

Capnocytophaga canimorsus

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Wim Gaastra, Len J.A. Lipman. Capnocytophaga canimorsus. Veterinary Microbiology, 2010, 140 (3-4), pp.339. 10.1016/j.vetmic.2009.01.040 . hal-00556047

HAL Id: hal-00556047 https://hal.science/hal-00556047

Submitted on 15 Jan 2011

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Accepted Manuscript

Title: Capnocytophaga canimorsus

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PII:	S0378-1135(09)00076-5
DOI:	doi:10.1016/j.vetmic.2009.01.040
Reference:	VETMIC 4355
To appear in:	VETMIC
Received date:	5-11-2008
Revised date:	22-1-2009
Accepted date:	27-1-2009



Please cite this article as: Gaastra, W., Lipman, L.J.A., Capnocytophaga canimorsus, *Veterinary Microbiology* (2008), doi:10.1016/j.vetmic.2009.01.040

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1	Type of article: Review
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19 Abstract

20 *Capnocytophaga canimorsus* is a commensal bacterium in the oral flora of dogs and cats. The 21 bacterium is a zoonotic agent and has been isolated from humans, infected by dog or cat bites, 22 scratches, licks or simply exposure to dogs or cats. Here the infectious agent, its pathogenicity and 23 potential virulence factors, infection in animals and humans, diagnostic methods, prevalence, therapy 24 and prevention are described. Suggestions for future research are given.

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Keywords: Capnocytophaga canimorsus, zoonosis, sepsis, virulence, risk factors, therapy.

27

28 Introduction

Worldwide millions of people are bitten by animals each year. It is estimated that 50% of all Americans will be bitten at least once in their lifetime. Ninety percent of these bites are caused by dogs and cats. The majority of dog and cat bite wounds is minor and no medical attention is sought by the victim. Occasionally a dog or cat bite is complicated and an overwhelming systemic infection occurs. In the USA the annual mortality rate due to dog and cat bites is 6.7 per 10⁸ persons, albeit that not all these fatalities are caused by infections (Griego et al., 1995).

35 The genus *Capnocytophaga* consists of seven species which inhabit the oral cavity of humans or domestic animals. Capnocytophaga gingivalis, Capnocytophaga ochracea, Capnocytophaga 36 37 sputigena, Capnocytophaga granulosa and Capnocytophaga haemolytica are inhabitants of the human oral cavity and have been associated with peridontitis. Capnocytophaga cynodegmi and 38 *Capnocytophaga canimorsus* (latin canis = dog and latin morsus = bite) are part of the oral microbiota 39 of canines and more rarely of cats. Eight percent of 50 dogs tested positive for C. canimorsus in the 40 41 first report on the prevalence of C. canimorsus (Bailie et al., 1978). In a later report C. canimorsus 42 could be cultured from 26% of the dogs tested and from 18% of the tested cats. The agent could not be cultured from hamsters and humans (Blanche et al., 1998). In our laboratory, a PCR test with primers 43 for the 16S rRNA gene of C. canimorsus was positive for 41% of the dogs (de Poel et al., unpublished 44 results). An even higher percentage was observed with a similar PCR by van Dam et al (manuscript 45 submitted for publication). In a single report, the isolation of C. canimorsus (DF-2) from sheep and 46

47 cattle (25-30% of the animals tested) but not from pigs was reported (Westwell et al., 1989). To our knowledge no attempt to confirm this observation has ever been made. C. canimorsus can cause 48 49 zoonotic infections ranging from very mild flu like symptoms to fatal sepsis (de Boer et al., 2007, Pers et al., 1996). C. canimorsus infections are associated with dog and cat bites (54% of cases), dog and 50 cat scratches (8.5% of cases) or close animal contact (27% of cases), such as licking of human wounds 51 (Lion et al., 1996:). Consequently human to human transmission of C. canimorsus has not been 52 53 reported apart from one case where it could not be excluded 100% (Risi and Sprangler 2006). Studies 54 on C. canimorsus and C. cynodegmi infections have mainly been initiated from an interest in human medicine for rapid diagnosis and antibiotic treatment, prompted by the often dramatic outcome of an 55 infection with C. canimorsus. Since its first description in 1976 approximately 200 human cases of C. 56 canimorsus infection have been reported worldwide (Macrea et al., 2008). Surprisingly little research 57 on this zoonotic agent has been performed from a veterinary background. Regarding the millennia old 58 59 bond between men and dog, this infection is probably not new, but only now it can be recognized due to the advances in modern medicine over the last four decades. 60

