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Waiting Time Distributions For Clusters of IP₃ Receptors

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Abstract

Inositol 1, 4, 5-trisphosphate (IP₃R) receptors control release of Ca^{2+} from the endoplasmic reticulum into the cytosol of a cell. The binding of both 1, 4, 5trisphosphate (IP₃) and activating Ca^{2+} is required for the receptor to open. At high Ca^{2+} concentrations, IP₃Rs are inhibited. IP₃Rs are composed of four identical subunits and form in clusters. Many models have been proposed to describe how the binding of IP₃ and Ca^{2+} to subunits results in the opening and closing of IP₃Rs. Here we compare the opening and closing probability distributions for clusters of IP₃Rs, resulting from three different models. The distributions are calculated both analytically, using a method we have developed, and with simulations. We found significant differences in the behaviour of the three models as the Ca^{2+} and IP₃ concentrations are varied.

Running title: Waiting Time Distributions

Keywords: Inositol 1, 4, 5-trisphosphate receptor, calcium dynamics, ion channels

1 Introduction

The inositol 1, 4, 5-trisphosphate receptor (IP₃R) acts as a channel which controls release of Ca^{2+} from the endoplasmic reticulum (ER) into the cytosol of a cell. The IP₃Rs release Ca^{2+} in response to an increase in the concentration of inositol 1, 4, 5trisphosphate (IP₃) in the cytosol, which results from a stimulus, such as the binding of a hormone to the cell membrane. IP₃Rs open and close randomly, and the open and closing probabilities are determined by the binding of IP₃ and Ca²⁺. A small increase in the concentration of cytosolic Ca²⁺ stimulates channel activation, whereas a large increase causes inhibition of the IP₃R. This results in a bell-shaped dependence of the open probability on the cytosolic Ca²⁺ concentration. Activation of the channels takes place on a faster time scale than inhibition. IP₃Rs are found in clusters, and so the release of Ca²⁺ resulting from the opening of one channel in a cluster, stimulates the opening of other channels in the cluster. The release of Ca²⁺ from the cluster is referred to as a Ca²⁺ puff. Puffs are the elemental events of intracellular Ca²⁺ release.

Various models have been proposed for describing the IP₃R [15, 7]. Here we have compared the opening and closing probability distributions resulting from three different models. These include the model proposed in Rüdiger et al. and Shuai et al. [10, 12] (model 1), the same model modified to include sequential binding of IP₃ and Ca²⁺ as suggested by Adkins and Taylor [1] (model 2), and the model developed by Sneyd and Dufour [14] (model 3) as given in reference Ullah et al. [19]. The IP₃R is comprised of four subunits, which are assumed to be identical, and these three models each describe the transitions between subunit states, most of which result from the binding and release of IP₃R and Ca²⁺. Model 1 [12] is closely based on the DeYoung-Keizer model [3], and assumes that a subunit has binding sites for IP₃, activating Ca²⁺ and inhibitory Ca²⁺. The model by Sneyd and Dufour [19, 14, 7] was designed to agree with several experimental observations.

While it has been known for about a decade now that puffs are random release events, we recently showed that spike sequences of intracellular Ca^{2+} oscillations are random events also [13]. The random opening and closing of individual channels, i.e.

the random state transitions of their subunits, is believed to be the most important source of noise causing the random behavior of the whole system [6]. That requires stochastic theory across all structural levels from a channel subunit up to cell level for modelling. However, existing subunit models typically comprise 6 or more subunit states, leading to hundreds of channel states and many thousands of channel cluster states. That makes stochastic theory on cluster level already almost impossible, not to mention cell level. Thus theoretical studies on puff characteristics used either extremely simplified channel models (with two subunit states [11], two or four channel states [9]) or relied on simulations [19].

Recently we presented a method which circumvents writing down the master equation of the channel molecule with its huge number of states and gets by with the subunit master equation only in calculations of waiting time distributions for channel clusters [18]. Here we follow a different approach. Channel states are defined by the number of subunits in subunit states. We derive the channel Master equation from the subunit Master equation. Subsequently, we lump channel states into aggregates based on time scale separation. Standard methods for calculating cluster properties can then be applied to the Master equation defined for the lumped states.

Using these approximations, we compared the probability distributions for the first opening time of a cluster and the first closing time of a cluster, at various IP₃R and Ca²⁺ concentrations. These distributions were calculated analytically as well as through simulations, and assume a cluster consists of five channels. The three models exhibit significantly different behaviour. The sequential binding of IP₃ and Ca²⁺ imposed in model 2 results in longer expected opening times compared to model 1 and longer expected closing times compared to both models 1 and 3. Model 3 has a shorter expected closing time than models 1 and 2, but has a much longer expected opening time at low IP₃ concentrations and high Ca²⁺ concentrations. These probability distributions are calculated assuming the Ca²⁺ concentration in the vicinity of the cluster remains constant. That is, we use one fixed value for the Ca²⁺ concentration when the cluster is closed, and another fixed value for Ca²⁺ when the cluster is open, i.e., when at least one channel is open. We have also looked at the effect of using a

variable Ca^{2+} concentration while the cluster is open, whereby a closed channel sees a lower concentration than an open channel. The resulting probability distributions are very similar to when a fixed value is used, in all three models.

2 Single Channel Activation

We would like to calculate F(A, t|I, 0), the probability distribution for a channel first being activated at time t, given it is in state I at time t = 0. A denotes the set of active channel states, and we say a channel is activated when it is in one of these states. We are assuming we have the subunit transition matrix W, where $w_{i,j} dt$ gives the probability for a subunit to move from state x_j to state x_i in infinitesimal time dt, for $i \neq j$. $w_{j,j} = -\sum_{i=1, i\neq j}^n w_{i,j}$ where n is the number of subunit states. The probability for a subunit to stay in state x_i during time dt is $1 + w_{i,i} dt$.

