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Poriferan survivin exhibits a conserved regulatory role in the interconnected pathways of cell cycle and apoptosis

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Running title: Conserved dual role of survivin

Abbreviations: aa, amino acid; ABTS, 2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid); Bcl-2, B-cell lymphoma-2; BIR, baculoviral IAP repeat; CDE, cell cycle dependent elements; Ced-9/-3, cell death abnormality -9/-3; CHR, cell cycle homology regions; CPC, chromosomal passenger complex; CRB-1, crumbs homolog 1; CRM1, chromosome region maintenance 1; DIG, digoxigenin; DMF,

dimethylformamide; dys-1, dystrophin-1; ELISA, enzyme-linked immunosorbent assay; IAP, inhibitors of apoptosis protein; LMB, leptomycin B; LSM, laser scanning microscopy; ML, maximum likelihood; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NES, nuclear export signal; NJ, neighbor-joining; ORF, open reading frame; PBS, phosphate buffered saline; PFA, paraformaldehyde

The following sequences from *S. domuncula* have been deposited in the EMBL/GenBank: SDSURVL (accession number FM210333 and FM210334, cDNA and genomic sequence respectively) and SDCASL2 (FM210332).

Abstract

Survivin orchestrates intracellular pathways during cell division and apoptosis. Its central function as mitotic regulator and inhibitor of cell death has major implications for tumor cell proliferation. Analyses in early-branching Metazoa so far propose an exclusive role of survivin as a chromosomal passenger protein, whereas only later during evolution a complementary anti-apoptotic function might have arisen, concurrent with increased organismal complexity. To lift the veil on the ancestral function(s) of this key regulator, a survivin-like protein (SURVL) of one of the earliest-branching metazoan taxa was identified and functionally characterized. SURVL of the sponge *Suberites domuncula* shares considerable similarities with its metazoan homologues, ranging from conserved exon/intron structure to presence of protein-interaction domains. Whereas sponge tissue displays a low steady-state level, SURVL expression was significantly up-regulated in rapidly proliferating primmorph cells. In addition, challenge of tissue and primmorphs with heavy metal or lipopeptide stimulated SURVL expression, concurrent with the expression of a newly discovered caspase. Complementary functional analyses in transfected HEK-293 cells revealed that heterologous expression of a SURVL-EFGP fusion not only promotes proliferation but also enhances resistance to cadmium-induced cell death. Taken together, these results suggest both a deep evolutionary conserved dual role of survivin and an equally conserved central position in the interconnected pathways of cell cycle and apoptosis.

Introduction

Apoptosis plays a crucial role not only in countless physiological processes (e.g., elimination of redundant cells during vertebrate embryonic development) but also during pathogenesis. Since apoptosis relies on the clear-cut function of numerous pro- and anti-apoptotic factors, any malfunction on their part or disruption of their respective interplay inevitably leads to dysfunctional cellular metabolism. The ensuing delayed or premature cell death often manifests in severe pathologies, including carcinogenesis and neurodegenerative diseases.

The groundbreaking discovery of the *Caenorhabditis elegans* proteins Ced-9, homologue of human anti-apoptotic Bcl-2, and Ced-3, homologue of human caspase-1, has demonstrated the molecular conservation of the apoptotic cell death program along a broad range of Metazoa, from nematodes to vertebrates.¹ Since then, the fundamental role of apoptosis has been increasingly recognized in various physiological processes. In addition, several invertebrate model organisms revealed a significant potential for analyses of apoptotic mechanisms, not only under an evolutionary point of view. Thus, the identification of the human crumbs homologue (CRB-1) and its causative role in the manifestation of retinitis pigmentosa was based on the discovery of mutational variants of the *Drosophila melanogaster* protein crumbs.² In addition, loss of function mutations of the *C. elegans* dys-1 gene confirmed the role of human dystrophin in the manifestation of the neuromuscular disease of Duchenne.³ During the last years a novel invertebrate model system has emerged: sponges (phylum Porifera), which are among the earliest-branching non-bilaterian animals.⁴ Recent biomarker analyses date the occurrence of probable stem-group poriferans back before the end of the Marinoan glaciation (~635 Myr ago)⁵ and molecular clock analyses estimated that the main poriferan lineages

diverged in the Cryogenian/Ediacaran⁶, well before the rapid appearance of major metazoan groups (Cambrian Explosion).

Until today, the model system "Porifera" has revealed several characteristics that have proven valuable for evolutionary studies within a comparative context. The major avail is the phylogenetically basal position of Porifera allowing a unique vantage point on the hypothetical common ancestral taxon, the Urmetazoa. Moreover, comparative transcriptomics and proteomics divulged several examples of poriferan molecules, genetic features, protein domains, etc. that remained conserved throughout evolution, but otherwise have been lost in invertebrate model systems traditionally used, *C. elegans* and *D. melanogaster*.⁷ Furthermore, the majority of poriferan genes display a closer sequence similarity to their human homologues than to the respective molecules of worm and fly (similar to some anthozoans), probably caused by an accelerated evolution of the latter animal lineages.^{8,9}

The fine-tuned balance between apoptosis and proliferation is upheld via numerous regulatory molecules that maintain tissue integrity and, ultimately, ensure survival of the organism. The inhibitors of apoptosis (IAP) family comprises structurally related proteins (the presence of at least one baculoviral IAP repeat (BIR) domain is mandatory) that were originally discovered in baculovirus. In addition to their cell death regulating activities several IAP have a complementary role in cell division, in particular attributed to survivin. Accordingly, the dual role of human survivin is based upon its function as (i) a chromosomal passenger protein that controls segregation of chromatin/regulation of cytokinesis and (ii) a caspase inhibitor that prevents processing/activation of caspases.¹⁰ Since transcription from its single gene locus is up-regulated in most tumor cell types, survivin is a well-established analytical marker of neoplasms.^{11,12} The function of survivin homologues

in non-bilaterian Metazoa is much less well-defined. Accordingly, similar to vertebrate survivin the *D. melanogaster* homologue deterin deters cells from caspase-induced apoptosis and some studies propose an additional pro-proliferative effect.¹³ On the other hand, the *C. elegans* survivin homologue BIR-1 appears to be exclusively involved in cytokinesis, in particular during embryogenesis.¹⁴

Here we report the discovery and functional characterization of a survivin-like protein (SDSURVL) from a member of the ancestral-like clade Porifera, *Suberites domuncula*. SDSURVL displays both significant sequence homologies to human survivin (in particular concerning the BIR domain) and a conserved genetic structure. Complementary studies in vivo and in transfected cells suggest a conserved dual role of survivin and, concurrently, indicate a loss of function in certain rapidly evolving and highly adapted metazoan taxa.

Results

Characterization of the poriferan survivin-like protein SDSURVL. The complete *SDSURVL* cDNA was isolated from a *S. domuncula* cDNA library by PCR and degenerate primers that were directed against a conserved region within the BIR domain. *SDSURVL* consists of 620 nt (excluding the poly(A) tail) that encompass an open reading frame (ORF) of 447 nt (excluding the first stop codon), beginning at nt_{43–45} (Met_{start}). During Northern blot analyses, a DIG-labeled probe detected a transcript whose size was consistent with that of the cDNA (\approx 650 nt; Figure 4A). The deduced protein, SDSURVL, comprises 149 aa with an expected size of 17,003 Da. Analyses of SDSURVL revealed significant sequence similarity to vertebrate survivin homologues, such as *Gallus gallus* survivin isoform 1 (with an expect value [*E* value]¹⁵ of 2 e-20; 32% identical and 52% similar aa) and human survivin (8 e-18; 30%/49%) (Figure 1A). Considerably lower was the similarity to survivin homologues of more basal Metazoa and fungi, e.g., *D. melanogaster* deterin (7 e-11; 45%/24%), placozoan (*Trichoplax adhaerens*) survivin-like protein (2 e-11; 19%/35%), and *Schizosaccharomyces pombe* survivin homolog (5 e-11; 34%/48% with respect to the BIR domain [aa₁₁₇₋₁₉₄]). Viral genomes often encode anti-apoptotic or cell cycle promoting proteins in order to bypass the host defense system. Viral survivin-like proteins generally display lower sequence homology to their metazoan counterparts and in particular to *S. domuncula* survivin, e.g., *Epiphyas postvittana* nucleopolyhedrovirus IAP-3 (4 e-07; 17% aa similarities) and *Cydia pomonella* granulosis virus ORF17 IAP-3 (3 e-06; 16%).

Since SDSURVL features a single BIR domain, a putative nuclear export signal (NES), and a coiled-coil region, but is missing a RING domain (characteristic of non-survivin IAP), the molecule represents a novel member of the survivin

subfamily of IAP. The poriferan BIR domain (aa₂₀₋₉₃) has an *E* value of 4.85 e-22 and is consistent with the PROSITE consensus sequence PS01282 with two exceptions (marked in bold): [HKEPILVY]-x(2)-R-x(3,**8**)-[FYW]-x(11,14)-[STAN]-G-[LMF]-x-[FYHDA]-x(4) | **x(7)**]-[DESL]-x(2,3)-C-x(2)-C-x(6)-[WA]-x(9)-H-x(4)-[PRSD]-x-C-x(2)-[LIVMA]. The putative NES (aa₉₅₋₁₀₃) loosely fits a consensus sequence proposed by ¹⁶, L-x(2,3)-[FILVMP]-x(2)-[LI]-x-[LIV]. Furthermore, COILS algorithms indicate the typical COOH-terminal coiled-coil conformation (aa₁₂₇₋₁₄₇), usually present in survivin but missing in other IAP family members. Moreover, the conserved threonine residue, whose phosphorylation by p34cdc2 mediates human survivin activation,¹⁷ is present in SDSURVL (aa₂₆). Additionally, a ubiquitin ligase complex-binding destruction motif (APCC-D box; aa₁₃₆₋₁₄₁) was predicted.

To determine the phylogenetic relationship of survivin-like proteins, molecular phylogenetic analyses were performed by neighbor-joining, maximum likelihood, and Bayesian methods, including the so far known sequences of metazoan taxa as well as related BIR-containing sequences of viruses and yeast. Accordingly, the resulting composite consensus tree shows the poriferan SDSURVL protein at the base of a monophyletic metazoan clade, whereas viral sequences form a separate monophyletic group (Figure 1B).

Characterization of the poriferan survivin-like gene (*SDSURVL*). The *S. domuncula* gene comprises four exons and three introns (Figure 2A). Each exon/intron border displays typical splicing signals (GT-intron-AG). The phases of introns vary: the first and third introns are in phase 0, while the second intron is in phase 2. The human survivin gene shares the same features (Figure 2B) except for the borders of the third intron (AT-intron-GG). Parts of the first and third exons and

the complete second exon are encoding the BIR domain (Figure 2C). Analyses of the 5'-flanking region (Figure 2D) of human (1) and murine (2) survivin genes reveal the presence of a TATA-less promoter, containing a canonical CpG island and several transcription factor binding sites.¹⁸ However, in silico analyses of the putative poriferan promoter region (3) predicted the presence of a TATA box (nt₆₄ to ₋₇₃) in addition to other cis-regulatory elements, CAAT box (nt₁₀₉ to ₋₁₁₃), and ribosomal binding site (nt₅ to ₋₈). Moreover, similar to the human and murine promoter two putative Sp1 sites (nt₋₅₁, ₋₂₉₂) have been predicted, although in contrast to the former ones no cell cycle dependent elements (CDE) or cell cycle homology regions (CHR) have been found in the poriferan non-coding region.