61

62 The Infectious Agent

63 Historical perspective

64 In 1976 a patient with meningitis and sepsis, caused by an unidentified Gram-negative 65 bacillus, after a recent dog bite was first described (Bobo and Newton, 1976). At that time, the phenotypic as well as the biochemical characteristics of the agent also became known. We now know 66 that this Gram-negative bacillus probably was a member of the *Capnocytophaga* family, very likely *C*. 67 *canimorsus*-(CDC group DF-2 or dysgonic fermentor 2, dysgonic = slow and relatively poor growth of 68 69 a bacterial culture). One year later, 17 cases of infection after recent dog bites, the majority caused by 70 an unidentified Gram-negative rod were described (Butler et al., 1977). The nature of the infection varied from cellulitis to meningitis, endocarditis to organ failure and fatal sepsis. The organism was 71 given its current name in 1989, based on its CO₂ requirement and usual mode of transmission (Brenner 72 et al., 1989). Since then, C. canimorsus infections were found to occur worldwide and have been 73 reported from the United States, Canada, Europe, Australia and South Africa. 74

75 Phenotypic and Genotypic Characteristics

Capnocytophaga canimorsus are capnophilic (greek:carbon dioxide loving) facultative anaerobic, fastidious Gram-negative rods (1-4µm long, Fig.1). They are fusiform or filamentous gliding bacteria and closely related to *Fusobacterium* and *Bacteroides* species (Andersen and Pedersen, 1992, Deshmukh et al., 2004). However, the gliding motility is not always apparent. The G+C content of *C. canimorsus* DNA is 35% (Speck et al., 1987; Brenner et al., 1989).

81

82 Cultural properties

Classically, the organism was regarded difficult to culture because of its specific requirements 83 for nutrients and thus may have easily escaped detection. C. canimorsus has been shown to require 84 85 large amounts of exogenous iron for growth (Brenner et al., 1989). It grows slowly on blood (5% sheep blood in Columbia agar) or chocolate agar in 10% CO₂ (Ciantar et al., 2001). Media had to be 86 incubated at 37°C for at least 5 days. Colonies could be not visible for 2-7 days, on blood agar. 87 Initially they are pin point in size and later become larger, convex and smooth. Colonies are non 88 89 hemolytic with the characteristic shiny spreading edge and finger-like projections of bacteria with gliding motility (Andersen and Pedersen, 1992, Desmukh et al., 2004). Although the color of colonies 90 91 may vary from pink to yellow, the colonial mass of all isolates is yellow when scraped from the agar 92 plate (Forlenza, 1991). Horse blood agar plates were found to be superior to sheep blood but do not seem to be used often (Westwell et al., 1989; Pers et al., 1996). The organism does not grow on 93 MacConkey agar. Recently, the growth medium was optimized (Heart Infusion agar Difco with 5% 94 sheep blood) and the bacteria were shown to grow in 2 days at 37°C in 5 % CO₂ (Shin et al., 2007, 95 Mally et al., 2008). To our knowledge, these growth conditions have only been used under 96 97 experimental conditions and not yet in a clinical setting. It is to be expected that faster growth of C. 98 *canimorsus* will be reflected in the speed with which diagnostics can be performed and therefore lead 99 to less fatalities due to this organism.

