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▶ To cite this version:

Suneeta Soni, Rajaventhan Srirajaskanthan, Sebastian Lucas, Sarah Alexander, Terry Wong, et al.. Lymphogranuloma venereum masquerading as Inflammatory Bowel Disease in 12 homosexual men. Alimentary Pharmacology and Therapeutics, 2010, 10.1111/j.1365-2036.2010.04313.x. hal-00552553

HAL Id: hal-00552553

https://hal.science/hal-00552553

Submitted on 6 Jan 2011

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Alimentary Pharmacology & Therapeutic

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Journal:	Alimentary Pharmacology & Therapeutics				
Manuscript ID:	APT-0165-2010.R1				
Manuscript Type:	Original Scientific Paper				
Date Submitted by the Author:	23-Mar-2010				
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Keywords:	Inflammatory bowel disease < Disease-based, Large intestine < Organ-based, Endoscopy < Topics, Histopathology < Topics, Microbiology < Topics				



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- disease in 12 homosexual men.
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SUMMARY

- 26 Background: Lymphogranuloma venereum(LGV) is a recognised cause of proctitis.
- 27 Symptoms, endoscopy and histology findings are similar in IBD and LGV proctitis.
- Aims: To characterise the clinical, endoscopic and histological features seen in men
- 29 diagnosed initially with IBD and subsequently with LGV proctitis. To attempt isolation
- of *Chlamydia trachomatis* DNA from the stored rectal biopsy specimens of these
- 31 patients using real-time PCR.
- 32 Methods: Clinical data were collated from confirmed or suspected cases of LGV
- 33 proctitis where endoscopy and biopsy had been performed as part of the
- investigation of clinical symptoms. LGV was confirmed by the detection of LGV-
- 35 specific DNA from rectal swab specimens, with supportive evidence from Chlamydial
- 36 serology. Stored histological specimens from rectal biopsies were analysed
- 37 retrospectively for LGV-specific DNA with molecular techniques.
- 38 Results: Rectal biopsies had been obtained from twelve cases of LGV proctitis.
- 39 Mucosal ulceration, cryptitis, crypt abscesses and granulomas were common
- 40 histological findings. Extraction of LGV-specific DNA from rectal biopsy specimens
- 41 enabled confirmation of three suspected cases.
- 42 Conclusions: During the recent LGV proctitis epidemic among UK men who have sex
- 43 with men, it has become apparent that this infection may closely resemble IBD.
- 44 Gastroenterologists should remain alert to LGV as a cause of proctitis in this group.

INTRODUCTION

Proctitis has several infectious and non-infectious aetiologies. In men who have sex with men (MSM) rectal infection with sexually transmitted pathogens such as herpes simplex virus (HSV), *Neisseria gonorrhoeae* (GC), *Treponema pallidum and Chlamydia trachomatis* (CT) are more common and important to consider. Lymphogranuloma venereum (LGV) proctitis is caused by L1, L2 and L3 serovars of CT and has evaded detection in recent decades in developed countries.

Following the report of an outbreak of LGV proctitis among MSM in the Netherlands in 2004 (1), the Health Protection Agency (HPA) in the UK introduced a national surveillance program and diagnostic support service in an attempt to enhance case ascertainment and to help control the spread of this sexually transmitted infection (STI) in the UK (2). Prior to October 2004, routine rectal Chlamydia testing had not been recommended in the UK for patients describing symptoms of proctitis. By December 2008, 849 UK cases of LGV had been confirmed (3). Where full genotyping has been possible, all cases have been confirmed as the L2 serovar of CT (4). This epidemic has been characterised by a preponderance of proctitis, rather than the classical LGV genital ulcer-adenopathy syndrome seen in tropical regions and affecting primarily heterosexuals. In the UK, LGV proctitis has been detected almost exclusively among men who have sex with men (MSM), many of whom are both HIV-positive and report high risk anal sexual practices with multiple and anonymous partners (5).

