as cells were activated. An early event in satellite cell activation is induction of MyoD, which can bind in the promoter of *Bmpr-1a* to enhance its expression, and MyoD over-expression leads to BMPR-1A induction (33). Thus up-regulation of BMPR-1A is part of the response of satellite cells to serum stimulation, and siRNA-mediated knockdown of BMPR-1A induced premature differentiation, showing that BMP signalling operates through BMPR-1A. Consistent with our observations, a dominant-negative BMPR-1A mutant enhanced myogenic differentiation in C2 cells maintained in high-serum medium, but a dominant-negative BMPR-1B mutant did not (23). By contrast, we found that a constitutively active BMPR-1A (28) reduced the activity of a *CKM* reporter that is normally up-regulated during myogenic differentiation.

BMPs are known to control the ability of MyoD to transactivate its target genes by Smad-mediated regulation of Id proteins (6-8), negative regulators of MyoD function and terminal differentiation (9, 27). Proliferating satellite cell-derived myoblasts contain both MyoD and Id1, with Id1 presumably curtailing MyoD function to prevent premature differentiation. Id1 levels are then dramatically reduced as satellite cells undergo differentiation (25). Consistent with these observations, while Id1 and Id3 are undetectable in

muscle lysates from uninjured muscle, they are up-regulated during muscle regeneration, peaking around day 3 (30). In satellite cells, we found that inhibiting BMP signalling reduced Id1 levels, promoted differentiation and increased *CKM* promoter activity.

Delta/Notch signalling permits satellite cell expansion and prevents precocious differentiation (34). Notch signalling is required for BMP4 to impede myogenic differentiation in C2 cells: inhibitors of γ -secretase prevent cleavage of Notch to release the active intracellular domain, and can partially reverse the BMP-induced differentiation block (18). The catalytic subunit of the γ -secretase complex is presenilin-1, and we recently showed that presenilin-1 can inhibit satellite cell differentiation via both γ -secretase-dependent and γ -secretase-independent mechanisms (25). The γ -secretase-dependent effects of presenilin-1 on impeding differentiation are likely through controlling Notch signalling. Crucially however, the γ -secretase (Notch)-independent presenilin-1 mechanism operates by augmenting Pax7 and controlling Id1 levels (25), and intriguingly, Id1, Id2 and Id3 are direct transcriptional targets of Pax7 (29). This γ -secretase-independent mechanism is unlikely to work through BMP signalling though, since there is no difference in pSmad1/5/8 levels between *presenilin-1*-null or control embryonic fibroblasts transfected with MyoD, even though only the MyoD-expressing *presenilin-1*-null cells readily differentiate in high serum medium (unpublished observations). Furthermore, over-expression of Wnt3a in C2 cells is capable of overcoming the BMP2-mediated block of differentiation by down-regulating Id1 (35). Thus γ -secretase-independent presenilin-1 signalling and the BMP pathway both appear to separately converge on Ids to regulate satellite cell function.

Antagonism of BMP signalling by Noggin is employed during development to protect pre-myogenic cells, and expansion of the Noggin expression domain promotes ectopic myogenesis (11, 12). We found that Noggin-mediated interference with BMP signalling was also redeployed during adult myogenesis. Noggin was up-regulated as satellite cells differentiated, and knocking-down Noggin resulted in more cells proliferating, while fewer differentiated. Since Noggin is a secreted signalling molecule, it may also act in a paracrine manner to reduce BMP signalling in proliferating cells to potentiate and augment timely differentiation. Noggin expression in C2 cells is up-regulated by exogenous BMP2 (36) and administration of Noggin to regenerating muscle reduces both pSmad1/5/8 and Id1 levels (30). In addition, rhabdomyosarcoma cells express myogenic genes including MyoD and myogenin but form myotubes poorly, and have relatively high levels of *BMP4* and lower expression of *Noggin*, than primary myoblasts (37).

Furthermore, specification of myogenic cells is unaffected in *Noggin*-null embryos, but differentiation is clearly perturbed, with meagre myotube formation during foetal myogenesis (38), again suggesting a common role for Noggin in developmental and adult myogenesis.

We observed that inhibition of BMP signalling during muscle regeneration resulted in smaller regenerated myofibres. Similarly, *Id1*^{-/+}*Id3*^{-/-} mice also exhibit a muscle regeneration defect, with the cross-sectional area of regenerated myofibres only ~50% of wild-type (30). These mice also had fewer proliferating satellite cells during regeneration (30).

In conclusion, BMP signalling is part of the program that regulates routine satellite cell function. BMPs stimulate proliferation and prevent precocious differentiation, to allow amplification of satellite cell-derived myoblasts. Noggin is then up-regulated as satellite cell progeny differentiate, to antagonise BMP signalling and facilitate coordinated differentiation (**Fig. 9**).

Material and methods

Isolation and culture of primary satellite cells and myoblasts

Adult (8-12 weeks old) C57BL10 mice were killed by cervical dislocation, and the EDL (or masseter for Fig. 3B only) muscle isolated and digested in collagenase type1 as previously described (25). Most satellite cells are mitotically quiescent in healthy adult muscle, so must first be activated and will then enter the cell cycle (2). To induce satellite cell activation *in vitro*, myofibers and associated satellite cells were cultured in plating medium [DMEM supplemented with L-glutamine or GlutaMax (Sigma), 10% horse serum (PAA Laboratories), 0.5% chicken embryonic extract (ICN Flow) and 1% penicillin-streptomycin (Sigma)] at 37°C in 5% CO₂, as described previously (24). To prepare plated satellite cells, either cells were removed from myofibers by enzymatic treatment with 0.125% trypsin-EDTA (Sigma) for 10 min at 37°C, or allowed to migrate from isolated myofibres cultured on Matrigel. Satellite cells were then re-plated on Matrigel at equal density and maintained in proliferation medium (DMEM supplemented with L-glutamine or GlutaMAX, 20-30% foetal bovine serum, 1% chicken embryo extract, 10 ng/ml bFGF and 1% penicillin-streptomycin). Satellite cells were treated with either 100 ng/ml recombinant BMP4 protein, (Peprotech), 50 ng/ml recombinant Noggin protein (Peprotech), 200 ng/ml sBMPR-1Af (R&D systems) or 0.1- 3 µM Dorsomorphin (also known as Compound C; Calbiochem) administered to the culture medium. In other experiments, satellite cells were switched from proliferation medium to differentiation medium (DMEM with L-glutamine or GlutaMax and supplemented with 2% horse serum and 1% penicillin-streptomycin) and cultured for 48 hours with or without BMP4 (200 ng/ml). Myofibres and associated satellite cells or plated satellite cells were fixed with warm 4% paraformaldehyde (PFA).

Antibodies used in the study

Antibodies were obtained from the following sources. Rabbit anti-BMPR-1A from Orbigen and a gift from Professor B. Kay (39); rat anti-Ki67 from DAKO; goat anti-Smad5, mouse anti-Smad4, mouse anti-MyoD and rabbit anti-MyoD antibodies were obtained from Santa Cruz (Santa Cruz); goat anti-Noggin antibody was obtained from R&D systems; mouse anti-MyHC (MF20), anti-Pax7, anti-myogenin (F5D) and anti-β-tubulin antibodies (E7) were supplied by the DSHB (Iowa City, IA); rabbit anti-pSmad1/5/8 antibody was from Cell

Signalling Technology (Beverly, MA); goat anti-collagen type 1 antibody was acquired from Southern Biotech (Birmingham, AL). Mounting medium containing 4,6-diamidino-2-phenylindole (DAPI) to counterstain all nuclei and the M.O.M kit were both purchased from Vector Laboratories.

Immunoblot analysis

Immunoblot analysis was performed as previously described (25). Anti-BMP-1A, anti-MyHC, anti-Pax7, anti-Myogenin or anti- β -tubulin antibodies were applied in PBS containing 0.1% Tween20 and incubated at 4°C overnight. HRP-conjugated secondary antibodies were used for visualisation by chemiluminescence (Fujifilm).

Immunostaining

Immunocytochemistry was as previously described (25). Anti-BMP-1A, anti-Ki67, anti-Smad5, anti-Smad4, anti-MyoD, anti-Noggin, anti-MyHC, anti-Pax7, anti-myogenin, anti-Id1, or anti-pSmad1/5/8 antibodies were applied at 4°C overnight. For co-immunostaining sections, frozen regenerating muscle cross-sections were fixed with 4% PFA, blocked using the M.O.M kit and incubated with either anti-pSmad1/5/8 and anti-MyoD antibodies, or with anti-MyHC and anti-collagen type 1 antibodies. Immunostained myofibres, plated cells or cryosections were viewed on a Zeiss Axiophot 200M using Plan-Neofluar lenses, or on a Nikon C1si confocal using Plan-Fluor lenses. Digital images were acquired with a Zeiss AxioCam HRm Charge-Coupled Device using AxioVision software version 4.4. Images were optimised globally and assembled into figures using Adobe Photoshop. The Click-iT TUNEL Alexa fluor imaging assay (Invitrogen) was used as per the manufacturer's instructions on satellite cells cultured for 24 hours in proliferation medium containing either 1 μ M or 5 μ M Dorsomorphin (n= 3 mice).

CKM luciferase reporter Assay

Plated satellite cells were co-transfected with the *CKM firefly Luciferase* reporter plasmid (*CKM-LUCpCS2*) and either *pcDNA3-Cre*-expressing control vector, *pcDNA3-mId1*-expressing vector (27) or *pcDEF3-constitutively active BMP-1A*-expressing vector (7), using Lipofectamine LTX according to the manufacturer's instructions. The *Renilla* luciferase plasmid *pRL* (Promega), driven by a minimal *tk promoter*, was included as an internal

control. Dorsomorphin was added to a final concentration of 5 μ M. 48 hours post-transfection, the dual-Luciferase reporter Assay (Promega) was performed according to the manufacturer's instructions and measured on an Anthos Lucy3 (JENCONS-PLS).

siRNA-mediated gene knockdown

The transfection of siRNA (Invitrogen) into primary satellite cell-derived myoblasts was performed using Lipofectamine 2000 reagent (Invitrogen) or Lipofectamine RNAiMAX (Invitrogen) as previously described (25). Transfection of siRNA into single myofibers was carried out 12-16 hours after isolation with Lipofectamine RNAiMAX (Invitrogen) (25). All samples were examined 1-3 days after the transfection. The following siRNA sequences were used: BMPR-1A siRNA: 5'-GAAGUGCUGGAUGAAAGCCUGAAUA-3', Smad4 siRNA: 5'-GGAGAGACGUUUAAGGUCCCUCAA-3', Smad5 siRNA: 5'-CCUGGGAUUGUUGUCAAAUGUUAAU-3' and Noggin siRNA: 5'-CGACCCGGGCUUUAUGGCUACUUCG-3'. Control siRNAs were supplied by Invitrogen.