Characterization of the poriferan caspase-like protease 2 (SDCASL2). Caspases (cysteine-dependent aspartyl-specific proteases) are synthesized as zymogens that become activated by scaffold-mediated transactivation or by cleavage via upstream proteases.¹⁹ Since the anti-apoptotic properties of vertebrate survivin have been functionally linked to inhibition of caspase-7 and -3, a *S. domuncula* cDNA library was screened for homologous molecules. The correct size of the resulting clone, *SDCASL2*, then was verified on Northern blots (Figure 4A). *SDCASL2* (1,827 nt, excluding poly(A)) comprises an ORF between nt₁₇₅₋₁₇₇ (Met_{start}) and nt₁₇₃₈₋₁₇₄₀ (stop), coding for a putative protein of 521 aa that was termed caspase-like protease 2 (SDCASL2; calculated Mr 58,427). SDCASL2 revealed sequence homology not only to SDCASL (*E* value 7 e-31; 33% aa identical and 52% similar aa) but also to members of the caspase-7 subfamily, e.g., *Xenopus laevis* and human representatives (*E* value 3 e-16 [21%/33%] and 3 e-14 [19%/35%] respectively) (Figure 3).

All caspases contain a CASc domain, which is cleaved into two subunits (p10, p20) during activation. SDCASL2 represents a putative procaspase, comprising subunits p10 (aa₁₉₁₋₂₆₇) and p20 (aa₂₂₋₁₅₄), the latter containing the characteristic Cys and His active sites (Figure 3). The Cys active site consensus pattern is conserved (PROSITE pattern PS01122) with two exceptions, marked in bold ([**KL**]-P-K-[LIVMF]-[LIVMFY]-[LIVMF](2)-[**QPD**]-[AF]-C-[RQG]-[GE]), whereas the histidine active site (PS01121) features three alterations, [**HL**]-x(2,4)-[SC]-x(1,2)-{A}-x-[LIVMFY](2)-[ST]-**G**-H-G.

The absence of a large NH₂-terminal prodomain, characteristic of initiator caspases, suggests a classification to the group of effector caspases. However, similar to SDCASL, SDCASL2 carries an unusual long COOH-terminal stretch of 254 aa, containing a putative double-stranded RNA-binding motif (aa₄₄₁₋₅₁₄; PF0035 [Pfam]).

Evolutionary conserved function(s) of poriferan survivin. To elucidate the controversially discussed ancestral function of survivin (or if indeed its dual role is conserved through metazoan evolution), functional analyses were performed in sponge tissue and primmorphs (aggregates of proliferating sponge cells). Moreover, functional conservation was assessed in a heterologous vertebrate cell model (SDSURVL-expressing transfected HEK-293 cells) that has been previously used to explore expression and functional mechanisms of survivin.^{20,21}

Regulation of proliferation. Comparative Northern blot analyses were performed, showing an increasing level of *SDSURVL* transcripts during the first two weeks of primmorph growth, whereas the control (taken immediately after primmorph

formation) revealed a considerably lower expression level (Figure 4A). After 14 d, survivin transcription reached the steady-state. On the other hand, in adult sponge tissue survivin transcripts were barely detected. Additionally, *SDCASL2* expression was very low throughout the experiments, in both primmorphs and sponge tissue (Figure 4A). Subsequently, *SDSURVL* and *SDCASL2* expression was investigated in aging primmorphs (two weeks of age) via complementary in situ hybridization studies (Figure 4B). For this purpose, primmorph cryosections were incubated with labeled, single-stranded sense (control) or antisense probes. Both sense probes did not produce specific signals. In contrast, the antisense *SDSURVL* probe detected survivin transcripts abundantly and homogenously throughout the sections, whereas only few *SDCASL2* copies were identified, mainly in the surface cell layer and in cells lining the aquiferous canal system within the mesohyl (internal part of the sponge) (Figure 4B).

In parallel, a construct encoding a fusion of *SDSURVL* and COOH-terminal *EGFP* (*SDSURVL-EGFP*) was introduced into HEK-293 cells. Expression was confirmed microscopically (Figure 5B) and by immunodetection, using anti-EGFP antibodies (Figure 8; expected size of 45 kD corresponding to the *SDSURVL-EGFP* fusion). The transfection efficiency was determined by the percentage of EGFP-positive HEK-293 cells via fluorescence microscopy. An average transfection efficiency of 94.6% was calculated.

Subsequently, the putative regulatory function of *SDSURVL* in cytokinesis was explored by monitoring the concentrations of *SDSURVL-EGFP* transfected (HEK293-*SDSURVL*) and mock-transfected HEK-293 cells, 48 h post-transfection (Figure 5A). Whereas mock-transfection provoked no differences in cell concentration compared to the non-transfected control, *SDSURVL-EGFP* expression caused a 2.3-fold

increase in number of HEK293-*SDSURVL* cells (significance level $P \leq 0.05$). In addition, trypan blue staining did not display altered cell viability between control and *SDSURVL-EGFP* expressing populations.

Finally, subcellular localization and nucleo-cytoplasmic trafficking of *SDSURVL-EGFP* in transfected cells was analyzed by confocal laser scanning microscopy (LSM). For this purpose, HEK293-*SDSURVL* cells were incubated with the nuclear export inhibitor leptomycin B (LMB) for 30 min or remained untreated (control). Following counterstaining of dsDNA with the far-red fluorescent dye DRAQ5, LSM showed that in the controls *SDSURVL-EGFP* predominantly localizes in the cytoplasm and, to a lesser degree in the nucleus (Figure 6A). Consequently, the fluorescence pattern of the merge revealed a barely detectable co-localization of fusion (green) and nuclei (red). Upon LMB treatment, *SDSURVL-EGFP* accumulated in the nucleus. The merged channel clearly depicted a significant co-localization of nuclei and *SDSURVL-EGFP* (Figure 6B; yellow), whereas the staining intensity of cytoplasmic *SDSURVL-EGFP* is decreased, compared to the untreated control.

Regulation of apoptosis. To explore the putative implementation of *SDSURVL* as a pro-survival factor, *SDSURVL* and *SDCASL2* expression was analyzed in sponge tissue and primmorphs challenged with pro-apoptotic stimulants. Moreover, the viability of mock-transfected and HEK293-*SDSURVL* cell lines was assessed following exposure to the same stimuli. Pam₃Cys-Ser-(Lys)₄ and cadmium were employed for their selective ability to trigger extrinsic (the former compound) or intrinsic (latter) cell death pathways.^{22,23} Both lipopeptide and heavy metal stimulated the expression of *SDSURVL* and *SDCASL2* in sponge tissue and primmorphs alike, already after 6 h (Figure 7). With prolonged incubation time, the expression level of

both transcripts further increased; however, this effect was more pronounced in primmorphs than in tissue samples.

To assess putative effects of *SDSURVL* expression, mock-transfected HEK-293 and HEK293-*SDSURVL* cell lines (95.5% transfection efficiency) were exposed to Pam₃Cys-Ser-(Lys)₄ and cadmium. Then, MTT assays, cell death detection ELISA, and Western blot analyses were performed. MTT assays revealed that the lipopeptide did not alter cell viabilities, neither in mock-transfected nor in HEK293-*SDSURVL* cells, compared to untreated control cells (Figure 8A). Additionally, Pam₃Cys-Ser-(Lys)₄-treatment did not change the amount of survivin (endogenous as well as poriferan). In contrast, both cell lines were sensitive to cadmium. Thus, MTT assays showed that 46.7% of mock-transfected cells underwent cell death after 12 h of exposure. Nevertheless, *SDSURVL*-expression rescued HEK293-*SDSURVL* since only 25.6% cells were non-viable (Figure 8B; significance level $P \leq 0.05$). Furthermore, Western blot analyses displayed an additional effect of cadmium, since increased immunodetection was observed of both, human survivin and its poriferan homologue after 6-12 h of challenge.

Concurrently, cell death detection ELISA could not reveal altered formation of mono- and oligonucleosomes, neither in mock-transfected nor HEK293-*SDSURVL* cells following Pam₃Cys-Ser-(Lys)₄-treatment, compared to untreated control cells (Figure 9A). In contrast, exposure to cadmium significantly induced apoptotic DNA fragmentation in both cell lines. However, *SDSURVL*-expression caused a reduced DNA fragmentation by 51% (Figure 9B; significance level $P \leq 0.05$).

Discussion

Survivin represents a unique protein within the IAP family, containing a single BIR domain and a COOH-terminal coiled-coil structure. Thus, it differs from all other family members that comprise 2-3 BIR domains and a canonical COOH-terminal RING finger.¹¹ In vertebrates, survivin not only controls cell cycle checkpoints and correct completion of cytokinesis but also regulates activation of caspases.²⁴ However, the evolutionary conservation of this dual role remains inconclusive since in Nematoda the survivin homologue appears to be implicated exclusively in cytokinesis.²⁵ In addition, even though a fungal (*S. pombe*) survivin-like protein was proposed to be required for efficient cell division and exhibits anti-apoptotic activity,²⁶ it is considerably larger in size than other survivin proteins.

Key regulators of apoptosis and cell cycle progression have already been discovered in the phylum Porifera,²⁷ which appears to be the earliest-branching metazoan taxon.⁴ However, identification of poriferan survivin provides an important tool not only to investigate the primordial function(s) of this versatile regulator protein, but also to elucidate the putative ancient and intimate link between apoptosis, proliferation, and morphogenesis.

The newly discovered poriferan survivin homologue demonstrates a significant degree of sequence conservation. A consensus tree resulting of NJ, ML, and Bayesian analyses places SDSURVL at the base of a metazoan clade, confirming the common ancestry of metazoan survivin-like proteins, whereas viral sequences form a well-supported separate group.

SDSURVL features all characteristics of survivins that facilitate protein-interaction and mediate both anti-apoptotic activity and proper kinetochore attachment.²⁸ However, it displays several additional putative domains that are

known to be involved in cell cycle regulatory functions: (i) APCC-D box, required for ubiquitination and proteasome-mediated degradation via the anaphase-promoting ubiquitin ligase complex (APC/C); (ii) LIG_FHA1/2 motifs (aa₉₈₋₁₀₄; aa₁₀₀₋₁₀₆) that bind regulators of the G2/M checkpoint; (iii) USP7-binding motif (aa₃₃₋₃₇) that is a target of the UPS7 deubiquitinating enzyme, whose substrates are regulators of cell survival pathways. So far, no such feature has been described for other survivin molecules.

The gene structure (intron/exon positions, intron phases) is conserved between poriferan and vertebrate (human, murine) survivin,¹⁸ but not within the *D. melanogaster deterin*,¹³ which possesses only two introns. However, some differences to the vertebrate promoter region are noticeable: the TATA-less vertebrate promoter includes CpG islands and several Sp1, CDE, and CHR sites. By analogy with other cell cycle-regulated genes it was postulated that Sp1 regulates basal transcriptional expression of the survivin gene, further modulated by CDE/CHR elements, thus imparting cell cycle periodicity of survivin expression.¹⁸ In contrast, the poriferan promoter features a canonical TATA box, two Sp1 sites, and is missing CDE/CHR elements. Therefore, considering the aforementioned protein interaction motifs, regulation of the poriferan survivin pool might also occur on a posttranscriptional/proteasomal level, e.g., via (de)ubiquitination or complex formation with other regulatory proteins.