100

101 Biochemical Characteristics

102 Biochemical tests are difficult to perform due to the slow growth of the organism. It is 103 catalase, oxidase, ONPG (O-nitrophenyl-β-D-galactoside) and arginine dihydrolase positive and 104 negative for urease, nitrate, indole, DNase, gelatin, lysine and ornithine (Hicklin et al., 1987; Brenner 105 et al., 1989). Members of the genus Capnocytophaga can use various carbohydrates like glucose, dextran, glycogen, inulin or starch as fermentable substrates and energy source. For a comparison of 106 107 the biochemical characteristics of C. ochracea, sputigena, gingivalis, canimorsus and cynodegmi see 108 Forlenza (1991). Gas-liquid chromatography of cellular fatty acids provides an additional rapid test for 109 differentiation of Gram-negative bacteria associated with dog-bite infections, including C. canimorsus (Dees et al., 1981). 110

111

112 Pathogenicity and Virulence Factors

Little is known about the pathogenesis of C. canimorsus infection. The bacterium was 113 reported to multiply in mouse macrophages and to be cytotoxic probably because it produces a toxin 114 (Fisher et al., 1995). In vitro, a very low level of cytokine production i.e. in comparison to N. 115 116 meningitidis was observed (Frieling, et al., 1997). Cytotoxicity could not be demonstrated by Shin et al., (2007) in ten strains of C. canimorsus tested, including the same strain as in the original study. 117 These strains also did not affect the viability of macrophages from different origin (mouse and 118 119 human). The clinical course of C. canimorsus infection strongly indicates that the organism is able to 120 avoid the immune system (at least in the early stages of infection). The absence of a proinflammatory 121 response is due to the fact that C. canimorsus does not interact with human Toll-like receptor 4 (TLR4). The effector for TLR4 is the lipid A moiety of LPS, which indicates that lipid A of C. 122 123 canimorsus is different from that of other pathogens. Capnocytophaga canimorsus is able to down 124 regulate TLR4 and the proinflammatory signaling cascade (Shin et al., 2007). Not only is C. 125 *canimorsus* resistant against phagocytosis and killing, it also blocks the killing of unrelated bacteria by macrophages (Meyer et al., 2008). Despite being classified as a fastidious grower, C. canimorsus 126 exhibits robust growth when it is in direct contact with mammalian cells, including phagocytes (Mally 127 et al., 2008). This is dependent on a surface located sialidase, which allows the bacterium to use 128 internal amino-sugars from glycan chains of host cell glycoproteins (Mally et al., 2008). The 129

molecular biology of *C. canimorsus* is still in its infancy, but promising genetic tools for this bacterium have recently been developed using a naturally occurring plasmid found in one of eight strains tested (Mally and Cornelis, 2008).

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134 Infection in Animals

The clinical syndrome in rabbits after experimental infection with *C. canimorsus* was characterized by disseminated intravascular coagulation, cellular necrosis in organs like kidneys and adrenal glands, cutaneous gangrene, thrombocytopenia, hypotension, hemorrhagic diathesis with purpuric lesions and petachiae and renal failure and is similar to that observed in man (Piccininno et al., 1984).

Only two reports have appeared in species other than humans that suffered from a dog 140 141 inflicted C. canimorsus infection. In one case the bacterium was cultured from an infected, bite wound in a pet rabbit (van Duijkeren et al., 2006) (Fig.2). In the other C. canimorsus was isolated from an 142 infected bite wound in a dog (Meyers et al., 2008). The dog did not have bacteremia, but C. canimorsus 143 144 could still be isolated from the wound two days after the primary isolation (J. Picard, personal communication). Curiously both bite wounds were on the head. No C. canimorsus infections of cats 145 after a bite incidence have been reported, but there is a case report of a cat with chronic sinusitis and 146 147 rhinitis due to a Capnocytophaga infection. The organism was identified by a 97% identity with C. canimorsus of its 16S rDNA sequence (Frey et al., 2003). The incidence of bite wounds inflicted in 148 dogs and cats by dogs or cats is not well known. It is estimated that they comprise between 10-15% of 149 all trauma related emergencies (Kolata et al., 1974). Like with human bite wounds many wounds are 150 not reported and owners do not seek veterinary attention. In the mobile, elastic skin of dogs and cats, 151 152 bite injuries may be less noticeable or may migrate through deeper tissues and escape detection. Dogs 153 and cats may also be less prone to C. canimorsus infection, which might explain the very low number of reported C. canimorsus infection in animals. 154