Rectal STIs can lead to histological changes of proctitis that may mimic IBD (6). The severe inflammatory changes seen with chlamydial infection are usually caused by the LGV-associated serovars (L1-3) whereas the oculogenital strains of CT (A-K) tend to produce milder proctitis histologically (7), and this infection is usually asymptomatic. Histological diagnosis of infective-type colitis, particularly in the acute phase, relies heavily on the lack of features usually seen in IBD, especially the crypt architectural distortion and the presence of a prominent increase in the cellularity of the lamina propria. Yet these features that usually predominate in IBD biopsies are also seen with LGV infection, including the development of well-formed granulomas. With more advanced LGV infection, transmural inflammation can ensue, further resembling Crohn's disease (8).

The aims of this study were firstly, to characterise the clinical, endoscopic and histological features seen in men diagnosed initially with IBD and subsequently with LGV proctitis, and secondly, to describe attempts to detect the causative organism from stored histological specimens with the use of advanced molecular techniques.

MATERIALS AND METHODS

106 cases of LGV proctitis were identified between October 2004 and August 2009 at our centre, a combination of Genitourinary medicine (GUM) clinic and HIV clinic patients. 78/106 (73.6%) were HIV-positive. All patients had rectal swab specimens taken for detection of *C. trachomatis* and *N. gonorrhoeae* DNA by strand displacement amplification (ProbeTec, Becton Dickinson, Sparks, MD) as well as culture for *Neisseriae*. After retrospective case note review of all HIV clinic patients it

was noted that 12 of these men had been referred to either gastroenterologists or colorectal surgeons for evaluation of their symptoms and had undergone rectal biopsy at some stage prior to their eventual diagnosis of LGV. None of these patients had undergone rectal testing for CT beforehand since this was not routine clinical practice outside of GUM clinics. Initial histological reports were reviewed and these stored slides were re-read by an expert pathologist experienced in reporting IBD specimens (SBL). Endoscopic findings were also reviewed. It is possible and likely that some HIV-negative men similarly may have undergone endoscopy and biopsy but such referrals would have been made through primary care and these reports would not be available to our GUM department.

Recently, Martin *et al.* described a technique of DNA extraction and identification of L2 serovar from biopsy specimens (8). They confirmed the presence of LGV DNA in one rectal biopsy specimen and one lymph node from two patients whose diagnoses had been uncertain but in whom LGV had been suspected (8). We approached the Sexually Transmitted Bacteria Reference Laboratory (STBRL) to attempt isolation of CT DNA from the stored rectal biopsy specimens of our 12 patients to help confirm our four suspected LGV diagnoses and explore further the validity of this technique. Rectal tissue was excised from 20 histology specimen blocks (sourced from 12 patients) and excess paraffin removed by heating at 56°C. The tissues were then digested using proteinase K and the DNA subsequently extracted using the DNeasy Blood & Tissue Kit (Qiagen). The presence of CT DNA (all serovars) was confirmed using a plasmid-based real-time polymerase chain reaction (PCR) assay (9). Any

specimen confirmed as CT-positive was then examined using an LGV-specific realtime PCR assay (10).

RESULTS

Median age of cases was 41.5 yrs (range 27-55). Two of the 12 had newly-diagnosed hepatitis C infection and four men had concurrent rectal GC. Median time to LGV diagnosis from time of initial rectal biopsy was 6 months (range 0-23).

Clinical symptoms and endoscopic findings

All 12 men described altered bowel habit and 11 described mucous discharge per rectum; further details regarding the presenting symptoms are listed in table 1. All underwent lower gastrointestinal endoscopy (either colonoscopy or flexible sigmoidoscopy). The findings at endoscopy are also listed in table 1: these varied from normal mucosal appearances to a moderately active proctitis (figure 1). In one case there was mucosal inflammation extending to the distal sigmoid colon.