We demonstrate how to build the subunit transition matrix W with model 1 (see section 5 for a detailed description of the model). First, we order the subunit states: (110 + Active, 111, 010, 011, 100, 101, 000, 001). For instance the transition rate from state 100 to 110+Active is a_5c where c denotes the Ca²⁺ concentration. Now we can build the 8 × 8 subunit transition matrix

$$W = \begin{pmatrix} -\sum_{1} & b_{2} & a_{1} \cdot p & 0 & a_{5} \cdot c & 0 & 0 & 0 \\ \frac{b_{0}a_{2}}{a_{0}+b_{0}} \cdot c & -\sum_{2} & 0 & a_{3} \cdot p & 0 & a_{5} \cdot c & 0 & 0 \\ \frac{b_{0}b_{1}}{a_{0}+b_{0}} & 0 & -\sum_{3} & b_{4} & 0 & 0 & a_{5} \cdot c & 0 \\ 0 & b_{3} & a_{4} \cdot c & -\sum_{4} & 0 & 0 & 0 & a_{5} \cdot c \\ \frac{b_{0}b_{5}}{a_{0}+b_{0}} & 0 & 0 & 0 & -\sum_{5} & b_{2} & a_{1} \cdot p & 0 \\ 0 & b_{5} & 0 & 0 & a_{2} \cdot c & -\sum_{6} & 0 & a_{3} \cdot p \\ 0 & 0 & b_{5} & 0 & b_{1} & 0 & -\sum_{7} & b_{4} \\ 0 & 0 & 0 & b_{5} & 0 & b_{3} & a_{4} \cdot c & -\sum_{8} \end{pmatrix}$$

All entries have dimension s^{-1} , p denotes the IP₃ concentration. $-\sum_i$ represents the negative sum of the *i*th column.

We can use W to form the channel state transition matrix P. An entry in P, $p_{i,j} dt$, gives the probability for a channel to move from state X_j to state X_i in infinitesimal

time dt, for $i \neq j$. The diagonal elements are given by $p_{j,j} = -\sum_{i=1, i\neq j}^{r} p_{i,j}$. The off-diagonal elements of P are found as follows. A channel state is represented by a vector V where an element of the vector, v_i , gives the number of subunits in subunit state x_i . Thus the length of V is n, and the sum of the elements in V is the number of subunits in each channel, h. The transition probability $p_{i,j}$ is 0, if X_j and X_i differ by more than one subunit and $p_{i,j} = \phi w_{k,l}$, if X_i can be reached from X_j by one subunit in state x_l moving to state x_k , and X_j has ϕ subunits in state x_l .

To find F(A, t|I, 0) we first remove all rows and columns in P which correspond to active channel states and denote the new matrix by \overline{P} . That is, if $X_i \in A$ then we remove row i and column i. This means if the system reaches an active state, it cannot leave it. Let $y(X_i, t|I, 0)$ be the probability of being in nonactive state X_i at time t and let $y = \{y(X_i, t|I, 0), X_i \notin A\}$. Then y can be found by solving the master equation

$$\dot{y} = \bar{P}y.$$

Let f(A, t|I, 0) be the probability that an activated state is reached in the time interval [0, t], let r be the total number of channel states and let r_a be the number of open channel states. Then

$$f(A,t|I,0) = 1 - \sum_{i:X_i \notin A} y(X_i,t|I,0) = 1 - \sum_{i=1}^{r-r_a} \sum_{k=1}^{r-r_a} c_k V_{k,i} e^{\lambda_k t}$$

where λ_k and $V_k, k = 1...r - r_a$ are the eigenvalues and eigenvectors respectively, of \bar{P} and where $c_k, k = 1...r - r_a$ are determined by the initial conditions. Then

$$F(A,t|I,0) = \frac{df(A,t|I,0)}{dt} = -\sum_{i=1}^{r-r_a} \sum_{k=1}^{r-r_a} c_k \lambda_k V_{k,i} e^{\lambda_k t}.$$

3 Cluster Activation

A cluster is first activated when any of the channels in the cluster are first activated. We assume the initial state of a channel is unknown, so we use the weighted average over all possible initial channel states, where the weights are given by the probabilities of each channel state occurring at rest. Let $F_{ch}(A, t)$ denote the probability distribution for a channel to first open at time t given that the initial state is unknown

and let $F_{\text{open}}(A, t)$ denote the probability distribution for a cluster of channels to first open at time t (opening time distribution). Then $F_{\text{open}}(A, t)$, for t > 0, is given by

$$F_{\text{open}}(A,t) = NF_{ch}(A,t)G(A,t)^{N-1}$$

where

$$G(A,t) = 1 - \int_0^t F_{ch}(A,\tau) d\tau$$

gives the probability that a channel has not been activated by time t and N is the number of channels in the cluster. That is, the probability for the cluster to be first activated in the time interval [t, t + dt] is given by the probability that one of the N channels is first activated in the time interval [t, t + dt] and all other channels have not yet been activated.

When t = 0, if we used the method above to calculate $F_{\text{open}}(A, 0)$, this would mean that the probability for the cluster to open in the interval [0, dt] is given by the probability that one of the N channels opens in [0, dt], while the other N - 1 do not. However, this is not correct. The probability that the cluster opens in [0, dt] is given by the probability that at least one of the N channels opens in [0, dt], while the other N - 1 channels can either open or not open. $F_{\text{open}}(A, 0)$ can be calculated using

$$F_{\text{open}}(A,0) = 1 - (1 - F_{ch}(A,0))^{N}.$$

That is, the probability that the cluster opens instantly is the negation of no channel opening instantly.

