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Technical Report

Real time PCR based determination of gene copy numbers in *Pichia pastoris*

Sandra Abad [1,*], Kerstin Kitz [1,*], Astrid Hörmann [2], Ulrike Schreiner[1], Franz S. Hartner [1], Anton Glieder [1,2 §].

[1]Applied Biocatalysis Research Centre, Petersgasse 14, 8010 Graz, Austria

[2]Institute of Molecular Biotechnology, Graz University of Technology, Petersgasse 14, 8010 Graz, Austria

[*]equally contributing authors

[§]Corresponding author

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Correspondence: Dr. Anton Glieder

Petersgasse 14/II, 8010 Graz, Austria

email: anton.glieder@a-b.at

tel: +43 316 8734074

fax: +43 316 8739302

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ABSTRACT

Pichia pastoris is a prefered host for heterologous protein production. Expression cassettes are usually integrated into the genome of this methylotrophic yeast. This manuscript describes a method for fast and reliable gene copy number determinations for *P. pastoris* expression strains. We believe that gene copy number determinations are important for all researchers working with *P. pastoris* and also many other research groups using similar gene integration techniques for the transformation of other yeasts. The described method uses Real Time PCR to quantify the integrated expression cassettes. Similar methods were employed previously for other host systems such as animal and plant cells but no such method comparing different detection methods and describing details for yeast analysis by quantitative PCR is known to us especially not for methylotrophic yeasts such as *P. pastoris*. Neglecting gene copy numbers can easily lead to false interpretations of experimental results from codon optimization or promoter studies and coexpression of helper proteins as demonstrated in an application example, which is also described in this manuscript.

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INTRODUCTION

The methylotrophic yeast *Pichia pastoris* has emerged to a frequently used protein expression host. High product titers have been obtained with the standard expression system [1, 2] employing the methanol inducible *AOX1* promoter (P_{AOX1}). Nevertheless, recent studies reported even higher expression rates when the target gene was under the control of deletion variants of the *AOX1* promoter [3] or in some cases also with the constitutive glyceraldehyde-3-phosphate dehydrogenase promoter (P_{GAP}) [4]. Depending on the target gene, high copy number expression strains do not necessarily lead to optimal expression rates. For example, for some secreted proteins rising numbers of expression cassettes resulted in a decreased expression due to other bottlenecks than transcript levels [5, 16]. Other approaches such as co-expression of helper proteins e.g. chaperones or proteins which initiate the unfolded protein response have shown reasonable expression improvements [6-9]

Although many studies describe improved expression due to different promoters variants, codon optimization or co-expression of helper proteins, hardly any information of copy numbers of the expression cassettes is given. Consequently, false interpretation of experimental results is possible, leaving the question whether improvements either correlated with a certain promoter, co-expressed chaperone or an accompanying copy number effect. Inan *et al.* pointed out the importance of the copy number determination for *P. pastoris* expression strains, reporting the correlation between increased protein secretion and the copy number of the co-expressed chaperone protein disulfide isomerase from *P. pastoris*, *Pp*PDI. Copy numbers were determined by southern blot analysis, which was so far the method of choice for this question. However, it is a time-consuming and laborious method, and requires large amounts of genomic DNA. In addition, restriction site loss during

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integration is possible, complicating the molecular analysis. To overcome these drawbacks and also in order to deal with increasing numbers of expression strains from high throughput experiments, an additional reliable and faster method for copy number determination of integrated expression cassettes was needed.

Quantitative PCR emerged as an important and widely used analytical tool (reviewed in [7]). But, there are only few studies reporting on real time PCR based copy number determination in human-, animal- and plants cells [10, 11]. No detailed method for yeast could be found, probably also since for many yeasts plasmid systems are still quite common. In general, assay design, template preparation and analytical methods are essential for accurate quantitative gene amplification [12, 13] and were especially adapted for *P. pastoris* in this study as a fast and simple alternative to southern blotting.