Muscle regeneration studies

Animal husbandry, breeding and experimental procedures were passed by the Ethical Review Process Committee of King's College London, and were carried out in accordance with British law under the provisions of the Animals (Scientific Procedures) Act 1986. To assay pSmad1/5/8 expression, the gastrocnemius muscle of 10-week old male C57BL10 mice was injected with 50 μ l of 10 μ M cardiotoxin (Sigma) using a 29 G 1/2 insulin syringe. Muscles were removed 3 days later and immediately frozen in isopentane cooled in liquid nitrogen, and stored at -80°C before being cryosectioned and immunostained for pSmad1/5/8 and MyoD. To assay regenerative capacity in the presence of perturbed BMP signalling, the belly of both the left and right gastrocnemius muscles of 7-9 week old male C57BL10 mice were injected with 25 μ l of 10 μ M cardiotoxin (Sigma). 1 and 3 days after cardiotoxin injection, 50 μ l of 10 μ M Dorsomorphin/DMSO dissolved in saline or 50 μ l of 100 μ g/ml sBMPR-1Af dissolved in saline, were injected intramuscularly into the right gastrocnemius muscle at the same site as the cardiotoxin. Control solutions (same concentration of DMSO dissolved in saline for Dorsomorphin or BSA dissolved in saline for sBMPR-1Af) were injected into the left muscle as a control. Muscles were removed 8 days after cardiotoxin injection and immediately frozen in isopentane cooled in liquid nitrogen and stored at -80 °C.

Transverse sections of muscle were cut with a cryostat and co-immunostained for MyHC and collagen type 1. Measurement of centrally nucleated regenerating myofibres and the area of endomysial fibrosis was performed using Image J software (developed by NIH and available for free download at <http://rsb.info.nih.gov/ij>).

Statistic analysis

Data are presented as mean±standard deviation (SD) or mean±standard error (SEM). Immunostained satellite cells from at least ten myofibres were scored from each of at least three mice for each experiment. Significant differences between test conditions and controls were determined using Student *t-test*, where *p* values of < 0.05 were considered to be statistically significant.

Acknowledgements

We would like to thank Dr. Stephen J. Tapscott for providing the *CKM-LUCpCS2* reporter plasmid; Dr. Robert Benezra for generously sharing *pcDNA3-mId1* construct; Dr. Naohiro Hashimoto for helpful discussions and Paul Knopp for much help. We gratefully acknowledge colleagues who shared antibodies, including through the Developmental Studies Hybridoma Bank developed under the auspices of the NICHD and maintained by the University of Iowa. Y.O. was funded by the Muscular Dystrophy Campaign (RA3/737). F.C. is sponsored by the Association of International Cancer Research (07-0151). J.E.M. is backed by a Wellcome Trust university award. The laboratory of P.S.Z. is also supported by The Wellcome Trust, The Medical Research Council, the MYORES Network of Excellence (contract 511978) from the European Commission 6th Framework Programme and the collaborative Project OPTISTEM (Grant Agreement number: 223098) from the European Commission 7th Framework Programme.

Conflict of interest

Authors declare no conflict of interest.

References

1. Zammit PS. (2008) All muscle satellite cells are equal, but are some more equal than others? *J Cell Sci.* 121: 2975-82.
2. Zammit PS, Golding JP, Nagata Y, Hudon V, Partridge TA and Beauchamp JR. (2004) Muscle satellite cells adopt divergent fates: a mechanism for self-renewal? *J Cell Biol.* 166(3): 347-57.
3. Halevy O, Piestun Y, Allouh MZ, Rosser BW, Rinkevich Y, Reshef R, Rozenboim I, Wleklinski-Lee M and Yablonka-Reuveni Z. (2004) Pattern of Pax7 expression during myogenesis in the posthatch chicken establishes a model for satellite cell differentiation and renewal. *Dev Dyn.* 231: 489-502.
4. Collins CA, Olsen I, Zammit PS, Heslop L, Petrie A, Partridge TA and Morgan JE. (2005) Stem cell function, self-renewal, and behavioral heterogeneity of cells from the adult muscle satellite cell niche. *Cell.* 122(2): 289-301.
5. Feng XH and Derynck R. (2005) Specificity and versatility in tgf-beta signaling through Smads. *Annu Rev Cell Dev Biol.* 21: 659-93.
6. Hollnagel A, Oehlmann V, Heymer J, Ruther U and Nordheim A. (1999) Id genes are direct targets of bone morphogenetic protein induction in embryonic stem cells. *J Biol Chem.* 274(28): 19838-45.
7. Katagiri T, Imada M, Yanai T, Suda T, Takahashi N and Kamijo R. (2002) Identification of a BMP-responsive element in Id1, the gene for inhibition of myogenesis. *Genes Cells.* 7(9): 949-60.
8. Korchynski O and ten Dijke P. (2002) Identification and functional characterization of distinct critically important bone morphogenetic protein-specific response elements in the Id1 promoter. *J Biol Chem.* 277(7): 4883-91.
9. Jen Y, Weintraub H and Benezra R. (1992) Overexpression of Id protein inhibits the muscle differentiation program: in vivo association of Id with E2A proteins. *Genes Dev.* 6(8): 1466-79.
10. Wozney JM, Rosen V, Celeste AJ, Mitsock LM, Whitters MJ, Kriz RW, Hewick RM and Wang EA. (1988) Novel regulators of bone formation: molecular clones and activities. *Science.* 242(4885): 1528-34.
11. Reshef R, Maroto M and Lassar AB. (1998) Regulation of dorsal somitic cell fates: BMPs and Noggin control the timing and pattern of myogenic regulator expression. *Genes Dev.* 12(3): 290-303.
12. Re'em-Kalma Y, Lamb T and Frank D. (1995) Competition between noggin and bone morphogenetic protein 4 activities may regulate dorsalization during *Xenopus* development. *Proc Natl Acad Sci U S A.* 92(26): 12141-5.
13. Hirsinger E, Duprez D, Jouve C, Malapert P, Cooke J and Pourquie O. (1997) Noggin acts downstream of Wnt and Sonic Hedgehog to antagonize BMP4 in avian somite patterning. *Development.* 124(22): 4605-14.
14. Amthor H, Christ B, Weil M and Patel K. (1998) The importance of timing differentiation during limb muscle development. *Curr Biol.* 8(11): 642-52.
15. Wada MR, Inagawa-Ogashiwa M, Shimizu S, Yasumoto S and Hashimoto N. (2002) Generation of different fates from multipotent muscle stem cells. *Development.* 129(12): 2987-95.
16. Katagiri T, Yamaguchi A, Komaki M, Abe E, Takahashi N, Ikeda T, Rosen V, Wozney JM, Fujisawa-Sehara A and Suda T. (1994) Bone morphogenetic protein-2 converts the differentiation pathway of C2C12 myoblasts into the osteoblast lineage. *J Cell Biol.* 127(6 Pt 1): 1755-66.
17. Asakura A, Komaki M and Rudnicki M. (2001) Muscle satellite cells are multipotential stem cells that exhibit myogenic, osteogenic, and adipogenic differentiation. *Differentiation.* 68(4-5): 245-53.
18. Dahlqvist C, Blokzijl A, Chapman G, Falk A, Dannaeus K, Ibanez CF and Lendahl U. (2003) Functional Notch signaling is required for BMP4-induced inhibition of myogenic differentiation. *Development.* 130(24): 6089-99.
19. Kaplan FS, Shen Q, Lounev V, Seemann P, Groppe J, Katagiri T, Pignolo RJ and Shore EM. (2008) Skeletal metamorphosis in fibrodysplasia ossificans progressiva (FOP). *J Bone Miner Metab.* 26(6): 521-30.
20. Lounev VY, Ramachandran R, Wosczyzna MN, Yamamoto M, Maidment AD, Shore EM, Glaser DL, Goldhamer DJ and Kaplan FS. (2009) Identification of progenitor cells that contribute to heterotopic skeletogenesis. *J Bone Joint Surg Am.* 91(3): 652-63.
21. Yu PB, Deng DY, Lai CS, Hong CC, Cuny GD, Bouxsein ML, Hong DW, McManus PM, Katagiri T, Sachidanandan C, Kamiya N, Fukuda T, Mishina Y, Peterson RT and Bloch KD. (2008) BMP type I receptor inhibition reduces heterotopic [corrected] ossification. *Nat Med.* 14(12): 1363-9.
22. Namiki M, Akiyama S, Katagiri T, Suzuki A, Ueno N, Yamaji N, Rosen V, Wozney JM and Suda T. (1997) A kinase domain-truncated type I receptor blocks bone morphogenetic protein-2-induced signal transduction in C2C12 myoblasts. *J Biol Chem.* 272(35): 22046-52.

