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11 **Marine Aggregates Facilitate Ingestion of Nanoparticles by Suspension-Feeding Bivalves**
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Abstract

As the application of nanomaterials to science and technology grows, the need to understand any ecotoxicological effects becomes increasingly important. Recent studies on a few species of fishes and invertebrates have provided data which suggest that harmful effects are possible. The way in which nanoparticles are taken up by aquatic organisms, however, has been little studied. We examined uptake of nanoparticles by two species of suspension-feeding bivalves (mussels, *Mytilus edulis*; oysters, *Crassostrea virginica*), which capture individual particles $< 1 \mu\text{m}$ with a retention efficiency of $< 15\%$. Given this limitation, it would appear that nanoparticles could not be ingested in large numbers. During certain times of the year, however, $> 70\%$ of suspended particles are incorporated within aggregates that are $> 100 \mu\text{m}$ in size. Therefore, we delivered bivalves fluorescently labeled, 100-nm polystyrene beads that were either (1) dispersed or (2) embedded within aggregates generated in the laboratory. Results indicate that aggregates significantly enhance the uptake of 100-nm particles. Nanoparticles had a longer gut retention time than 10- μm polystyrene beads suggesting that nanoparticles were transported to the digestive gland. Our data suggest a mechanism for significant nanoparticle ingestion, and have implications for toxicological effects and transfer of nanomaterials to higher trophic levels.

Keywords: nanoparticles, blue mussel, eastern oyster, aggregates, ingestion

1. Introduction

Manufactured nanoparticles (NPs) and nanotubes are at the forefront of nanotechnology and are being used in a variety of applications including cosmetics, electronics, drug delivery, manufacturing technologies, molecular biology, and paints (e.g., Harrison et al., 2000, Daniel and Astruc, 2004; Royal Society and Royal Academy, 2004; Kim et al. 2005; Engheta 2007; Schmid and Riediker, 2008). Nanomaterials with different compositions are being manufactured in increasing amounts, including those composed of carbon (e.g., single-walled nanotubes; fullerenes), germanium, transition metals (e.g., gold, palladium, platinum, silver), metal oxides (titanium dioxide, zinc oxide), polystyrene, and silica. Such widespread use will likely lead to their increased release into the environment, either directly via spills and use of products (direct emissions), or indirectly via landfills, waste incineration plants, and wastewater treatment facilities. Currently there are few data on environmental loads, but some estimates of use and release have been calculated. In Switzerland, 43 companies used quantities $> 1000 \text{ kg year}^{-1}$ per company (median quantity), with silver (Ag), aluminum oxide (Al_2O_3), iron oxides (e.g., Fe_2O_3), silicon dioxide (SiO_2), titanium dioxide (TiO_2), and zinc oxide (ZnO) being the most commonly used NPs (Schmid and Riediker, 2008). In Australia, more than 300 registered sunscreen products contain either nanoscaled TiO_2 or ZnO (Australian Gov., 2006). World-wide, some reports estimate that over 800 consumer products contain nanomaterials (Project on Emerging Nanotechnologies, 2008). Predicted environmental concentrations of nano-Ag and nano- TiO_2 range from 0.03 to $16 \mu\text{g L}^{-1}$, respectively (Mueller and Nowack, 2008). The concentration of TiO_2 NPs (20 & 300 nm) in runoff water that contacted painted facades was reported to be as high as $3.5 \times 10^8 \text{ particles L}^{-1}$ (Kaegi et al., 2008). Actual transport and retention of NPs in soils and wastewater treatment plants likely depends on properties of the surrounding media, including pH, ionic strength, and the concentration of organics and surfactants (LeCoanet et al., 2004; Limbach et al., 2008; Wang et al., 2008).