We use the non-open states weighted according to their resting probabilities as initial condition for calculating F(A, t) for t > 0. The resting probabilities for the initial channel states are given by the normalized eigenvector of P corresponding to the eigenvalue $\lambda = 0$.

4 Cluster Closure

The cluster of channels is considered closed when all channels in the cluster are closed. Let $F_{\text{close}}(A, t)$ denote the probability distribution for a cluster of channels

to first close at time t if it has opened at time t=0 (closing time distribution). To calculate $F_{\text{close}}(A, t)$, we find the cluster state transition matrix, M, from the channel state transition matrix P, in an analogous way to how we found P from the subunit state transition matrix, W. That is, a cluster state is represented by a vector with length given by the number of channel states. An entry in the vector gives the number of channels in the corresponding channel state. Thus, the sum of the elements in the vector is the number of channels in the cluster, N. The probability of a transition between cluster states occurring which involves more than one channel changing state instantaneously is infinitely small compared to the probability of a transition where only one channel changes state, and so for such transitions, the entry in M is 0. If cluster state \mathcal{X}_i can be reached from state \mathcal{X}_j by one channel in state X_l moving to state X_k , and \mathcal{X}_j has ϕ channels in state X_l , then the entry in M, $m_{i,j} = \phi p_{k,l}$. The diagonal entries of M are such that the columns of M sum to 0.

Once M has been constructed, $F_{\text{close}}(A, t)$ can be found analogously to the way in which F(A, t|I, 0) is found in section 2. That is, let \bar{M} be the cluster state transition matrix with rows and columns corresponding to closed cluster states removed. Let ρ be the total number of cluster states and let ρ_a be the number of closed cluster states. Let μ_k and $U_k, k = 1...\rho - \rho_a$ be the eigenvalues and eigenvectors respectively, of \bar{M} . Then

$$F_{\text{close}}(A,t) = -\sum_{i=1}^{\rho-\rho_a} \sum_{k=1}^{\rho-\rho_a} d_k \mu_k U_{k,i} e^{\mu_k t}.$$

where $d_k, k = 1...\rho - \rho_a$ are determined by the initial conditions.

We choose as the initial probabilities for a channel to be in each of the channel states the probabilities at the expected cluster opening time. That is, we begin with the resting probabilities, then solve the master equation,

$$\dot{z}(t) = P_0 z(t)$$

where z(t) gives the probabilities of being in the channel states at time t, and P_0 is found from P by setting transition rates out of open states to 0, because we ave interested in those trajectories only, which did not go through open states. z(t) is then evaluated at the expected opening time for the cluster, which is calculated from

 $F_{\text{open}}(A, t)$. Initially, we assume one channel in the cluster is open, while the others are closed. To enforce this condition, we find from z(t) the conditional probability of being in each state where exactly one channel is open, given that the cluster is in one such state. All other cluster states have an initial probability of zero.

The large number of cluster states presents a problem when calculating $F_{\text{close}}(A, t)$. The number of channel states, r is given by $r = \binom{h+n-1}{n-1}$. Recall, n is the number of subunit states and h is the number of subunits per channel. The number of cluster states, ρ , is given by $\rho = \binom{N+r-1}{r-1}$. Recall, N is the number of channels in the cluster. So ρ is very large for the values of n, h and N we require, which results in the above method for finding $F_{\text{close}}(A, t)$ being too slow and requiring too much memory to be feasible.

To overcome the problem of there being too many cluster states, we reduce the number of channel states before finding M. We developed a reduction method motivated by and similar to that described in Huisinga et al. [4]. It is based on the theory of weakly coupled Markov chains. The weakly coupled state aggregates are identified by a Perron cluster analysis in the study by Huisinga et al. We replace cluster identification by a simpler method. In one case we compared our results with those of the method used in [4] and found them to be identical.

We identify aggregates by repeatedly finding the largest transition rate in the channel state transition matrix P, then combining the two states connected by the transition, into one aggregate state. In general we combine two states X_i, X_j to an aggregate state $\{X_i, X_j\}$ and after repeating the reduction process we get a set of aggregate states of the form $\{X_{a_i}, i = 1 \dots m\}$. The transition rate for moving from the aggregate state $\{X_{a_i}, i = 1 \dots m\}$ to the aggregate state $\{X_{b_i}, i = 1 \dots m_2\}$ is then given by

$$\sum_{j=1}^{m_2} \left[\left(\sum_{i=1}^{m_1} p_{b_j, a_i} V_0(a_i) \right) / \sum_{i=1}^{m_1} V_0(a_i) \right].$$

 V_0 represents the stationary solution of the whole un-reduced system and p_{b_j,a_i} denotes the transition probability from state a_i to b_j . This is repeated until the number of aggregate channel states is sufficiently small that the number of cluster states is small

enough for computations to be feasible or till the stop criterion has been reached (see section 7).

This process of combining channel states results in open and closed channel states being combined. However, as we require open states to be distinct from closed states, we divide the aggregate states which contain both open and closed states into two separate states, one containing only the closed states and one containing only the open states. After these aggregate states are split, transition rates to and from the new aggregate states must be found.

5 Subunit Models

We have compared results using three models of IP_3 channel subunits. These include the nine state model of Rüdiger et al. [10] which we have reduced to eight states, the same model but with conditions of sequential binding of Ca^{2+} and IP_3 imposed, and the model of Sneyd and Dufour [14] modified and reduced to six states as given by Ullah and Jung [19].

