

# 17q21.31 microduplication patients are characterized by behavioural problems and poor social interaction.

Bernard Grisart, Lionel Willatt, Anne Destrée, Jean-Pierre Fryns, Katrina Rack, Thomy de Ravel, Jill Rosenfeld, Joris Robert Vermeesch, Christine Verellen-Dumoulin, Richard Sandford

# ▶ To cite this version:

Bernard Grisart, Lionel Willatt, Anne Destrée, Jean-Pierre Fryns, Katrina Rack, et al.: 17q21.31 microduplication patients are characterized by behavioural problems and poor social interaction.. Journal of Medical Genetics, 2009, 46 (8), pp.524. 10.1136/jmg.2008.065367 . hal-00552684

# HAL Id: hal-00552684 https://hal.science/hal-00552684

Submitted on 6 Jan 2011

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

# 1 **17q21.31** microduplication patients are characterized by behavioural 2 problems and poor social interaction.

3

Bernard Grisart<sup>1,\*,§</sup>, Lionel Willatt<sup>2,\*</sup>, Anne Destrée<sup>1</sup>, Jean-Pierre Fryns<sup>3</sup>, Katrina Rack<sup>1</sup>,
Thomy de Ravel<sup>3</sup>, Jill Rosenfeld<sup>4</sup>, Joris Robert Vermeesch<sup>3</sup>, Christine Verellen-Dumoulin<sup>1</sup>,
Richard Sandford<sup>2</sup>.

7

<sup>1</sup> Centre de Génétique Humaine, Institut de Pathologie et de Génétique, 25 Avenue Georges
<sup>9</sup> Lemaître, B-6041 Charleroi, Belgium.

10<sup>2</sup> East Anglian Medical Genetics Service, Cytogenetics Laboratory, Box 108, Level 6,

11 Addenbrooke's Treatment Centre, Addenbrooke's Hospital, Cambridge CB2 0QQ, United 12 Kingdom.

<sup>3</sup> Center for Human Genetics, University Hospitals Leuven, Herestraat 49, 3000 Leuven,
 Belgium.

<sup>4</sup> Signature Genomic Laboratories, LLC, 2820 North Astor Street, Spokane, WA 99207 USA.

- 16 \* Both authors equally contributed to this work.
- 17 § Corresponding author : Phone number ##/32/71/447185, Fax Number : ##/32/71/471520, E-
- 18 mail : bernard.grisart@ipg.be.
- 19

20 **KEYWORDS**: 17q21.31 microduplication, MAPT, CRHR1, autism, behaviour.

- 21
- 22 Word count : 3434.

## 2 ABSTRACT.

Background : Microdeletions at 17q21.31 have recently been shown to cause a novel syndrome. Here we identify the reciprocal 17q21.31 duplication syndrome in four patients.
 Method : Patients with the 17q21.31 duplication were indentified by screening a large cohort of patients (n=13070) with mental retardation and congenital malformation by CGH microarray. Parental origin was investigated in three patients by quantitative PCR and microsatellite genotyping.

9 **Results**: In three cases it was possible to show that duplication arose *de novo*. Intellectual 10 skills range from normal to mild mental retardation. Patients are characterized by poor social interaction, with relationship difficulties, reminiscent of autistic spectrum disorders. Other 11 12 features are rather variable with no striking common phenotypic features. Parental origin was 13 investigated for 3 patients. In all cases duplication was of maternal origin either through 14 interchromosomal (2 cases) or interchromatid (1 case) rearrangement. The three mothers are 15 all carriers of the inverted H2 haplotype emphasizing the role of local genomic architecture alteration as a predisposing factor for this duplication. 16

17 **Conclusion :** Autistic features observed in our patients suggest that genes in the duplicated 18 interval should be considered as candidates for disorders in the autistic spectrum. Other 19 phenotypic observations are rather variable or aspecific. This adds 17q21.31 duplications to a 20 growing group of recently identified genomic disorders with variable penetrance and 21 expressivity.

22

## 23 INTRODUCTION.

24 The widespread use of CGH microarray has resulted in the identification of many new 25 genomic disorders in patients with mental retardation with, or without, multiple congenital 26 anomalies (MR/MCA) due to recurrent microdeletions. Many of these new syndromes 27 including the 17q21.31 microdeletion were recently reviewed by Slavotinek et al. [1]. The 28 17q21.31 microdeletion, with a critical region of 424 kb in size, was the first to be identified 29 by screening large cohorts of patients by CGH microarrays even prior to the detailed 30 characterization of the associated clinical features. The clinical picture was recently 31 documented for 22 patients. It included developmental delay, hypotonia, facial dysmorphism 32 and usually a friendly and amiable disposition [2]. The microdeletion encompasses 6 genes : 33 2 ORF and 4 known genes (MAPT, IMP5, CRHR1 and STH). Three of these are good 34 candidates to explain the neurological disturbance observed in these patients : MAPT (OMIM 35 157150), IMP5 (OMIM 608284) and CRHR1 (OMIM 122561). The microdeletion 36 breakpoints were recurrently mapped within a low copy repeat [2-6]. Even if there is no 37 formal proof that the underlying mechanism is non allelic homologous recombination 38 (NAHR), it seems reasonable to postulate that NAHR between two directly oriented repeats is 39 actually involved. Morever, an inversion within 17q21.31, present in the Western European population at a frequency of 20% [7], was shown to be closely associated with this 40 41 rearrangement. Inverted haplotypes (H2) differ notably from normal haplotypes (H1) by the 42 orientation of critical repeat sequences. Such an orientation allows NAHR to occur either 43 between H1 and H2 or between two H2 haplotypes.

The frequency of this microdeletion syndrome was estimated to be around 1 in 16000 live births [2]. Given the mechanisms involved, the reciprocal duplication is expected to occur in a ratio of roughly 1 duplication for 2 deletions [8]. However, to our knowledge only one patient with a 17q21.31 microduplication has been reported to date [9].