155

156 Human Infections and Clinical Symptoms

157 The incubation period from the dog bite to the onset of systemic symptoms is about 5 days. On average seven days pass between the time of the bite and hospitalization (range 1-14 days) (LeMoal et 158 159 al., 2003). Patients who arrive at the emergency room within 8-12 hour after a dog bite may show 160 local lesions without significant signs of inflammation. At later stages, the infection may have symptoms as: localized cellulitis, pain at the site of injury, a purulent discharge, lymphangitis and 161 regional lymphadenopathy. The initial symptoms of septicemia are fever (78% of patients), chills 162 163 (46%), myalgia (31%), vomiting (31%), diarrhea (26%), abdominal pain (26%), malaise (26%), dyspnea (23%), mental confusion (23%) and headache (18%) (Pers et al., 1996). A fulminant and 164 165 severe course of the infection may occur in immuno-compromised persons, characterized by sepsis, meningitis, osteomylitis, peritonitis, endocarditis, pneumonia, purulent arthritis, disseminated 166 167 intravascular coagulation (DIC) and fulminant purpura but has also been observed in previously healthy persons (Hantson et al., 1991). DIC can lead to peripheral gangrene of the upper and lower 168 extremities, lips nose and ears (often resulting in amputation). Purpura is a group of blood disorders 169 170 that cause bleeding in the tissue from the small blood vessels, shock and multi organ failure. They are 171 often under or in the skin as tiny pinpoint petechiae (point-shaped skin bleedings or larger blue spots (ecchymosis). Hemolytic uremic syndrome and Waterhouse-Friderichsen syndrome have also been 172 reported (Tobé et al., 1999; Mirza et al., 2000; Mulder et al., 2001; Macrea, et al., 2008). C. 173 174 canimorsus septicemia has been associated with up to 30% mortality (Lion et al., 1996) and significant 175 morbidity, including amputation secondary to gangrene, myocardial infarction and renal failure. Curiously, Lion et al. (1996) report 22% mortality in asplenic individuals, 28% in individuals with 176 other predisposing conditions (alcohol abuse, immunosuppression) and 32% in individuals with no 177 risk factor. The mortality rate from C. canimorsus meningitis is low (5%) compared to that from C. 178 179 canimorsus septicemia (LeMoal et al., 2003; Janda et al., 2006). In a review of 12 cases of C. 180 canimorsus endocarditis a mortality rate of 25% was noted (Sandoe, 2004). A fatal outcome of the infection in non-immunocompromised individuals occurs with lower frequency than in 181 immunocompromised persons. Four of these cases were reported until 1991 and several others 182 between 1991 and 2008 (Hicklin et al., 1987, Hantson et al., 1991, Deshmukh et al., 2004). Ocular 183

infections associated with cat scratches or cat bites have been reported several times (Glasser, 1986,
Zimmer-Galler et al., 1996, Papadaki et al., 2008).