5.1 Model 1

Model 1 is based on the DeYoung-Keizer model [3], whereby the subunit has an IP₃ binding site, an activating Ca^{2+} binding site and an inhibitory Ca^{2+} binding site. Subunit states are represented by the triplet (ijk), where i, j and k represent these three binding sites respectively. An occupied site is represented with a 1 and an unoccupied site with a 0. In the DeYoung-Keizer model [3], the subunit is considered active when it is in state (110). That is, when the IP₃ and activating Ca^{2+} binding sites are occupied but the inhibitory Ca^{2+} binding site is not occupied. Rüdiger et al. [10] have added an extra state labelled 'Active' with transitions to and from state (110). The subunit is only active when it is in this state. This model is illustrated in Fig. 1. We have combined the 'Active' state and state (110) into one state by assuming the transition rates between the two states are fast, and so reduced the model to eight states. We have used the transition rates given by Shuai et al. [12],

which are listed in the Appendix.

A channel contains four such subunits, and is considered open when any three of the four subunits are in the active state.

5.2 Model 2

It had been suggested by Adkins and Taylor [1] that the binding of Ca^{2+} to the activating and inhibitory binding sites may be affected by whether or not the IP₃ binding site is occupied. Specifically, if IP₃ is bound, then Ca^{2+} cannot bind to the inhibitory Ca^{2+} binding site, and if IP₃ is not bound, then Ca^{2+} cannot bind to the activating Ca^{2+} binding site. Thus, model 2 uses the same scheme as model 1, but the activating Ca^{2+} binding rate when IP₃ is not bound, and the inhibitory Ca^{2+} binding rate when IP₃ is not bound, and the inhibitory Ca^{2+} binding rate when IP₃ is not bound, and the inhibitory Ca^{2+} binding rate when IP₃ is bound, are made small (they are set to $1.0 \times 10^{-3} \,\mu \mathrm{M}^{-1} \mathrm{s}^{-1}$). In order to maintain the condition of detailed balance, the corresponding reverse transition rates (the b_i values) have also been made small, so that the dissociation constants, $K_i = b_i/a_i$ remain the same. The transitions that are made slow are indicated by dotted lines in Fig. 1.

5.3 Model 3

Sneyd and Dufour [14] describe a 10 state model of the IP₃R subunit. It includes one IP₃ binding site, two binding sites for Ca²⁺ activation and one binding site for Ca²⁺ inactivation. The Sneyd and Dufour [14] model was designed to produce agreement with a number of results from experimental data. Falcke [7] modified the model to overcome a problem involving a lack of Ca²⁺ conservation in triangular motifs employed to achieve saturating binding kinetics. These triangular motifs are replaced by square motifs Fig. 2 B, and the full model contains then thirteen states. The resulting transition diagram is shown in Fig. 2. We have used the parameter values given by Ullah and Jung [19], but changed them slightly in order to obey detailed balance. They can be found in the Appendix.

A channel consists of four of these subunits, and is considered open when all four

are in state O', \tilde{O} , \bar{O} (the open states) or state \tilde{A} , \bar{A} (the activated states) or an intermediate combination, such as one subunit in state O' and three in state \bar{A} .

6 Results

We show results for a cluster consisting of five channels, using various values for the Ca^{2+} concentration when the cluster is closed, c_{close} , the Ca^{2+} concentration when the cluster is open, c_{open} , and the IP₃ concentration, p. When finding the steady channel state probabilities, we use $p = 10^{-7}$ and the Ca^{2+} concentration c_{close} . When the cluster is open, we use the Ca^{2+} concentration c_{open} and it remains fixed regardless of the number of open channels. When c_{close} , c_{open} and p are not being varied, their values are 0.1 μ M, 100 μ M and 0.15 μ M respectively. Figures 3-9 show the effect of changing c_{close} , c_{open} and p when using models 1, 2 and 3. We present the probability distribution for the opening time $F_{open}(A, t)$ and closing time $F_{close}(A, t)$ of a cluster of channels and the expected value of the opening time distribution T_{open} and closing time distribution T_{close} .

In Fig. 3 A,B ,we see that the opening distributions for models 1 and 2 are very similar when $c_{\text{close}} < 0.1 \,\mu\text{M}$. At large Ca²⁺ concentrations, model 2 has a much longer expected opening time compared to model 1 (see Fig. 4). At high Ca²⁺ concentrations, there is a high probability of a subunit being in state (011) at rest. In model 1, the fastest route from (011) to (110) is through the binding of IP₃ followed by the release of inhibitory Ca²⁺. However, in model 2 this route is not fast, resulting in a longer expected opening time. When the Ca²⁺ concentration is low (~ 0.1 μ M) subunits are more likely to initially be in state (000), from which transitions to the active state are very fast in both models. However, if the Ca²⁺ concentration is very low (0.01 μ M for example), the binding of Ca²⁺ to the activating Ca²⁺ site is slow, resulting in the increase in the expected opening times seen in Fig. 4 A,B.

In model 3, the single Ca^{2+} dependent transition rate into an active state depends on the concentration of IP₃ too, which is slow for small IP₃ concentration. Therefore, the effect of increasing the Ca^{2+} concentration is to increase the transition rates into

the inactivated or shut states. That slows down activation (Fig. 3 C, Fig. 4) and the expected opening time does not exhibit a minimum in dependence on the Ca^{2+} concentration.

In Fig. 5 A-C and Fig. 6 A,B we see that increasing p results in an initial rapid decrease in the expected opening time. All three models exhibit high sensitivity to the IP₃ concentration at low concentrations, and low sensitivity at high concentrations. Model 2 again has a long expected opening time compared to model 1 as the fastest routes to the active state in model 1 are made slow in model 2.