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2 Nonetheless, the above data suggest that freshwater and coastal environments are at risk of being
3
4 contaminated with manufactured NPs because of their proximity to human populations, industry
5
6 and wastewater discharge.
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9 Most research on the ecotoxicity of NPs has focused on mammals, and various effects have
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11 been reported and debated (e.g., Colvin, 2003; Hoet et al., 2003; Lam et al., 2006; Handy and Shaw,
12
13 2007). Surface chemistry, physiochemical properties, and reactivity of NPs, as well as
14
15 environmental factors, can affect interactions with cells and tissues (Sayes et al., 2004; Hardman,
16
17 2006). Even particles made of low-toxicity material (e.g., polystyrene, TiO₂) can be harmful when
18
19 delivered in their nano form, presumably due to the concomitantly high surface area (Oberdörster et
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21 al., 1994; Brown et al., 2001). Fewer studies have examined the effects of nanomaterials on aquatic
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23 organisms (Baun et al., 2008; Handy et al., 2008). Fullerenes (C₆₀) and single-walled carbon
24
25 nanotubes have been shown to produce adverse effects on the water flea (*Daphnia magna*;
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27 Oberdörster et al., 2006; Roberts et al., 2007), zebrafish embryos (Zhu et al., 2007; Usenko et al.,
28
29 2008), fathead minnow (Zhu et al., 2006), and rainbow trout (Smith et al., 2007). Using similar
30
31 concentrations, however, other workers have found no effects of these nanomaterials on freshwater
32
33 amphipods, harpacticoid copepods, or meiobenthic copepods (Oberdörster et al., 2006; Templeton
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35 et al., 2006). Nanoparticles containing metals (i.e., copper, gold, cadmium-telluride, silver) have
36
37 also been shown to have deleterious effects on zebrafish (Griffitt et al., 2007; Asharani et al., 2008),
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39 on two species of daphnids (Griffitt et al., 2008), and on two species of bivalves (Gagné et al., 2008;
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41 Tedesco et al., 2008). Nanoparticles of TiO₂ and ZnO can be toxic to *D. magna* (Adams et al.,
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43 2006), and TiO₂ NPs can cause oxidative stress in the rainbow trout (Federici et al., 2007).
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51 Additionally, because of their relatively high surface area, NPs are prime targets for adsorption and
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53 concentration of soluble pollutants such as arsenic (As), hydrocarbons, and metals (Cheng et al.,
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55 2004; Moore, 2006; Sun et al., 2007). Therefore, NPs have the potential to impact organisms
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2 directly through their inherent properties and also indirectly as a vehicle for other anthropogenic
3 materials. Consequently, particulate material from nanotechnology is currently under scrutiny as a
4 potential problematic anthropogenic pollutant (Hoet et al., 2003; Dreher, 2004; Royal Society and
5 Royal Academy, 2004; Hardman, 2006; SCENIHR, 2006).
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11 Although a growing number of studies are addressing the toxicity of manufactured
12 nanomaterials on aquatic organisms, most have lacked an ecological and physiological approach.
13
14 Almost nothing is known about the mode of uptake or ingestion rates of NPs, so the actual internal
15 exposure concentration is unknown. This fact may explain some of the discrepancies in reported
16 effects of NPs on aquatic organisms. Another gap in our current knowledge is the lack of data on
17 organ retention times. Knowledge of these processes would provide valuable information on
18 bioavailability of NPs and *in vivo* exposure times in the natural environment, and allow for more
19 realistic toxicity studies. Additionally, for commercially harvested aquatic organisms such
20 information is critical for an understanding of the potential transfer of NPs to humans.
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33 In this study we examined the ingestion rate and egestion dynamics of polystyrene NPs by two
34 species of bivalve molluscs. Suspension-feeding bivalves use their gills to capture particulate
35 matter. Particles greater than ca. 6 μm are captured with an efficiency $> 90\%$, whereas smaller
36 particles are captured with an efficiency that decreases asymptotically with decreasing size
37 (Riisgård, 1988). Although a few bivalve species can capture 1- μm particles with 50% efficiency
38 (e.g., *Geukensia demissa*), most species exhibit capture efficiencies of $< 15\%$ for particles $< 1 \mu\text{m}$ in
39 size (Ward and Shumway, 2004; Kach and Ward, 2008). Given such limitations of the feeding
40 organs, it would appear that bivalves would be at little risk of accumulating significant amounts of
41 NPs. However, particulate material in aquatic environments rarely exists as individual particles.
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43 Physical and biological processes operate to aggregate particles into larger masses (Alldredge and
44 Silver, 1988). At certain times of the year, a large proportion ($> 70\%$) of natural particulates of
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2 varying size and quality can form aggregates mixed with high molecular weight substances (flocs,
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4 marine snow; Alldredge et al., 1993; Crocker and Passow, 1995). Flocculation of phytoplankton,
5
6 detritus and other particles into rapidly sinking aggregates is an important mechanism of vertical
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8 transport of material to the benthos (Kjørboe et al., 1990; Passow and Wassmann, 1994; Crocker
9
10 and Passow, 1995). The aggregate pathway can facilitate the trophic transfer of dissolved and
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12 microparticulate material to bivalves (Alber and Valiela, 1994, 1996; Kach and Ward, 2008). We
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14 propose that this pathway also increases accessibility of NPs.
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21 **2. Materials and Methods**