186

187 Risk groups

At special risk are animal keepers, veterinarians, breeders and pet owners. A survey of 188 veterinarians in the US showed that 65% sustained a major animal related injury of which animal bites 189 190 and scratches accounted for 38% of the traumas. Dogs and cats were involved in 24% and 10% of the 191 injuries. In their entire careers, 92% of the veterinarians was bitten by a dog, 81% by a cat and 72% was scratched by a cat (Landercasper et al., 1988). Yet only few reports of persons in this risk group 192 193 infected with C. canimorsus are available: an employee in an animal shelter (Deshmukh et al., 2004), a 194 pet shop worker (Hantson et al., 1991), three veterinarians (Job et al., 1988; de Smet et al., 1990; 195 Chodosh, 2001), a kennel operator (Tison and Latimer, 1986.) and a veterinary worker (Bobo and Newton, 1976), which is much lower than for rat bite fever, for which the same group is also at risk 196 197 (Gaastra et al., 2009). Two cases in veterinarians were not caused by bites or scratches, but by fracture 198 of a tooth during manual extraction which struck the veterinarians in the eye (de Smet et al., 1990; 199 Chodosh, 2001). In one case it was the tooth of a cat, in the other of a dog. C. canimorsus is primarily an opportunistic pathogen that infects individuals weakened by a compromised immunity. Most 200 infections in humans occur among people older than 40 years, but infections have been reported in a 4-201 202 month-old and 83 year-old patient, the ratio male to female is 3:1 (Danker et al., 1987; Pers et al., 1996, Deshmukh et al., 2004; Sandoe, 2004; Janda et al., 2006). Sixty percent of the people that get an 203 infection with this organism have predisposing conditions. Those who have had a splenectomy (33% 204 of cases) comprise a high-risk group with a 30-60 times increased risk of fatal septicemia (Singer, 205 1973, Deprés-Brummer et al., 2001). Often, already within 24 hours of symptom onset, overwhelming 206 207 postsplenectomy infections (OPSI) can result in a rapidly deteriorating clinical course that progresses to multiple organ system failure and death (Band et al., 2008; Sawmiller et al., 2008). In the United 208 States each year 25.000 splenectomies are performed (Sumaruju et al., 2001). Together with the 209 thousands of functionally hyposplenic or asplenic patients a significant number of people is therefore 210 at marked risk for OPSI, which has an estimated mortality rate of 70% (Brigden and Pattullo, 1999). 211

212 Asplenic individuals suffer deficient IgM and IgG production and delayed macrophage mobilization. They also produce less tuftsin, a protein derived from IgG that stimulates phagocytosis (August, 213 214 1988). Liver disease caused by alcohol abuse (24% of cases) and the use of cortisone (3% of cases) are other predisposing factors for the infection (Lion et al., 1996). Hematologic malignancy, neutropenia 215 and chronic lung disease were also considered as predisposing conditions (Dire et al., 1994). Up to 216 40% of cases however occur in previously healthy adults (LeMoal,et al., 2003). Alcohol abuse, 217 218 asplenia, hemochromatosis, β-thalassemia major and tobacco smoking lead to absorption of twice the 219 amount of alimentary iron as in normal persons. In two cases of immune-competent persons with C. canimorsus infection, both women, heavy smoking was explicitly mentioned (Finn et al., 1996; 220 Aslam, 1999). An alternative explanation for why these conditions form a risk for C. canimorsus 221 infection might be that this two-fold increase in serum iron values much better supports the growth of 222 the poor iron scavenger C. canimorsus (Weinberg, 2000). It was suggested that the presence of 223 detectable amounts of C. canimorsus in only one quarter of dogs and cats might be due to the fact that 224 positive animals have insufficient amounts of lactoferrin and other iron binding factors in their oral 225 226 cavity (Weinberg, 2000). Since the 25% is likely to be an underestimation based on data with improved selective cultivation conditions (see above) and PCR (see above), the suggested lack of 227 lactoferrin in C. canimorsus positive animals is unlikely. However, iron overload was suggested as a 228 229 risk factor in a case of C. canimorsus infection in a patient receiving numerous blood transfusions over 230 a two year period (Pletschette et al., 1992).

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232 **Diagnostics**

The diagnosis *C. canimorsus* infection is usually made based on the bacterial culture of blood (88% of cases), other body fluids (cerebrospinal fluid, 7% of cases) or less frequently from the bite wound or tissue from the bitten individual (Janda et al., 2006). Polyanethole-sulphonate, an anticoagulant frequently present in automatic blood culture systems, inhibits the growth of *C. canimorsus* (Sowden et al., 1995). Swabs of the mouth of the dog that bit the patients have been taken in a number of cases, but growth of *C. canimorsus* was only obtained in half of them (Martone et al., 1980; Howell and Woodward, 1990; Hantson et al., 1991; Valtonen et al., 1995; Finn et al., 1996;