Fig. 7 A,B and Fig. 8 show that changing the Ca^{2+} concentration has a strong effect on the closing behavior of model 1, but has very little effect on the closing time distribution of model 2. In model 2, the Ca^{2+} dependent transition rate out of the open state is very low, so the Ca^{2+} concentration has little effect on the expected closing time. The most prominent closing transition is IP_3 dissociation. At high Ca^{2+} concentrations, model 1 closes quickly due to the binding of inhibitory Ca^{2+} . However, in model 2 inhibitory Ca^{2+} cannot bind quickly until IP₃ has first been released, resulting in longer expected closing times. We increased the rate of IP_3 release in model 2 by a factor of 10 (that is, $K_1 = 0.036 \ \mu\text{M}$ and $K_3 = 8.0 \ \mu\text{M}$), and this decreased the expected closing time. Changing the Ca^{2+} concentration still has little effect on the expected closing time when the IP_3 release rate is increased. Model 3 does neither exhibit any noteworthy dependence of the cluster closing rate on the Ca^{2+} concentration. Closing is obviously dominated by the Ca^{2+} -independent transitions with the rate $4(l_{-4} + k_{-2})$ (see Fig. 2). Fig. 9 A-C and Fig. 10 A,B show that increasing the IP_3 concentration increases the expected closing time in all three models.

In Fig. 11 A-C we show results using both a fixed and variable value for the Ca²⁺ concentration, c_{open} . When c_{open} is fixed, it has a value of 100 µM and applies to all channels in the cluster as soon as at least one channel is open. That corresponds to very strong spatial coupling by Ca²⁺ diffusion. When c_{open} is variable it takes on different values depending on whether an individual channel is open or closed corresponding to weaker spatial coupling. For an open channel $c_{\text{open}} = 0.1 + 99.9 =$

100 μ M as before. This corresponds to a base level concentration of 0.1 μ M and an increase of 99.9 μ M resulting from the opening of the cluster. For a closed channel the increase is reduced. We use $c_{\text{open}} = 0.1 + 0.1 \times 99.9 = 10.09 \ \mu$ M and $c_{\text{open}} = 0.1 + 0.01 \times 99.9 = 1.099 \ \mu$ M. We see that making c_{open} variable in this manner has very little effect.

Variable $c_{\rm open}$ has of course no affect on the expected opening time, but could change the time course of a puff and therefore the expected closing time. Model 1 shows significant dependence of the closing time distribution on the Ca²⁺ concentration and so we may expect that making the Ca²⁺ concentration variable would have a more significant effect than seen here. A possible explanation is that channels still open fast with the smaller concentrations and channel reopening is very slow compared to channel closing. Thus if a channel closes, it is very likely that the rest of the channels will close before it has reopened, whether it sees a Ca²⁺ concentration of 1.099 μ M, 10.09 μ M or 100 μ M.

In Fig. 12 A-C and 13 A-C we compare the analytic results with the simulation results, for $F_{\text{open}}(A, t)dt$ and $F_{\text{close}}(A, t)dt$ resp., using the fixed values of c_{close} , c_{open} and p. Because our method for finding $F_{\text{close}}(A, t)dt$ relies on an approximation of the channel state transition matrix, there is potential for there to be significant errors in the analytic results. For models 1 and 2 the number of channel states is reduced from 330 to 12 and for model 3 from 1820 to 11. Comparison of the expected values found using the analytic results and the simulation results (see Fig 8 and Fig. 10 A,B) show the agreement for model 1 and model 3 is not good for high values of IP₃. Model 1 has a large expected closing time compared to the other two models which is partly due to a comparatively high open probability at large time values. Fig. 12 and 13 show that the agreement of the analytical and numerical distributions for small times is rather good. However, if the distribution is more spread out, a small error in the tail of the analytic distributions may be causing the bad agreement.

7 Stop criterion for Reduction

Fig. 14 A-C show how the expected closing time for a single channel varies as the number of aggregate states is reduced. When there are 330 aggregate states for models 1 and 2 and 1820 states for model 3, the full model is being used. The calculations are done for a single channel here, as it is too computationally expensive to use more channels with a large number of aggregate states. The vertical lines mark the number of aggregate states used in the results presented in this study. Note, the horizontal axes give the number of states prior to the open states being separated out. We use 12 aggregate states for model 1, 2 and 11 for model 3 here (after separating the open states). This is the maximum number for which results could be computed due to limitations set by computational efficiency.¹ We see also from Fig. 14 that this is close to the minimum value for which we can expect reasonably accurate results, when using a single channel.

It can be seen in Fig. 14 A-C that the expected closing time changes abruptly as the number of aggregate states changes. The abrupt changes in the expected closing time correspond exactly to changes in the number of open aggregate states. There is a dramatic change for all three models and all values of c_{open} when the number of open aggregates gets below a critical value. That critical value provides the stop criterion for cluster state reduction.

Fig. 15 compares exact analytic results with simulations. The error does not vary greatly as the number of channels increases from 1 to 5, suggesting that if the results are reasonably accurate when using a single channel, they will also be reasonably accurate when using 5 channels. Thus, the method we suggest for predicting the accuracy of results after aggregation, is to compute the expected closing time for a single channel using the full model and to compare this with the expected closing time computed after the number of states has been reduced. The minimum number of aggregate states for which reasonable approximations can be expected is the number just above the dramatic change of results.

¹Several hours using a UniServer 3346 with 4Dual-Core AMD Opteron 800 series CPU's

8 Discussion

Many models describing the kinetics of IP_3R subunits have been proposed. We have compared the probability distributions for the expected opening and closing times of a cluster of five IP₃Rs resulting from three different subunit models. The first model is that of Shuai et al. [12], which is similar to the DeYoung-Keizer model [3]. Here there are binding sites for IP₃, activating Ca^{2+} and inhibitory Ca^{2+} , resulting in eight subunit states. Shuai et al. [12] add an extra 'Active' state reachable only from the state where IP_3 and activating Ca^{2+} are bound, and inhibitory Ca^{2+} is not bound. We have reduced these two states to one state by assuming the transitions between them are fast. The parameter values are such that the binding of IP₃ reduces the affinity of the inhibitory Ca^{2+} site, and Ca^{2+} binding to the inhibitory site reduces the affinity of the IP_3 binding site. The second model used is a modification of the first suggested by Adkins and Taylor [1]. Here the inhibitory Ca^{2+} binding rate when IP_3 is bound is made very small, as is the activating Ca^{2+} binding rate when IP_3 is not bound. The reverse transition rates are also made small to maintain detailed balance. The third model is a thirteen state model given by Sneyd and Dufour [14] and extended by Falcke [7] designed to accommodate various experimental findings.