Survivin is expressed in many human tumors and in rapidly dividing cells of fetal tissue during the G2/M phase, but is not detectable in terminally differentiated cells. It was proposed that survivin exerts its anti-apoptotic function via inhibiting caspases, particularly caspase-7 and -3,²⁹ though other studies argue against a direct mechanism of caspase inhibition by survivin, see e.g.,³⁰.

To approach the enigmatic role of survivin in non-bilaterian Metazoa, comparative functional studies of poriferan survivin were performed. Accordingly, the implementation of SDSURVL in cell proliferation and cell death was studied in sponges as well as in a human cell model (HEK-293; as SDSURVL-EGFP fusion). This strategy had already been successfully adopted to assess the anti-apoptotic potential of deterin and poriferan Bcl-2.^{31,32} Moreover, survivin-EGFP fusions have been widely used to study both the bifunctional role of survivin and its dynamic intracellular localization and appear fully functional.³³⁻³⁵ In contrast to sponge tissue that displays a very low steady-state expression level, proliferating cells of primmorphs contained a considerable amount of *SDSURVL*, indicative of its involvement in cell division. This assumption was supported by a significantly increased proliferation rate of HEK-293 cells expressing SDSURVL-EGFP. In addition, application of two inducers of cell death (cadmium, Pam₃Cys-Ser-(Lys)₄) indicated an anti-apoptotic function of SDSURVL since both compounds triggered the expression of *SDSURVL* in sponge tissue and primmorphs, possibly a countermeasure to the induced expression of the caspase-7 homologue *SDCASL2*. Concurrently, cadmium induced apoptosis in HEK-293 cells, although concomitant expression of SDSURVL-EGFP drastically reduced the occurrence of cell death ($\approx 50\%$). However, Pam₃Cys-Ser-(Lys)₄ incubation did not alter HEK-293 cell viability, probably due to the different repertoire of Toll-like receptors (mediating lipopeptide-induced innate immunity and apoptosis) found within HEK-293 and sponge cells. Future studies will investigate the effect of further inducers of cell death on sponge and HEK293-*SDSURVL* cells.

Cytotoxicity mediated by cadmium is associated with the release of mitochondrial pro-apoptotic factors into the cytosol.²³ These factors cause activation

of caspase-3/7,³⁶ which is considered as a biochemical hallmark of the apoptotic cascade. However, survivin was proposed to interact with various caspases and, consequently, to inhibit further progression of the apoptotic cascade. The aforementioned observations in combination with significant similarities of sequence and structure between SDSURVL and its vertebrate counterparts propose a common regulatory function in the interconnected pathways of cell cycle and apoptosis. In HEK-293 cells, poriferan survivin possibly complements the endogenous molecule by sequestering pro-apoptotic binding partners, consequently delaying cell death. In addition, the presence of putative target sites for ubiquitination might explain the elevated levels of human and poriferan survivin detected in challenged transfected cells. Accordingly, this effect is not the result of an induced expression but is indicative of a decreased proteasomal degradation of either protein. Therefore, the poriferan molecule replenishes the pool of endogenous survivin, concomitantly preventing degradation of its human counterpart by engaging/silencing elements of the proteasome.

Furthermore, concurrent with previous observations during which elevated levels of survivin occur in most human tumor cell types and overexpression of survivin in transfected HEK-293 increases cell proliferation,^{20,21} SDSURVL-EGFP expression stimulates cell division. In addition, LMB treatment of SDSURVL-EGFP expressing HEK-293 cells causes significant nuclear accumulation of the fusion protein. This is consistent with previous studies in which CRM1 (chromosome region maintenance 1)-mediated, NES-dependent nuclear export of human survivin was blocked by LMB.^{10,16} Survivin is a subunit of the chromosomal passenger complex (CPC), consisting of Aurora-B, Borealin and INCENP, and is essential for proper chromosome segregation and cytokinesis. Since Crm1 is involved in tethering the

CPC to the centromere by interacting with survivin's NES,¹⁰ SDSURVL might affect various stages of cell proliferation, from proper kinetochore attachment to spindle formation.

In conclusion, the identification of a bona fide poriferan survivin homologue represents a compelling opportunity to investigate the deep evolutionary original function(s) of survivin, while concurrently avoiding limitations of classical invertebrate model organisms. The data presented propose an evolutionary ancient dual regulatory role of survivin, implemented in cell cycle control and programmed cell death.

Materials and Methods

Preparation of *S. domuncula* tissue and primmorphs. Specimens of the marine sponge *Suberites domuncula* (Porifera, Demospongiae, Hadromerida) were collected near Rovinj (Croatia) in the northern Adriatic Sea and then kept in aquaria in Mainz (Germany) at 17°C.

Primmorphs were obtained after tissue dissociation into single cells as described earlier.³⁷ They were cultured in natural sterile-filtered seawater (Sigma-Aldrich; Taufkirchen, Germany), supplemented with 0.2% [v/v] RPMI-1640 medium (Biochrom AG; Berlin, Germany) and 60 µM silicate. Cultures were shaken for 5 d to assist the formation of aggregates, consisting of proliferating cells. Then, two sets of experiments were performed. For the first set, primmorphs were exposed to cadmium (CdCl₂, Sigma-Aldrich; 50 µM) or the synthetic lipopeptide (S)-[2,3-Bis(palmitoyloxy)-(2-RS)-propyl]-N-palmitoyl-(R)-Cys-(S)-Ser-(S)-Lys₄-OH·3HCl (Pam₃Cys-Ser-(Lys)₄; Axxora; Grünberg, Germany; 40 µg/mL) for 6 and 9 h, or they remained untreated as a control. For the second set of experiments, samples were taken 0 (control), 7, 14, and 21 d after primmorph formation. All experiments were performed under mild shaking.

In a similar way, *S. domuncula* tissue was exposed to cadmium (50 µM) or Pam₃Cys-Ser-(Lys)₄ (40 µg/mL) for up to 12 h.

RNA extraction and Northern blotting analyses. Total RNA was isolated through lysis of homogenized tissue/primmorphs using TRIzol Reagent (Invitrogen; Karlsruhe, Germany) as described earlier.³⁸ Subsequently, 5 µg of total RNA was size-separated and blotted on Hybond N1 membranes (Amersham; Freiburg, Germany). Labeled probes were generated with DIG-11-deoxyuridine triphosphate

through the PCR-DIG-Probe Synthesis Kit (according to the manufacturer's instructions (Roche Applied Science; Mannheim, Germany)). Hybridization was performed with a *S. domuncula* survivin-like probe (*SDSURVL*, nt₁₋₃₅₇) or a caspase-like protease 2 probe (*SDCASL2*, nt₈₁₀₋₁₄₈₀). Furthermore, a probe was designed to detect the expression of the housekeeping gene β -tubulin (*SDTUB*, nt₈₃₋₄₈₃; NCBI accession number AJ550806) as internal reference. Hybridized probes were detected with anti-DIG Fab fragments (conjugated to alkaline phosphatase) and ultimately visualized by chemiluminescence, using the 1,2-dioxetane compound CDP Star (Roche) as substrate.

Cloning of *S. domuncula* survivin-like protein and caspase-like protease 2.

cDNA coding for the survivin-like protein *SDSURVL* was isolated from a *S. domuncula* cDNA library³⁹ by polymerase chain reaction (PCR) with degenerate primers that were designed against the conserved BIR domain (forward primer, 5'-GAT GGC IGA GGC T/CGG CTT-3'; reverse primer, 5'-CT/CA TGT CCA GTT TT/CA AG/AA-3'; where I = inosine). PCR was carried out at an initial denaturation at 95°C for 5 min, followed by 35 amplification cycles at 95°C for 30 s, 54°C for 45 s, and 72°C for 1 min, and terminated with a final extension at 72°C for 10 min. Thus, a fragment of \approx 209 bp was isolated and subsequently sequenced using an automatic DNA sequencer (Li-Cor 4300). Ultimately, the *SDSURVL* sequence was completed through primer walking. Subsequently, genomic sequences were extracted from a *S. domuncula* genomic DNA library.⁴⁰

Similarly, *SDCASL2* was isolated from the *S. domuncula* cDNA library: application of the degenerate forward primer 5'-T/CTI ATT/C/A CAA/G GCI TGT/C A/T/CG/TI GG-3' (directed against the caspase Cys active site L/F-I-D-A-C-R/L-G)

combined with a library-specific primer resulted in a fragment of $\approx 1,200$ bp. PCR was carried out at an initial denaturation at 95°C for 5 min, followed by 35 amplification cycles at 95°C for 30 s, 54°C for 45 s, and 72°C for 2 min, and terminated with a final extension at 72°C for 10 min. The complete cDNA was isolated by using a combination of sequence- and library-specific primers.

Sequence analyses. Putative promoter regions were analyzed with TESS (<http://www.cbil.upenn.edu/cgi-bin/tess/tess?>), TFSEARCH (<http://www.cbrc.jp/research/db/TFSEARCH>), TATA Signal Prediction in Eukaryotic Genes (http://zeus2.itb.cnr.it/~webgene/wwwHC_tata.html), and CpGProD (http://pbil.univ-lyon1.fr/software/cpgprod_query.html). Coiled-coil conformations were studied via COILS (http://www.ch.embnet.org/software/COILS_form.html). Homology searches were performed via the servers at the European Bioinformatics Institute (Hinxton, United Kingdom (<http://www.ebi.ac.uk/inc/head.html>)) and the National Center for Biotechnology Information (NCBI; Bethesda, MD (<http://www.ncbi.nlm.nih.gov/BLAST/>)). Phylogenetic and molecular evolutionary analyses were conducted using MEGA version 4 for neighbor-joining analyses or were obtained through the phylogeny.fr server (http://www.phylogeny.fr/version2_cgi/programs.cgi) for maximum likelihood analyses (PhyML) and Bayesian inference analyses (MRBAYES).^{41,42} For NJ and ML analyses the degree of support of internal branches was assessed by bootstrapping. For Bayesian analyses posterior probability values were estimated. Potential subunits, domains, patterns, and transmembrane regions were predicted after searching databases of Pfam (<http://www.sanger.ac.uk/Software/Pfam/>), SMART (<http://smart.embl-heidelberg.de/>), and ELM (<http://elm.eu.org/>).

Vector constructs, transfections, and leptomycin B treatment. The forward primer 5'-GAA TTC ATG GCT AAT ACG TAT GAG A-3' (*EcoRI* site is underlined) and the reverse primer 5'-GGA TCC CGA TGT TCG TGT CTT TTC A-3' (*BamHI*) were used to amplify the ORF of *SDSURVL* (nt₁₋₄₄₇), including M_{start} and excluding the first stop codon. The cDNA was ligated in frame into the eukaryotic expression vector pEGFP-N2 (Clontech; Heidelberg, Germany). The recombinant protein, thus, contained a COOH-terminal EGFP-tag.

Human embryonic kidney (HEK-293; ATCC CRL-1573) cells were obtained from the American Type Culture Collection (LGC Promochem; Wesel, Germany) and grown in Dulbecco's Modified Eagle Medium (high glucose and L-glutamine; GIBCO; Karlsruhe, Germany) with 10% [v/v] fetal bovine serum. Cell counting was performed using a Neubauer chamber and trypan blue staining, three times for each sample.