Treatment

All cases were seen in either gastroenterology or colorectal surgical outpatient clinics where initial diagnoses of IBD proctitis or "non specific" proctitis were made. Table 1 summarises the different treatments that patients were initially and subsequently prescribed. Three patients were initially treated with metronidazole and 5 patients were commenced on oral 5-aminosalicylates and hydrocortisone enemas prior to

diagnosis of LGV proctitis, all with no effect on symptoms. Two patients were given reducing courses of oral prednisolone, leading to a transient partial improvement in symptoms. Longer times to LGV diagnosis were associated with increased numbers and types of treatments prescribed. Once their LGV diagnoses were made all patients were treated with doxycycline 100mg bd for three weeks with prompt and complete resolution of symptoms in all cases and at 12-month follow-up only one patient was having recurrent symptoms and was undergoing further investigations for possible irritable bowel syndrome.

Histology

Cryptitis and crypt abscesses were common histological findings but distortion of the crypt architecture was not prominent, more consistent with an infective aetiology (11). Granulomas were seen in biopsies from 5 of the 12 cases. Table 2 summarises the histological features and the diagnostic timeline for each case. Where biopsies from the same patient were obtained on more than one occasion for the same symptomatic episode the histological findings have been combined (in most cases the biopsy findings were identical). Representative images demonstrating some of the histological features are presented (figure 2).

Confirmation of LGV cases

Eight of the 12 men had the diagnosis of LGV confirmed by detection of CT DNA from rectal swab specimens, confirmed to be L-serovar DNA by the STBRL. In the remaining cases, LGV was suspected on the basis of clinical proctitis in the presence of a positive CT DNA test from a rectal swab specimen (not forwarded for

 LGV typing) and/or a high titre anti-Chlamydial group antibody response detected with the use of either Chlamydia Group antibody serology or a Whole Immunofluorescence (WIF) test that employs LGV L2 strain as the source antigen (12).

In total, 17 biopsy specimens from the 12 patients were submitted to STBRL.

Evidence of LGV-specific CT DNA was found in 7 of 17 biopsy specimens from 5

different patients, 3 of whom had suspected but unconfirmed diagnoses of LGV. Of

the remaining 10 specimens, 7 tested negative for CT plasmid DNA and hence did

not undergo further testing for LGV-specific DNA. Two specimens gave equivocal

concentrations and therefore testing for LGV was not performed. One specimen was

results on initial CT plasmid DNA testing, which is consistent with low CT DNA

repeatedly inhibitory thus further molecular analysis was not possible.

DISCUSSION

The clinical, endoscopic and histological findings in LGV can be indistinguishable from those of IBD. The potential for misdiagnosis of LGV and subsequent inappropriate management has been well described (13-17). At our centre between 2002 and 2006, twelve HIV-positive men underwent extensive investigation for symptoms of proctitis, including lower gastrointestinal endoscopy and biopsy. Some patients were treated for IBD, with repeated courses of metronidazole, 5-aminosalicylates and topical and oral steroids to which they failed to respond either clinically or histologically.

Rectal biopsy specimens from these 12 patients with LGV proctitis demonstrated features of acute and chronic inflammation with less distortion of crypt architecture than is usually seen in IBD, though there were no other distinguishing features or patterns. Granulomas and ulceration were present in 5 of 12 and 7 of 12 biopsy samples, respectively. These findings serve as a reminder that histologically LGV can be confused with IBD as well as other infective and non-infective causes of proctitis.

De la Monte *et al.* commented on the distribution of lesions in the colons of 28 subjects at autopsy in whom LGV had been diagnosed previously (18). They observed that the rectum was uniformly involved and colonic lesions in LGV were likely to affect the distal left side whereas Crohn's colitis can have a more varied colonic distribution. Other attempts have been made to distinguish between LGV and Crohn's or ulcerative colitis but often a considerable degree of overlap between the two is found histologically (11,18,19) and thus far, no pathognomonic features of LGV have been described. Recognition of the disease has relied on clinical suspicion in a patient with risk factors, though formal molecular or microbiological identification of the causative organism remains the only definitive diagnostic technique. Although not currently licensed for use at extra-genital sites (throat and rectum), commercially available nucleic acid amplification tests (NAATs) have shown high sensitivity and specificity for the detection of CT in the rectum and are now used routinely in most GUM clinics (20).