We have compared the opening and closing time distributions resulting from the three models, at various Ca^{2+} and IP_3 concentrations, as well as the expected opening and closing times. We found the three models gave significantly different results. At low Ca^{2+} concentrations (< 0.1 μ M), models 1 and 2 have similar expected opening times, but at larger Ca^{2+} concentrations model 2 has a much longer expected opening time. When the Ca^{2+} concentration is large, both the activating and inhibiting Ca^{2+} binding sites are likely to be occupied prior to the addition of IP_3 . Model 1 then opens comparatively quickly due to the binding of IP_3 followed by the release of inhibitory Ca^{2+} . This route to activation is slow in model 2 as the rate for the release of inhibitory Ca^{2+} when IP_3 is bound is small.

The models show also differences in closing behavior. Using model 1 there is a decrease in the expected closing time of a cluster, as the Ca^{2+} concentration increases

from 20 μ M to 200 μ M. This is due to an increased rate of Ca²⁺ inhibition. In model 2 the rate for Ca²⁺ inhibition while IP₃ is bound is very slow, resulting in less sensitivity of the expected closing time to the Ca²⁺ concentration. Model 3 exhibits only a very weak dependence of the expected closing time on Ca²⁺. Using all three models, the expected cluster closing time increases monotonically with the IP₃ concentration.

We have used a fixed value for the Ca^{2+} concentration when the cluster is open. However, the concentration around the cluster may vary depending on whether individual channels are open or closed. In Fig. 11 we use a variable Ca^{2+} concentration, so that channels in the cluster which are closed see a lower Ca^{2+} concentration than those that are open. Making the Ca^{2+} concentration variable has had very little impact. Fraiman et al. [8] note that the time taken for a single inactivated IP₃R to become uninhibited is much longer than a typical puff duration. Our results support this statement. Consequently, the Ca^{2+} concentration seen by a closed IP₃R has little effect, as all channels are likely to close before it has time to reopen. These results imply that puffs with these models are insensitive towards spatial arrangement of individual channels within the cluster as long as the distance is not too large. We mimicked a decay of the Ca^{2+} concentration on the channel distance corresponding to 3-4.6 diffusion lengths of free Ca^{2+} .

Our method for calculating the closing time distribution of a cluster requires the number of channels in the cluster to be small (≤ 5) so that the number of possible cluster states is not too large. There have been various estimates of the number of channels in a cluster. Some estimate it to be in the range of 20–30 [2, 5, 16], while Shuai et al. [12] estimate 40–70. However, Fraiman et al. [8] estimate 4 or 5, which agrees with an estimate by Swillens et al. [17] and Demuro et al. of about 10 channels in a cluster [20]. Very recent experiments suggest an average number of 4 [21].

In order to calculate the closing time distribution of a cluster, we reduced the number of channel states so that the number of cluster states was not too large. We tried different methods to reduce the system among them a rapid-equilibrium approximation: We identified the states connected by the fastest transition rates, assumed equilibrium just between these two states, lumped them accordingly and

repeated the procedure with the new set of states. With respect to accuracy, that method was as good as the one we used here. Its only disadvantage was that it did not allow for re-separating closed and open states once the aggregates had been found.

The state aggregation introduces error to the calculation. This is particularly apparent in the poor agreement between the analytic and simulation estimates of the expected cluster closing times for model 2. There is a loose relationship between the expected opening time, and the agreement between simulation and analytic results, such that if the expected opening time is long it is likely the agreement will be worse than if the expected opening time is short. Thus, it seems that the error results from the smallest eigenvalues of the coefficient matrix.

We tried a variety of criteria for the degree of state aggregation still providing good approximations. We required for instance the inverse of the smallest transition rate lumped to be much smaller than moments of the waiting time distributions or residence times of the system in aggregates. These are criteria suggested by the use of time scale separation as the basis for state aggregation. Surprisingly, meeting conditions of that type did not give conclusive predictions on the quality of the approximation. The criterium to stop lumping states into aggregates when analytic single channel results get bad compared to single channel simulations was the most predictive and feasible criterium.

We have not related our results to experimental observations in this study yet. Fitting models to experimental data will be the subject of future research. Puffs can be found in a very small range of IP_3 concentration only below which no release is observed and above which spiking or sustained release is found. Only recently, the lab of I. Parker's developed a method to observe puffs over a larger range of IP_3 concentrations by decoupling puff sites by the buffer EGTA [22]. This method will produce data with which we can meaningfully compare our results.