On the eve of transfection, cells were seeded to allow for \approx 60% confluency on the next day. The transfection technology was provided by IBA (Göttingen, Germany) and based on the combination of cationic liposomes and magnetic DNA-binding nanoparticles. Transfections were performed in 12-wells plates with 1.2 μ g DNA per well. For constitutive expression, the cells were split 48 h post-transfection into selective medium, containing G418 at the appropriate concentration (Carl Roth; Karlsruhe, Germany; 200 μ g/mL). *SDSURVL* expression was confirmed microscopically and immunologically, using anti-EGFP antibodies (Miltenyi Biotec GmbH; Bergisch Gladbach, Germany) and an AHBT3 light microscope with AH3-RFC reflected light fluorescence attachment (Olympus; Hamburg, Germany).

The nucleo-cytoplasmic trafficking of the *SDSURVL* fusion protein was analyzed through treatment of transfected cells with the nuclear export inhibitor

leptomycin B (LMB), according to ¹⁶. In short, 48 h post-transfection cells were treated with LMB (6 nM, 30 min), washed in phosphate buffered saline (PBS), and fixed in 4% [w/v] paraformaldehyde (PFA; 5 min, RT). Following additional washing of cells with PBS and subsequent counterstaining of nuclei with DRAQ5 (5 µM; Biostatus Ltd.; Shephed, UK), images were acquired on a Zeiss 710 confocal LSM (Zeiss; Göttingen, Germany). The argon laser line of 488 nm was used to excite EGFP and a 633 nm HeNe laser was employed to excite DRAQ5. For illustration purposes, green and red channels were extracted from the merged pictures with the ZEN software (Zeiss).

In situ localization studies. The in situ hybridization method applied was based on a procedure described by ³⁹. In short, frozen sections (8 µm) of primmorph samples were fixed in 4% paraformaldehyde, then incubated with proteinase K (1 µg/mL), and fixed again in paraformaldehyde. Hybridization was performed with single-stranded *SDSURVL* DNA probes, overnight at 45°C in 2x SSC (sodium chloride/sodium citrate, supplemented with 50% formamide). The probes (236 nt each) were generated during two separate PCR reactions, either with forward primer 5'-TAT GAG AGT TGT GAC AGA GTA AA-3' (sense/control probe) or with reverse primer 5'-TGT TCA TCA CGG GGA TTG TCT GA-3' (antisense probe), using *SDSURVL* cDNA as template. During the linear amplification the probes were labeled with the DIG-Probe Synthesis Kit. In a similar way *SDCASL2* probes (292 nt) were obtained, using forward primer 5'-GGT GTG TGG CTT ACA AAA CTT TC-3' or reverse primer 5'-GAA CAG ACT CAG TTG GAG ATG CT-3'. Following hybridization the sections were washed at 50°C at decreasing salt concentrations (1x-0.2x SSC), blocked, and then incubated

with anti-DIG Fab fragments, conjugated to alkaline phosphatase. Through addition of NBT and BCIP hybridized probes were visualized.

Western blot analyses. Mock-transfected (Mock) and SDSURVL-expressing (HEK293-SDSURVL) cells were exposed to Pam₃Cys-Ser-(Lys)₄ (40 µg/mL, 6 and 12 h) or cadmium (CdCl₂, 50 µM, 6 and 12 h). Cells were then lysed with an adequate volume of NP40 lysis buffer (50 mM Tris-HCl, 150 mM NaCl, 1% [v/v] NP-40, pH 8.0; 20 min on ice). After centrifugation, the total protein concentration was determined in the supernatant with BCA Protein Assay Reagent (Pierce; Rockford, IL), according to the manufacturer's protocol. 20 µg of total proteins were subjected to electrophoresis through 14% [v/v] polyacrylamide gels, containing 0.1% [w/v] SDS. Following electroblotting, PVDF membranes (Invitrogen) were incubated with rabbit anti-EGFP (1:1,000), conjugated with horseradish peroxidase. Immune complexes were visualized with Supersignal West Pico Chemiluminescent substrate (Pierce) on X-ray films. Other antibodies were acquired from Serotec (Raleigh, NC), including anti-survivin (1:500; directed against aa₁₋₁₂ of human survivin, a region without homology to SDSURVL) and anti-tubulin (1:1,000; human).

MTT assays and cell death detection ELISA. In MTT assays the activity of mitochondrial succinate dehydrogenase is measured as indicator of cellular metabolic rate and viability. To determine the effect of SDSURVL expression, transfected and mock-transfected HEK-293 cells were seeded into 96-well plates (0.5x10⁴ cells/mL) and exposed to Pam₃Cys-Ser-(Lys)₄ (40 µg/mL, 12 h) or cadmium (50 µM, 12 h). Afterwards, 10 µL of an MTT solution (Sigma-Aldrich; 5 mg/mL in PBS) were added to each well for 4 h (37°C). The resulting formazan crystals were

dissolved in a stop solution (40% [v/v] DMF, 10% [w/v] SDS). Absorbances were determined at 570 nm with a Bio-Rad 3550 ELISA plate reader.

Additionally, the apoptotic response was measured in HEK293-*SDSURVL* and mock-transfected HEK-293 cells. Since fragmentation of genomic DNA represents a hallmark of apoptosis, a cell death detection ELISA (enzyme-linked immunosorbent assay) was applied, based on the immunochemical detection of mono- and oligonucleosomes in the cytoplasmic fractions of cell lysates. For this purpose, lysates of cells that had been exposed to Pam₃Cys-Ser-(Lys)₄ (40 µg/mL, 12 h) or cadmium (50 µM, 12 h) were transferred to streptavidin-coated 96-well plates and incubated with biotin-labeled anti-histone and peroxidase-conjugated anti-DNA antibodies, according to the manufacturer's instructions (Roche). Following extensive washing, immobilized antibody-histone complexes were visualized colorimetrically through incubation with peroxidase substrate ABTS (2,2'-azino-bis(3-ethylbenzthiazoline-6-sulfonic acid)). Finally, absorbances were determined at 405 nm and 490 nm (reference).

Statistical analyses. Statistical analyses were performed with a Mann-Whitney U test, a non-parametric statistical test for assessing whether the difference in medians between two samples is statistically significant. The level of significance was set at $P \leq 0.05$.

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Titles and legends to figures

Figure 1 *Suberites domuncula* survivin-like protein SDSURVL. (A) The deduced amino acid sequence (SUBDO) was aligned with survivin homologues of *D. melanogaster* (DROME, NCBI accession number NP_650608), *Anopheles gambiae* (ANOGA, XP_317026), *Nematostella vectensis* (NEMVE, XP_001624746), human (HUMAN, CAG46540), *G. gallus* (CHICK, NP_001012318), *Danio rerio* (DANRE, NP_919378), and *S. pombe* (SCHPO, NP_587866). Conserved residues (identical or similar with respect to physicochemical properties) in all sequences are shown in white letters on black; those in 80% (60%) are in white on gray (black on light gray). The following features are marked, BIR domain, NES, coiled-coil region, APCC-D box (underlined) as well as the putative phosphorylation site (star). (B) Composite consensus tree of the phylogenetic analyses, depicting the evolutionary relationship of *S. domuncula* SDSURVL to survivin homologues of Metazoa, fungi, and viruses. The tree was generated after alignment of the aforementioned sequences, further integrating sequences from: *T. adhaerens* (TRIAD, EDV24865), *Schistosoma japonicum* (SCHJA, AAW27178), *Felix catus* (FELCA, NP_001009280), mouse (MOUSE, NP_033819), *Xenopus laevis* (XENLA, BAD98266), *Pan troglodytes* (PANTR, XP_512010), *Sus scrofa* (PIG, NP999306), *Canis lupus* (CANFA, NP_001003348) as well as BIR-containing viral sequences of *Helicoverpa armigera nuclear polyhedrosis virus* (NPVHA, NP_075172), *Buzura suppressaria NPV* (NPVBS, AAC34373), *Epiphyas postvittana NPV* (NPVEP, NP_203195), *Choristoneura fumiferana NPV* (NPVCF, NP_848342), *Amsacta moorei entomopoxvirus* (AMEPV, NP_064803), and *Cydia pomonella granulosus virus* (GVCP, NP_148801). Close to each node are the bootstrap values in the order,

Neighbor-Joining, Maximum Likelihood, and the posterior probability estimates of the Bayesian inference.

Figure 2 *Suberites domuncula* survivin-like gene. (A) Exon/intron architecture. Each exon/intron border is marked (boxed) as well as introns, start and stop codon (underlined). (B) Alignment of SDSURVL and human survivin exons with correlating intron phases. Sizes of introns are indicated. (C) Schematic representation of *SDSURVL* mRNA and its translational product, drawn to scale. The BIR domain and its contributing exons are marked, as well as untranslated regions. (D) Schematic representation of survivin promoter regions from representative species: murine (1), human (2), and poriferan (3). The different elements: Sp1 (sequence-specific DNA-binding protein-1), CDE (cell cycle dependent element), CHR (cell cycle homology region), CAAT and TATA boxes, and RBS (ribosomal binding site) are drawn to scale (first base of Met_{start}, +1 nt). Regions of high CpG island density are also indicated. The promoter region is depicted up to -400 nt.

Figure 3 *Suberites domuncula* caspase-like protease 2 SDCASL2. The CASc domain of the deduced poriferan protein (CASL2_SUBDO; aa₂₂₋₂₆₇) was aligned with the corresponding domain of *S. domuncula* caspase-like protein (CASL_SUBDO; CAL36107; aa₁₇₋₂₇₄); *D. melanogaster* DCP-1 (DCP1_DROME; AAB58237; aa₇₇₋₃₁₅), *Culex quiquefasciatus* (CASP_CULQU; XP_001842236; aa₆₀₋₃₀₄), and human caspase-7 (CAS7_HUMAN; BAG62964; aa₇₄₋₃₁₁). The putative subunits contained within (p20 and p10) are marked as well as the Cys and His active sites.

Figure 4 Expression of *SDSURVL* and *SDCASL2* in *Suberites domuncula* primmorphs and adult tissue. (A) Northern blotting analyses. *SDSURVL* and *SDCASL2* transcription was monitored in aging primmorphs and in adult sponge tissue (control, 7, 14, and 21 d). Tubulin transcription was used as an internal control. (B) In situ hybridization analyses. Cryosections of primmorphs (two weeks old) were subjected to in situ hybridization analyses, using labeled *SDSURVL* (1, 2) or *SDCASL2* (3, 4) probes (1/3, sense (negative control) or 2/4, antisense probe). Hybridized probes were detected through anti-DIG Fab fragments, conjugated to alkaline phosphatase, and subsequent NBT/BCIP treatment. The sense probes (1, 3) did not produce any signal, whereas the *SDSURVL* antisense probe (2) abundantly and homogeneously detected transcripts. The *SDCASL2* antisense probe (4) identified few transcripts in the surface cell layer (S) and in cells lining the aquiferous canal system (Ca) within the mesohyl, the internal part of the sponge. Sponge spicules (Sp) are marked. Bar, 1 mm.