240 Phipps et al., 2002; van de Ven et al., 2004). An indication of a C. canimorsus infection can already be obtained when a Gram stain of the buffy coat of peripheral blood shows many extracellular Gram-241 242 negative rods and several intracellular copies of the same rods in almost all neutrophils (Pedersen et al., 1993; Sawmiller et al., 1998; Mirza et al., 2000). In cases of overwhelming C. canimorsus 243 septicemia, whole blood smears have also been reported positive for Gram-negative rods in the Gram 244 stain (Holmes and Kozinn, 1986; Ndon et al., 1992; Newton, 2006, Kleijnen-Grebien et al., 2008). 245 246 Examination of the buffy coat by Gram stain in splenectomized patients is particularly useful when C. 247 *canimorsus* infection is suspected, since several days may pass before the organism is identified in the laboratory (Hicklin et al., 1987). The slow growth of C. canimorsus may lead to diagnostic problems 248 since culture plates are routinely discarded after five days and some strains do not grow at all. A Gram 249 stain of the cerebrospinal fluid showed Gram-negative bacilli in 65% of 20 cases of C. canimorsus 250 meningitis (de Boer et al., 2007). 251

The diagnosis of C. canimorsus infection was evaluated by Janda et al., (2006) with isolates 252 collected over a 30 year period. It was concluded that many laboratories are unable to identify C. 253 254 canimorsus isolates and report them as (fastidious) Gram-negative rods. In their identification procedure the authors use a combination of biochemical tests, fatty acid methyl ester analysis and 16S 255 rDNA sequencing (Janda et al., 2006). The number of cases in which broad range PCR followed by 256 DNA sequencing is used for diagnostic purposes is increasing (Ciantar et al., 2005; Gotwein et al., 257 258 2006; Janda et al., 2006; Meybeck et al., 2006; Wareham et al., 2007; Le Meur et al., 2008; Kleijnen-259 Grebien et al., 2008; Papadaki et al., 2008). It should be noted that in one case, amplification of C. canimorsus DNA from an infected valve was obtained while attempts to amplify the DNA from the 260 positive blood culture of the same patient was unsuccessful, probably due to the presence of 261 polyanethylsulphonate which inhibits the PCR reaction (Wareham et al., 2007). 262

263

264 Therapy and Prevention

265 The first choice antibiotic for infection with *C. canimorsus* is penicillin G, although resistance 266 of isolates has been mentioned (Meybeck et al., 2006). After investigation of the available literature 267 we came to the same conclusion as LeMoal et al. (2003), that no β -lactamase producing *C. canimorsus*

strain has been reported yet. There are however, several reports on β-lactamase producing 268 Capnocytophaga species (Arlet et al., 1987; Roscoe et al., 1992; Jolivet-Gougeon et al., 2007). With 269 270 broth or agar dilution techniques, susceptibility for penicillins, imipenem, clindamycin, chloramphenicol, third generation cephalosporins, fluoroquinolones, erythromycin, doxycycline and 271 metranidazole has been demonstrated. Aztronam, polymyxin, fosfomycin, trimethoprim / 272 sulfamethoxazole and aminoglycosides are not effective against this bacterium (Verghese et al., 1988; 273 274 Bremmelgaard et al., 1989; Desmukh et al., 2004). However, in the literature disagreement on the 275 results for trimethoprim / sulfamethoxazole, aminoglycosides and vancomycin can be found, which seem to depend on the method used. Routine prophylaxis with amoxicillin-clavulanic acid has been 276 proposed for all immunocompromised patients and in cases seen within 8 hours after moderate to 277 severe crush injuries or when the bones and joints are involved (Goldstein, 2005). Immediate cleaning 278 of the wound and disposal of scrap fabric filler are important during the antibiotic therapy. Although 279 systemic administration of antimicrobial agents is controversial for healthy persons, those from whom 280 the spleen was removed should be given prophylactic antibiotics against this organism after a dog bite 281 282 or in the case of an open wound which has been in contact with the saliva of a dog. In several countries systemic antibiotic treatment is recommended for all persons (not only immuno-283 compromised patients) after a dog bite. The clinical course of a patient with C. canimorsus sepsis, 284 285 which presented as an acute abdomen described by Sawmiller et al., (1998) clearly demonstrates the 286 importance of early diagnosis and treatment of an overwhelming postsplenectomy infection. One may 287 wonder whether there is an indication to test healthy pets for C. canimorsus, in households with highrisk individuals and if there is an indication to treat colonized pets in these households. Given the high 288 prevalence of dogs when tested by PCR for C. canimorsus, it is very likely that dogs in high risk 289 290 households will test positive. It is unclear what the advance of treating the dogs in these households 291 will be since a treated "C. canimorsus free" dog can readily be colonized again after contact with other 292 dogs. There are no reports on dogs that have been tested several times over a period of time. So it is not known whether a negative dog will stay negative during its whole life or whether a positive dog 293 will always test positive a next time. Furthermore, since a PCR test for C. canimorsus has not been 294