23. Kodaira K, Imada M, Goto M, Tomoyasu A, Fukuda T, Kamijo R, Suda T, Higashio K and Katagiri T. (2006) Purification and identification of a BMP-like factor from bovine serum. *Biochem Biophys Res Commun.* 345(3): 1224-31.
24. Ono Y, Boldrin L, Knopp P, Morgan JE and Zammit PS. (2010) Muscle satellite cells are a functionally heterogeneous population in both somite-derived and branchiomic muscles. *Dev Biol.* 337(1): 29-41.
25. Ono Y, Gnocchi VF, Zammit PS and Nagatomi R. (2009) Presenilin-1 acts via Id1 to regulate the function of muscle satellite cells in a gamma-secretase-independent manner. *J Cell Sci.* 122(24): 4427-38.
26. Yu PB, Hong CC, Sachidanandan C, Babbitt JL, Deng DY, Hoyng SA, Lin HY, Bloch KD and Peterson RT. (2008) Dorsomorphin inhibits BMP signals required for embryogenesis and iron metabolism. *Nat Chem Biol.* 4(1): 33-41.
27. Benezra R, Davis RL, Lockshon D, Turner DL and Weintraub H. (1990) The protein Id: a negative regulator of helix-loop-helix DNA binding proteins. *Cell.* 61(1): 49-59.
28. Akiyama S, Katagiri T, Namiki M, Yamaji N, Yamamoto N, Miyama K, Shibuya H, Ueno N, Wozney JM and Suda T. (1997) Constitutively active BMP type I receptors transduce BMP-2 signals without the ligand in C2C12 myoblasts. *Exp Cell Res.* 235(2): 362-9.
29. Kumar D, Shadrach JL, Wagers AJ and Lassar AB. (2009) Id3 is a direct transcriptional target of Pax7 in quiescent satellite cells. *Mol Biol Cell.* 20(14): 3170-7.
30. Clever JL, Sakai Y, Wang RA and Schneider D. (2010) Inefficient Skeletal Muscle Repair In Inhibitor of Differentiation (Id) Knockout Mice Suggests a Crucial Role for BMP Signaling During Adult Muscle Regeneration. *Am J Physiol Cell Physiol.* 298(5): C1087-99.
31. Sterrenburg E, van der Wees CG, White SJ, Turk R, de Menezes RX, van Ommen GJ, den Dunnen JT and t Hoen PA. (2006) Gene expression profiling highlights defective myogenesis in DMD patients and a possible role for bone morphogenetic protein 4. *Neurobiol Dis.* 23(1): 228-36.
32. Xiao Q, Du Y, Wu W and Yip HK. (2010) Bone morphogenetic proteins mediate cellular response and, together with Noggin, regulate astrocyte differentiation after spinal cord injury. *Exp Neurol.* 221(2): 353-66.
33. Liu R, Ginn SL, Lek M, North KN, Alexander IE, Little DG and Schindeler A. (2009) Myoblast sensitivity and fibroblast insensitivity to osteogenic conversion by BMP-2 correlates with the expression of *Bmpr-1a*. *BMC Musculoskelet Disord.* 10: 51.
34. Conboy IM and Rando TA. (2002) The regulation of Notch signaling controls satellite cell activation and cell fate determination in postnatal myogenesis. *Dev Cell.* 3(3): 397-409.
35. Nakashima A, Katagiri T and Tamura M. (2005) Cross-talk between Wnt and bone morphogenetic protein 2 (BMP-2) signaling in differentiation pathway of C2C12 myoblasts. *J Biol Chem.* 280(45): 37660-8.
36. Takayama K, Suzuki A, Manaka T, Taguchi S, Hashimoto Y, Imai Y, Wakitani S and Takaoka K. (2009) RNA interference for noggin enhances the biological activity of bone morphogenetic proteins in vivo and in vitro. *J Bone Miner Metab.* 27(4): 402-11.
37. Goldstein M, Meller I and Orr-Urtreger A. (2007) FGFR1 over-expression in primary rhabdomyosarcoma tumors is associated with hypomethylation of a 5' CpG island and abnormal expression of the *AKT1*, *NOG*, and *BMP4* genes. *Genes Chromosomes Cancer.* 46(11): 1028-38.
38. Tylzanowski P, Mebis L and Luyten FP. (2006) The Noggin null mouse phenotype is strain dependent and haploinsufficiency leads to skeletal defects. *Dev Dyn.* 235(6): 1599-607.
39. Kariyawasam HH, Xanthou G, Barkans J, Aizen M, Kay AB and Robinson DS. (2008) Basal expression of bone morphogenetic protein receptor is reduced in mild asthma. *Am J Respir Crit Care Med.* 177(10): 1074-81.

Titles and legends to figures

Fig. 1. BMPR-1A, pSmad 1/5/8, Smad4 and Smad 5 are up-regulated during satellite cell activation

Isolated EDL myofibres with their associated satellite cells were either immediately fixed (T0) or cultured in plating medium for either 48 hours (T48) or 72 hours (T72) before fixation and immunostaining. **(A)** BMPR-1A was undetectable on the majority of Pax7⁺ quiescent cells at T0, but robustly expressed in both Pax7⁺ and Pax7⁻ cells at both T48 and T72. **(B)** Quiescent Pax7⁺ satellite cells (T0) did not contain pSmad1/5/8, whereas pSmad1/5/8 became readily detectable in the nuclei of proliferating Pax7⁺ cells at T48, and both Pax7⁺ and Pax7⁻ cells at T72. **(C)** In accordance with the dynamics of pSmad1/5/8 expression, Smad5 levels were very low/absent in quiescent satellite cells at T0, but up-regulated in both proliferating Pax7⁺ and MyoD⁺ cells at T48, as was Smad4. Arrows indicate the same satellite cell at each time point to aid comparison. Representative images from at least three independent experiments are shown. Scale bar equals 30 μm .

Fig. 2. Noggin is highly expressed in satellite cell progeny committing to myogenic differentiation

Isolated EDL myofibres with their associated satellite cells were either immediately fixed (T0) or cultured in plating medium for 72 hours (T72) before fixation and immunostaining. **(A)** Noggin was not expressed in quiescent Pax7⁺ satellite cells (arrows) at T0. **(B)** After culture, Noggin was highly expressed in both Pax7⁻ and Myogenin⁺ cells committed to myogenic differentiation (arrowheads) at T72, but at much lower levels in Pax7⁺ cells (arrows). **(C and D)** Plated satellite cell-derived myoblasts were cultured in differentiation medium for 2 days and immunostained for Noggin and Pax7. **(C)** Pax7⁻ differentiating cells (arrowheads) and **(D)** myotubes both expressed high levels of Noggin protein, while Noggin levels in Pax7⁺ cells were low (arrows). Representative images from at least three independent experiments are shown. Scale bar equals 30 μm .

Fig. 3. Inhibition of BMP signalling promotes myogenic differentiation in satellite cells

Isolated myofibres associated with satellite cells, or plated satellite cell-derived myoblasts, were cultured in the presence of recombinant BMP4 protein (100 ng/ml), recombinant Noggin protein (50 ng/ml) or sBMPR-1Af (200 ng/ml). **(A)** EDL myofibres were cultured in plating medium supplemented with BMP4, Noggin, or

sBMPR-1Af, then fixed after 24 (T24), 48 (T48) or 72 (T72) hours and immunostained for Pax7 and MyoD. BMP4 reduced the number of Pax7⁻ MyoD⁺ cells. Both Noggin and sBMPR-1Af reduced the mean total number of cells per myofibre and promoted myogenic differentiation (increase of the Pax7⁻ MyoD⁺ phenotype). **(B)** Co-immunostaining isolated myofibres for Pax7 and myogenin confirmed that BMP4 inhibited myogenic differentiation, while Noggin and sBMPR-1Af promoted it, as shown by the increase of satellite cell progeny with the Pax7⁻ myogenin⁺ phenotype. **(C)** Treatment of masseter-derived satellite cells with BMP4 or Noggin for 72 hours produced the same result as for EDL cells, i.e. BMP4 inhibited, while Noggin induced, myogenic differentiation. **(D-F)** To examine the effects of these recombinant proteins on the later stages of myogenesis, EDL satellite cells were plated and cultured in the presence of recombinant BMP4, Noggin or sBMPR-1Af in plating medium for 4 days (with the medium changed every other day). **(D)** Co-immunostaining for Pax7 and MyoD again showed a dramatic increase in the proportion of cells with the differentiating Pax7⁻ phenotype when exposed to either Noggin or sBMPR-1Af. **(E and F)** Immunostaining for MyHC revealed that exposure to either Noggin or sBMPR-1Af resulted in precocious differentiation and formation of large multi-nucleated myotubes. **(G-J)** To better examine the effects of BMP on satellite cell proliferation and myogenic progression, plated satellite cells were also cultured in differentiation medium supplemented with recombinant BMP4 protein (200 ng/ml) and immunostained for MyoD and myogenin **(G, quantified in H-J)**. Exposure to BMP4 significantly reduced the proportion of cells expressing myogenin **(H)** and fusing into large myotubes **(I)**, but stimulated continued cell proliferation, resulting in a 1.46±0.2-fold increase in total cell number **(J)**. Data is mean±SEM **(A-C)** or mean±SD **(F-J)** from at least three independent experiments. An asterisk indicates that data are significantly different from controls ($p<0.05$) using Student's T-test. Scale bar equals 100 µm for **(E)**.

Fig. 4. BMP signalling operates through BMPR-1A

We used siRNA-mediated protein knockdown to explore if BMP signalling operates through BMPR-1A. **(A and B)** Control (Cont si) or BMPR-1A (BMPR-1Asi) siRNA were transfected into satellite cells retained in their niche on isolated EDL myofibers and cultured for ~2.5 days before fixation and co-immunostaining for Pax7 and MyoD. BMPR-1A knockdown reduced the mean total number of cells and especially the number of Pax7⁺MyoD⁺ cells. **(C-G)** Control siRNA or BMPR-1A siRNA duplexes were also transfected into plated satellite cell-derived myoblasts. **(C)** Immunoblot analysis clearly demonstrated efficient knockdown of

BMPR-1A protein levels, which resulted in an up-regulation of MyHC, but a down-regulation of Pax7 (β -tubulin served as a protein loading control). **(D-G)** The effects of BMPR-1A knockdown were also examined by immunostaining for Pax7, Ki67 or MyHC in plated satellite cell-derived myoblasts cultured in proliferation medium for 3 days after transfection. **(D)** Representative images of Ki67 and MyHC immunostaining show that BMPR-1A knockdown inhibited cell proliferation and promoted myogenic differentiation. **(E-F)** There was a significant decrease in the proportion of cells containing Pax7, or Ki67 (proliferating) after BMPR-1A siRNA-transfection. **(G)** In contrast, there was a significant increase in the proportion of MyHC⁺ differentiated cells and myotubes. Data is either mean \pm SEM **(B)**, or mean \pm SD **(E - G)** from at least three independent experiments. An asterisk denotes that data are significantly different from control ($p<0.05$). Scale bar equals 50 μ m.

Fig.5. Inhibiting intracellular BMP signal mediation promotes myogenic differentiation.