Figure 5 *SDSURVL* expression in HEK-293 cells. (A) Effect of *SDSURVL* expression on cell proliferation. Cells were transfected either with empty vector (Mock) or with a *SDSURVL* construct (HEK293-*SDSURVL*) or remained untransfected (HEK293). Cell concentrations were measured 48 h post-transfection. Proliferation rates of Mock and HEK293 were comparable, demonstrating that the vector itself had no effect. However, HEK293-*SDSURVL* revealed a significantly higher proliferation rate ($P \leq 0.05$). Bars represent mean \pm SD (standard deviation) of three independent experiments, each with $n = 5$. (B) Histological detection of the *SDSURVL*-EGFP fusion protein in transfected HEK-293 cells. Mock-transfected HEK-293 cells (1, 2) and HEK293-*SDSURVL* (3, 4) under visible (1/3) or UV light (2/4). Bar, 100 μ m.

Figure 6 Subcellular localization of SDSURVL-EGFP in transfected HEK-293 cells. SDSURVL-EGFP expressing cells either remained untreated (A, control) or were incubated with the nuclear export inhibitor LMB (6 nM, 30 min; B). The localization of the fusion protein (green) was analyzed by laser scanning microscopy. Nuclei were DRAQ5-stained (red). The merge pictures revealed a predominantly cytoplasmic localization of SDSURVL-EGFP in the control, whereas upon LMB treatment the EGFP-tagged protein accumulated in the nucleus (yellow). The panels show representative examples of SDSURVL-EGFP expressing cells. Bars, 10 μ m.

Figure 7 Gene expression analyses of challenged sponge tissue and primmorphs. Following exposure of sponge tissue and primmorphs to Pam₃Cys-Ser-(Lys)₄ (A) or cadmium (B) for the indicated time, RNA was isolated, size-separated (equal amounts of RNA were loaded), and blotted. The blots were probed for transcripts of poriferan survivin-like protein (*SDSURVL*), caspase-like protease 2 (*SDCASL2*), and tubulin (*SdTUB*). The latter one was used as an internal control.

Figure 8 SDSURVL-EGFP mediated effects on Pam₃Cys-Ser-(Lys)₄- and cadmium challenged HEK-293 cells. Viability (MTT assays, left; bars represent mean \pm SD of three independent experiments, each with n = 5) and protein detection (Western blot analyses, right) of HEK293-*SDSURVL* and mock-transfected (Mock) cells, following exposure to Pam₃Cys-Ser-(Lys)₄ (PAM; A) or cadmium (B). SDSURVL-EGFP expression was detected with anti-EGFP antibodies while expression of endogenous survivin was detected with anti-survivin antibodies. Co, untreated cells. Whereas PAM altered neither cell viability nor survivin/SDSURVL-EGFP concentrations,

cadmium provoked significantly decreased cell viability. However, HEK293-*SDSURVL* cells were less sensitive to cadmium than mock-transfected controls ($P \leq 0.05$) and demonstrated an increased accumulation of both endogenous survivin and SDSURVL-EGFP.

Figure 9 Susceptibility of HEK-293 cells to Pam₃Cys-Ser-(Lys)₄- and cadmium. Following exposure to Pam₃Cys-Ser-(Lys)₄ (PAM; A) or cadmium (B), apoptotic fragmentation of genomic DNA was assessed immunologically in HEK293-*SDSURVL* and mock-transfected HEK-293 (Mock) cells. In this cell death detection ELISA, immobilized cytoplasmic histone-associated-DNA-fragments were detected through consecutive addition of peroxidase-conjugated anti-DNA antibodies and ABTS and, then, quantified photometrically. Co, untreated cells. Data represent mean \pm SD of four independent experiments in triplicate. Whereas PAM did not cause internucleosomal fragmentation of genomic DNA (neither in HEK293-*SDSURVL* nor in mock-transfected HEK-293), cadmium induced apoptotic degradation of DNA. However, expression of SDSURVL-EGFP significantly reduced this effect compared to mock-transfected controls ($P \leq 0.05$).

A

DROME MES-----PVVNEVAASLGGEKLEVFRKLNLEQHRVESYK--SWPFPE--TASCSISKMAEAGFYWTGT 61
 ANOGA MES-----THNSAKIYLLQEDREKSEFK--HWPFSD--DKQCSIQKMAEAGFYWHGT 47
 NEMVE MDS-----TEYSFEMNMEKKRLETFK--DWPFNHM-DCKCTAEKMAAAGFYHCET 47
 HUMAN MG-----APTLPPAWQPFLKDHRISTFK--NWPFFE--GCACTPERMAEAGFIHCPT 48
 CHICK MAA-----YAEMLPKEWLVLVSTRATFR--NWPFFE--GCACTPERMAAAGFVHCPS 50
 DANRE MD-----LASDDQTKMYFYENRLOTFV--GWPFFE--GCVCTPENMAKAGFIHTPS 47
 SUBDO MAN-----TYESCGRVKEKGALYLVEDRAOTFVGSNWPFFES--GPCTPLKMAEAGFYCYGS 54
 SCHPO MKP-----PSCPWAYILSSKNNPNQNPQAAALTKCREQTFVDKVPYPTNRPDYHCEPSVMAASGFVYNPT 154

[89 aa]

★

NES BIR

DROME KRENDTATCFVCGKTLDGWEPEDDPWKEHVKAPOCEFAK-----LSCPERNLTVSQFLEILGTVVKGSI 126
 ANOGA ETEIDIAACFVCGKELDGWEESDDPWSEHRKHAPOCFVK-----FGRPETHLTVM-----GKCYWLND 107
 NEMVE DDDPDVARCHVCFKELDGWEPEDDPWQEHKKHSPKCFVK-----LNKSSNEITVQOFLELEANRQANRM 112
 HUMAN ENEPDLAQCFCFKELEGWEPEDDPIEHHKHSAGCAFLS-----VKKQFEELTLGFEFLKLDREKAKNKI 113
 CHICK ENSPDVVQCFCFKELEGWEPEDDPIEHHKHSAGCAFAA-----LQKDPSNLTVQEFKLDKKRRTKNVI 115
 DANRE ENSPDIAQCFCFKELEGWEPEDDPEKEHKAHSPSCDFIL-----LKKTVDSLTVVEEFLKQKERQKFI 112
 SUBDO AQTEDWVRCAVCHQDMEGWEESDNPRDEHKRCQPGCDYLK-----LKDPYT-ITVGDILDLEKAAAYEPI 118
 SCHPO ADAKDAAHCLYCDINLHDWEPEDDPYTEHKRRRACVFT-----NNLNARPLTMSLYTNTSEEKDSQPT 273

BIR

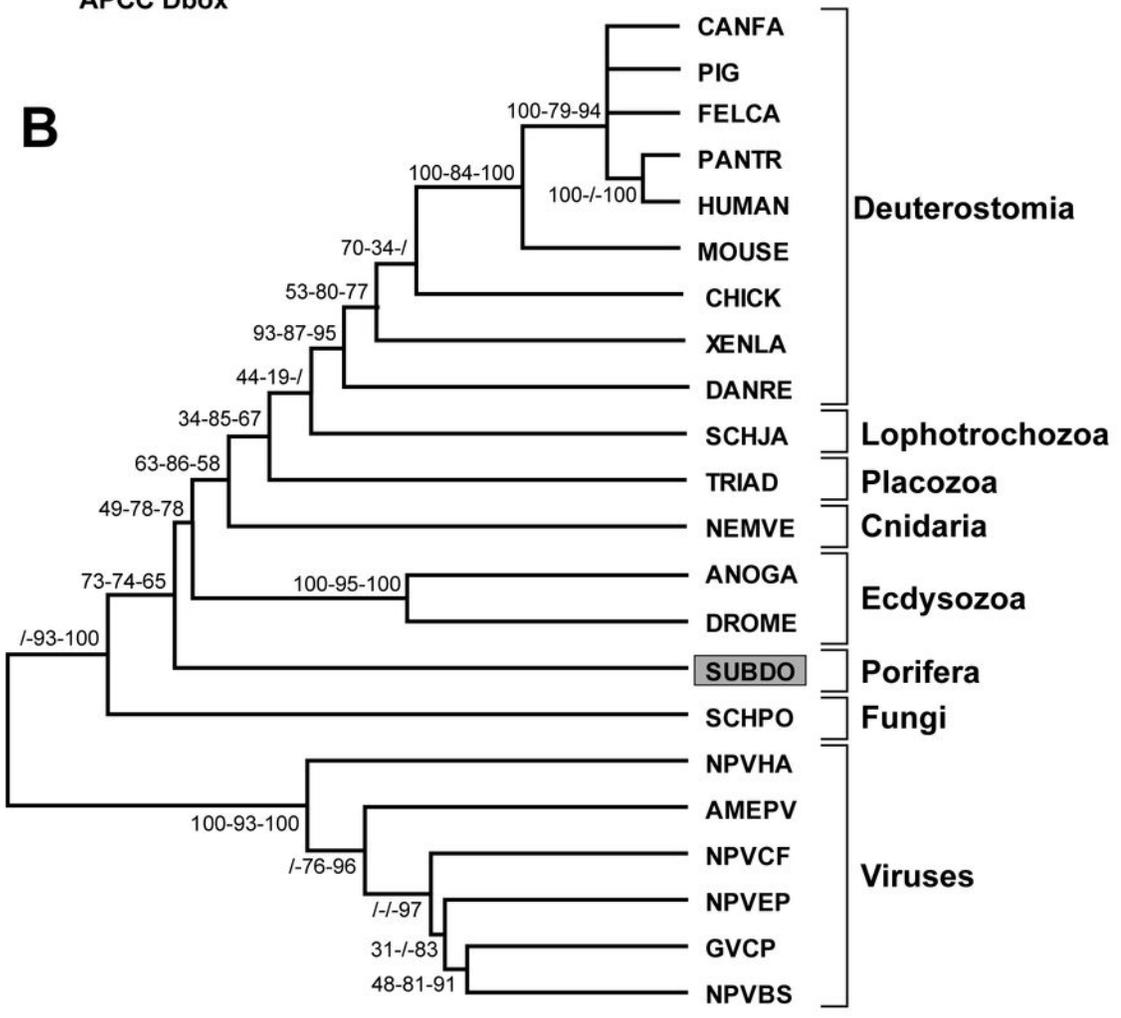
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DROME EKTCKAFKSSFVRENEKRLDEFTRNQK----- 153
 ANOGA EEKIKLTDITFALN--LQDEFVTIMMAT----- 134
 NEMVE RKYVDANLKD FEHMRDQAREEMMKLVPEP----- 141
 HUMAN AKETNNKKKEFEETVKKVRRATEQLAAMD----- 142
 CHICK KKAI SQKETDIEDVAKGRHA TENMGP----- 142
 DANRE KKSCGHAIDKFEDAVKLRNQLIQ TAMGEE----- 142
 SUBDO RQEGDKLSADLDEMVGDMREQLVNAEKTRTS----- 149
 SCHPO RAPQSPTKPVLLTAPRRKNKSPKSKPAVFKPVKPIFSDDEDDEDDDL----- 997

Coiled-coil region
APCC Dbox

[677aa]