48 Here we report four additional patients with a 17q21.31 microduplication with extensive 49 molecular and clinical data.

#### 1 MATERIAL AND METHODS.

#### 2 Patient clinical data.

3 Patient 1 (Figure 1 and Table 1).

4 This girl is the first child of unrelated parents. She was born at term after a pregnancy 5 complicated by placental detachment in the first month of gestation. The birth weight was 6 3.570 kg (P50). She started to walk at 12 months but with a tendency to tiptoe-walking 7 resulting in tendinous retraction. Language onset was normal but she developed difficulties in 8 sentence formation and pronunciation which persist at 6 years of age. She was referred for 9 genetic consultation for behavioural problems, hyperactivity and difficulties with social 10 interaction. The intellectual quotient (IQ) was evaluated to be 105 (WPPSI-R : Wechsler 11 Preschool and Primary Scale of Intelligence-Revised) with a verbal IQ of 108 and performance IQ of 100. The developmental quotient was 91 for fine mobility and 73 for 12 global mobility. Brain magnetic resonance imaging was performed with normal results. 13

At six years, growth parameters were in the normal range : height was at the 90<sup>th</sup> centile (126 14 cm) and weight at the 50<sup>th</sup> centile (23.3 kg). Microcephaly was observed with head 15 circumference at -2SD (49 cm). Some facial dysmorphic features were noticed with 16 synophrys, puffy eyelids, short philtrum, thin upper lip, retrognathia and dysplastic ears. 17 18 Slight hirsutism was observed on the back. Clinodactyly of the fifth finger, single palmar crease on right hand and partial bilateral syndactyly between the  $2^d$  and  $3^d$  and  $3^d$  and  $4^{th}$  toes 19 20 was also noted. Hormonal blood measurements (FSH, LH, DHEA sulphate, 17-21 hydroxyprogesterone) were in the normal range. These patient data were submitted in 22 DECIPHER database (https://decipher.sanger.ac.uk/application/) with identification number 23 LEM249002.

24

25 Patient 2 (Table 1).

26 This girl is the only child of unrelated parents. The pregnancy was uncomplicated. Birth 27 weight was 3.5 kg (P50), length 53 cm (P90) and head circumference 34.2 cm (P25-50). 28 Meconium aspiration occurred during delivery which resulted in no apparent sequelae. 29 Feeding and early milestones were normal. She started to walk at the age of 2 when 30 hypotonia was noticed. At age 3 she demonstrated obsessive behaviour characterized by an 31 adherence to ritual in performing various common tasks. She had difficulty with toilet 32 training. She was very passive, often played by herself and did not demand affection. First 33 clinical evaluation was carried out at age of 6. She had significant behavioural problems at 34 school and home, poor social interaction with peers, disruptive behaviour and occasional 35 aggressive outbursts. Her IQ was evaluated at 78 (WPPSI-R). Verbal skills and vocabulary 36 were normal. She has normal reasoning and comprehension but she has some difficulty 37 following instructional language. Fine motor skills, spatial awareness and non-verbal 38 communication were evaluated as poor. She was very rule driven preferring routine tasks. At 39 the age of 11 some dysmorphic features were noted including epicanthal folds, large 40 posteriorly rotated ears, short upturned nose, short philtrum, flat midface, high arched palate, 41 prominent upper incisors, low posterior hairline, thick hair, mild truncal obesity and tapering 42 fingers.

The behavioural problems persisted with difficulties maintaining friendships and occasional inappropriate responses to other people. Fine motor skills remain poor and her obsessional behaviour is still problematic. Growth parameters are in the normal range with height and weight at the 91<sup>th</sup> centile and head circumference between the 50<sup>th</sup> and the 75<sup>th</sup> centile.

The common 17p11.2 deletion responsible for hereditary neuropathy with liability to pressure palsies (HNPP, OMIM 162500) was also identified but she is asymptomatic for this syndrome. This rearrangement was inherited from her mother. Therefore patient 2 has both an HNPP deletion and a 17q21.31 duplication with a maternal origin for both (see FISH 1 analysis in results section). DECIPHER identification number for this patient is 2 CAM249012.

- 3
- 4 Patient 3 (Figure 1 and Table 1).

5 This boy is the second child of non-consanguineous healthy parents. His sister is healthy. The 6 pregnancy and delivery occurred without complications and he was born at term with a birth 7 weight of 2.890 kg (P3), length of 50 cm (P50) and head circumference of 34 cm (P10). He 8 underwent bilateral inguinal herniorraphy between the ages of 10 and 12 months. He also had 9 an orchidectomy due to torsion. His early milestones were considered normal : he could walk 10 independently at the age of 14 to 16 months and said his first words at 12 to 14 months. He 11 received speech therapy at the age of 3.5 years as he was socially anxious and his verbal 12 communication was poor at nursery school. He also began tiptoe-walking and physiotherapy 13 was initiated. Assessment with the WPPSI-R at the age of 4 years and 9 months indicated a 14 total IQ of 81 (verbal IQ = 80, performance IQ = 87). His auditory memory, visuomotor 15 interaction and word finding capacities were poor. Due to his socio-communicative difficulties, certain elements of the autism spectrum disorder (ASD) were recognised in him. 16 17 Investigations showed a normal hearing test, normal magnetic resonance imaging of the brain 18 and normal electromyogram. Clinical examination at the age of 7 years and 4 months revealed height of 127 cm (50<sup>th</sup> centile), weight of 25.3kg (50<sup>th</sup> centile) and head 19 20 circumference of 50.7cm (10<sup>th</sup> centile). He has brachycephaly, an open square facies with a 21 pointed chin. A congenital hypopigmented 3x3 cm mark was found on his left flank and there 22 was no hirsutism. He has normal symmetric reflexes and shortened gastrocnemius muscles 23 (right more than left). DECIPHER identification number for this patient is CHG249083.