validated it is not clear what the value of a negative PCR is with respect to that particular dog beingcolonized or not

297

298 Prevalence

Worldwide millions of people are bitten by animals each year (one out of every two persons 299 will be bitten by an animal at some point in his lifetime (Greene and Goldstein, 2006). Ninety percent 300 301 of the bites are by dogs and cats (Griego et al., 1995, Matter and Sentinella, 1998). Less than half of the bites are reported and 18% of these persons seek medical attention (Overall and Love, 2001). Most 302 bites are minor and occur (in descending order of incidence) on the extremities (hands and feet), head 303 and neck and the fuselage. Although 80% of dog bite wounds inflicted on people culture positive for 304 bacteria, the chance that a dog bite gets infected is between 3 and 20 % (Underman, 1987, Goldstein, 305 1992). For cat bites, this is between 20 and 50% (Talan et al., 1999). Usually the infection is caused by 306 a combination of aerobic and anaerobic bacteria. In the Netherlands yearly between 50.000 and 307 100.000 people get bitten by a pet animal. The number of registered bite incidents by dogs or cats in 308 309 children under the age of ten in The Netherlands is about 4800 per year (Deprés-Brummer et al., 2001). Since 1976, worldwide approximately 200 cases of septicemia, meningitis or gangrene caused 310 by C. canimorsus have been reported in humans bitten, scratched or licked by a dog or cat (Brenner et 311 al., 1989, Blanche et al., 1998, Tierney et al., 2006). Even though almost equal percentages of dogs 312 313 and cats (24% vs. 18%)(Blanche et al., 1998) carry C. canimorsus in the oral cavity, the majority of cases is caused by dog bites or dog contact. Cat bites, scratches or contact with cats has been reported 314 in less than 10 % of the cases. The deep, sharp, needle like punctures with relatively little tissue 315 damage through cat bites are probably creating less favorable conditions for the growth of the 316 bacterium than the larger superficial wounds with a lot of tissue damage caused by dog 317 318 bites.(Carpenter et al., 1987; Scarlett et al., 1991; Mahrer and Raik, 1992; Valtonen et al., 1995; Zimmer-Gallen et al., 1996; McLean et al., 2004; Le Meur, et al., 2008). The risk of infection depends 319 on adequate wound treatment and host factors like age, health status. 320

Infection with *C. canimorsus* is not a notifiable disease. The number of reports on *C. canimorsus* infection is increasing, but it is still a relatively rarely reported disease. The frequency of

323 C. canimorsus infection is very likely underestimated (Janda et al., 2006). Presently, not even all severe and lethal infections are reported but only those cases where the clinical picture has not been 324 325 seen before. Additionally, patients who are supplied with antibiotics in time do not develop a C. canimorsus infection. This may also influence the infection rate. The bacteria may be transmitted in 326 various ways by close contact between pets and their owners. Transmission is not necessarily followed 327 by multiplication of the bacteria in the human body (infection). Infection does not necessarily lead to 328 329 serious clinical symptoms and humans with subclinical infections will not report to the physician. If 330 the incubation period extends to several weeks and clinical symptoms are aspecific, C. canimorsus infection is probably not considered when bites by dogs or cats or contact with these animals are not 331 explicitly mentioned in the anamnesis. 332

Patients suspected of *C. canimorsus* infection will often be treated with antibiotics most of which will be active against the causative bacteria. The number of cases in which laboratory diagnostic examinations are carried out is therefore limited to very severe cases or when antibiotic therapy fails.