B



A

ATGGCTAATACGTATGAGAGTTGTGACAGAGTAAAGGAGAAGGGAGCGCTCTATCTTGTGGAGGACAGAGCACAAACGTTTCGTTGGAAGTAATTGGCCT
Met

TTTGAGAGTGGGCCATGCACTCCTCTGAAA**GT**AAGATGATTGGACATCCAGTGTTCATGTCAGAGTGTGGATGTGAGTCATGCAATGGTTTGTGGCT
Intron 1

CTGTTGTT**AG**ATGGCAGAGGCTGGGTTCTATTATTGTGGCAGTGTCAAACCTCCAGACTGGGTTTCGTTGTGCTGTGTGCATCAAGACATGGAGGGTT

GGGAAGAATCAGACAATCCCC**GT**ATGTGTATGTTGTAATATTGAAATTGTCCTTGAGTTGGCTAAATCTACACGTT**AG**TGATGAACACAAGCGTTGT
Intron 2

CAGCCTGGATGTGATTATCTCAAATTGAAAGACCCGTACACCATCACTGTGGGGGACATCTTAGACTTGGAGAAGGCAGCGTATGAGCACTATATT**GTG**

AGTGACAGATAGTTTTGACTGAACATATAAAGCTATAATCGCTGTTTTCTTT**AG**AGACAAGAGGGTGACAAGTTGTCTGCAGATCTAGATGAGATGGT
Intron 3

TGGTGATATGAGGGAACAGCTGGTGAATGCTGAAAAGACACGAACATCT**TGA**
Stop

B

SURV_HUMAN -----ATGGCTGCCCGACGTTGCCCTGCCTGGCAGCCCTTT--CTCAAGGACCACCGCATCTCTACATTCA--AGAACT 73
 SDSURVL_SUBDO ATGGCTAATACGTATGACACTTGTGACAGAGTAAAGGAGAAGGGAGCGCTCTATCTTGTGGAGGACAGAGCACAAACGTTTCGTTGGAAGT 90

SURV_HUMAN GGCCTTC--TTGGAGGCTGGCCTGCACCCCGGAGCGG-----253nt-----ATGGCCGAGGCTGGCTTC 382
 SDSURVL_SUBDO AATTGGCCATTTGAGATGGGCCATGCACTCCTCTGAAA-----80nt-----ATGGCAGAGGCTGGCTTC 227

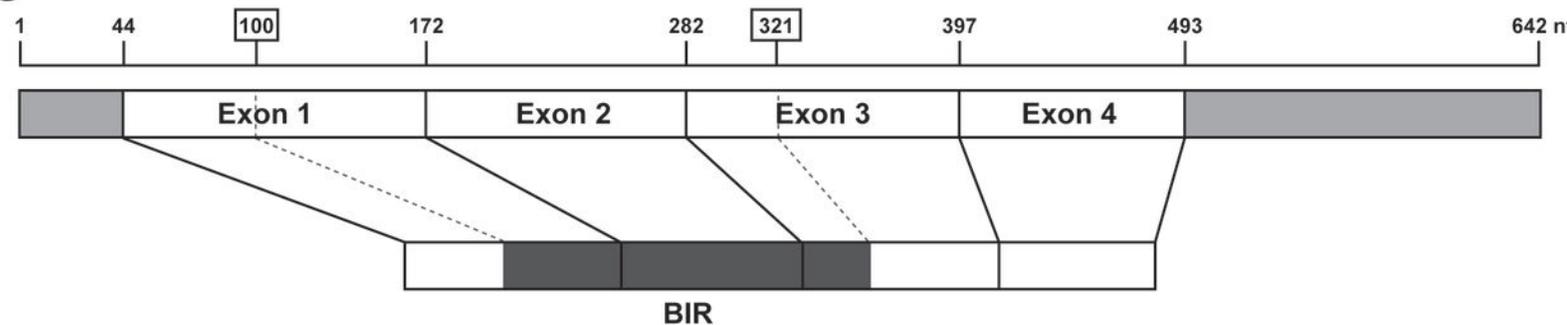
SURV_HUMAN ATCCACTGCCCACTGAGAACGAGCCAGACTTGGCCAGTGTCTTCTGCTTCAAGGAGCTGGAAGGCTGGGAGCCAGATGACGACCCC 472
 SDSURVL_SUBDO TATTATGCTGGCAGTCTCAAACCTCCAGACTGGGTTCTGTGTGCTGTGTGTCATCAAACATGGAGGGTTGGGAAGAATCAGACAATCCC 317

SURV_HUMAN AT-----1875nt-----AGAGGAACATAAAAAGCATTCGTCCGGTTGCGCTTCTCTTCTGTCAAGAAGCAGTTTGA 2409
 SDSURVL_SUBDO CG-----58nt-----TGATGAACAACAAGCTTGTGAGCCTGGATGTGATTATCTCAAATTCAAAGACCCGT---A 434

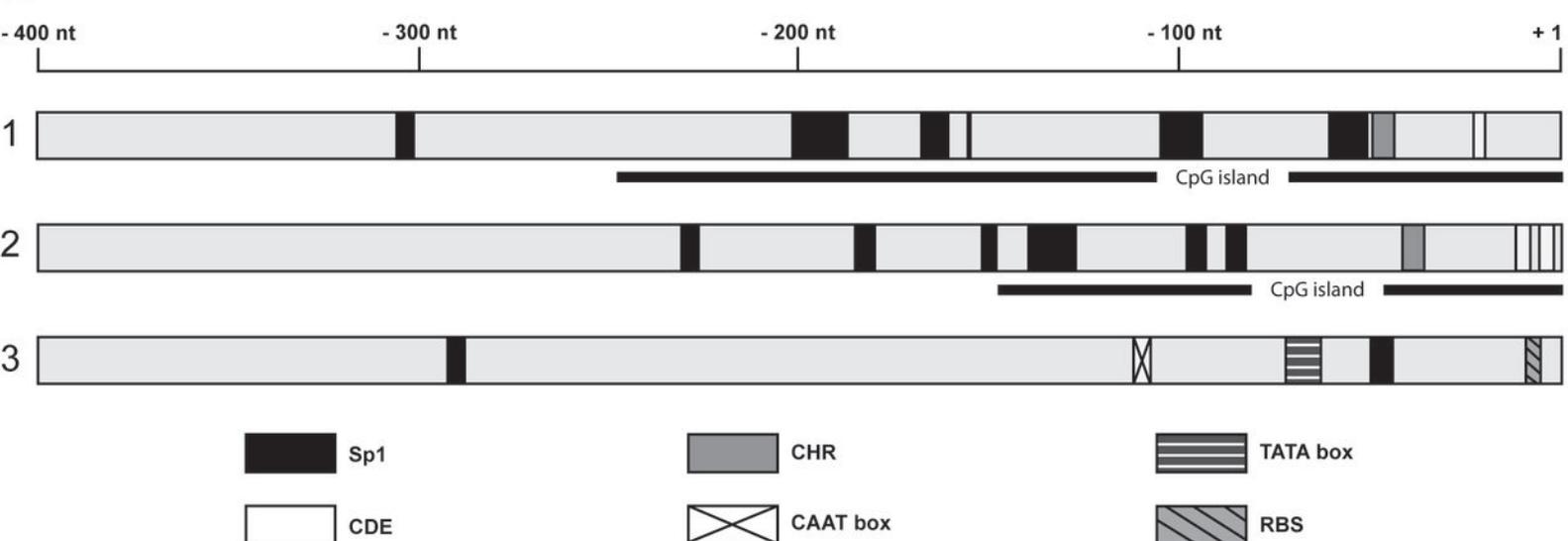
SURV_HUMAN AGAATTAACCCCTGGTGAATTTTTGAAACTGGAACAGAGAAAGAGCCAAAGAACAAAATT-----6685nt-----GCAAAAG 9158
 SDSURVL_SUBDO CACCATCACTGTGGGGACATCTTAGACTTGGAGAAGCCAGCGTATGAGCACTATATT-----58nt-----AGACAA 556

SURV_HUMAN GAAACCAACAATAAGAAGAAAGAATTTGAGGAAACTCCGGAGAAAGTCGGCCGT--GCCATCGAGCAGCTGGCTGCCATGGA----TTGA 9242
 SDSURVL_SUBDO GAGGGTGACAAGTTGTCTGCAGATCTAGATGAGATGCTTGGTGAATCAGGGAACAGCTGGTGAATGCTGAAAAGACACGAACATCTTGA 646

C



D



CASP_CULQU	KRGLALIFNHEHFE-IPQLKSRAGTINVD	CENLASTLKHLD	EDVVRVYKDMKLRDMQKEVEKVAKMD	----HSDNDCILITILSHG	-----	79	
DCP1_DROME	HRCVALIFNHEFFD-IPSLKSRGTINVDAQELKKA	FENLGF	AVSVHKDCKLRDILKHV	GKAAELD	----HTDNDCLAVAILSHG	-----	79
CAS7_HUMAN	KLCKCI IINNKNFDKVTGMGVRNGTDKDAEALFKC	FRSLG	FDVIVYND	CSCAKMQDLLKKASEED	----HTNAACFACILLSHG	-----	80
CASL2_SUBDO	ARGLAIIVTNDYDSPEQELTPLRGTVCDGTNM	TRAFKNLHFAV	HAEHNIGIHMLQQLIRETTQFDAYHKLKNHKCVAFV	FSGHG	-----	84	
CASL_SUBDO	GYGLAI IITNDYRR-APDLSALTE	THNDGKRIADV	FRELNYAVFHM	YNVSEHQLDQLLYEATQKSTYP--RCYQSI	VVVYSGHG	SIQVIN	87