References

- Adkins, C. E., and C. W. Taylor. 1999. Lateral Inhibition of inositol 1,4,5trisphosphate receptors by cytosolic Ca²⁺. Curr. Biol. 9:1115–1118.
- [2] Callamaras, N., J. S. Marchant, X.-P. Sun, and I. Parker. 1998. Activation and co-ordination of InsP₃-mediated elementary Ca²⁺ events during global Ca²⁺ signals in Xenopus oocytes. J. Physiol. 509:81–91.
- [3] De Young, G. W., and J. Keizer. 1992. A Single-pool inositol 1,4,5-trisphosphatereceptor-based model for agonist-stimulated oscillations in Ca²⁺ concentration. Proc. Natl. Acad. Sci. USA. 89:9895–9899.
- [4] Deuflhard, P., W. Huisinga, A. Fischer, Ch. Schütte. 2000. Identification of almost invariant aggregates in reversible nearly uncoupled Markov chains. Linear Algebra and its Applications. 315:39–59
- [5] Dupont, G., and S. Swillens. 1995. Quantal release, incremental detection, and long-period Ca²⁺ oscillations in a model based on regulatory Ca²⁺-binding sites along the permeation pathway. Biophys. J. 71:1714–1722.
- [6] Falcke, M. 2003. On the role of stochastic channel behavior in intracellular Ca²⁺ dynamics. Biophys. J. 84:42–56
- [7] Falcke, M. 2004. Reading the patterns in living cells the physics of Ca²⁺ signaling. Adv. Phys. 53:255–440.
- [8] Fraiman, D., B. Pando, S. Dargan, I. Parker, and S. P. Dawson. 2006. Analysis of Puff Dynamics in Oocytes: Interdependence of Puff Amplitude and Interpuff Interval. Biophys. J. 90:3897–3907.
- [9] Nguyen V., R. Mathias, G. Smith. 2005. A stochastic automata network descriptor for markov chain models of instantaneously-coupled intracellular Ca²⁺ channels. Bull. Math. Biol. 67:393–432

- [10] Rüdiger, S., J. W. Shuai, W. Huisinga, C. Nagaiah, G. Warnecke, I. Parker, M. Falcke. 2007. Hybrid Stochastic and Deterministic Simulations of Calcium Blips. Biophys. J. 93:1847–1857.
- [11] Shuai, J.W., P. Jung. 2002. Stochastic properties of Ca²⁺ release of inositol 1,4,5trisphosphate receptor clusters. Biophys.J. 83:87–97
- [12] Shuai, J., J. E. Pearson, J. K. Foskett, D. D. Mak and, I. Parker. 2007. A Kinetic Model of Single Clustered IP₃ Receptors in the Absence of Ca²⁺ Feedback. Biophys. J. 93:1151–1162.
- [13] Skupin, A., H. Kettenmann, U. Winkler, M. Wartenberg, H. Sauer, St. C. Tovey, C. W. Taylor, M. Falcke. 2008. How does intracellular Ca²⁺ oscillate: by chance or by the clock?. Biophys. J. 94:
- [14] Sneyd, J., and J. F. Dufour. 2002. A dynamic model of the type-2 inositol trisphosphate receptor. Proc. Natl. Acad. Sci. USA, 99:2398–2403.
- [15] Sneyd, J., and M. Falcke. 2005. Models of the inositol trisphosphate receptor. Prog. Biophys. Mol. Biol. 89:207–245.
- [16] Sun, X.-P., N. Callamaras, J. S. Marchant, and I. Parker. 1998. A continuum of InsP₃-mediated elementary Ca²⁺ signalling events in Xenopus oocytes. J. Physiol. 509:67–80.
- [17] Swillens, S., G. Dupont, L. Combettes, and P. Champell. 1999. From calcium blips to calcium puffs: theoretical analysis of the requirements for interchannel communication. Proc. Natl. Acad. Sci. USA. 96:13750–13755.
- [18] Thul, R., and M. Falcke. 2007. Waiting time distributions for clusters of complex molecules. Europhysics Letters 79:38003.
- [19] Ullah, G., and P. Jung. 2006. Modeling the Statistics of Elementary Calcium Release Events. Biophys. J. 90:3485–3495.

- [20] Demuro, A. and I. Parker. 2008. Multi-dimensional resolution of elementary Ca²⁺ signals by simultaneous multi-focal imaging, Cell Calcium 43(4):367–374
- [21] Taufiq-Ur-Rahman, A. Skupin, M. Falcke, C.W. Taylor. 2009. Clustering of IP₃ receptors by IP₃ returnes their regulation by IP₃ and Ca²⁺, Nature, in press
- [22] Smith, F., S.M. Wiltgen, and I. Parker. 2009. Localization of puff sites adjacent to the plasma membrane: Functional and spatial characterization of Ca^{2+} signaling a Jalein Accepteonnanus Accepteon in SH-SY5Y cells utilizing membrane-permeant caged IP₃, Cell Calcium 45:65–76

Parameter	Value	Parameter	Value
a_0	$550.0 \ s^{-1}$	b_0	$80.0 \ s^{-1}$
K_1	$0.0036 \ \mu M$	a_1	$60.0~\mu M^{-1} s^{-1}$
K_2	$16.0~\mu\mathrm{M}$	a_2	$0.2 \ \mu M^{-1} s^{-1}$
K_3	$0.8~\mu\mathrm{M}$	a_3	$5.0 \ \mu M^{-1} s^{-1}$
K_4	$0.072~\mu\mathrm{M}$	a_4	$0.5 \ \mu M^{-1} s^{-1}$
K_5	0.8 µM	a_5	150.0 $\mu M^{-1} s^{-1}$
	ed		

Table 1: Parameter values for Model 1

Parameter	Value	Parameter	Value
k_1	$2.0 \ \mu M^{-1} s^{-1}$	k_{-1}	$0.04 \ s^{-1}$
k_2	$37.4 \ \mu M^{-1} s^{-1}$	k_{-2}	$1.4 \ s^{-1}$
k_3	$0.11 \ s^{-1}$	k_{-3}	$29.8 \ s^{-1}$
k_4	$4.0 \ \mu {\rm M}^{-1} s^{-1}$	k_{-4}	$0.37 \ s^{-1}$
k_5	$2.0 \ \mu M^{-1} s^{-1}$	l_1	$10.0~\mu{\rm M}^{-1}s^{-1}$
l_3	100.0 $\mu M^{-1} s^{-1}$	l_5	$0.1 \ \mu M^{-1} s^{-1}$
L_1	0.12 μΜ	L_3	$0.025~\mu\mathrm{M}$
L_5	38.2 µM	l_2	$1.7 \ s^{-1}$
l_{-2}	$0.8s^{-1}$	l_4	$37.4 \ \mu M^{-1} s^{-1}$
l_{-4}	$2.5 \ s^{-1}$	l_6	$4707.0 \ s^{-1}$
l_{-6}	$11.4 \ s^{-1}$		

Table 2: Parameter values for Model 3

Figure 1: The nine state model of an IP₃ receptor channel subunit proposed by Rüdiger et al. [10]. The Ca²⁺ concentration is denoted by c and the IP₃ concentration is denoted by p. The dotted lines indicate the transitions that are made small in model 2.