C. canimorsus may be difficult to grow in primary culture after antibiotic therapy and detection by broad range PCR and nucleotide sequence determination is operational in a limited number of diagnostic laboratories only. *C. canimorsus* strains may be misidentified despite the fact that the bacteriologic characteristics are rather typical.

341

342 Conclusions

C. canimorsus is a commensal in the oral cavity of dogs ands cats and is considered to be of low virulence under normal circumstances. The bacterium can cause severe illness in predisposed patients (i.e. those who have had a splenectomy, who are alcohol abusers, suffer from chronic lung or liver diseases or are on immunosuppressive medication). However, 40% of the reported cases occurred in previously healthy people.

The true number of *C. canimorsus* infections each year probably is largely underestimated due to the fastidious growth of the organism. The isolation from animal saliva is difficult and the organism is seldom isolated from bite wounds. The relatively more successful growth from blood culture can explain why the bacterium is only detected in cases of septicemia. Penicillin is considered the drug of choice in the case of *C. canimorsus* infection. The length of the treatment varies between reports (14-21 days). (Le Moal et al., 2002).

In the past, diagnosis and identification have been difficult due to the fastidious growth of the 354 organism, which has lead to fatal outcomes in cases where the patient probably could have survived 355 since the bacterium is susceptible to a whole range of commonly used antibiotics. With the 356 357 introduction of optimized growth conditions on selective media and PCR as a diagnostic tool this type of fatalities is expected to decrease. Because of the seriousness of C. canimorsus infection, taking this 358 bacterium into account in the management of animal bites, is necessary. Moreover, high risk patients 359 should be informed about the risks of keeping of pets. Not only do bites and scratches pose a risk to 360 these patients but other scenario's one not normally thinks of are possible. This can be illustrated by a 361 case report where an 18 year old person on automated peritoneal dialysis developed C. canimorsus 362 peritonitis following the puncture of his dialysis tubing by a domestic cat that slept in his bed (Chada 363 and Warady, 1999). 364

Administration of antimicrobial agents is controversial for healthy persons, but it is justifiable for the following groups of bite victims: those with deep wounds, particularly to the face or hands and those at high risk for *C. canimorsus* infection e.g. immuno-compromised patients.

368

369 Future research

More insight in the genetic properties and the molecular biology of C. canimorsus is a 370 prerequisite for understanding the overwhelming sepsis, this organism can cause. In this respect whole 371 genome sequencing of strains from different origin seems relevant. Questions to be answered are: are 372 isolates from humans and dogs different? Or are the former a more virulent subset of the latter? Is 373 374 there a difference between dog and cat isolates? Once the whole genome of C. canimorsus has been sequenced this might allow the identification of the genetic basis of virulence factors and the reason 375 for the low virulence normally seen in immunocompetent bite victims. To improve diagnostics, media 376 should be developed that allow rapid growth of the organism. For the following rapid identification the 377 development of a species specific PCR is needed. This can eventually be followed by sequencing of 378

the amplicon for confirmation. A systematic search for the presence of *C. canimorsus* in the oral cavity of other animals than cats and dogs is also needed to determine the potential risk with bites from these animals. So far there are no indications that point to a host specificity that is limited to dogs and cats.

383

384 Acknowledgements

The authors are indebted to Chantal van Mourik, Engeline van Duijkeren (Veterinary Microbiological Diagnostics Centre, Veterinary Faculty, Utrecht University) and Marcel Leuven (Animal Clinic Leuven, Heerlen) for Fig. 1 and Fig. 2.

388

389 **Conflict of interest:**

None of the authors (Wim Gaastra, Len J. A. Lipman) has a financial or personal relationship
with other people or organizations that could inappropriately influence or bias the paper
entitled "Capnocytophaga canimorsus".

393

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- 606

607 Figure Legends

- **Fig.1.** Gram stain of *C*.*canimorsus* isolate from a rabbit bitten by a dog.
- **Fig.2**. Rabbit with *C. canimorsus* infection after a dog bite in the head.