To inhibit BMP signal mediators, we used either Dorsomorphin, an inhibitor of Smad1/5/8 phosphorylation, or siRNA to knockdown the R-Smad - Smad5 or Co-Smad - Smad4, levels. **(A)** EDL myofibres and their associated satellite cells were cultured with or without 1 μ M Dorsomorphin and the plating medium changed daily. Dorsomorphin markedly decreased the mean total number of cells per fibre, and particularly the number of cells with the Pax7⁺MyoD⁺ phenotype. **(B)** Satellite cell-derived myoblasts were plated at high cell density (80-90% confluency) and cultured in proliferation medium for 2 days with or without Dorsomorphin. Immunostaining for MyHC showed that Dorsomorphin promoted myogenic differentiation and fusion in a dose-dependent manner. **(C and D)** Efficient siRNA-mediated knockdown of Smad5 or Smad4 was confirmed by immunostaining of plated satellite cells 24 hours after transfection. **(E-G)** Co-immunostaining for MyHC and Ki67 after Smad5 or Smad4 silencing for 3 days in proliferation medium, was sufficient to promote cell cycle exit and induce precocious differentiation, characterised by a decreased proportion of cells with Ki67⁺ expression and an increase in MyHC⁺ cells and myotubes. Data is mean \pm SEM **(A)**, or mean \pm SD **(F and G)** from at least three independent experiments. An asterisk indicates that data are significantly different from control conditions ($p<0.05$). Scale bar equals 50 μ m.

Fig. 6. BMP signalling regulates the MyoD-controlled *CKM* promoter

Id1 is a target gene of BMP signalling, and a negative regulator of MyoD activity. **(A)** Exposure to 3 μ M Dorsomorphin for 6 hours to inhibit BMP signalling resulted in a down-regulation of Id1 expression in satellite cells associated with a myofibre. **(B)** Similarly, immunostaining of plated satellite cells after exposure to 3 μ M Dorsomorphin for 18 hours in proliferation medium also resulted in down-regulation of Id1 protein. Scale bar equals 20 μ m. **(C)** In order to investigate the ability of MyoD to activate its transcriptional targets, plated satellite cell-derived myoblasts were transfected with a *CKM* (muscle creatine kinase) Luciferase reporter plasmid (*CKM-LUCpCS2*), whose activity is regulated by MyoD, together with a *Renilla* luciferase plasmid *pRL* driven by a minimal *tk promoter*, as an internal control. Plated satellite cells were co-transfected with either *Cre*-expressing control plasmid or *Id1*-expressing plasmid or constitutively active *BMPR-1A* (*caBMPR-1A*)-expressing plasmid, together with *CKM-LUCpCS2*. After transfection, the cells were cultured in proliferation medium with or without 5 μ M Dorsomorphin for 24 hours before reporter assay. Id1 or caBMPR-1A reduced *CKM-LUCpCS2* activity. By contrast, inhibition of BMP signalling with Dorsomorphin had the opposite effect, and increased *CKM-LUCpCS2* activity. Both Id1 and caBMPR-1A were able to reverse this increase in *CKM-LUCpCS2* activity produced by exposure to Dorsomorphin. Data is represented as mean \pm SD from at least three independent experiments. An asterisk denotes that data are significantly different from control conditions, while a cross indicates that data is different from the level of *CKM-LUCpCS2* activity in the presence of Dorsomorphin ($p < 0.05$), using Student's T-test.

Fig. 7. Reducing Noggin levels enhances satellite cell proliferation but inhibits differentiation

To further understand the role of Noggin in myogenic progression, we used siRNA-mediated knockdown of Noggin protein. **(A)** Efficient Noggin knockdown in plated satellite cell-derived myoblasts was confirmed by immunostaining for Noggin 3 days after siRNA (Noggin si) transfection. **(B-E)** The effects of Noggin knockdown were analysed by co-immunostaining for MyHC and Ki67, which revealed an increase in myoblast proliferation, but a reduction in differentiation and fusion into large myotubes. **(F)** Western blotting also showed that Noggin siRNA reduced myogenin levels in plated satellite cells. **(G and H)** The effect of Dorsomorphin (1 μ M) treatment to inhibit BMP signalling on Noggin siRNA transfected cells was analysed by immunostaining for MyHC. Dorsomorphin reversed the fusion defect induced by Noggin knocked-down.

Mean \pm SD from at least three independent experiments is shown. An asterisk indicates that data are significantly different from control ($p<0.05$) using paired one-tail T-test. Scale bar equals 30 μ m.

Fig. 8. BMP signalling operates during normal muscle regeneration *in vivo*.

Regeneration was induced in mouse gastrocnemius muscles by injection of cardiotoxin. **(A)** Satellite cell-derived myoblasts, identified by MyoD expression, also expressed pSmad1/5/8 (arrows) in regenerating muscle on day 3 of regeneration. **(B-G)** To examine the effects of blockade of BMP signalling, 50 μ l of 10 μ M Dorsomorphin/DMSO in saline or 50 μ l of 100 μ g/ml sBMPR-1Af were intramuscularly injected into the right gastrocnemius 1 and 3 days following cardiotoxin. Regenerating left gastrocnemius served as a control and received vehicle alone (DMSO/saline for Dorsomorphin or BSA solution for sBMPR-1Af). Muscle regeneration was assayed by immunostaining cryosections for MyHC and collagen type 1. **(B, C, E and F)** Treatment with either Dorsomorphin or sBMPR-1Af resulted in a significant decrease in the mean perimeter of centrally nucleated regenerating myofibres, compared to controls (100 regenerating fibres in the regenerating area of each animal were measured, and the mean size after exposure to Dorsomorphin or sBMPR-1Af expressed as the proportional change compared to the mean size of control regenerating myofibres). **(B, D, E and G)** Immunostaining for collagen type 1 was used to measure the degree of endomysial fibrosis and a significant increase in the fibrotic index was observed in muscles exposed to either Dorsomorphin or sBMPR-1Af, compared with controls (the area of collagen type 1 expression in the regenerating area was measured in 1-3 sections for each animal, using ImageJ and changes with exposure to Dorsomorphin or sBMPR-1Af expressed as fold-difference compared to control). Six mice were used for each condition. Data is mean \pm SEM, where an asterisk denotes a significant difference from control ($p<0.05$). Scale bars equal 100 μ m.

Fig. 9. Role of BMP signalling during myogenic progression in satellite cells

During muscle repair and regeneration, quiescent Pax7⁺ satellite cells are activated to co-express Pax7 and MyoD. Satellite cell-derived myoblasts proliferate extensively before many then down-regulate Pax7 and differentiate to either repair damaged muscle fibres, or fuse together to generate new myofibres. Other satellite cells maintain Pax7 expression but lose MyoD, and self-renew to maintain a viable stem cell

compartment. Normal BMP signalling via BMPR-1A and Smad1/5/8 phosphorylation is required during muscle repair/regeneration to allow the satellite cell-derived myoblast population to expand, by preventing precocious differentiation. BMP signalling operates through controlling Id levels, a negative regulator of MyoD and myogenic differentiation. As satellite cell progeny differentiate, Noggin is up-regulated to antagonise the BMP signal to facilitate the differentiation programme.