24 25

#### 26 Patient 4 (Table 1).

27 This man was born after an uncomplicated pregnancy by caesarean section. He is the second child of unrelated healthy parents. Birth weight was 5.1 kg (>97<sup>th</sup> centile) and length 58 cm 28 29 (>97<sup>th</sup> centile). Mild hypotonia and nasal congestion was noted but feeding was normal. An 30 inguinal hernia was repaired at age 6 months. All major developmental milestones were 31 delayed. He walked at 27 months and speech was significantly delayed. A muscle biopsy 32 performed for hypotonia was reported as normal. A diagnosis of Asperger's Syndrome was 33 made by a clinical psychologist at age 11 years. The history also included a diagnosis of 34 depression and attention deficit hyperactivity disorder for which he receives long-term 35 medication. He required educational support in all subjects and a formal IQ was in the low-36 normal range. He was also described as a loner and established no close friendships. Eye 37 contact and social interaction were poor. His behaviour was occasionally complicated by 38 rages but these were not associated with self-harm. His height and weight were noted to be 39 <3<sup>rd</sup> centile but no cause was identified. Puberty was delayed. LH/FSH and testosterone levels were all reported as low. A brain MRI was also reported as normal. At age 18, height 40 and head circumference were between  $10^{\text{th}} - 25^{\text{th}}$  centiles and weight remained  $<3^{\text{rd}}$  centile. 41 He was dysmorphic with thin arched eyebrows, deep-set eyes and down slanting palpebral 42 fissures. Mild malar hypoplasia was also noted. The ears were simple and elongated 43 (>+2SD). Mild pectus excavatum, 5<sup>th</sup> finger clinodactyly, 2-3 toe syndactyly and a sandal gap 44 45 were also present. Secondary sexual characteristics were absent and he had a slender body 46 habitus with a generalised lack of body fat. The remainder of the physical examination was 47 normal. DECIPHER identification number for this patient is LEM249014.

- 48
- 49

## 1 Microarray analysis

Patients' DNA were analysed with 4 different CGH technologies. For patient 1, CGH
microarray analysis was performed using a 44K oligonucleotide microarray (Agilent, Santa
Clara, CA, USA). Patient 2 was analysed using a 250K Affymetrix Nsp SNP array and
Nexus software. Patient 3 was investigated using a 1 Mb BAC array as previously reported
[10] and confirmed using a whole genome tiling-path array. Patient 4 was studied using the
Signature Genomics BAC/PAC array.

8

# 9 Parental origin of the duplication

10 Quantitative PCR analysis.

Quantitative PCR was used to confirm the rearrangement. Primer design and real-time PCR 11 12 amplification were performed as described by Hoebeeck et al. [11]. ABgene SYBRgreen 13 mastermix was used according to the manufacturers (ABgene, Epsom, UK) recommendations 14 with 250 nM final primers concentration and 10 ng of DNA in a volume of 25 µl. Ct values 15 obtained from PCR amplification of exon 1 of MAPT (GCCGTCTTCCGCCAAGAG, GGAGCCGATCTTGGACTTGACATT) were normalized by two reference genes (primers 16 for GPR15 : GGTCCCTGGTGGCCTTAATT and TTGCTGGTAATGGGCACACA and 17 18 primers for P53 CCCAAGCAATGGATGATTTGA and • 19 GAGCTTCATCTGGACCTGGGT).

- 20
- 21 <u>Microsatellite and haplotype analysis.</u>

Microsatellites were used to determine the parental origin of the duplication. Primers are reported in supplementary Table 1. Two microsatellites (D17S810 and DG17S920) located outside the duplication were genotyped as well as 3 microsatellites (VNTR17\_1, VNTR17\_4 and D17GS142) inside the duplication. PCR were performed using AmpliTaq Gold (Applied Biosystems, Foster City, CA, USA) in semi-quantitative conditions by running only 25 cycles of amplification to allow semi-quantitative analysis of number of inherited alleles based on the analysis of relative height of alleles observed on the electrophoretic data.

29 specific 238 deletion [12] The H2 bp was genotyped using primers 30 GGAAGACGTTCTCACTGATCTG and AGGAGTCTGGCTTCAGTCTCTC, followed by 31 electrophoresis on a 1% agarose gel.

32

33 <u>FISH analysis.</u>

34 BAC probes were used to confirm the rearrangement observed in patient 2. BAC RP11-

- 35 64B12 at PMP22 locus (17p12) was used in conjunction with RP11-100C5 and RP11-413P22
- 36 at the 17q21.31 microduplication locus.

2	Table 1 : List of clinical observations reported for 5 patients with 17q21.31 duplic	ation
4	Table 1. List of childen observations reported for 5 patients with 17921.51 dupite	auon.