We were able to confirm LGV in three of our patients with suspected diagnoses by extracting DNA from rectal biopsy specimens and identifying the presence of LGV-specific CT DNA. This technique may provide a useful way of confirming LGV retrospectively in cases where isolation of the organism prior to treatment was not performed yet the diagnosis was suspected based on sexual history, clinical presentation and/or histological findings. In one case of LGV (case 4), the diagnosis was based on a typical clinical presentation and a high titre of anti-Chlamydial antibody, though treatment was commenced prior to the attempted detection of CT through molecular testing. Although serology cannot provide a definitive diagnosis of LGV, in our experience, a high WIF titre of 1:4000 has only been observed in confirmed cases of LGV infection.

Notably, the majority of LGV diagnoses made at our centre were in HIV-positive individuals, many with concomitant STIs and hepatitis C and some testing positive for HIV at the same time, reflecting a pattern seen across Western Europe within the present LGV epidemic. The exact route of transmission and mechanism of spread of LGV remain unclear; however MSM with untreated LGV remain at risk of infecting others or being infected with HIV and hepatitis C due to the inflammation and ulceration associated with the condition. Targeted interventions in this group are warranted.

After reaching a peak in mid-2005, symptomatic cases of LGV proctitis and occasional ulcer adenopathy disease continue to be diagnosed in the UK. Once established among MSM populations, infections such as gonorrhoea and syphilis

have remained endemic in recent years and it is likely that LGV will follow a similar course. From Table 2 it is evident that, at least in our GUM/HIV clinics, diagnosis and treatment of affected individuals has become timelier since the first cases were seen. Nevertheless, affected individuals may not consider their symptoms to be sexuallyrelated and thus may present to gastroenterology and colorectal surgery clinics, many via general practice or emergency department referrals. During the last twelve months we have encountered patients in our GUM clinic with sexually transmitted proctitis who have reported prior referral to gastrointestinal specialists via general practice. Relevant clinicians should remain alert to the possibility of LGV as a cause of proctitis, particularly in the presence of MSM sexual risk behaviour and HIVpositivity. Where NAAT-based testing is available through local laboratories, we recommend that all relevant clinicians send a rectal swab specimen for CT testing if suspicion of LGV is high and refer to GUM clinic for onward management, full STI screening and HIV testing. Histopathologists should also remember to include LGV in the differential diagnosis when reporting on inflammatory colorectal biopsy specimens, as MSM risk behaviour might not be provided in the clinical history accompanying the specimen.

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Figure 1: Endoscopic images from one patient with LGV proctitis demonstrates mild patchy colitis, with inflamed mucosa.

Figure 2: a and b) Rectal biopsies from Cases 7 and 1 showing multinucleate giant cells seen a) within a ruptured crypt abscess and b) between crypts – these were rarer findings. c) Rectal biopsy from Case number 6 showing surface ulceration with underlying densely inflamed granulation tissue. The large bowel mucosa shows crypt hyperplasia and the lamina propria is fibrotic and contains some smooth muscle fibres. There is acute and chronic inflammation with crypt abscesses and focal granuloma formation.