Figure 1

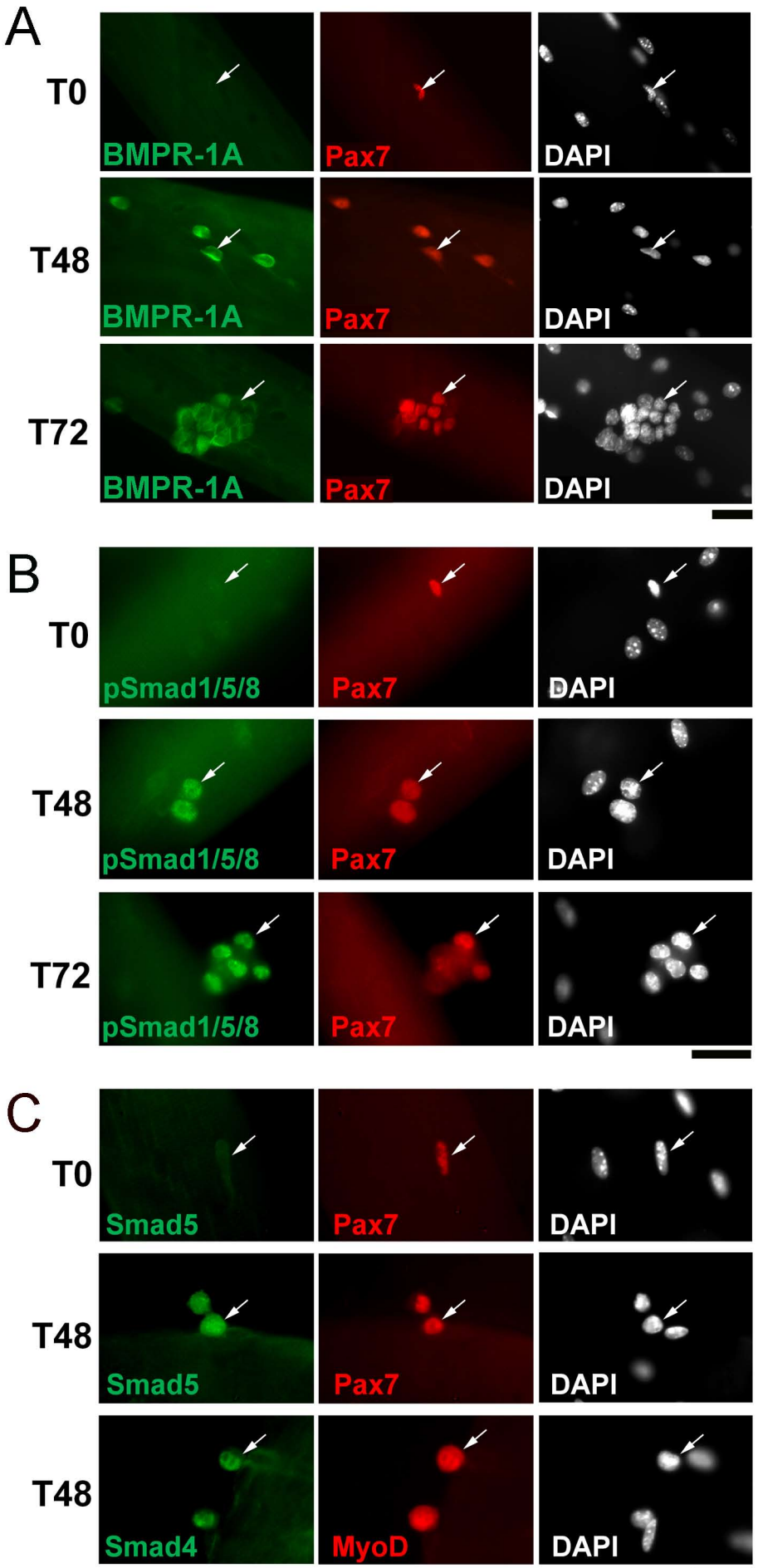


Figure 2

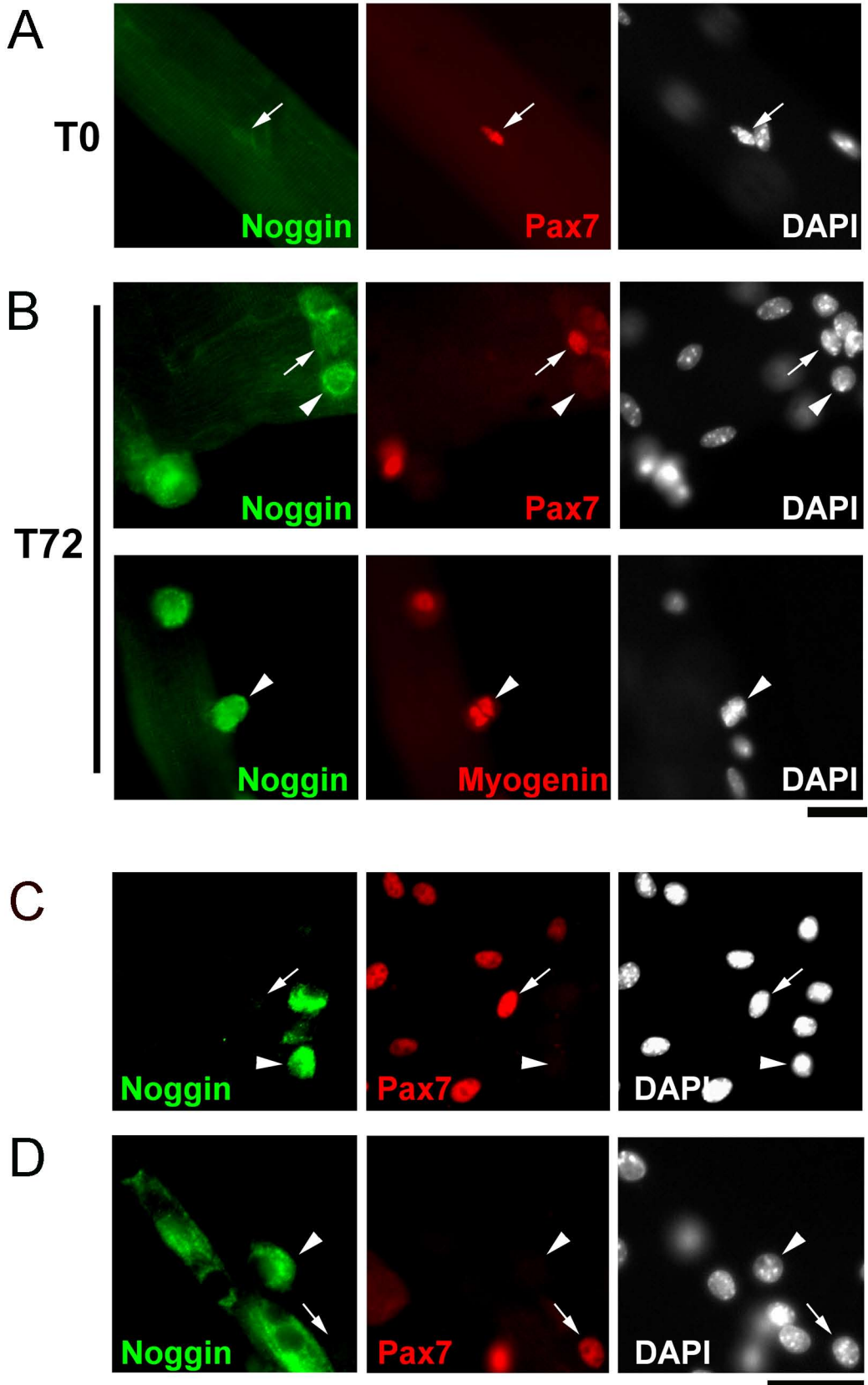


Figure 3

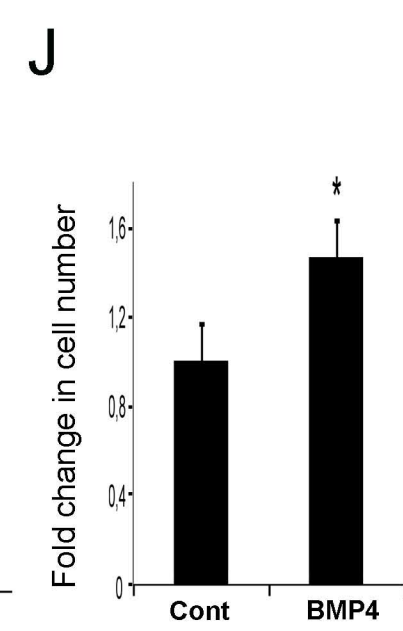
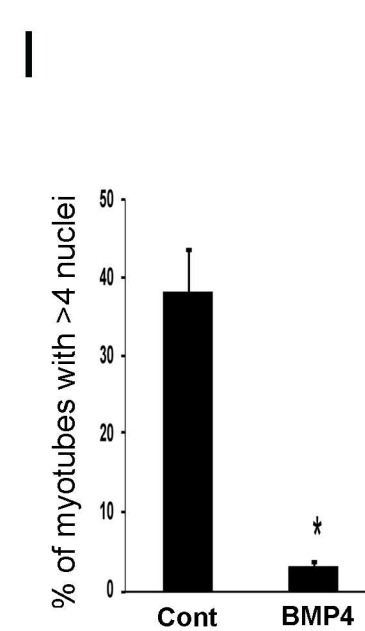
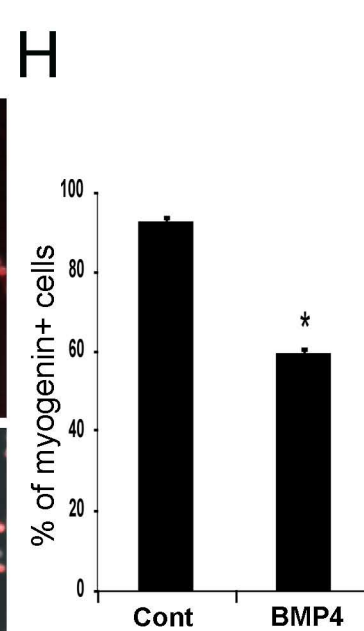
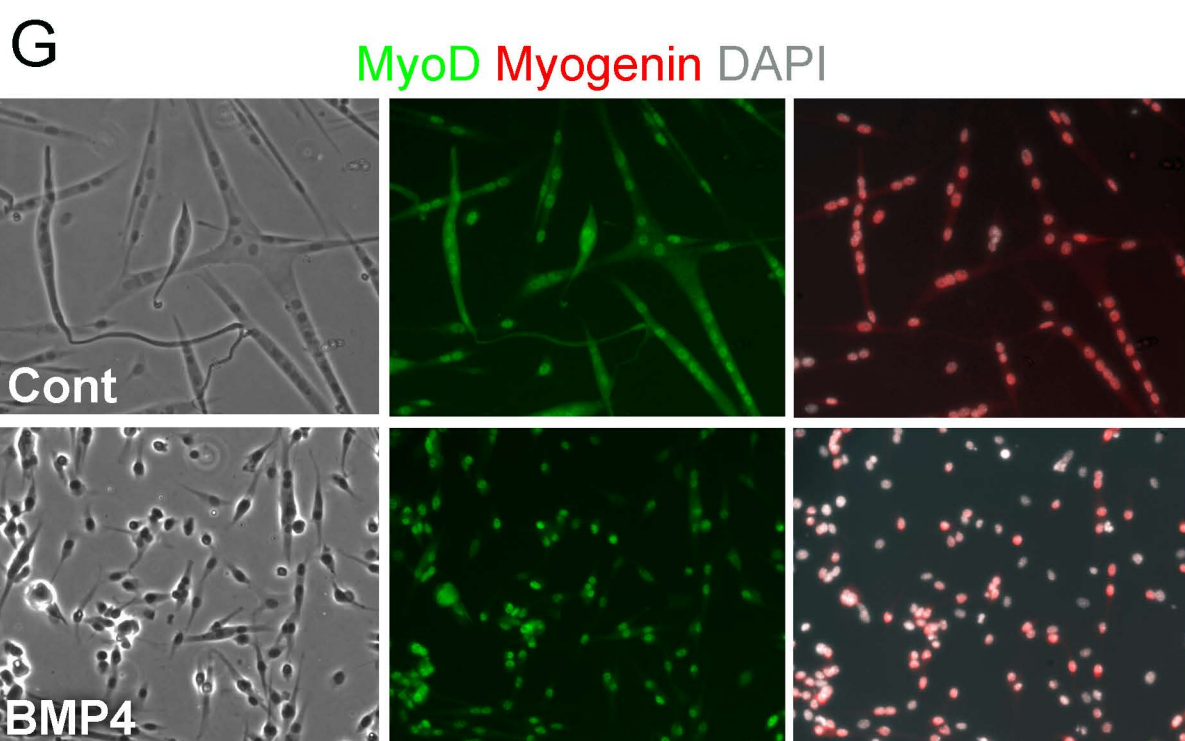
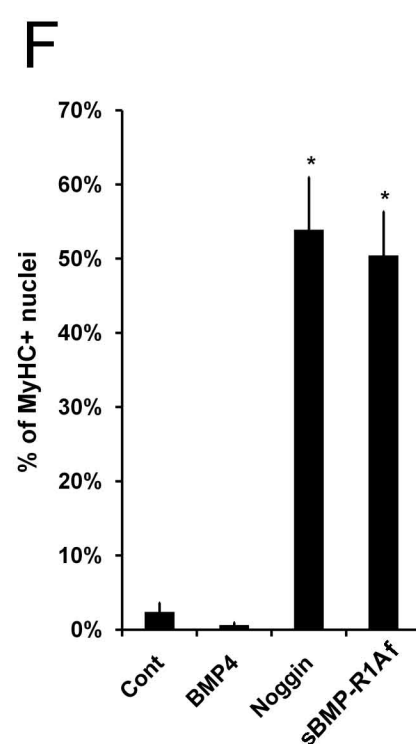