	Kirchhoff et	Patient 1	Patient 2	Patient 3	Patient 4	
Clinical observation	al., 2007	(female)	(female)	(male)	(male)	Frequency
Pregnancy	Uneventful	Placental detachment	Normal	Normal	Normal	
	42 weeks	At term	At term	At term	At term	
Birth						
Weight (kg)	3.070	3.570 (P50)	3.500 (P50)	2.890 (-2SD)	5.100 (+3SD)	
Length (cm)	50	ND	53 (P90)	50 (P50)	58 (+3SD)	
OFC (cm)	34	ND	34.2 (P25-50)	34 (P10)	ND	
Walk (month)	60	12	24	14-16	27	
Psychomotor development						
Hypotonia	+	-	+	-	+	3/5
Hyperactivity	+	+	-	ND	+	3/4
Passivity	ND	ND	+	ND	ND	
Obsessive behaviour	ND	ND	+	ND	-	1/2
Social interaction	ND	Poor	Poor	Poor, anxious	Poor	4/4
Aggressiveness	ND	ND	+	ND	+	2/2
(without self injury)						
Intelligence						
Psychomotor retardation	Severe	Mild	Mild	Mild	Mild	5/5
IO	ND	105	78	81	ND	5/5
Verbal skills	Very Poor	Limited	Normal	Poor	Normal	3/5
Motor skills	ND	Poor	Poor	ND	Normal	2/3
Tiptoe-walking	ND	+	-	+	-	2/4
Dysmorphism	THE .					2/1
Microcephaly	+	Mild	_	_	_	1-2/5
Synophrys	ND	+	Bushy eyebrows			2/4
Epicanthal folds	ND	_	+	_	_	1/4
Dysplastic ears	+	+	+	_	+	4/5
Nose	Short	Upturned	Short, Upturned	_	'	2-3/5
Philtrum	Smooth	Short	Short			2-3/5
Midface	ND	Short	Flat		Flat	2/4
Palate	High arched		High arched		1 Iat	2/4
Prominent incisor	+	+	+	_		3/5
Micrognathia	+	ND	-	ND	_	1/3
Retrognathia	ND	+	ND	ND		1/3
Finger	Broad	Clinodactyly of 5 <sup>th</sup>	Tapering	ND	Clinodactyly 5th	2/4
Palmar crease	ND	Single on right hand	r apering	ND	ND	1/2
Feet	Broad	Partial syndactyly	-	ND	2-3 syndactyly	2/3
Others	Dioau	i ai tiai syndaetyly	-	nD	2-5 Syndactyly	212
Hirsutism	On the back	On the back	low post-hairline	_		2-3/5
Failure to thrive	At 7 months	On the back	low post-main the	-	ND	2-3/3
		ND	ND	ND		
Cysts on liver & spleen	+ ND		ND ND	ND ND	ND	
Hypogonadism	ND	ND	ND	ND	+	

3 ND : not determined : no data reported in the clinical file.

4 "+" : Trait present in the patient.

5 "-" : Trait not observed in the patient.

6

#### 7 **RESULTS.**

#### 8 Microarray results.

9 The size of the duplication in the three patients investigated with non-targeted array (patient 10 1-3) ranged from 585 kb to 763 kb. An overview of the breakpoints is given in Figure 2 11 where the duplications are aligned with the recurrent deletion [2] and CNV of region 12 17q21.31. Exact breakpoint positions are listed as a supplementary Table 2. The difference 13 in the breakpoints likely reflects the different resolution of the four array types used.

14

## 15 **Parental origin of the duplication.**

## 16 Quantitative PCR analysis.

17 Quantitative PCR was performed on patient and parental DNA to determine the origin of the 18 duplication using a PCR system developed in exon 11 of MAPT. In patients 1 to 3 the

19 duplication appeared de novo (data not shown). De novo occurrence in patient 4 could not be

- 20 determined due to lack of parental DNA.
- 21

1 <u>Microsatellite and haplotype analysis</u>

2 To investigate the origin of the duplication, the probands and their parents were genotyped for 3 5 microsatellites which were phased in order to identify transmitted haplotypes (see 4 supplementary Table 1). In all cases the duplication occurred on the maternal chromosome. 5 Detailed analysis of genotypic data shows that the duplication occurred by interchromosomal rearrangement (probably NAHR) between H1 and H2 haplotypes in patient 1 and 2 whereas it 6 7 occurs by interchromatid rearrangement (probably NAHR) between two H2 haplotypes in 8 patient 3. An illustration of electrophoretic data obtained from a capillary sequencer, as well 9 as detailed interpretation of these microsatellite data, is given in supplementary Figure 1. An H2 specific deletion of 238 bp was also genotyped by PCR (supplementary Table 2 and 10 supplementary Figure 2). Patient 4 could not be tested for parental origin because parental 11 12 DNAs were unavailable.

13

14 FISH analysis

Patient 2 was investigated by FISH given the presence of two rearrangements on chromosome
17. Co-hybridization of HNPP locus specific probes (RP11-64B12) and 17q21.31 specific
probes (RP11-100C5 and RP11-413P22) demonstrated that the 17q21.31 duplication occurs
on the same chromosome as the HNPP microdeletion (data not shown).

19

## 20 **DISCUSSION.**

21 The 17q21.31 microduplication described here is a novel syndrome characterized by features 22 of the autism spectrum disorder. All patients have behavioural problems with social 23 interaction and communication difficulties and mild psychomotor retardation suggesting that 24 some of the genes within the duplication interval may be candidates for the autistic spectrum, 25 even if at this stage this hypothesis requires formal testing. A linkage study of autistic 26 families [13] has identified strong linkage within 17q21. However this association was not 27 confirmed by a larger linkage study reported by the Autism Consortium [14] nor in the CNV 28 (copy number variant) analysis of autistic families reported by Sebat *et al.* [15]. The 29 duplicated interval contains 6 genes (2 ORFs and 4 known genes). Some of these are good 30 candidates for the impaired cognitive function or motor skills observed in our patients. Some 31 mutations of MAPT (microtubule associated protein TAU) altering the biochemistry of tau 32 protein have been reported to cause a greater abundance of unbound tau species and formation 33 of insoluble tau inclusions in frontotemporal dementia and parkinsonism (reviewed in Pittman 34 et al. [16]) which could therefore be considered as a "toxic gain of function mutation". 35 Increased accumulation could be expected from dosage sensitive overexpression of the 36 duplicated MAPT gene observed in our patients. However MAPT gene duplications have not 37 been identified in cases of frontotemporal lobar degeneration [17]. Patients 1, 2 and 4 have 38 had a brain scan with no sign of neurodegeneration. However it should be mentioned that our 39 patients are between 6 and 20 whereas the first signs of degeneration in frontotemporal 40 dementia usually becomes apparent around 45-55 years of age [16]. Then a follow up of 41 these patients for this particular aspect is certainly advisable to determine their evolution with 42 age. CRHR1 (Corticotropin Releasing Hormone Receptor 1) is expressed in the embryonic 43 and postnatal cerebellum and could play a role in central nervous system development by 44 affecting neurone survival through a depolarization mechanism [18]. Duplication of the gene 45 coding for CRHR1 could explain the impaired motor skills reported for at least two patients (patient 1 and 2). Moreover experimental evidence suggests that CRHR1 is involved in stress 46 response and anxiety related behaviour [19]. CRHR1 is one of the preferred targets for 47 48 depression, anxiety and stress related therapy [20]. Again duplication of this gene could 49 contribute to the behavioural troubles and poor social interaction observed in 4 of the 5 50 patients reported with 17q21.31 microduplication. Finally, IMP5 encodes a gene highly

homologous to the presenilin 1 and 2 genes, which are involved in APP (Amyloid Precursor
 Protein) cleavage. Interaction of IMP5, or other presenilin analogues, with APP has not been

3 formally established but experimental evidence suggests that protease others than presenilin 1

4 and 2 are involved in APP cleavage [21, 22] since APP cleavage still occurs in presenilin 1

5 and 2 double knock-out mouse fibroblasts.