Two settled detection methods, namely TaqMan and SYBR Green were compared, as well as approaches for absolute and relative quantification. For a simple visualization of our results and for a correlation between copy numbers and expression, green fluorescent protein (GFP) fusion with a Zeocin resistance marker has been chosen as a simple intracellular reporter system [14]. Furthermore the importance of expression cassettes quantitation in *P. pastoris* was emphasized in an industrially relevant example. Real time PCR was already used for copy number determination in *P. pastoris* in our group and independently also by others before [3, 15, 16]. A demonstration of the importance of copy number determination as a routinely performed experiment for molecular strain characterization and a first detailed protocol especially adapted for *P. pastoris* and including the comparison of two different detection methods, were the main goals of this work.

MATERIALS AND METHODS

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Chemicals and materials

Sterile water was purchased from Fresenius Kabi Austria (Graz, Austria). Unless otherwise stated, all chemicals were purchased from Carl Roth GmbH (Karlsruhe, Germany), Becton, Dickinson and Company (Franklin Lakes, NJ, USA) and Sigma-Aldrich (St Louis, MO, USA). DNA modifying enzymes were supplied by Fermentas GmbH (Burlington, Ontario, Canada). All PCRs were performed with Phusion HF according to the producer's manual (Finnzymes Oy, Espoo, Finland).

GFP reporter expression cassette

P_{AOX1} and the *AOX1* terminator were amplified from vector pPICZ-B (Invitrogen, Carlsbad, USA), and cycle-3-GFP-Zeocin from pTracer™-CMV2 [14, 17]. All PCR products were purified by agarose gel electrophoresis prior to overlap extension PCR [18].

The obtained PCR product displayed 2.4 kb and was purified by agarose gel electrophoresis and gel extraction using Wizard® SV Gel and PCR Clean-Up system (Promega, Madison, WI, USA). pAOX was constructed by digesting the P_{AOX1}-GFP-Zeo-*AOX*1TT fragment with *Kpn*I and *Not*I and ligation with *Kpn*I/*Not*I-digested pBlueScript® SK^- vector (Stratagene, La Jolla, CA, USA).

The *P. pastoris* P_{AOX1}d6* promoter variant was excised from vector pPICZ-GFP-d6* [3] employing *Bgl*II/*Eco*RI restriction sites into the vector pAOX thereby generating vector pAOXd6*. The correct sequence was verified by sequencing (AGOWA Genomics, Berlin, Germany).

P. pastoris transformation

*Bgl*II and *Not*I linearized pAOX or pAOXΔ6* was transformed into electro-competent *P. pastoris* cells. Electro-transformation was performed as described in [19]. After a regeneration phase, aliquots were spread on MSM-Zeo agar plates (1.5% Bacto™

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Agar, 1.34% Difco™ Yeast Nitrogen Base w/o Amino Acids, $4 \cdot 10^{-5}$% d-Biotin, 2% Sorbitol, 1% methanol, 100 µg/mL Zeocin™) and incubated for 3-4 days at 30°C.

Pichia pastoris microscale cultivation and GFP reporter expression

P. pastoris cultivation for GFP expressing strains was performed as described in [20]. GFP intensity was detected with SPECTRA MAX Gemini XS plate reader (Molecular Devices Corp., Sunnyvale, CA, USA) as described in [3].

Generation of a _Pichia pastoris_ genomic DNA-library

Genomic DNA (gDNA) from *P. pastoris* X-33 was isolated employing the Easy-DNA™-Kit (Invitrogen). 10µg from the DNA was partially digested with *Sau*3AI. Gel purified fragments (6-9 kb) were ligated into *Xho*I digested pGAPZ plasmid (Invitrogen) which was partially filled with Klenow fragment. Ligation was transformed into *E.coli* Top 10 F' (Invitrogen). Transformation was plated into LB-Zeocin (25 mg/mL). Transformants were pooled and cultivated for plasmid isolation with Wizard® Plus SV (Promega) according to manufacturer's protocol. The library contained 10,500 clones and the average insert size of 14 independent transformants was estimated to 4.5 kb. The probability to find any given unique sequence in this library is 99%.