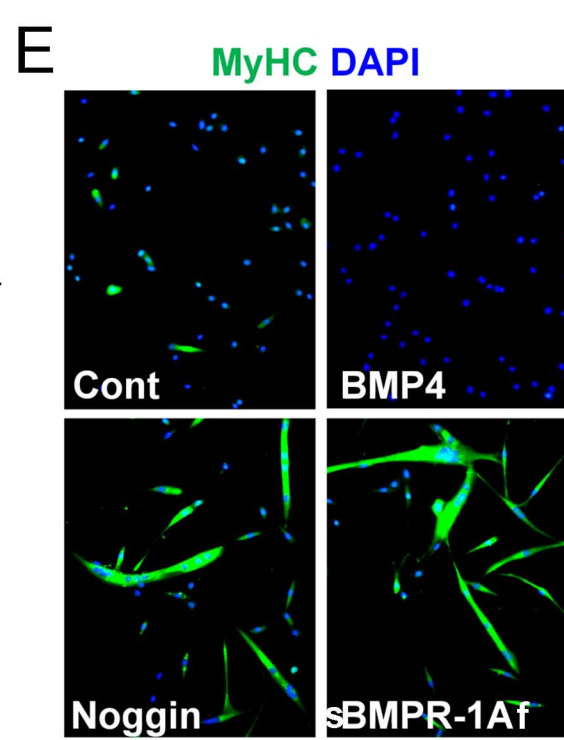
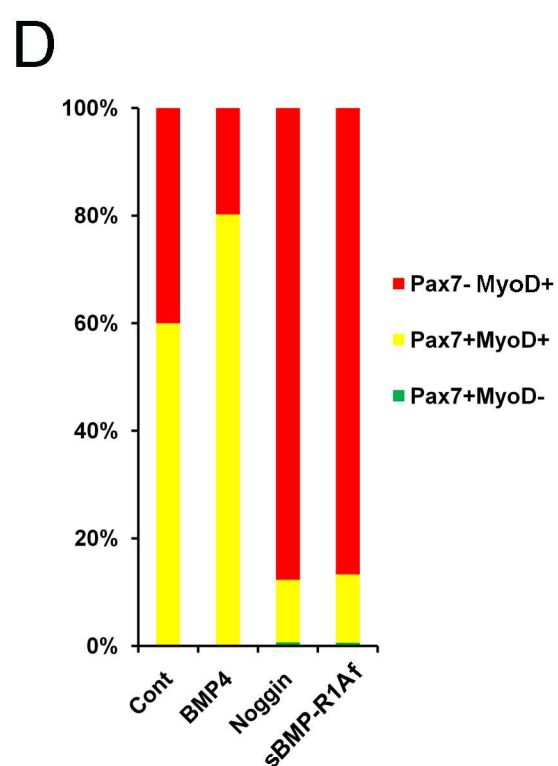
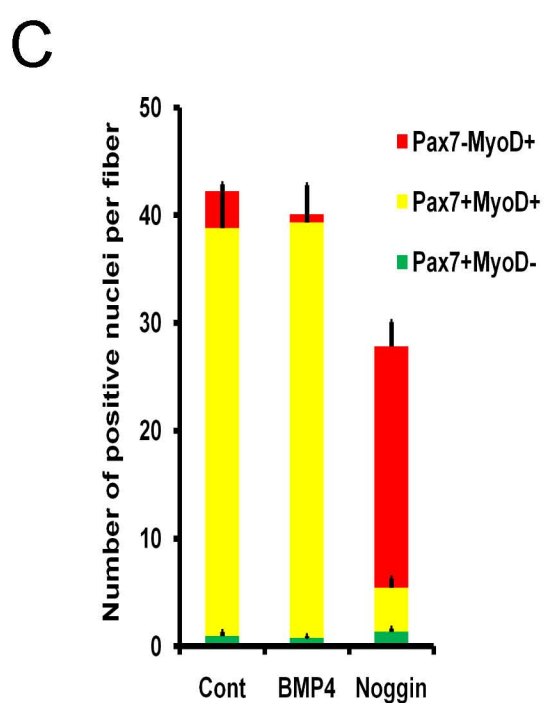
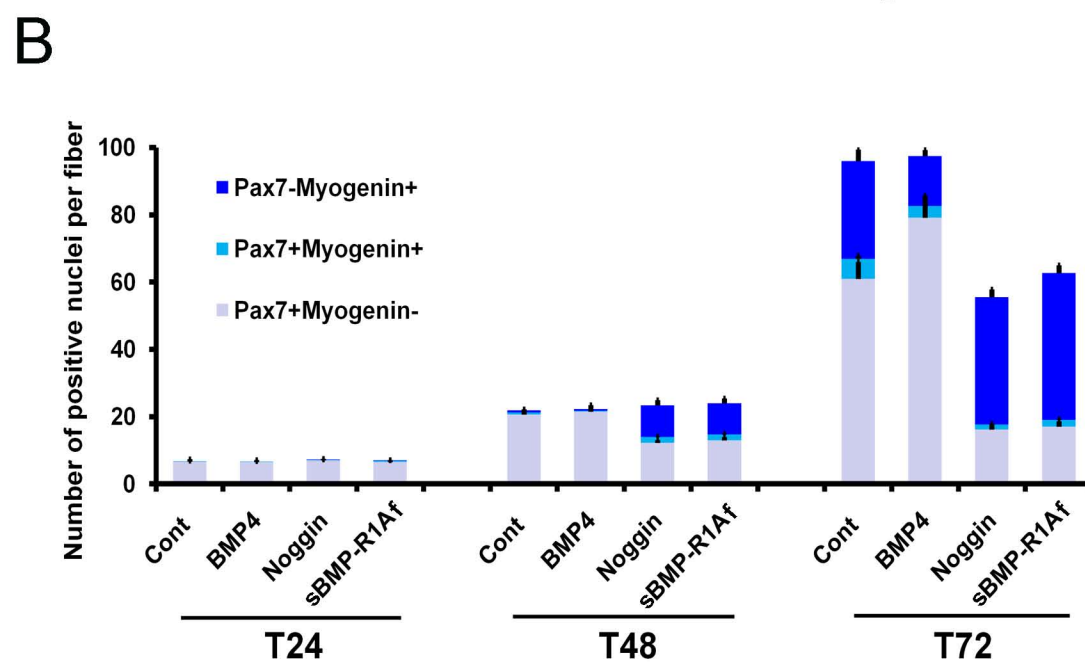
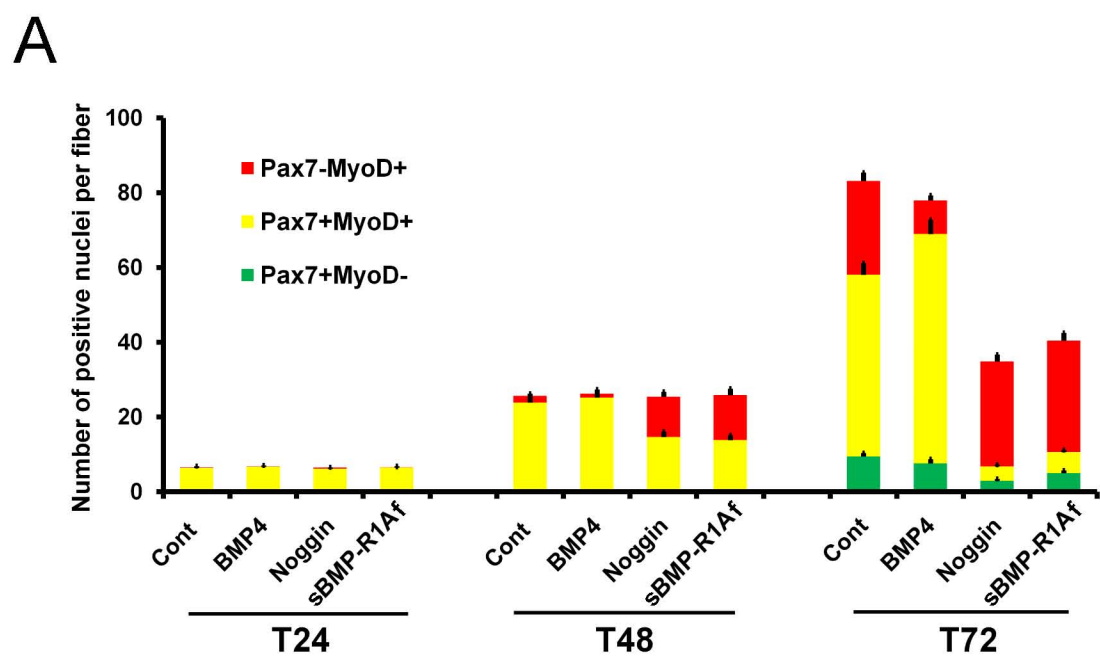
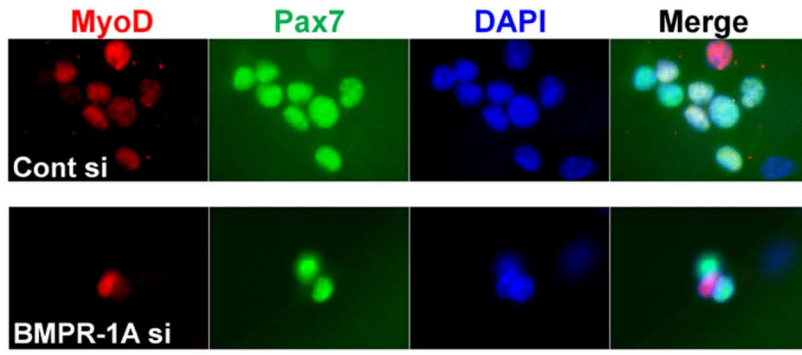
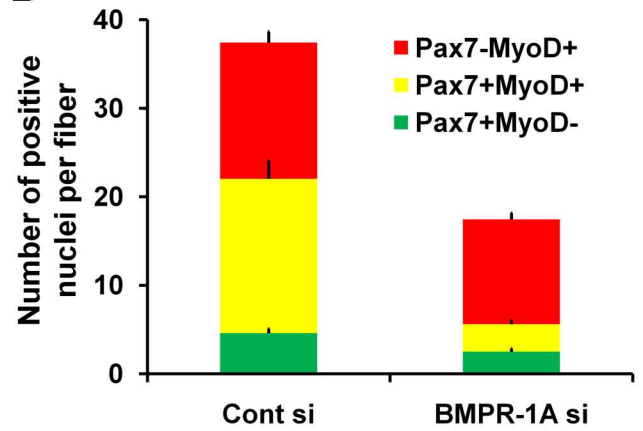