6 The other phenotypic aspects are very variable and much milder. Other common features displayed by these four cases and by the patient reported by Kirchhoff et al. [14] include : 7 8 dysplastic ears (4/5), hypotonia (3/5), hyperactivity (3/4), poor verbal skills (3/5), prominent 9 incisors (3/5), poor motor skills (2/3), tiptoe-walking (2/4) and short nose (2/5). More cases 10 are required to accurately define the clinical spectrum associated with this microduplication as many features described are mild and non-specific. Patient 2 was noted to share a number of 11 dysmorphic features with the case described by Kirchhoff including hypotonia, poor motor 12 13 skills, dysplastic ears, short nose, short philtrum, high-arched palate, prominent incisors and 14 hirsutism. The role of the duplication in the aetiology of the hypogonadism in case 4 is 15 unclear and will await identification of further cases. However it suggests that all cases of 17q21.31 microduplication should be assessed for this potential complication. A patient 16 17 identified by Hunter Genetics in Australia has been recently reported in the DECIPHER 18 database (https://decipher.sanger.ac.uk/application/). This patient was reported to have 19 mental retardation, abnormal shaped teeth and tapering fingers.

No phenotypic similarities or differences between deletion and duplication carriers are obvious. Deletion carrier seems to be more affected (moderate to severe mental retardation) than duplication carriers (mild mental retardation in our 4 patients). It's also interesting to note that, whereas deletion carriers have a friendly and amiable behaviour, duplication carriers have on contrary some traits reminiscent of the autistic spectrum. This opposite phenotypic situation should certainly be kept in mind in the future when additional patients are clinically assessed.

27 The phenotypic variability is striking and adds this syndrome to a growing group of 28 chromosomal imbalances with variable expressivity. Such variability is observed especially 29 in association with microduplication genomic disorders : duplications of the SMS [23], 30 Williams-Beuren [24], 22q11 [25] and NF1 [26] loci result in a heterogeneous phenotype 31 ranging from normal to severe. Distal deletion and duplication of the 22q11 locus has been 32 reported in patients with highly variable symptoms especially for the duplication carriers [25, 33 27]. Another recently identified microdeletion/duplication genomic disorder of 1.35 Mb at 34 1q21.1 also escapes syndromic classification due to large phenotypic variability such that 35 identification of carriers probably precludes a phenotypic-based evaluation [28].

36 The broad variability of phenotypes associated with specific chromosomal rearrangements 37 suggests that many mildly affected individuals remain undiagnosed. This generates an 38 ascertainment bias in the evaluation of the frequency of such rearrangements. The cohort 39 investigated here was made of 13070 patients. A total of 24 deletion carriers were observed. 40 Based on a frequency 2% for individuals presenting mental retardation in the human 41 population, the prevalence of 17q21.31 deletion could be evaluated to 1 in 55000 in our 42 cohort, as explained in Koolen et al. [2], which is lower that the value of 1 in 16000 reported 43 by Koolen *et al.* [2]. This certainly reflects a bias in the selection of patients. Similarly the 44 prevalence of duplication can be evaluated to 1 in 327000 which is less than half the 45 prevalence of deletion which would be mechanistically expected from the model proposed by 46 Turner et al. [8] for deletion and duplication resulting from NAHR.

47 Koolen et al. [2] were able to map the proximal breakpoint to a 500 bp interval (position

48 41046729-41047168 bp on chromosome 17) corresponding to a L2 LINE element. This is

49 comparable to the proximal breakpoints in the four patients with the microduplication.

50 However, as mentioned by Koolen et al. [2], the distal breakpoint falls within a CNV region,

1 known to be polymorphic in normal individuals. Interestingly, each patient with the 2 duplication investigated with non-targeted arrays (Patients 1-3 and Kirchhoff's patient) show 3 an amplified copy number for this CNV indicating either that one of the parents is a carrier of 4 a multi-copy allele for this CNV or that the duplication involves a more distal breakpoint.

5 In three of four cases reported here, the microduplication could be demonstrated to be of 6 maternal origin whereas a paternal origin was established for the first patient described by 7 Kirchhoff *et al.* [9]. In the three cases tested, the parent of origin was either heterozygote or 8 homozygote for the H2 haplotype. In patient 3 the duplication arose through interchromatid 9 rearrangement whereas, for the 2 others, an interchromosomal mechanism was discovered. 10 This again emphasizes the role of the inverted H2 haplotype in this microdeletion /

11 microduplication syndrome as recently investigated in detail in the human and great apes 12 lineage [29].

In conclusion, we report four additional cases with 17q21.31 microduplication (for 3 of which *de novo* occurrence could be assessed) which contribute to documenting the molecular and clinical features associated with this recently discovered syndrome. Whilst the microdeletion associated phenotype is now clearly established, the microduplication is associated with a very variable phenotype, of which behavioural problems and poor social interaction seem to be the most consistent.

- 19
- 20

# 21 ACKNOWLEDGMENT.

We would like to thank Mélanie D'Amico and Dr Pascale Hilbert for their support with 22 23 microsatellites analysis. This work was funded by a grant from the "Institute de Recherche Scientifique en Pathologie et en Génétique" and by grants to JV from the Catholic University 24 25 of Leuven (GOA/2006/12 and Centre of Excellence SymBioSys, Research Council 26 K.U.Leuven EF/05/007) and the IWT (IWT 60848). We are grateful to the MicroArray 27 Facility, Flanders Interuniversity Institute for Biotechnology (VIB) for their help in the 28 spotting of the arrays and the Mapping Core and Map Finishing groups of the Wellcome Trust 29 Sanger Institute for the initial clone supply and verification. This work was also funded by a 30 Biomedical Research grant from the National Institute for Health Research (NIHR), UK.

- 31
- 32 Competing interests : J Rosenfeld is employee of Signature Genomic.
   33
- 34 **Patient consents :** Obtained.
- 35

36 Statement : The Corresponding Author has the right to grant on behalf of all authors and does grant on behalf of all authors, an exclusive licence (or non exclusive for government 37 38 employees) on a worldwide basis to the BMJ Publishing Group Ltd to permit this article to be 39 published in Journal of Medical Genetics and any other BMJPGL products and sublicences 40 exploit all subsidiary rights, as set out in our such use and licence 41 (http://JMG.bmj.com/misc/ifora/licenceform.shtml).

42 43

# 44 <u>**REFERENCE.</u>**</u>

- I Slavotinek AM. Novel microdeletion syndromes detected by chromosome microarrays.
   Hum Genet. 2008;124:1-17.
- 48
- 2 Koolen DA, Sharp AJ, Hurst JA, Firth HV, Knight SJL, Goldenberg A, Saugier-Veber P,
   Pfundt R, Vissers LELM, Destrée A, Grisart B, Rooms L, Van der Aa N, Field M,

- 1 Hackett A, Bell K, Nowaczyk MJM, Mancini GMS, Poddighe PJ, Schwartz CE, Rossi E, 2 De Gregori M, Antonacci-Fulton LL, McLellan II MD, Garrett JM, Wiechert MA, Miner TL, Crosby S, Ciccone R, Rauch A, Zenker M, Aradhya S, Manning MA, Strom TM, 3 Wagenstaller J, Krepischi-Santos AC, Vianna-Morgante AM, Rosenberg C, Price SM, 4 Stewart H, Shaw-Smith C, Brunner HG, Wilkie AOM, Veltman JA, Zuffardi O, Eichler 5 EE, de Vries BBA. Clinical and molecular delineation of the 17q21.31 microdeletion 6 7 syndrome. Genet. Published Online First: 17 J Med July 2008. 8 doi:10.1136/jmg.2008.058701.
- 3 Koolen DA, Vissers LE, Pfundt R, de Leeuw N, Knight SJ, Regan R, Kooy RF, Reyniers E,
  Romano C, Fichera M, Schinzel A, Baumer A, Anderlid BM, Schoumans J, Knoers NV,
  van Kessel AG, Sistermans EA, Veltman JA, Brunner HG, de Vries BB. A new
  chromosome 17q21.31 microdeletion syndrome associated with a common inversion
  polymorphism. Nat Genet 2006;**38**:999-1001.
- 15

- 4 Sharp AJ, Hansen S, Selzer RR, Cheng Z, Regan R, Hurst JA, Stewart H, Price SM, Blair E,
  Hennekam RC, Fitzpatrick CA, Segraves R, Richmond TA, Guiver C, Albertson DG,
  Pinkel D, Eis PS, Schwartz S, Knight SJ, Eichler EE. Discovery of previously
  unidentified genomic disorders from the duplication architecture of the human
  genome.Nat Genet 2006;**38**:1038-1042.
- 21
- 5 Shaw-Smith C, Pittman AM, Willatt L, Martin H, Rickman L, Gribble S, Curley R,
  Cumming S, Dunn C, Kalaitzopoulos D, Porter K, Prigmore E, Krepischi-Santos AC,
  Varela MC, Koiffmann CP, Lees AJ, Rosenberg C, Firth HV, de Silva R, Carter NP.
  Microdeletion encompassing MAPT at chromosome 17q21.3 is associated with
  developmental delay and learning disability. Nat Genet 2006;**3**:1032-1037.
- 6 Varela MC, Krepischi-Santos AC, Paz JA, Knijnenburg J, Szuhai K, Rosenberg C,
  Koiffmann CP. A 17q21.31 microdeletion encompassing the MAPT gene in a mentally
  impaired patient. Cytogenet Genome Res 2006;114:89-92.
- 31

- 7 Stefansson H, Helgason A, Thorleifsson G, Steinthorsdottir V, Masson G, Barnard J, Baker
  A, Jonasdottir A, Ingason A, Gudnadottir VG, Desnica N, Hicks A, Gylfason A,
  Gudbjartsson DF, Jonsdottir GM, Sainz J, Agnarsson K, Birgisdottir B, Ghosh S,
  Olafsdottir A, Cazier JB, Kristjansson K, Frigge ML, Thorgeirsson TE, Gulcher JR, Kong
  A, Stefansson K. A common inversion under selection in Europeans. Nat Genet
  2005;**37**:129-137.
- 38
- 8 Turner DJ, Miretti M, Rajan D, Fiegler H, Carter NP, Blayney ML, Beck S, Hurles ME.
  Germline rates of de novo meiotic deletions and duplications causing several genomic
  disorders. Nat Genet 2008;40:90-95.
- 43 9 Kirchhoff M, Bisgaard AM, Duno M, Hansen FJ, Schwartz M. A 17q21.31
  44 microduplication, reciprocal to the newly described 17q21.31 microdeletion, in a girl with
  45 severe psychomotor developmental delay and dysmorphic craniofacial features. Eur J
  46 Med Genet 2007;**50**:256-263.
- 47

<sup>10</sup> Menten B, Maas N, Thienpont B, Buysse K, Vandesompele J, Melotte C, de Ravel T, Van
Vooren S, Balikova I, Backx L, Janssens S, De Paepe A, De Moor B, Moreau Y, Marynen
P, Fryns JP, Mortier G, Devriendt K, Speleman F, Vermeesch JR. Emerging patterns of

1

cryptic chromosomal imbalance in patients with idiopathic mental retardation and multiple congenital anomalies: a new series of 140 patients and review of published reports. J Med Genet 2006;**43**:625-33.