22 *2.1 Production of aggregates*

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26 Aggregates were produced in the laboratory following the method outlined by Shanks and
27
28 Edmondson (1989). This roller-table method produces aggregates largely by differential settling,
29
30 one of several physical mechanisms that cause particles to collide in aquatic systems. The
31
32 procedures were modified slightly so that fluorescent NPs could be incorporated into the
33
34 aggregates. Briefly, natural seawater was passed through a 210- μm sieve to removed large
35
36 zooplankton and particles. Polystyrene NPs (red fluorescent, 100 nm; Thermo Fisher) were added
37
38 to the seawater at a concentration of ca. 1.3×10^4 particles ml^{-1} , along with $10 \mu\text{g l}^{-1}$ of hyaluronic
39
40 acid, a carbohydrate polymer that enhanced aggregation. The suspension was transferred to 12, 1-L
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42 plastic bottles. Six of the bottles were then placed on a roller table at 15 RPM for 4 days. The other
43
44 six bottles were placed next to the roller table to serve as controls (no aggregates). During the
45
46 rolling period, water samples were taken from the 1-L bottles to follow incorporation of NPs into
47
48 the aggregates. Incorporation was determined by allowing the aggregates in each rolled bottle to
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50 settle for 20 min and then taking a 20-ml sample from the center of the bottle. Bottles not being
51
52 rolled were inverted three times, allowed to stand for 20 min and then sampled. The peak
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2 fluorescence (PF) of these samples was measured on a fluorescence spectrophotometer (Hitachi F-
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4 2500; excitation = 303 nm, emission = 610 nm), and concentration of freely suspended NPs
5
6 determined using a standard curve generated by measuring the fluorescence of known
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8 concentrations of NPs (see 2.4 below). Incorporation of NPs into aggregates was then determined
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10 by calculating the percent difference from the start concentration.
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16 2.2 Feeding Experiments

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18 We examined ingestion of NPs by two species of bivalves, the mussel (*Mytilus edulis*) and
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20 the oyster (*Crassostrea virginica*). These two species possess ctenidia with a unique combination
21
22 of architecture and complexity of laterofrontal (LF) ciliary tracts, which aid in particle capture
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24 (Ward et al. 1998). Mussels (ca. 5.1 cm in shell length) and oysters (ca. 5.6 cm in shell height)
25
26 were collected three days before experimentation and their outer shells scrubbed to remove living
27
28 macro-organisms. Animals were then attached to wooden craft sticks using Velcro[®], so that they
29
30 could be suspended in the experimental bottles. Using marine epoxy, one side of the Velcro[®] was
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32 secured to the shell and the other to a craft stick. Animals were maintained in flowing-seawater
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34 tables in the Rankin Laboratory on the Avery Point campus, University of Connecticut (Groton, CT,
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36 USA), until they were used in the experiments.
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42 Experiments were conducted in an environmental chamber held at ca. $15^{\circ} \pm 0.5$ C, a
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44 temperature consistent with ambient conditions measured in the holding tanks in the Rankin
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46 Laboratory. Two different experiments were conducted for each species. In experiment 1, twelve
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48 individuals were used with six being exposed to NPs incorporated in aggregates and six to freely
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50 suspended NPs. In experiment 2, twelve individuals were exposed to NPs incorporated in
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52 aggregates and six to freely suspended NPs. For all experiments, each animal was introduced into
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54 either a bottle with aggregates or a bottle with NPs freely suspended (control), and its craft stick
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2 secured to the lip of the container with a wooden clip. A stir bar was added and the bottle placed on
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4 an electromagnetic stir plate. To ensure that aggregates and particles remained suspended, each
5
6 bottle was supplied with gentle aeration and stirred for 10 sec every 15 min. Just prior to the start of
7
8 a trial, 10- μm , non-fluorescent polystyrene beads were added to each bottle at a final concentration
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10 of 1000 beads ml^{-1} . These beads were in the size range that could be captured by each species with
11
12 an efficiency of about 100%, allowing for the quantification of feeding rates. Additionally, two
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14 animals of each species were delivered only 10- μm , non-fluorescent beads and served as
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16 experimental blanks. Animals were allowed to feed for 45 min, with start times beginning when the
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18 individual animal opened its shell and appeared to be feeding (siphons or mantle extended).
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26 *2.3 Feces collection and preparation*