_Ca_lB reporter strain construction

Zeocin resistance cassette from pPICZ-B (Invitrogen) was replaced with the KanMX6 cassette from PFA6a-kanMX6 [21], which confers resistance against Kanamycin in *E. coli* and Geneticin (G418) in yeasts. The resulting plasmid was named pPICK. Codon optimized *Candida antartica* lipase B (*Ca*lB, GenBank: ACIO6118) was assembled via oePCR. The resulting gene was *Eco*RI/*Not*I digested, gel purified and cloned into an equally digested pPICK-plasmid. *E.coli* Top 10 F' (Invitrogen) was transformed with this construct and plated onto LB-Kanamycin (50 mg/mL). The construct was analyzed by sequencing, and linearized with *Bgl*II for *P. pastoris*

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KM71H (Invitrogen) transformation as described before. After transformation the cell suspensions were plated on YPD-G418-plates (300 mg/L) and incubated at 28℃ for 2 days. Transformants were cultivated in deep-well plates as described before, and screened for *Ca*lB activity.

Transformation of X-33 gDNA library into *Ca*lB reporter strain

The *P. pastoris Ca*lB-H5 (multicopy *Ca*lB strain) was transformed with 3 µg of *Bgl*II linearized genomic DNA (gDNA) library. Cells were plated in YPD-Zeocin-plates (50 mg/L) and incubated at 28℃ for 2 days. Using a Genetix QPixII robot (Genetix Limited, Hampshire, UK) the transformants were picked into 384-plates filled with 50 µL BMD1%. Incubation was carried out at 28℃ for 5 days. Glycerol-stocks were prepared by addition of 30 µL of glycerol/BMD1% to a final concentration of 15% glycerol. The 384-plates were covered with foil seal (Silverseal Aluminium-tape, Greiner, Frickenhausen, Germany) and stored at -80℃. Transformants were replicated into deep-well plates and cultivated as described before with one minimal modification, cell harvest after 60 h of methanol induction.

***Ca*lB activity assay**

P. pastoris strains were cultivated in deep-well plates as described before. Esterase activity determination of *Ca*lB was performed according to [22] with minimal modifications. Briefly, 20 µL of the supernatant or a dilution of it was pipetted into a micro-titer plate well. The *Ca*lB esterase activity was measured following color development (due to p-nitro phenolate formation) at 405 nm for 5min at RT after addition of 180 µL of the assay solution. The assay solution consisted of 300 mM Tris/HCl pH7, 1% ethanol and 4 mM of *p*-nitrophenyl butyrate. One unit was defined as the formation of 1 µmol *para* nitrophenol per minute (ε= 17,700 $M^{-1}cm^{-1}$)

Recovery of the library-insert from the *Ca*lB-clones

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gDNA from selected strains was isolated employing the Easy-DNA™-Kit from Invitrogen. 500 ng gDNA were digested with *Bgl*II, after purification with Wizard® SV Gel and PCR Clean-Up system (Promega), self-ligation with T4-ligase was incubated at 16℃ over night. 2 µL of the ligation mixture were transformed into chemical competent One Shot® Top10 (Invitrogen). Resulting transformants were sent for sequencing (AGOWA genomics).

Isolation of genomic DNA for RT-PCR

The isolation procedure was performed as described by Hoffman and Winston [23]. The DNA concentration was measured at 260 nm. DNA quality was checked by gel electrophoresis and photometrically (SPECTRAmax PLUS plate reader, Molecular Devices Corp.). Samples displaying 260 nm/280 nm ratios higher than 1.8 were used for further analysis. Alternatively gDNA was isolated employing the E.N.Z.A.® Yeast DNA Kit (Omega Bio-Tek, Norcross, Georgia, USA) and the Easy-DNA™-Kit (Invitrogen) as described by the suppliers protocols. gDNA isolation was also performed with Y-PER Yeast Protein Extraction Reagent (Thermo Fisher Scientific Inc.,Rockford, Illinois, USA). Briefly, the cells of a 10-15 mL an overnight culture were lysed with 300 µL Y-PER (30 min, room temperature, mixed on an Eppendorf shaker with 600 rpm). Lysed cells were spun down at 13,000 rpm, 10 min at room temperature. The supernatant then was subjected to a chloroform/phenol/isopropanol extraction followed by ethanol precipitation. Finally, the DNA pellet was resuspended with 50 µL H_2O, dest.