p20

His active site

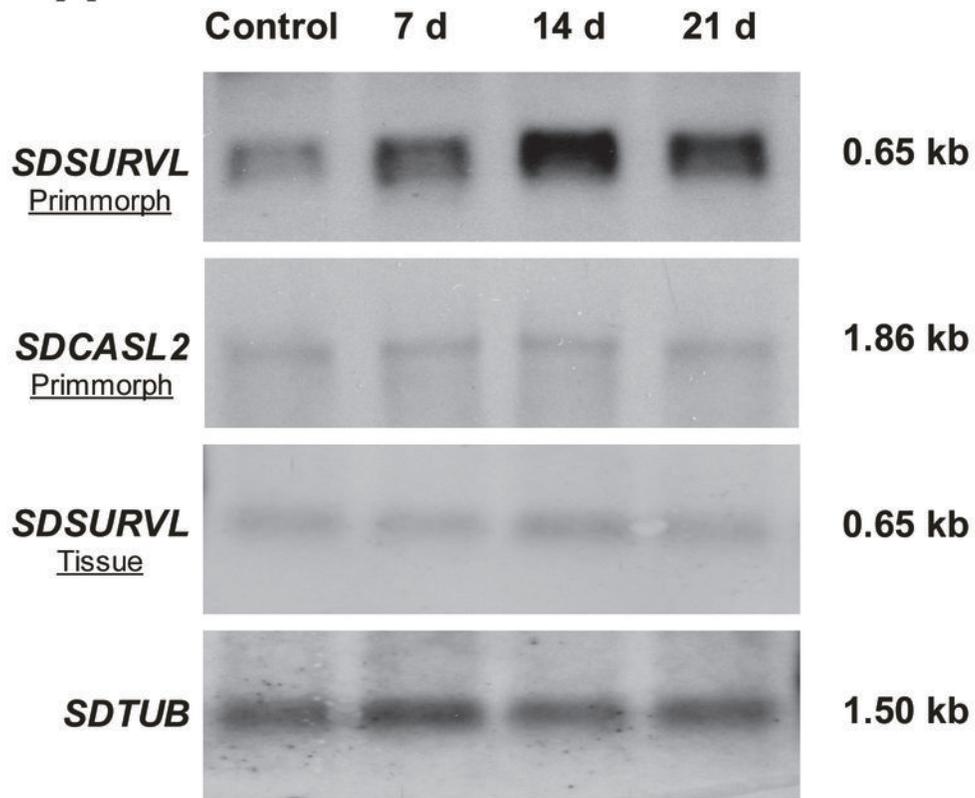
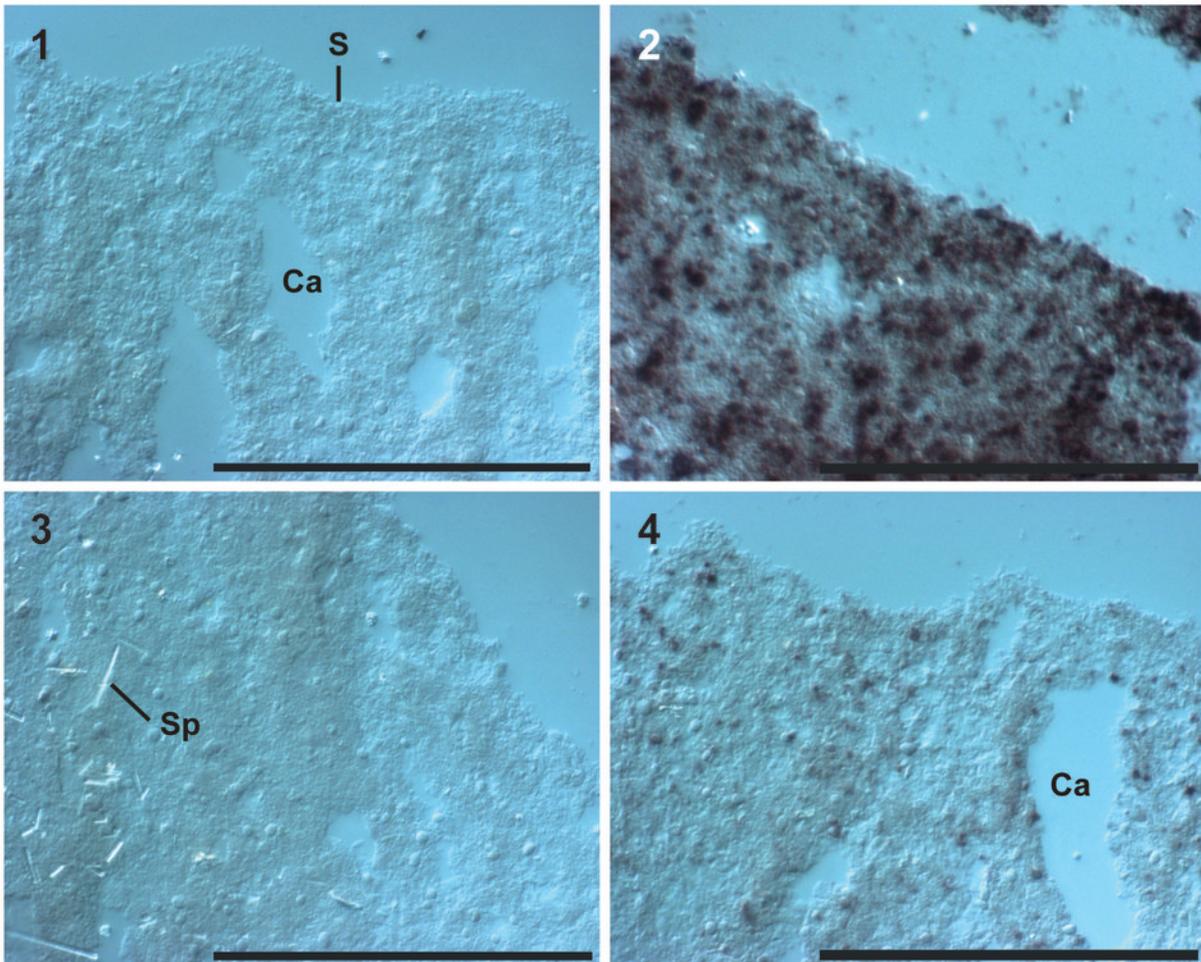
CASP_CULQU	---ELGYLYAKD--CQYKLDI	IWSYETA-NHCPTLAGKPKLFFIQACQGDQ--LDAGVTLSD	-----RTETDSAGSMSYKIPAH	150
DCP1_DROME	---EHGYLYAKD--TQYKLDNI	WHYETA-TFCPSLAGKPKLFFIQACQGDR--LDGGITILEKG	-----VTETDGESSTSYKIPAH	151
CAS7_HUMAN	---EENVIIYGKD--GVTPIKDLTAHERG-ARCKTLLEKPKLFFIQACRGTE--LDDGIQADS	-----GPINDTDANPRYKIPVE	151	
CASL2_SUBDO	--NSNHRLYTQDGNLVCIMDDIIKPFQPTTECP	SMGALPKLFFFDACRGKN--NMKGVYVPSGSVGKEGVIEPVLEPRGVRVMKKLVCPE	170	
CASL_SUBDO	DQNSLTFLYTQD-CFQYPIKEIVKCEMP-SYSPKIGAMPKVFLIDACLGD	KQMYTEAVTVPKSGGAEVRGLAMVSDKGGKEVPTIDVPE	175	

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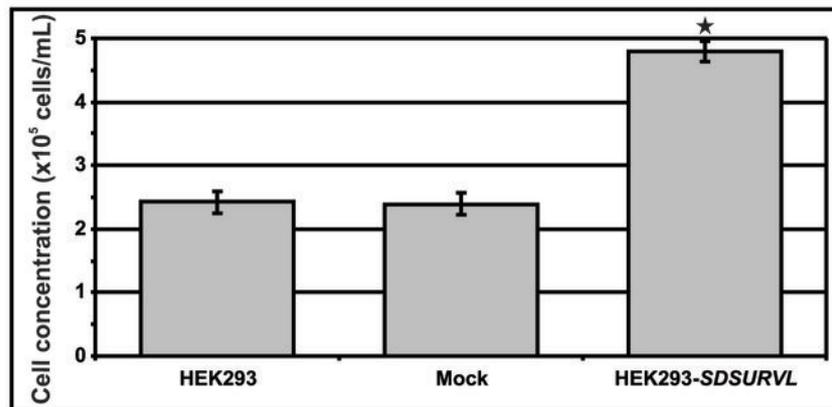
Cys active site

CASP_CULQU	ADFLIAYSTIPGFYSWRNTQKGSWF	MSL	CQELNQHCKKYDIL	TLLTFVAQ	RVAYDFESNTPDIPMMHQKQIPCVT	TMLTRL	LRFTD	238
DCP1_DROME	ADFLFSYSTIPGYFSWRNINNGSWYM	QSLIRELNANGKKYDLL	TLLTFVNQ	RVALDFESNVPATPMDRQKQIPCLT	SMLTRIL	REGD	239	
CAS7_HUMAN	ADFLFAYSTVPGYYSWRSPGRGSWFVQALCSILEEHGK	DLEIMQILTRV	NDRVARHFESQS-DDPHFHEKKQIPCVV	SMLTKELYFSQ	238			
CASL2_SUBDO	SNFLIAYSTMDSYRAWKPGDGGVWLT	TKLSERIQSRC--ESIDDVLT	NVNEKLADDLN-----VRMQQ-----PEKLS	TLNKVLYLHP	246			
CASL_SUBDO	GNYIVAHSTSVGYQSYEIKGS	GGVWIKRLAEKIRFKGRDESVMN	ILTEVSGELLQLYQNPSYRMAQA-----PYALCKLHGTLYFVP	258				

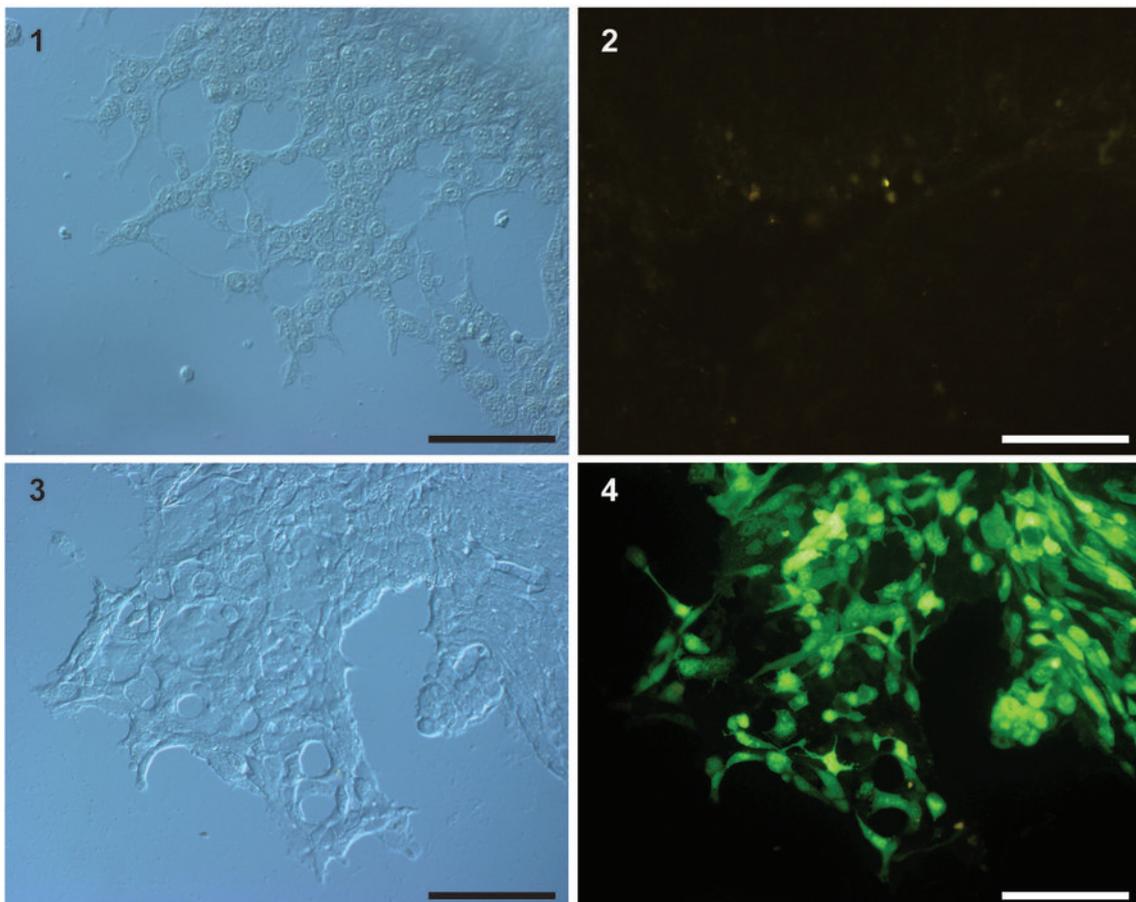
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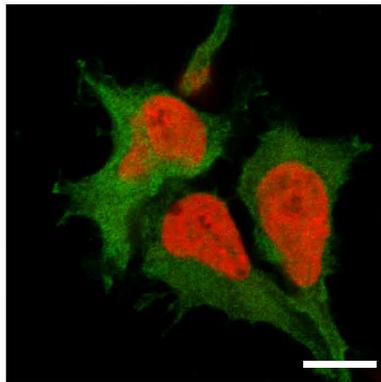
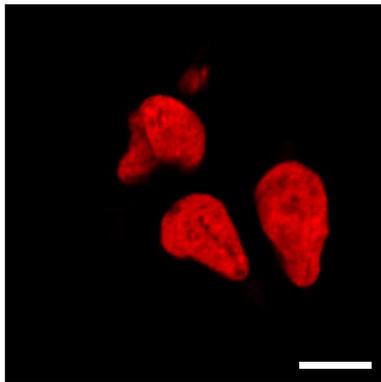
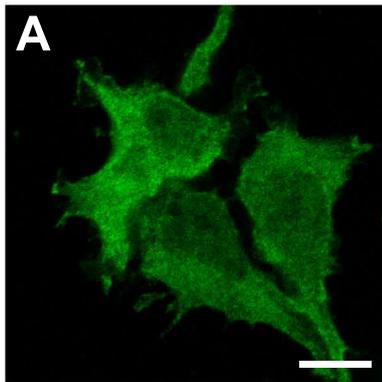
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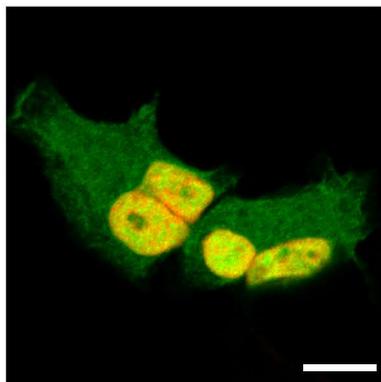
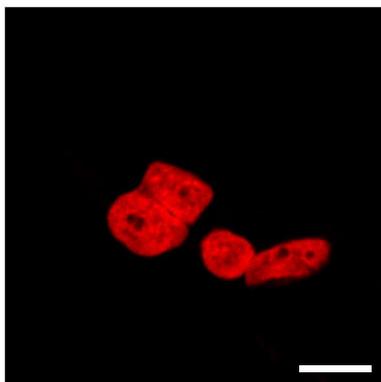
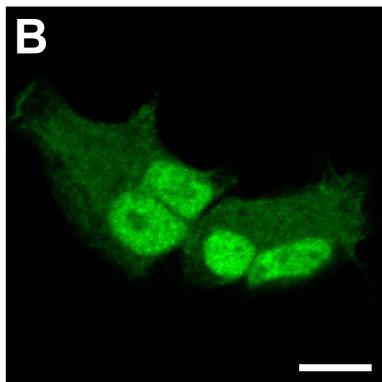