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28 Egestion dynamics were investigated after the exposure to NPs. Animals were removed
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30 from the bottles and placed in individual, 1-L beakers filled with filtered seawater. For experiment
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32 1, all feces produced over the next 72 hours were collected from each animal and placed in separate
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34 vials. For experiment 2, separate fecal collections were made for three animals after 6, 24, 48 and
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36 72 hours (individuals of each species being sacrificed at each time interval). Again, feces from each
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38 individual were placed in a separate vial. These samples served as a time series for egestion. The
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40 microalga *Tetraselmis* sp. was added to each beaker (final concentration = ca. 1×10^4 cells ml^{-1})
41
42 after 24 and 48 hours to stimulate transit of material through the digestive tract. The collected fecal
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44 samples were placed in individual, 15-ml falcon tubes and centrifuged for 10 minutes at 1320 rcf
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46 (g). The seawater supernatant was decanted and the pellet resuspended in 6 ml of deionized water
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48 (DI), spun for another ten minutes, and again decanted. This washing process was repeated two
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50 additional times to remove salts which react with sodium hydroxide (NaOH) to form a precipitate.
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52 After the final wash, 2.0 ml of 1N NaOH were added to each tube. Digestive-gland samples were
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2 patted with laboratory tissue, dried in a desiccator and weighed. Samples were placed in individual
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4 falcon tubes and treated with 2.0 ml of 1N NaOH. The fecal and digestive-gland samples were then
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6 sonicated for five minutes, resuspended by means of a Vortex Genie[®] and allowed to digest for
7
8 seven days. Samples were then diluted with 6.0 ml of DI water to bring the total volume of each to
9
10 ca. 8.0 ml.
11
12

13 14 15 16 *2.4 Spectrophotometric analysis* 17

18
19 Material in each falcon tube was re-suspended and a 2.0-ml sub-sample removed. The peak
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21 fluorescence (PF) of each sub-sample was measured by means of a fluorescent spectrophotometer
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23 (excitation = 304 nm, emission = 610 nm), and the concentration of NPs determined using a
24
25 standard curve. The 9-point, linear standard curve was generated by suspending known numbers of
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27 NPs in a fixed volume of DI water and determining PF on the fluorescent spectrophotometer. To
28
29 confirm the accuracy of the equations, feces and digestive glands were obtained from animals not
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31 used in the experiments and digested in NaOH. Known numbers of NPs were suspended in the
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33 resulting liquid and analyzed.
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38 Total number of NPs egested was then calculated for each animal, and ingestion rates
39
40 estimated by dividing by the pumping rate of the individual. To calculate pumping rate, a 1-ml sub-
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42 sample was removed from each falcon tube and the number of 10- μ m beads (non-fluorescent)
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44 counted on a hemocytometer. The total number of beads egested represents the total number
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46 ingested. As these beads could be captured at about 100% efficiency (Møhlenberg and Riisgård,
47
48 1978; Riisgård, 1988), pumping rate (PR) could be calculated using the following equation (Kach
49
50 and Ward, 2008):
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52

$$53$$
$$54 \text{ PR (ml h}^{-1}\text{)} = [\text{beads ingested (No.)} / \text{initial bead concentration (No. ml}^{-1}\text{)}] / \text{time (h)} \quad (1)$$
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3. Results

Visible aggregates ranging in size from $> 100 \mu\text{m}$ to several mm always formed in the rolled bottles. No such aggregates formed in the bottles that were not rolled, although some nanoparticle loss did occur ($< 2.5\%$). Additionally, data from preliminary trials indicated that some NPs ($< 1.8\%$) adhered to the sides of rolled bottles. Therefore, incorporation of NPs into aggregates was adjusted for these minor losses. For all experiments, the efficiency at which NPs were incorporated into aggregates ranged from 52 to 64%. Visual inspection of our laboratory-produced aggregates showed that they differed little from natural aggregates in size and shape (see Kach and Ward, 2008). These observations corroborate the study of Shanks and Edmondson (1989) who demonstrated that aggregates produced on a roller table were a good representation of natural aggregates.