Figure 4

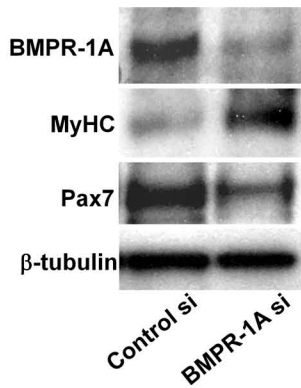
A



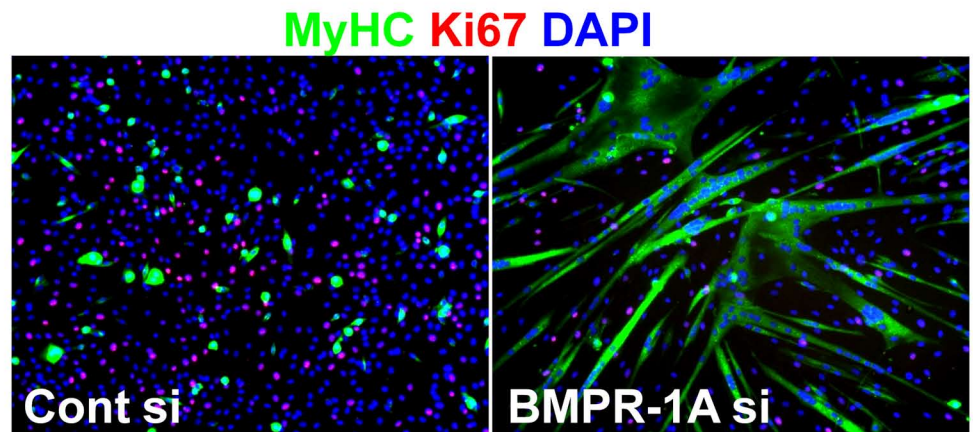
B



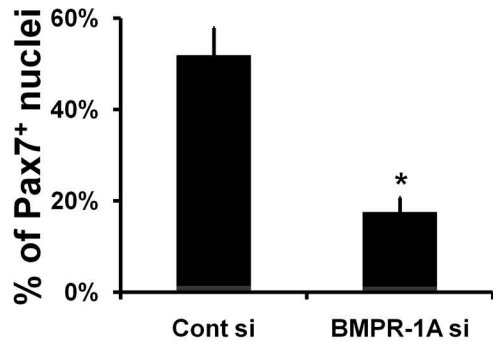
C



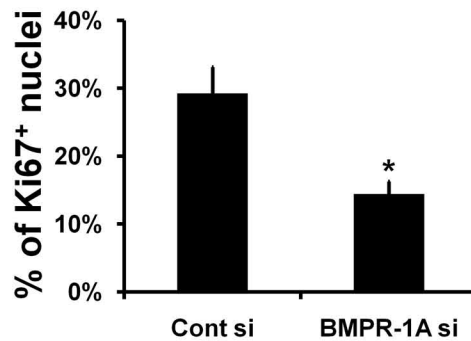
D



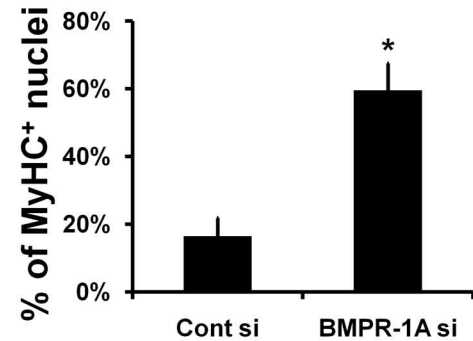
E



F



G



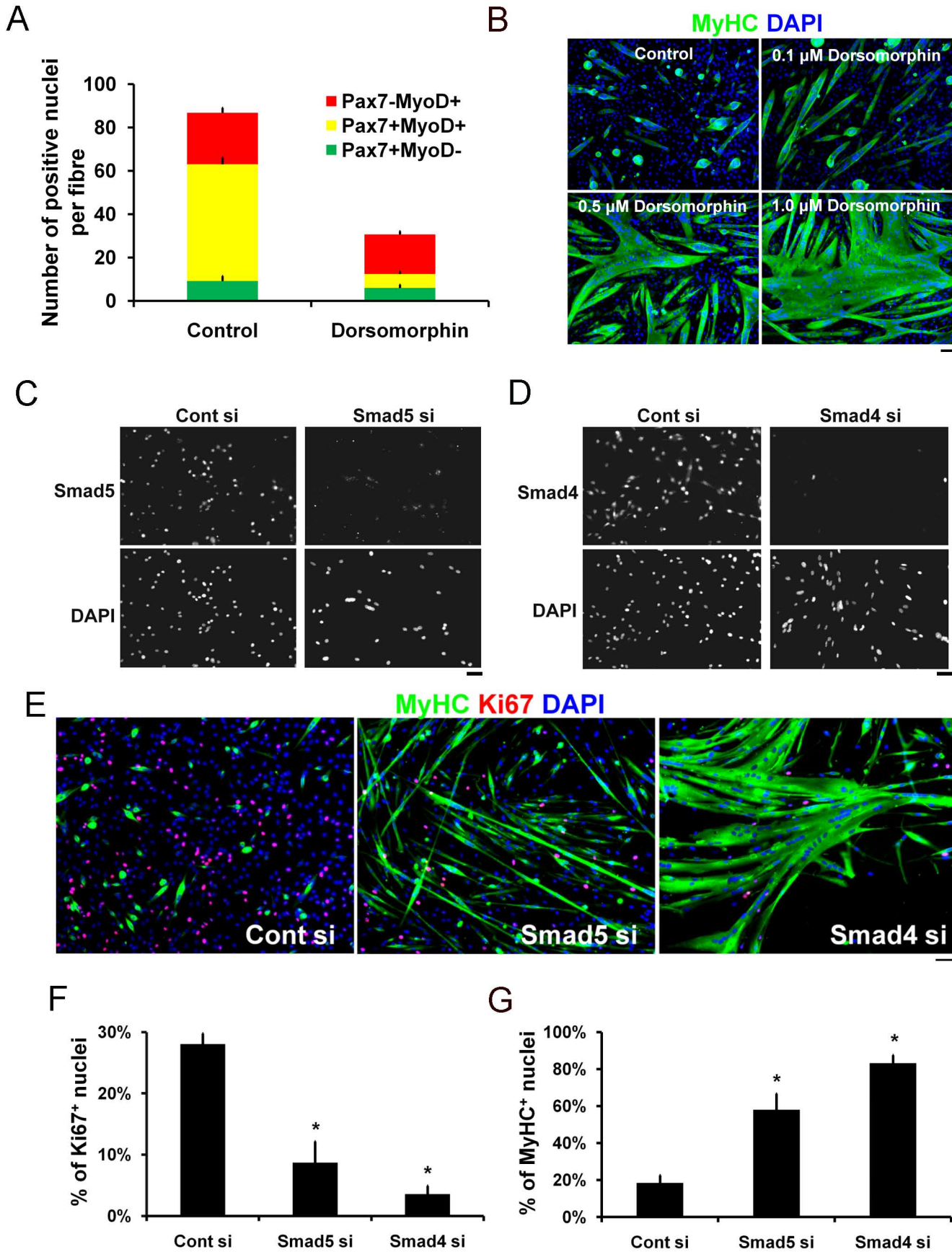
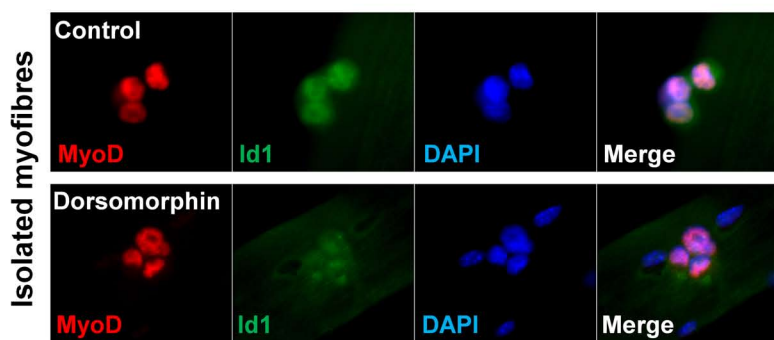
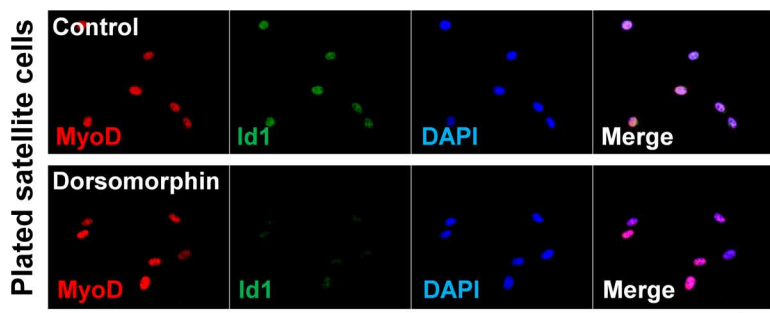