Figure 2: The model of an IP₃ receptor channel subunit given by Sneyed [14]. The channel states are R: unbound receptor, I_1 and I_2 : inactivated, O: open, A: activated, and S: shut. The Ca²⁺ concentration is denoted by c and the IP₃ concentration is denoted by p.

Figure 3: Open probability distribution $F_{\text{open}}(A, t)$ for various c_{close} values. A: $F_{\text{open}}(A, t)$ for model 1. B: $F_{\text{open}}(A, t)$ for model 2. C: $F_{\text{open}}(A, t)$ for model 3.

Figure 4: A: The expected opening times T_{open} for the three models. B: Enlarged plot of the expected opening times for the three models to show clearly the minimum of models 1 and 2 for small values of c_{open} .

Figure 5: Opening time distribution $F_{\text{open}}(A, t)$ when changing p for some representative IP₃ concentrations. A: $F_{\text{open}}(A, t)$ for model 1. B: $F_{\text{open}}(A, t)$ for model 2. C: $F_{\text{open}}(A, t)$ for model 3.

Figure 6: A: The expected opening times T_{open} for the three models as function of the IP₃ concentration. B: Enlarged plot of T_{open} for small concentrations.

Figure 7: Cluster closing time distribution $F_{\text{close}}(A, t)$ when changing c_{open} . A: $F_{\text{close}}(A, t)$ for model 1. B: $F_{\text{close}}(A, t)$ for model 2. C: $F_{\text{close}}(A, t)$ for model 3.

Figure 8: The expected closing times T_{close} for the three models. Model 2 is represented by the upper lines, where the dashed line with ×-symbols show the simulation results and dashed line without shows analytic results. Model 1 is shown by two

full lines, with +-symbols for simulation and without for analytic results. Fast IP₃ release in model 2 corresponds to $K_1 = 0.036 \ \mu\text{M}$ and $K_3 = 8.0 \ \mu\text{M}$. The simulation results are represented by the dashed line with \Box -symbols and the analytic results by the dashed line. Results for model 3, analytic and simulation, is shown by the dashed line with the *-symbols below. The analytic and simulation results can not be distinguished.

Figure 9: Cluster closing time distribution $F_{\text{close}}(A, t)$ when changing p. A: $F_{\text{close}}(A, t)$ for model 1. B: $F_{\text{close}}(A, t)$ for model 2. C: $F_{\text{close}}(A, t)$ for model 3.

Figure 10: A: The expected closing times T_{close} for models 1 and 2, from both the analytic and simulation results. Model 1 is represented by the upper dashed line (analytic results) and dashed line with ×-symbols (simulation results). Model 2 is represented by the lower full lines. There are few differences between analytic (full line) and simulation results (full line with +-symbols). B: The expected closing times T_{close} for model 3 only with analytic (full line) and simulation results (full line with +-symbols).

Figure 11: $F_{\text{close}}(A, t)$ using both a fixed and variable Ca²⁺ concentration. Here $c_{\text{close}} = 0.1 \,\mu\text{M}$ and $p = 0.15 \,\mu\text{M}$. $c_{\text{open}} = 100 \,\mu\text{M}$ when fixed. When c_{open} is variable, it has a value of 100 μM for a channel which is open and a value of 10.09 μM (labeled 'Variable c_{open} 1') or 1.099 μM (labeled 'Variable c_{open} 2') for a channel which is closed. A: Results for model 1. B: Results for model 2. C: Results for model 3.

Figure 12: Analytic and simulation results showing $F_{\text{open}}(A, t)$ for all three models. Here $c_{\text{close}} = 0.1 \ \mu\text{M}$, and $p = 0.15 \ \mu\text{M}$. The full lines represent the analytic results and the \Box -symbols the simulation results. A: Results for model 1. B: Results for model 2. C: Results for model 3.

Figure 13: Analytic and simulation results showing $F_{\text{close}}(A, t)$ for the three mod-

els. Here, $c_{\text{open}} = 100 \ \mu\text{M}$ and $p = 0.15 \ \mu\text{M}$. A: $F_{\text{close}}(A, t)$ for model 1. B: $F_{\text{close}}(A, t)$ for model 2. C: $F_{\text{close}}(A, t)$ for model 3.

Figure 14: The effect of the number of aggregate states on the expected closing time. These results are for a single channel. A-C show the expected closing time T_{close} as the number of aggregate states is reduced. This is shown for the three models at various parameter values. The vertical lines represent the number of aggregate states which are used in the results presented here. D-F show the number of open aggregates N_{open} as function of the aggregate states for all three models and various parameters.

Figure 15: Varying the cluster size. Graphs A, B and C show the expected closing time of the cluster T_{close} , using models 1 (A), 2 (B) and 3 (C) respectively, as the number of channels in the cluster is varied from 1 to 5. We use $p = 0.15 \,\mu\text{M}$, $c_{\text{open}} = 100 \,\mu\text{M}$ and $c_{\text{close}} = 0.1 \,\mu\text{M}$ (to compute the initial condition). Both analytic (full lines) and simulation results (dashed lines with +-symbols) are shown.







Figure 3:





Figure 5:





Figure 7:





Figure 9:





Figure 11:



Figure 12:



Figure 13:



Figure 14:



Figure 15: