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Altered thyrotropic and lactotropic axes regulation in Huntington's disease

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Abstract

Background: Recently a loss of hypothalamic dopamine D₂ receptors was demonstrated in Huntington's disease. Activation of dopamine D₂ receptors is known to inhibit both thyrotropic and lactotropic axes function. Objective: To assess whether thyrotropic and lactotropic axes activity are disturbed in HD patients, contributing to symptoms such as unintended weight loss. Participants & methods: In nine early-stage, medication-free HD patients (6 males, 3 females) and nine age-, sex- and body mass indexmatched controls, we measured serum levels of thyroid-stimulating hormone (TSH) and prolactin (males only) every 10 min for 24 h. Multi-parameter auto-deconvolution and approximate entropy analysis were applied to quantify basal, pulsatile and total TSH and prolactin secretion rates as well as the regularity of hormone release. Results: Compared with controls, TSH and prolactin secretion tended to be slightly, but not significantly, higher in HD patients (TSH: 1.13±0.14 vs. 0.91±0.19 mU/L, p=0.40; prolactin: 4.91±0.42 vs. 4.83±0.26 μg/L, p=0.87). However, in HD patients total T₃ levels were significantly higher $(1.60\pm0.05 \text{ vs. } 1.35\pm0.09, p=0.045), \text{ while } T_4 \text{ levels tended to be higher as well } (91.9\pm3.9 \text{ vs. } 81.3\pm3.1,$ p=0.085). Prolactin secretion was significantly more irregular in HD patients (ApEn; 1.06 ± 0.08 vs. 0.80 ± 0.09 , p=0.037). Total T₃ levels were negatively associated with motor impairment (r=-0.72, p=0.030), whereas increasing free T₄ levels were associated with a larger mutant CAG repeat size (r=+0.68, p=0.044). Conclusion: Our findings indicate a mild hyperactivity of the thyrotropic axis and a disturbed regulation of the lactotropic axis in early stage HD patients.

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Key words: Huntington's disease, thyroid-stimulating hormone, thyroid hormones, prolactin, weight loss

Introduction

Huntington's disease (HD) is a progressive, autosomal dominant neurodegenerative disorder caused by a CAG repeat expansion in exon 1 of the *HTT* gene, resulting in a long polyglutamine tract in the N-terminus of the encoded protein huntingtin. It is characterized by motor disturbances, cognitive decline and behavioral problems. Progressive weight loss and muscle wasting are also hallmarks of the disease, both in HD patients and several transgenic mouse models of the disease. Moreover, abnormalities in glucose homeostasis as well as a higher prevalence of diabetes mellitus have been reported in HD patients, which are also evident in the transgenic models. The cause of these peripheral signs is largely unknown, although hypothalamic dysfunction and subsequent endocrine alterations may be involved.

Both the thyrotropic and lactotropic axes are intimately involved in the complex neuroendocrine regulation of body weight and metabolism. ^{8,9} Relatively few studies have, however, evaluated hypothalamic-pituitary-thyroid axis function in HD patients. Although basal levels of total thyroxine (T₄), triiodothyronine (T₃), free T₄, and thyroid-stimulating hormone (TSH) have been reported to be similar between HD patients and normal controls, ¹⁰⁻¹² others have found an impaired TSH response to thyrotropin-releasing hormone (TRH) stimulation. ¹³ Moreover, in a retrospective chart review study of 97 HD patients residing in long-term care facilities, the most commonly prescribed drug for problems 'unrelated' to HD was found to be levothyroxine. ¹⁴ Compared to thyrotropic axis function, lactotropic axis activity in HD patients has been investigated more intensively. ¹⁵ Nevertheless, as prolactin levels in HD patients have been reported to be unchanged, ^{10,16} increased, ¹¹ or even decreased, ¹⁷ it still remains unknown whether the lactotropic axis is indeed affected in HD or whether altered prolactin levels are merely a consequence of anti-dopaminergic medication use in HD. The discordances in findings regarding thyrotropic and lactotropic axes functioning in HD are likely due to the use of a few baseline

measurements of hormone levels or long blood sampling intervals which are not adequate to assess either the pulsatile nature of TSH and prolactin secretion or their total daily production rates.¹⁸

Recently a loss of hypothalamic dopamine D_2 receptors was demonstrated in both early stage HD patients as well as premanifest HD mutation carriers.¹⁹ Activation of dopamine D_2 receptors is known to inhibit both thyrotropic and lactotropic axes function.^{20,21} Therefore, we postulated that loss of hypothalamic D_2 receptors in HD patients may give rise to disinhibited thyrotropic and lactotropic axes activity, contributing to the disrupted energy homeostasis in these subjects. We tested this hypothesis by deconvolution analysis of 24 h serum TSH and prolactin concentration profiles as well as assessment of thyroid hormone levels in both early stage, medication-free HD patients and healthy matched controls.

Subjects and Methods

Subjects

Nine early stage HD patients (six males and three females) and nine healthy control subjects, matched for age, sex, and body mass index (BMI), were enrolled in the study. Thyroid axis function was assessed in all participants. However, as estrogens can have a marked impact on prolactin secretion, lactotropic axis activity was assessed in male subjects only. Clinical details are summarized in **Table 1**. In the patient group, mutant CAG repeat size ranged between 41 and 50. The clinical diagnosis of HD was made by a neurologist specialized in movement disorders (R.A.C.R.). The Unified Huntington's Disease Rating Scale (UHDRS) was used to assess HD symptoms and signs. None of the subjects used medication, except one male HD patient who discontinued paroxetine ($t_{1/2} \approx 21$ h) use three weeks prior to study. Subjects were eligible for participation after exclusion of hypertension, any known (history of) pituitary or thyroid disease, a struma on physical examination, recent intentional weight change (>3 kg weight gain or

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loss within the last 3 months), and any other chronic conditions except HD as assessed by clinical examination and routine laboratory tests. Written informed consent was obtained from all subjects. The study was approved by the medical ethics committee of the Leiden University Medical Center.

Clinical protocol

Subjects were admitted to the Clinical Research Center for 24 h blood sampling. Two women (one patient and one control) were postmenopausal, the other women were studied in the early follicular phase of their menstrual cycle. A cannula was inserted into an antecubital vein 45 min before the start of blood sampling at 1630 h. Blood samples were collected with S-monovetten (Sarstedt, Etten-Leur, The Netherlands) from a three-way stopcock that was attached to a 0.9% NaCl and heparin (1 U/ml) infusion (500 ml/24 h) to keep the cannula from clotting. Sampling was performed through a long line to prevent sleep disruption by investigative manipulations. During 24 h, blood was collected in serum tubes at 10-min intervals.

Blood was allowed to clot and, within 60 min of sampling, all tubes were centrifuged at 1610 g at 4 °C for 20 min, and serum was stored at -80 °C until assay. Three standardized meals were served at 0900, 1300, and 1900 h (Nutridrink, 1.5 kcal/ml, 1500–1800 kcal/d; macronutrient composition per 100 ml: protein, 5 g; fat, 6.5 g; carbohydrate, 17.9 g; Nutricia, Zoetermeer, The Netherlands). Subjects remained sedentary except for bathroom visits. Twenty-four hour urine was collected for the determination of creatinine, catecholamines and cortisol concentrations. No daytime naps were allowed. Lights were switched off at 2300 h and, the next morning, subjects were awakened at 0730 h.

Body composition

Bioelectrical impedance analysis was used to assess lean body mass and fat percentage at 0800 h.

Assays

Serum TSH and prolactin levels were measured by time-resolved immunofluorometic assays (Delfia, Wallac Oy, Turku, Finland). The detection limit of the TSH assay was 0.01 mU/L, and the interassay variation ranged from 3.1 to 8.3% at very low levels. The detection limit of the prolactin assay was 0.05 µg/L, and the interassay variation ranged from 2.7 to 3.8%. Serum T₄ and T₃ levels were measured with Abbott Axsym (Abbott Laboratories, Abbott Park, IL). Free T₄ concentrations were estimated using electrochemiluminescence immunoassays (Roche Diagnostic Nederland BV, Almere, The Netherlands). Urinary epinephrine, norepinephrine and dopamine concentrations were assessed by high performance liquid chromatography with electron capture detection (ESTA-Coulochem, Chelmsford, MA, USA).

Calculations and statistics

Deconvolution analysis. A recently developed, fully automatic, multi-parameter deconvolution procedure, *AutoDecon*, was used to estimate various specific measures of secretion and disappearance rate of TSH and prolactin, considering all serum hormone concentrations and their dose-dependent intra-sample variance simultaneously.^{23,24} The standard deviation of the secretion events was initialized to 5-min. For TSH, a fixed two-component half-life was assumed with 18-min for the first component and 92-min for the second component, with a relative contribution of 32% of the slow component to the total elimination.²⁵ For prolactin, a starting one-component half-life of 45-min was assumed,²⁶ and the *AutoDecon* algorithm was then used to find the best fit. The following parameters of the TSH and prolactin time series were estimated: number of secretory bursts, secretory burst half-duration (duration at half-maximal amplitude), mean mass secreted per burst, hormone half-life, basal secretion rate, pulsatile secretion rate, and total secretion rate.

Diurnal rhythmicity analysis. Twenty-four-hour variations in serum hormone concentrations were assessed by cosinor regression, an algorithm that fits a cosine function to the data using repeated nonlinear regression. This analysis estimates an acrophase, which is the clock time during the 24 h period at which

hormone concentration is maximal; a mesor, which is the average value about which the diurnal rhythm oscillates; and an amplitude, which is half the difference between the peak and nadir values of the 24 h concentration series.

Approximate entropy (ApEn). ApEn is a model-independent statistic used to quantify the regularity of a time series, in which is measured, within a predefined tolerance r given a pattern of window length m, the likelihood of a similar pattern in the next incremental window.²⁷ Greater regularity yields smaller ApEn values, whereas greater independence among sequential values of a time series yields larger ApEn values. ApEn parameters of m = 1 and r = 20% of the intra-series standard deviation were used, the statistical suitability of which has been established previously.²⁷

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Statistical analysis. Results are expressed as mean \pm standard error (SE) unless otherwise specified. Non-parametric Mann-Whitney U tests were used to assess intergroup differences. Pearson's correlation coefficient was applied to assess all correlations. All tests were two-tailed and significance level was set at p < 0.05. Statistical analyses were performed using SPSS for Windows (release 16.0, SPSS, Inc., Chicago, IL).

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Subjects

Results

The HD and the control group did not differ with respect to age, sex, BMI, body fat or lean body mass (all $p \ge 0.27$, Table 1). There were also no significant differences in urinary creatinine, epinephrine, norepinephrine and dopamine levels (all $p \ge 0.10$).

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Deconvolution analysis of TSH time series

Mean 24 h TSH and prolactin concentrations tended to be slightly, but not significantly, higher in HD patients compared with controls (TSH: $1.13 \pm 0.14 \text{ vs. } 0.91 \pm 0.19 \text{ mU/L}$, p = 0.40; prolactin: 4.91 ± 0.42

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vs. $4.83 \pm 0.26 \,\mu\text{g/L}$, p = 0.87; Figure 1 and supplementary Figure 1). The number of TSH and prolactin pulses as well as their basal, pulsatile and total secretion rates were also similar in the patient and control group (all $p \ge 0.12$). Details of all deconvolution-derived TSH and prolactin secretory kinetics are presented in Table 2.

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Diurnal rhythmicity analysis

TSH and prolactin displayed significant diurnal variations, both in patients and in controls (Figure 1).

However, the acrophase, amplitude and mesor of the TSH and prolactin concentration series were not significantly different between HD patients and controls (all $p \ge 0.27$).

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Regularity of TSH and prolactin concentration time series

The ApEn values of the TSH time series were similar between HD patients and controls ($p \ge 0.83$). However, the ApEn values of the prolactin time series were significantly higher in HD patients compared with controls (1.06 \pm 0.08 vs. 0.80 \pm 0.09, p = 0.037), indicating significantly more irregular prolactin secretion in HD patients (**Figure 2**).

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Thyroid hormone levels

Fasting levels of both T_3 and T_4 were higher in HD patients compared with controls (T_3 : 1.60 \pm 0.05 vs.

 $1.35 \pm 0.09 \underline{\text{nmol/L}}$, p = 0.045; T_4 : $91.9 \pm 3.9 \text{ vs.} 81.3 \pm 3.1 \underline{\text{nmol/L}}$, p = 0.085). Free T_4 levels also tended to be higher in HD patients, but the difference was much less pronounced (15.1 $\pm 0.70 \text{ vs.} 14.2 \pm 0.46 \underline{\text{pmol/L}}$, p = 0.45).

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Thyrotropic and lactotropic axes activity in relation to clinical phenotype

In HD patients, total daily TSH and prolactin secretion rates were not significantly associated with BMI, motor score, total functional capacity or mutant CAG repeat size (all $p \ge 0.17$). However, total T_3 levels were significantly associated with motor impairment (r = -0.72, p = 0.030). Moreover, increasing mutant CAG repeat size was significantly related to higher free T_4 levels (r = +0.68, p = 0.044; supplementary Figure 2). Associations between total T_4 levels and mutant CAG repeat size (r = +0.63, p = 0.069), and total T_4 levels and BMI (r = -0.64, p = 0.064) did not reach statistical significance.

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Discussion

In this study we rigorously evaluated thyrotropic and lactotropic axes function in a group of early stage, medication-free HD patients. Although daily TSH production rates were similar between HD patients and matched controls, thyroid hormone levels were significantly higher in HD patients, consistent with a mild hyperactivity of the hypothalamic-pituitary-thyroid axis. Total daily prolactin production rates were also similar between HD patients and controls, however, prolactin secretion was significantly more irregular in HD patients.

Thyroid hormones (T₄ and T₃) are critically involved in the regulation of systemic energy homeostasis and their secretion is tightly regulated by a complex interplay of positive and negative feedback loops. Hypothalamic TRH induces pituitary TSH secretion which then stimulates the synthesis and release of thyroid hormones by the thyroid gland. Although TSH synthesis and secretion are primarily controlled by the stimulatory action of TRH and the negative feedback restraint by thyroid hormones, other factors such as dopamine exert important modulatory effects. Dopamine has a dual influence on TSH secretion: it inhibits TSH synthesis and release through D₂ receptor activation at the level of the pituitary thyrotropes, whereas it stimulates TRH secretion by the hypophysiotropic neurons located in the paraventricular nucleus. Using positron emission tomography with

et al.¹⁹ recently demonstrated a loss of D_2 receptors in the hypothalamus of both early stage HD patients and premanifest HD mutation carriers. Moreover, in the R6/2 transgenic mouse model of HD a loss of D_2 receptors has also been reported at the level of the anterior pituitary.²⁸ Therefore, our finding of a mild thyrotropic axis hyperactivity in HD patients may, at least partly, be attributed to a specific pattern of hypothalamic and pituitary D_2 receptor loss. The modest decrease of hypothalamic D_2 receptors (by about 28%) in early stage HD patients, ¹⁹ may explain the relatively mild increase in the activity of the thyrotropic axis in our cohort.

Altered hypothalamic-pituitary dopamine signaling may also underlie the significantly more irregular pattern of prolactin release in HD patients. There is now ample evidence that dopamine of tubero-infundibular origin, delivered through long portal vessels into the sinusoid capillaries of the anterior pituitary, is the major physiological regulator of prolactin release. Hypothalamic dopamine inhibits the basally high-secretory tone of pituitary lactotrophs by binding to D₂ receptors expressed on their cell membranes. Prolactin in turn regulates the activity of the tubero-infundibular neurons via a short-loop feedback mechanism. Hence, pathology of the tubero-infundibular dopaminergic system, located in the hypothalamic infundibular nucleus (i.e. the human homologue of the arcuate nucleus in rodents), or loss of pituitary D₂ receptor expression as described in the R6/2 mice²⁸ could both underlie the irregular pattern of prolactin secretion in HD patients. The diminished regularity of prolactin secretion even in our cohort of early stage, medication-free HD patients may also account for the inconsistencies in findings from previous studies on prolactin levels in HD subjects, ^{7,11,16,17} since due to the irregular pattern of prolactin release single or a few baseline measurements are likely to yield ambiguous outcomes. It

abnormal responses to physiological stimuli of prolactin release such as stress.8 Neuropathological

evaluation of the infundibular nucleus, as well as in vivo assessment of pituitary D_2 receptor binding in

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HD patients could provide more mechanistic insights into the basis of this abnormality in prolactin secretion.

Interestingly, when assessed in relation to clinical characteristics, higher free T₄ levels were associated with larger mutant CAG repeat sizes. In addition, there was an inverse trend for the relation between total T₄ levels and BMI in HD patients. As thyroid hormones are known to increase energy expenditure, elevated thyroid hormone levels in early stage HD patients that seem to increase with mutant CAG repeat size, may contribute to the lower BMI in HD mutation carriers,^{29,30} and possibly account for the association between mutant *HTT* CAG repeat size and weight loss in HD.² As mutant CAG repeat size was not associated with TSH secretion, its association with free T₄ levels is more likely to be mediated peripherally, for example, by a direct effect of mutant huntingtin on tissue deiodinases that are found throughout the body.^{9,31} On the other hand, total T₃ levels decreased with increasing motor impairment paralleling findings from a recent study.¹¹ The underlying mechanism is unclear, although it may result from a progressive dysfunction or loss of hypothalamic TRH producing neurons.⁷ However, larger scale studies in, especially early stage and neuroleptic-free, HD patients are needed to confirm these preliminary associations.

In conclusion, we found a mild hyperactivity of the hypothalamic-pituitary-thyroid axis, as well as a more irregular pattern of prolactin secretion in HD patients compared with matched controls. These findings are consistent with disrupted hypothalamic-pituitary dopamine signaling in HD. However, other mechanisms such as reduced differentiation and function of hypothalamic neurosecretory cells, as well as diminished expression of hypothalamic neuropeptides due to local mutant huntingtin expression may also be involved. 32 Further neuropathological, imaging and functional studies are necessary to unveil the cause of these abnormalities and provide rationale for potential endocrine-based therapies for HD.

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Figure 1. Mean serum TSH (A) and prolactin (B) concentrations in HD and control subjects.

Sampling started at 1630 h and was continued at 10-min intervals for 24 h. The acrophase, amplitude and mesor of the TSH and prolactin concentration series were not significantly different between HD patients and controls (all $p \ge 0.27$). Acrophase = the clock time during the 24 h period at which hormone concentration is maximal. Mesor = the average value about which the diurnal rhythm oscillates.

Amplitude = half the difference between the peak and nadir values of the 24 h concentration series. Error

bars indicate the standard error of the mean.

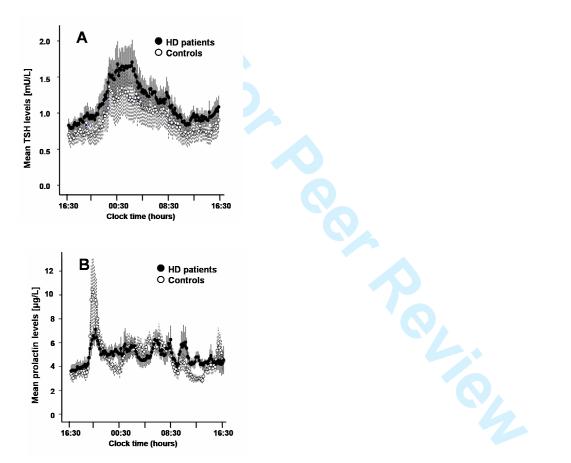
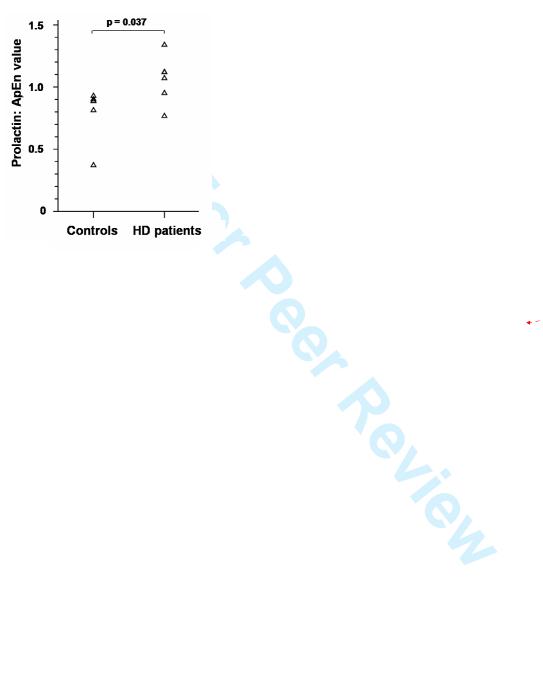


Figure 2. Prolactin secretion regularity in HD patients and controls. Compared with controls, the approximate entropy (ApEn) of prolactin secretion is higher in HD patients $(0.80 \pm 0.09 \text{ vs. } 1.06 \pm 0.08 \text{ p} = 0.037)$, indicating significantly more irregular prolactin release.