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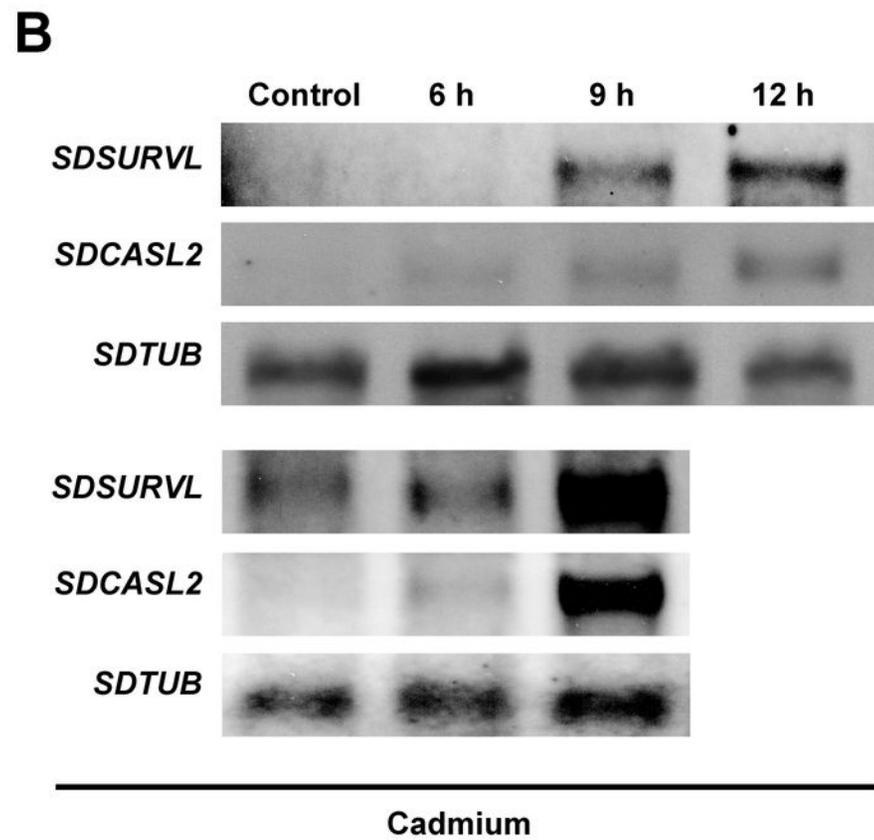
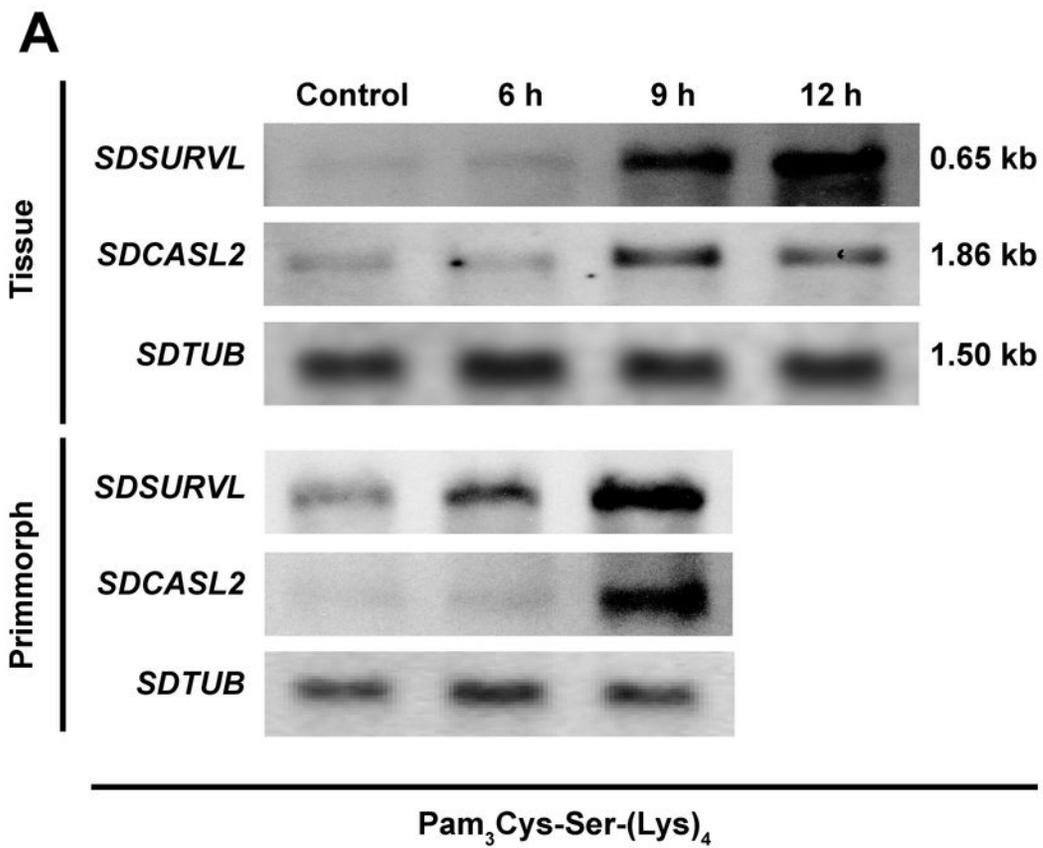


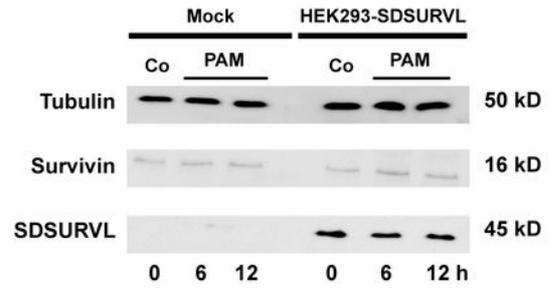
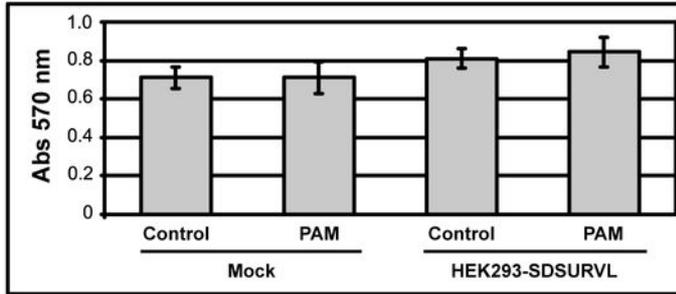
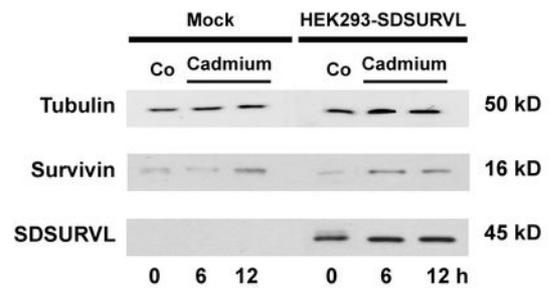
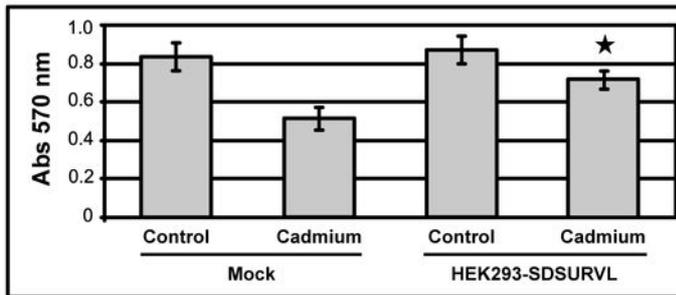


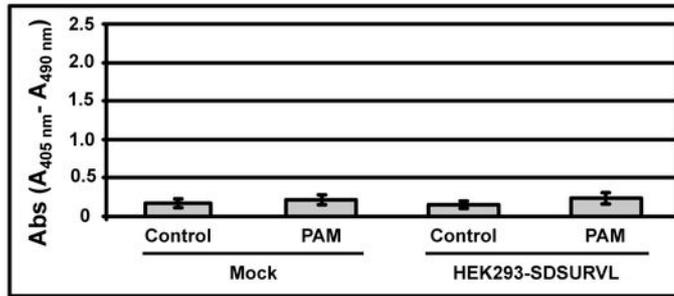
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