4 5

> 6 7

> 8

11 Hoebeeck J, Speleman F, Vandesompele J. Real-time quantitative PCR as an alternative to Southern blot or fluorescence in situ hybridization for detection of gene copy number changes. Methods Mol Biol 2007;**353**:205-226.

9 12 Baker M, Litvan I, Houlden H, Adamson J, Dickson D, Perez-Tur J, Hardy J, Lynch T,
10 Bigio E, Hutton M. Association of an extended haplotype in the tau gene with
11 progressive supranuclear palsy. Hum Mol Genet. 1999;8:711-5.

12

13 Cantor RM, Kono N, Duvall JA, Alvarez-Retuerto A, Stone JL, Alarcón M, Nelson SF,
 Geschwind DH. Replication of autism linkage: fine-mapping peak at 17q21. Am J Hum
 Genet 2005;76:1050-1056.

16

17 14 Szatmari P, Paterson AD, Zwaigenbaum L, Roberts W, Brian J, Liu XQ, Vincent JB, 18 Skaug JL, Thompson AP, Senman L, Feuk L, Qian C, Bryson SE, Jones MB, Marshall CR, Scherer SW, Vieland VJ, Bartlett C, Mangin LV, Goedken R, Segre A, Pericak-19 20 Vance MA, Cuccaro ML, Gilbert JR, Wright HH, Abramson RK, Betancur C, Bourgeron 21 T, Gillberg C, Leboyer M, Buxbaum JD, Davis KL, Hollander E, Silverman JM, 22 Hallmayer J, Lotspeich L, Sutcliffe JS, Haines JL, Folstein SE, Piven J, Wassink TH, 23 Sheffield V, Geschwind DH, Bucan M, Brown WT, Cantor RM, Constantino JN, Gilliam 24 TC, Herbert M, Lajonchere C, Ledbetter DH, Lese-Martin C, Miller J, Nelson S, 25 Samango-Sprouse CA, Spence S, State M, Tanzi RE, Coon H, Dawson G, Devlin B, Estes 26 A, Flodman P, Klei L, McMahon WM, Minshew N, Munson J, Korvatska E, Rodier PM, 27 Schellenberg GD, Smith M, Spence MA, Stodgell C, Tepper PG, Wijsman EM, Yu CE, Rogé B, Mantoulan C, Wittemeyer K, Poustka A, Felder B, Klauck SM, Schuster C, 28 29 Poustka F, Bölte S, Feineis-Matthews S, Herbrecht E, Schmötzer G, Tsiantis J, 30 Papanikolaou K, Maestrini E, Bacchelli E, Blasi F, Carone S, Toma C, Van Engeland H, 31 de Jonge M, Kemner C, Koop F, Langemeijer M, Hijmans C, Staal WG, Baird G, Bolton 32 PF, Rutter ML, Weisblatt E, Green J, Aldred C, Wilkinson JA, Pickles A, Le Couteur A, 33 Berney T, McConachie H, Bailey AJ, Francis K, Honeyman G, Hutchinson A, Parr JR, Wallace S, Monaco AP, Barnby G, Kobayashi K, Lamb JA, Sousa I, Sykes N, Cook EH, 34 35 Guter SJ, Leventhal BL, Salt J, Lord C, Corsello C, Hus V, Weeks DE, Volkmar F, 36 Tauber M, Fombonne E, Shih A, Meyer KJ. Mapping autism risk loci using genetic 37 linkage and chromosomal rearrangements. Nat Genet 2007;39:319-328.

38

15 Sebat J, Lakshmi B, Malhotra D, Troge J, Lese-Martin C, Walsh T, Yamrom B, Yoon S,
Krasnitz A, Kendall J, Leotta A, Pai D, Zhang R, Lee YH, Hicks J, Spence SJ, Lee AT,
Puura K, Lehtimäki T, Ledbetter D, Gregersen PK, Bregman J, Sutcliffe JS, Jobanputra
V, Chung W, Warburton D, King MC, Skuse D, Geschwind DH, Gilliam TC, Ye K,
Wigler M. Strong association of de novo copy number mutations with autism. Science
2007;**316**:445-449.

45

Pittman AM, Fung H-C, de Silva R. Untangling the tau gene association with
 neurodegenerative disorders. Hum Mol Genet 2006 ;15:R188-R195.

- 17 Lladó A, Rodríguez-Santiago B, Antonell A, Sánchez-Valle R, Molinuevo JL, Reñé R, Pérez-Jurado LA. MAPT gene duplications are not a cause of frontotemporal lobar degeneration. Neurosci Lett 2007;424:61-65.
   18 King JS, Bishop GA. Localization of the type 1 corticotropin releasing factor receptor (CRF-R1) in the embryonic mouse cerebellum. J Neurocytol 2003;32:305-316.
- 8 19 Hauger RL, Risbrough V, Brauns O, Dautzenberg FM. Corticotropin Releasing Factor
  9 (CRF) Receptor Signaling in the Central Nervous System: New Molecular Targets. CNS
  10 Neurol Disord Drug Targets 2006;5:453-479.
- 11