Symptoms										Treatment (Tx)			
Case No	Age & CD4 count (cells/mm³) at diagnosis	Date of Onset of symptoms	Per rectal bleeding	Mucous	Altered bowel habit	Tenesmus	Anal pain	Endoscopic findings	Tx initiated	Tx response	Tx change	Tx response	
1	41yrs, 856	Sep 2002			•			Moderate proctitis -circumferentially inflamed granular mucosa	MTZ, Ciprofloxacin	Nil	ASA, HC enema	Nil	
2	41 yrs, 432	Jan 2003			•	•	-	Moderate proctitis -patchy mucosal inflammation & small ulcers	ASA, HC enema	Nil	Oral pred.	Partial	
3	42 yrs, 335	Dec 2002				•		Mild proctitis patchy mucosal inflammation	MTZ x3 courses	Nil	ASA, HC enema	Nil	
4	45 yrs, 849	Aug 2003	•	•	•		•	Moderate proctitis inflamed granular mucosa and multiple ulcers	Acyclovir and Ciprofloxacin	Nil	Doxy.	Full	
5	27 yrs, 156	Mar 2004			•		•	Moderate proctitis -inflamed, granular and ulcerated mucosa	Erythromycin & fluconazole	Nil	MTZ, Doxy	Full	
6	55 yrs, 651	Aug 2004	•	•	•			Normal endoscopic findings	Doxy.	Full	- ,		
7	54 yrs, 487	Apr 2004		-	•	•		Moderate proctitis inflamed, granular and ulcerated mucosa	ASA, HC enema	Nil			
8	34 yrs, 572	Dec 2004	•	•	•			Moderate proctitis -patchy mucosal inflammation, small ulcers in rectum and distal sigmoid colon	Oral pred.	Partial			
9	42 yrs, 625	Jun 2005		•	•			Mild proctitis -inflamed, granular mucosa	Doxycycline	Full			
10	41 yrs, 625	Nov 2005	•	•	•		•	Normal endoscopic features	MTZ x3 courses	Nil	ASA, HC enema	Nil	
11	42 yrs, 600	Oct 2005	-	•	•			Mild proctitis -inflamed mucosa	Antibiotics x 2 courses	Nil			
12	39 yrs, 497	Aug 2006		•	•		•	Not recorded	Penicillin	Nil			

Table 1. Patient demographics, clinic symptoms, endoscopic findings and therapy commenced. MTZ = metronidazole, ASA = 5-aminosalicylate, HC = hydrocortisone, Pred = prednisolone.

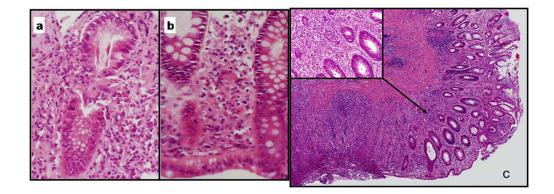
				Histo	logical fe	atures			Diagnosis			
Case No.	Time between biopsy & LGV diagnosis (months)	Mucosal Ulceration		Crypt abscess		Granuloma	Plasma Cell infiltrate	Giant Cells present	Initial Histological Diagnoses/ Suggestions	Mode of initial LGV diagnosis	LGV DNA results from biopsy(date of biopsy)	
1	23		•		■ mild			•	Uncertain aetiology Possible IBD	Rectal swab LGV Positive	LGV Positive (Sept 04) Negative (Dec 2002 & June 2003)	
2	22		-			•			?IBD ?infective	Rectal swab LGV Positive	LGV Positive (May 03)	
3	23				mild				?Due to prolapse ?IBD	Rectal swab LGV Positive	Negative	
4	12		•				•		Uncertain aetiology	Clinical & Serology WIF titre = 1:4000	Negative (Sept 2004) Inhibitory (Dec 2003)	
5	12					-			?Infective ?Crohn's	Rectal swab LGV Positive	Positive	
6	3	•				•			Exclude LGV	Clinical Rectal swab CT detected, not sent for LGV testing	LGV Positive	
7	9			•	■ minimal			-	Favours infective aetiology	Rectal swab LGV Positive	Negative (Sept 2004 and Jan 2005)	
8	2				■ mild		_		?IBD. Exclude CT/LGV	Clinical & Serology CFT titre = 1:512 WIF titre = 1:4000	LGV Positive (Mar 2005) Negative (Jun 2005)	
9	4								?Crohn's	Clinical & Serology CFT titre = 1:1280	LGV Positive (Nov 2005) Aug 2005 not tested	
10	0								Diagnosed Crohn's 1999, thought to be recurrence	Rectal swab LGV Positive	Negative (Nov 2005)	
11	0	•			■ mild	•			?IBD. Exclude LGV Concurrent Anal SCC present	Rectal swab LGV Positive	Negative (Nov 2005) Dec 2005 not tested	
12	6								?Early ulcerative colitis	Rectal swab LGV Positive	Negative	

Table 2. Summary of histological features of rectal biopsies and diagnostic timeline from 12 HIV positive men subsequently diagnosed with LGV proctitis





70x74mm (300 x 300 DPI)



160x60mm (300 x 300 DPI)