Figure 6

A



B



C

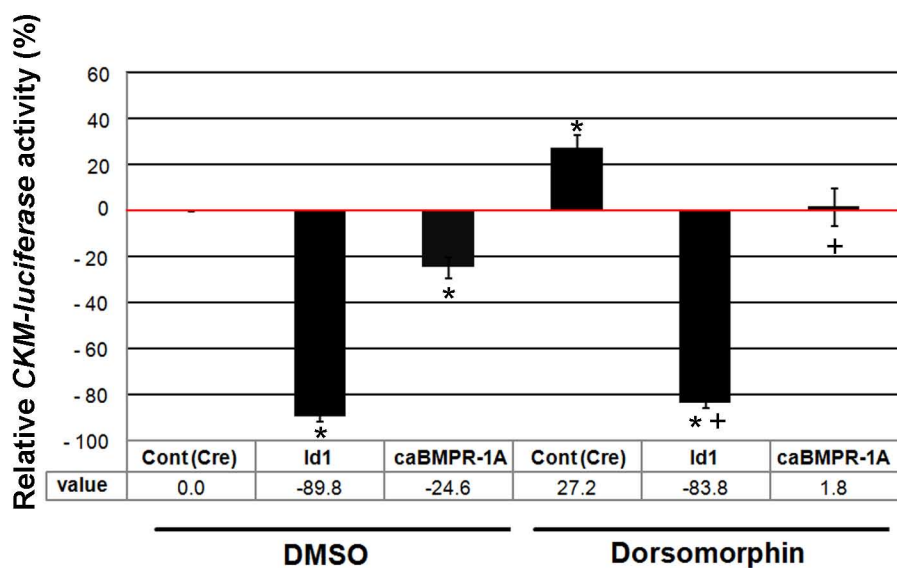


Figure 7

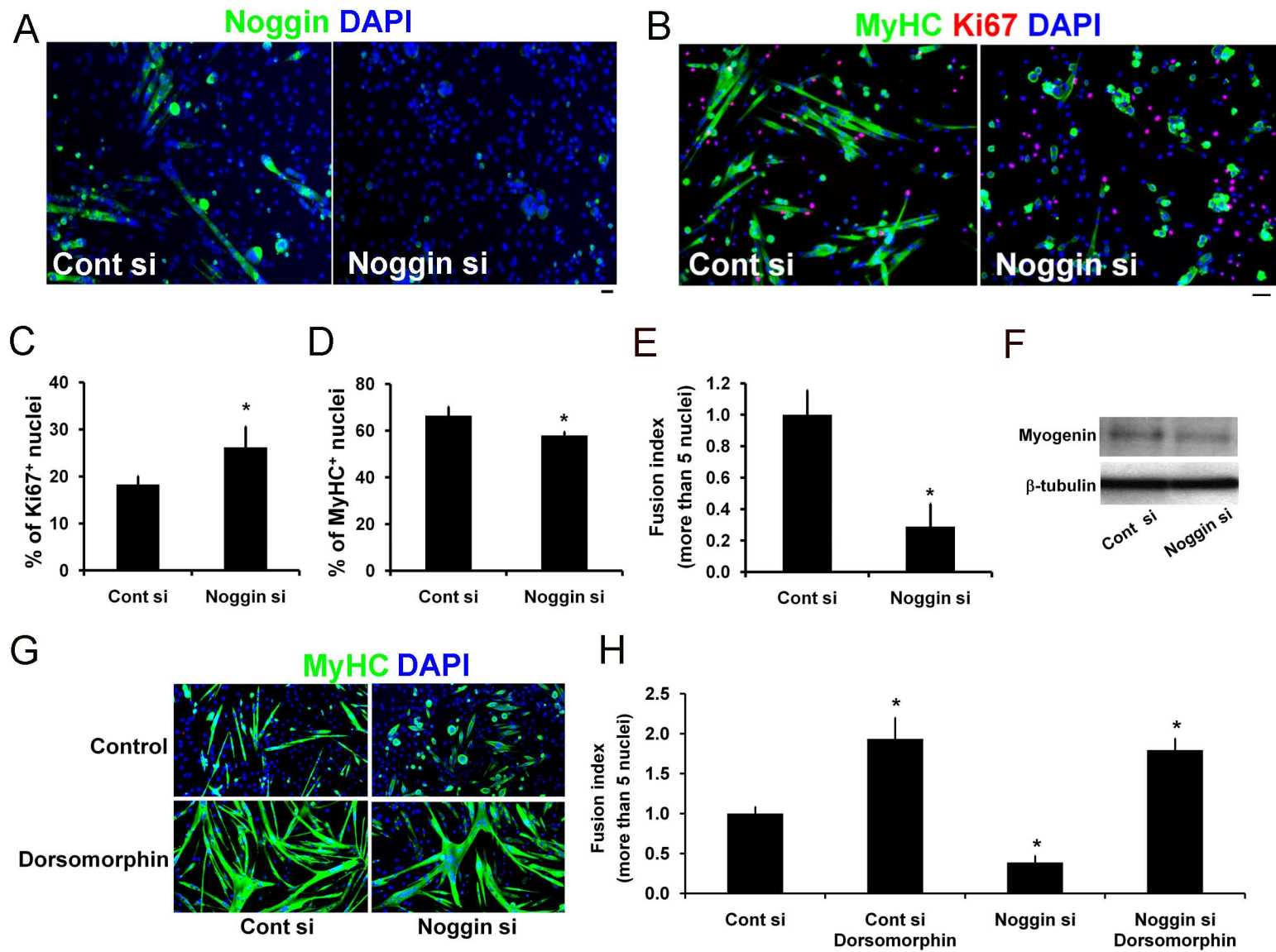


Figure 8

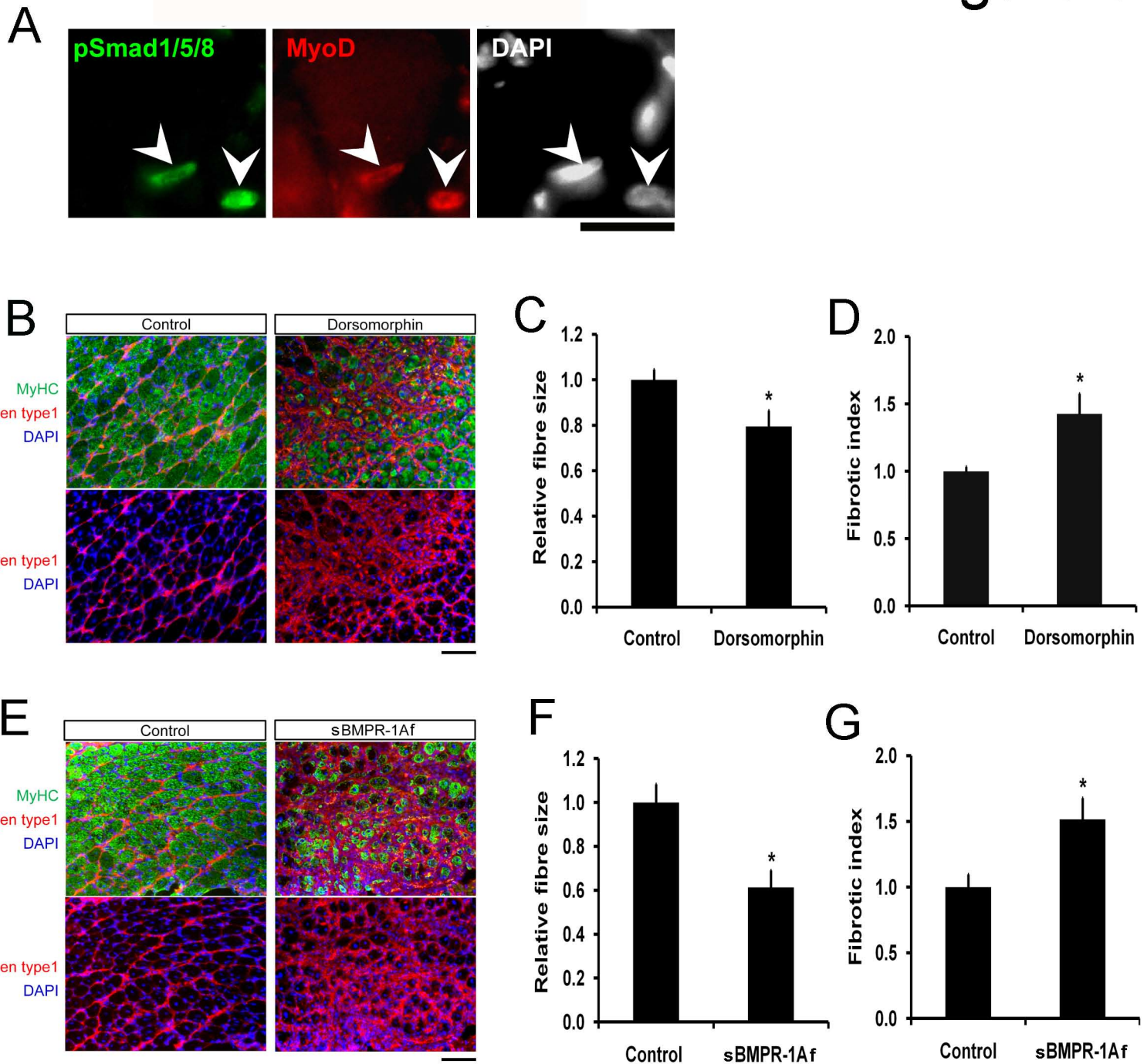
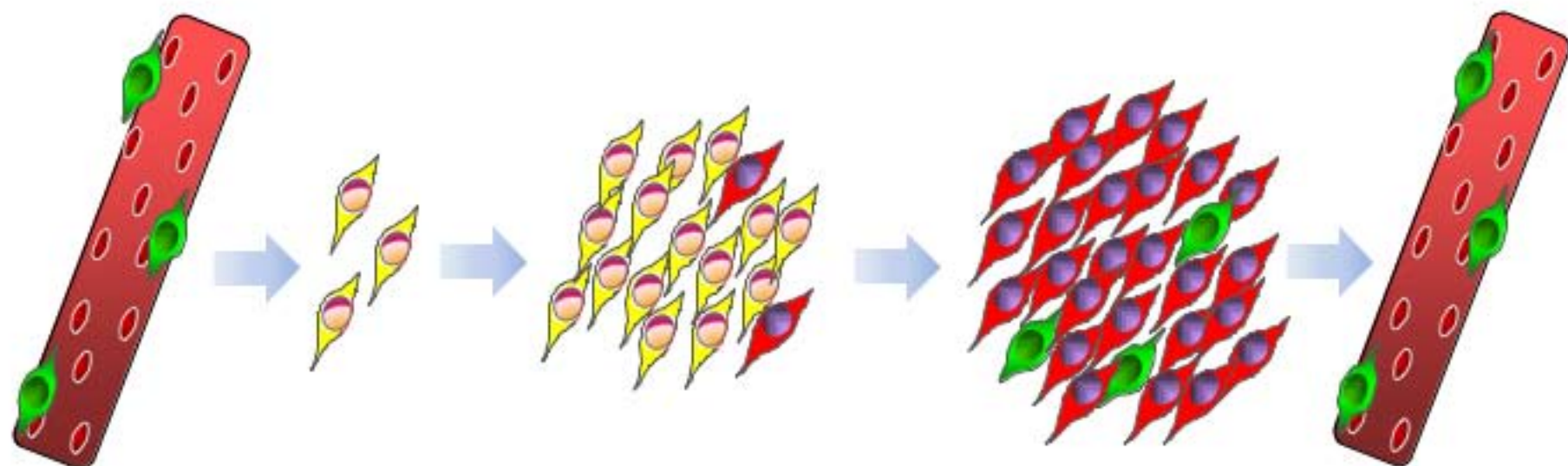


Figure 9






BMPR-IA expression

pSmad1/5/8 expression

Id1 expression

Noggin expression

BMP responsiveness

-  Quiescent or self-renewing cell (Pax7⁺MyoD⁻)
-  Proliferating cell (Pax7⁺MyoD⁺)
-  Differentiating cell (Pax7⁻MyoD⁺)