Both species of bivalves ingested NPs at a significantly higher rate when particles were incorporated in aggregates compared to when they were freely suspended (2-way ANOVA, GLM, $p < 0.01$; Fig. 1). In fact, the number of freely suspended NPs ingested by the bivalves was so low that no fluorescent activity could be detected. Pumping rate, however, for mussels in both treatments (aggregate = $0.360 \pm 0.018 \text{ L hr}^{-1}$; no aggregate = $0.356 \pm 0.018 \text{ L hr}^{-1}$) and oysters in both treatments (aggregate = $0.319 \pm 0.014 \text{ L hr}^{-1}$; no aggregate = $0.326 \pm 0.010 \text{ L hr}^{-1}$) were similar. For the aggregate treatment, mussels ingested NPs at a significantly higher rate than oysters (Bonferroni multiple comparison, $p < 0.01$).

For both species of bivalves, egestion of $10\text{-}\mu\text{m}$ beads over time displayed an opposite trend compared to egestion of NPs over time (Fig. 2). Whereas the highest number of $10\text{-}\mu\text{m}$ beads was egested after 6 hours, the highest number of NPs was egested after 72 hours. Number of $10\text{-}\mu\text{m}$ beads egested decreased over time, whereas number of NPs egested increased over time. No fluorescence was detected in fecal samples from the experimental blanks.

4. Discussion

Our data clearly indicate that mussels and oysters more efficiently capture and ingest NPs that are incorporated into aggregates compared to those freely suspended. As reported for mucus-bound food material (Ward et al., 1993; Ward et al., 1994), aggregates are likely broken down by the action of cilia on the gills and labial palps and the constituent particles ingested. Some aggregated material is likely to be rejected as pseudofeces, but our results demonstrate that a large fraction is ingested. Time series experiments show that polystyrene NPs have a longer gut retention time (GRT) compared to 10- μm polystyrene particles. Previously studies report that > 85% of > 5- μm polystyrene, silica, and aluminum beads are often egested after ca. 30 h (Ward and Target, 1989; Cranford et al., 1998; Brilliant and MacDonald, 2002). Bivalves are capable of particle selection in the gut (Brilliant and MacDonald, 2000, 2002, 2003), and longer GRT for food usually indicates that the material has undergone more extensive extracellular digestion and, perhaps, been transported to the digestive glands for more complete intracellular digestion. In the present case, the longer GRT suggests that most NPs were directed into the tubules of the digestive glands and potentially taken up by the digestive cells via endocytosis. Sucrose-polyester NPs can be taken up by isolated digestive-gland cells of blue mussels (*Mytilus edulis*) and by whole animals (Moore et al., 1997). Cadmium-telluride quantum dots can produce cellular damage in the digestive gland of the freshwater bivalve *Elliption complanata* (Gagne et al., 2008), and Au and SiO₂ NPs can produce harmful effects in the digestive gland of *M. edulis* (Koehler et al., 2008; Tedesco et al. 2008). Our data from analysis of the digestive gland and gut confirm the presence of fluorescent NPs in these tissue 72 hours after the experiments. Consequently, the calculated ingestion rates of NPs should be considered minimal values.

1
2 Results of the present study have implications for experimental as well as natural exposure
3 of bivalves to NPs. In laboratory assays that expose bivalves to a known concentration of freely
4 suspended NPs, the uptake and thus bioavailability of these particles will be very low. Even
5 aggregates of NPs up to 0.5 μm will have a low bioavailability (Kach and Ward 2008). Although
6 some particles could perhaps be taken up across the epithelium of the gill, mantle and labial palps
7 (Kashiwada, 2006; Koehler et al., 2008), the major route for internal exposure and potential effects
8 is via capture and ingestion. Consequently, the actual exposure levels will likely be much lower
9 than those calculated based on mass or number per volume of water delivered to the animals. In
10 fact, actual uptake will be a consequence of at least two factors: 1) the form in which the NPs are
11 delivered (e.g., monodispersed, slightly aggregated, highly aggregated), and 2) the capture
12 efficiency of the particular bivalve species for the delivered particle masses (e.g., nanoparticle
13 masses $< 5 \mu\text{m}$ will be captured and ingested with different efficiencies). These factors need to be
14 quantified for each experiment to allow comparison of results among studies that utilized bivalves.
15 For example, of the several studies that have examined the effects of manufactured NPs on bivalves
16 (Koehler et al., 2008; Tedesco et al., 2008), only Gagné et al. (2008) mention that aggregation of
17 particles occurred. In this study, the authors indicate that cadmium-telluride quantum dots tended to
18 aggregate at medium (4 mg L^{-1}) and high (8 mg L^{-1}) concentrations. If so, then the aggregated
19 quantum dots probably were ingested by freshwater mussels at a higher rate than those not
20 aggregated (i.e., at 1.6 mg L^{-1}). Thus, the dose-response trend observed by Gagné et al. (2008)
21 could have been confounded by the possibility that quantum dots at higher concentrations were
22 more available to mussels over the experimental period.