18

22

27

37

41

- 20 Kehne JH. The CRF1 receptor, a novel target for the treatment of depression, anxiety, and
   stress-related disorders. CNS Neurol Disord Drug Targets. 2007;6:163-182.
- Armogida M, Petit A, Vincent B, Scarzello S, Alves da Costa C, Checler F. Endogenous amyloid production in presenilin-deficient embryonic mouse fibroblasts. Nat Cell Biol
   2001;3:1030-1033.
- 19 22 Grigorenko AP, Moliaka YK, Korovaitseva GI, Rogaev EI. Novel Class of Polytopic
   20 Proteins with Domains Associated with Putative Protease Activity. Biochemistry
   21 (Moscow) 2002;67:826-834.
- 23 Potocki L, Chen KS, Park SS, Osterholm DE, Withers MA, Kimonis V, Summers AM,
  Meschino WS, Anyane-Yeboa K, Kashork CD, Shaffer LG, Lupski JR. Molecular
  mechanism for duplication 17p11.2-the homologous recombination reciprocal of the
  Smith-Magenis microdeletion. Nat Genet 2000;24:84-87.
- 24 Kriek M, White SJ, Szuhai K, Knijnenburg J, van Ommen GJ, den Dunnen JT, Breuning
  MH. Copy number variation in regions flanked (or unflanked) by duplicons among
  patients with developmental delay and/or congenital malformation; detection of reciprocal
  and partial Williams-Beuren duplication. Eur J Hum Genet 2006;14:180-189.
- 25 Ou Z, Berg JS, Yonath H, Enciso VB, Miller DT, Picker J, Lenzi T, Keegan CE, Sutton
  VR, Belmont J, Chinault AC, Lupski JR, Cheung SW, Roeder E, Patel A.
  Microduplications of 22q11.2 are frequently inherited and are associated with variable
  phenotypes. Genet Med 2008;10:267-277.
- 26 Grisart B, Rack K, Vidrequin S, Hilbert P, Deltenre P, Verellen-Dumoulin C, Destrée A.
   NF1 microduplication first clinical report: association with mild mental retardation, early
   onset of baldness and dental enamel hypoplasia? Eur J Hum Genet 2008;16:305-311.
- 42 27 Ben-Shachar S, Ou Z, Shaw CA, Belmont JW, Patel MS, Hummel M, Amato S, Tartaglia
  43 N, Berg J, Sutton VR, Lalani SR, Chinault AC, Cheung SW, Lupski JR, Patel A. 22q11.2
  44 distal deletion: a recurrent genomic disorder distinct from DiGeorge syndrome and
  45 velocardiofacial syndrome. Am J Hum Genet 2008;82:214-221.
- 47 28 Mefford HC, Sharp AJ, Baker C, Itsara A, Jiang Z, Buysse K, Huang S, Maloney VK,
  48 Crolla JA, Baralle D, Collins A, Mercer C, Norga K, de Ravel T, Devriendt K, Bongers
  49 EM, de Leeuw N, Reardon W, Gimelli S, Bena F, Hennekam RC, Male A, Gaunt L,
  50 Clayton-Smith J, Simonic I, Park SM, Mehta SG, Nik-Zainal S, Woods CG, Firth HV,

1 Parkin G, Fichera M, Reitano S, Lo Giudice M, Li KE, Casuga I, Broomer A, Conrad B, 2 Schwerzmann M, Räber L, Gallati S, Striano P, Coppola A, Tolmie JL, Tobias ES, Lilley C, Armengol L, Spysschaert Y, Verloo P, De Coene A, Goossens L, Mortier G, Speleman 3 F, van Binsbergen E, Nelen MR, Hochstenbach R, Poot M, Gallagher L, Gill M, 4 McClellan J, King MC, Regan R, Skinner C, Stevenson RE, Antonarakis SE, Chen C, 5 Estivill X, Menten B, Gimelli G, Gribble S, Schwartz S, Sutcliffe JS, Walsh T, Knight SJ, 6 7 Sebat J, Romano C, Schwartz CE, Veltman JA, de Vries BB, Vermeesch JR, Barber JC, 8 Willatt L, Tassabehji M, Eichler EE. Recurrent rearrangements of chromosome 1q21.1 9 and variable pediatric phenotypes. N Engl J Med. 2008;359:1685-1699. 10

10

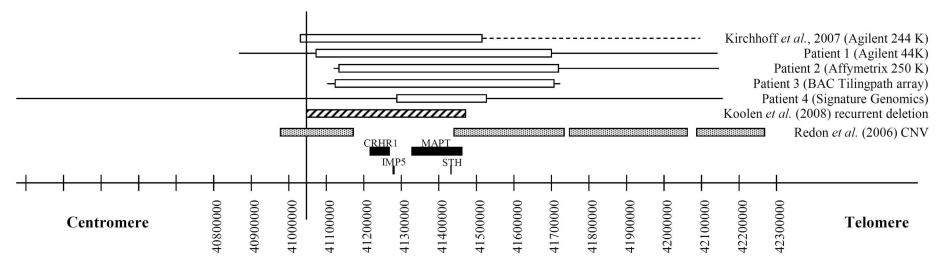
29 Zody MC, Jiang Z, Fung HC, Antonacci F, Hillier LW, Cardone MF, Graves TA, Kidd
JM, Cheng Z, Abouelleil A, Chen L, Wallis J, Glasscock J, Wilson RK, Reily AD,
Duckworth J, Ventura M, Hardy J, Warren WC, Eichler EE. Evolutionary toggling of the
MAPT 17q21.31 inversion region. Nat Genet 2008;40:1076-1083.

# 1 Figure legend

Figure 1 : Pictures of patient 1 (a) and patient 3 (b). Consents to publish these pictures were
obtained from the parents of the patients.

6 Figure 2 : Schematic representation of genomic duplication in Kirschhoff's patient [9] and in 7 the 4 patients presented here. Array used in each cases is noticed at the right. 8 Duplicated genomic regions are showed as white rectangle. A thin horizontal line on both side illustrate the break point regions. The exact distal position of the 9 duplication was not available in Kirschhoff's paper (thin dotted line). The regions 10 of overlap of normal and duplicated BAC in patient 3 are shown as a thin 11 12 horizontal line even if these regions do not actually correspond to breakpoints regions. The minimal recurrent deletion reported by Koolen et al. [2] is depicted 13 14 as a shaded rectangle. Dotted rectangles illustrates known CNV. Positions of the 15 4 genes mentioned in the discussion are depicted as black rectangles. The thin 16 vertical line localize the position of the recurrent proximal breakpoint [2].





Map position on hg18 assembly