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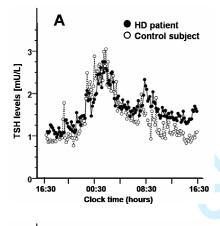
Deleted: $0.48 \pm 0.04 \ vs. \ 0.61 \pm 0.04$

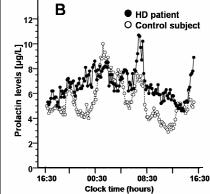
Deleted: Error bars indicate the standard error of the mean.



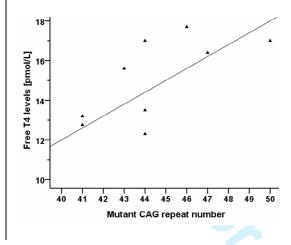
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Supplementary figure 1. Serum TSH (A) and prolactin (B) concentrations in a representative HD patient and his matched control subject. Sampling started at 1630 h and was continued at 10-min intervals for 24 h.





Supplementary figure 2. Association between mutant CAG repeat number and free T_4 levels in HD patients. Increasing mutant CAG repeat size was significantly associated with higher free T_4 levels (r = ± 0.68 , p = 0.044).



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Table 1. Characteristics of the study population

| | HD patients ¹ | Controls ¹ | p-value ² |
|---------------------------------|--------------------------|-----------------------|----------------------|
| Male/female | 6/3 | 6/3 | - |
| | | | |
| Age [y] | 47.1 (3.4) | 48.6 (3.3) | 0. <u>691</u> |
| | | | |
| BMI | 24.1 (1.0) | 24.3 (0.6) | 0. <u>691</u> , |
| | | | |
| Fat [%] | 25.5 (2.4) | 25.6 (2.4) | 0. <u>825</u> , |
| | | | |
| Lean body mass [kg] | 57.3 (3.2) | 56.2 (3.0) | 0. <u>691</u> , |
| | 0.00 (0.00) | 0.04 (0.00) | 0.450 |
| Waist-to-hip ratio | 0.89 (0.03) | 0.94 (0.02) | 0.270, |
| Mutant CAG repeat size | 44.4 (1.0) | | |
| Mutant CAG repeat size | 44.4 (1.0) | _ | - |
| Age of onset [y] | 41.4 (3.0) | - | = |
| rigo or onsee [,] | 1211 (210) | | _ |
| Disease duration [y] | 5.7 (1.1) | - | - |
| | | | |
| UHDRS motor score | 22.2 (6.0) | - 9 | - |
| | | | |
| TFC score | 11.7 (0.7) | - | - |
| 1) Values are indicated as made | | | |

¹⁾ Values are indicated as mean (SE).

Abbreviations: BMI = Body Mass Index; TFC = Total Functional Capacity; UHDRS = Unified Huntington's Disease Rating Scale.

²) Differences between groups were assessed by <u>Mann-Whitney *U*</u> tests.

Table 2. Deconvolution analysis of 24 h serum TSH prolactin concentrations.

| | TS | БН | Prolactin | | |
|---|--------------------------|-----------------------|--------------------------|-----------------------|--|
| | HD patients ¹ | Controls ¹ | HD patients ¹ | Controls ¹ | |
| Basal secretion rate ² | 16.8 (1.8) | 12.8 (2.6) | 75.4 (27.8) | 44.8 (13.1) | |
| Pulsatile secretion rate ² | 10.4 (1.9) | 9.2 (2.5) | 59 (11) | 62 (7) | |
| Total secretion rate ² | 27.2 (3.6) | 22.0 (4.7) | 134 (38) | 106 (17) | |
| Percent pulsatile [%] | 36.0 (3.0) | 37.8 (4.4) | 48.5 (4.5) | 61.5 (7.5) | |
| Pulse half-duration [min] | 50.3 (5.7) | 57.2 (9.7) | 41.5 (11.4) | 30.3 (9.0) | |
| Pulse frequency [no./24 h] | 11.4 (1.2) | 10.9 (1.4) | 15.7 (1.2) | 17.3 (1.2) | |
| Mean mass secreted per pulse ³ | 0.94 (0.20) | 0.87 (0.22) | 4.2 (1.2) | 3.8 (0.7) | |

¹⁾ Values are indicated as mean (SE). There were no significant differences between HD patients and controls (unpaired t-tests).

²) Secretion rates are in mU/L/24 h for TSH, and ug/L/24 h for prolactin.

³) Mean mass secreted per pulse is in mU/L for TSH, and ug/L for prolactin.