23
24 Under natural conditions, self-aggregation of NPs into larger particle masses and
25 incorporation of NPs into aggregated material will increase the bioavailability to suspension-feeding
26 bivalves. The adsorption of organic material by NPs can alter their physiochemical properties and
27

1
2 lead to enhanced aggregation (Brant et al., 2005; Moreau et al., 2007; Li et al., 2008; Xie et al.,
3
4 2008). Not only would incorporation of NPs into aggregates enhance uptake and internal exposure,
5
6 but aggregates would tend to sink faster than their constituent NPs (Kriest and Evans, 1999; Waite
7
8 et al., 2000). Higher sinking rates would increase the flux of NPs to the benthos, exposing
9
10 suspension and deposit feeders to a large concentration of these materials. Additionally,
11
12 nanomaterials are thought to be persistent in the environment (e.g., SCENIHR, 2006), so
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14 resuspension events, such as storms, could re-expose bivalves to previously deposited NPs, causing
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16 spikes of uptake and accumulation.
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23 **5. Conclusion**

24
25 The promise of nanomaterials for science, technology and the economy is great, but the
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27 ecotoxicological effects and pathological risks to animals and humans may be equally large (Colvin,
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29 2003; Royal Society and Royal Academy of Engineering, 2004). In order to define the effects of
30
31 nanoparticle exposure on benthic organisms and develop reliable biomarkers, the uptake and
32
33 ingestion dynamics by benthic animals, and visceral mass retention times need to be determined.
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35 Evaluating the environmental risk of these materials will require knowledge of the concentrations in
36
37 the environment as well as the form in which they exist (e.g., dispersed, attached to particles,
38
39 incorporated in aggregates) and their distribution in the pelagic vs benthic realms.
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44
45 Bivalves are ideal sentinel organisms because they process large volumes of water per unit
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47 time (e.g., 3-9 L h⁻¹ g dry mass⁻¹; Newell et al., 2005). The amount of suspended material removed,
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49 however, is a function of particle size and retention efficiency of the particular species. The
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51 aggregate pathway can facilitate the trophic transfer of dissolved and microparticulate material to
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53 bivalves (Alber and Valiella, 1994, 1996; Kach and Ward, 2008), and we have demonstrated that
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55 this pathway can also significantly enhance the ingestion of NPs. Our findings have implications
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1
2 for the accumulation of NPs in bivalves, and the transfer to higher trophic levels including humans.
3
4 Future studies should examine the incorporation rates of NPs into marine aggregates, as well as the
5
6 uptake dynamics of nanoparticle-laden aggregates by suspension and deposit feeding animals in
7
8 order to better understand the impacts of these materials on aquatic ecosystems.
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10

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14
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2 **Figure captions**
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7 **Figure 1.** Ingestion rate of polystyrene nanoparticles (NPs, 100 nm) by the mussel (*Mytilus edulis*)
8 and oyster (*Crassostrea virginica*). For both species, aggregate-bound NPs were ingested at a
9 significantly higher rate than freely suspended NPs (no aggregates). Mussels ingested aggregate-
10 bound NPs at a significantly higher rate than oysters. Bars with different letters are significantly
11 different at $p < 0.01$. Data are means \pm SD for 6 replicate animals. ND = not detected.
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21 **Figure 2.** Egestion of polystyrene nanoparticles (NPs, 100 nm) and 10- μ m beads by **A.** the mussel
22 (*Mytilus edulis*), and **B.** the oyster (*Crassostrea virginica*). Both particle types were delivered to
23 bivalves at the same time, but NPs were incorporated in aggregates. Notice that egestion of 10- μ m
24 beads decreases with time whereas egestion of NPs increases with time. End point for NP egestion
25 was not reached. Data are means \pm SD for 12, 9, 6, and 3 replicate animals at times 6, 24, 48 and 72
26 hours, respectively. ND = not detected.
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