Primer sets for real time PCR

The primer design was performed using *Primer Express v2.0* software (Applied Biosystems, Foster City, CA, USA). The primers had similar melting temperatures (T_m) (58 – 60℃) and the T_m of the probe was 10℃ higher. The amplicon size settings were defined between 80 and 150 bp. The nucleotide sequences are shown

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in Table 1. Primers were purchased from Invitrogen and probes were purchased from MWG-Biotech (Ebersberg, Germany).

The real time PCR assays were optimized with respect to the primer concentration by titration experiments. Thereby, combinations of different concentrations (100 nM – 300 nM) of forward and reverse primers for *GFP* and *ARG4* genes, respectively, were tested, using the same template amount per real time PCR reaction (see below). The primer combinations with lowest Ct values and no occurrence of by-products in the non-template control (NTC) reactions were considered as optimal.

Real time PCR using SYBR Green and TaqMan probes

Real-time PCR amplification was performed using an ABI 7300 instrument with Sequence Detection Software SDS v1.2 (Applied Biosystems). Real-time PCR mixtures were prepared using the TaqMan® Universal PCR Master Mix or Power SYBR® Green Master Mix (both Applied Biosystems), respectively.

For TaqMan probe based determinations, a single reaction contained 1x TaqMan® Universal PCR Master Mix, 200 nM of each primer and 125 nM of the corresponding FAM/TAMRA labeled probe. For SYBR Green examinations, 200 nM of each primer were added to the 1x Power SYBR® Green Master Mix. In both cases, reactions were performed in triplicate for the method establishment and in duplicate for faster routine strain analysis. 2 ng of genomic DNA were added in a reaction volume of 20 µL.

For each unknown strain, reactions with GFP and ARG4 primer sets were performed in separate tubes. Standard curves and NTCs for both genes were recorded in every plate. The reactions were done in MicroAmp® Optical 96-Well Reaction Plates sealed with MicroAmp® Optical Adhesive Covers (Applied Biosystems).

The thermal profile initiates with a 10 min step at 95°C followed by 40 cycles of 15 sec at 95°C and 1 min at 60°C. The data collection of the fluorescence signal was performed at the end of the elongation step. These settings were the same for both

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detection strategies. In case of the SYBR Green samples, the amplification period was followed by a melting curve analysis with a temperature gradient of 0.1 °C/s from 70 to 95 °C to exclude amplification of unspecific products.

Establishment of standard curves for copy number determination

The GFP-Zeo strain X-33 pAOX D2 was first identified as single copy strain via two colony PCRs. The first primer pair served as control and binds in the coding region of the GFP gene, the second pair was set on the origin of replication region (forward) of the integrated linearized plasmid and in the P_{AOX1} (reverse). One or multiple tandem integration of the expression cassette delivers a defined band with both primer combinations, whereas the single copy integration only displays a band with the control primer setting. The strain was confirmed by previously measured intracellular fluorescence level as a single copy strain due to its low value in the fluorescent/transformant landscape and selected as calibrator for the establishment of the *GFP* and *ARG4* standard curves. In addition the integration of further expression cassettes in other orientations or in other loci was excluded by southern blot analysis (data not shown). The standard curves cover a copy quantity range from 1.2×10^4 to 3.1×10^6 and 1.1×10^5 to 3.1×10^6 copies per reaction for SYBR Green and TaqMan detection, respectively.

For calculations with the Sequence Detection Software SDS v1.2 (Applied Biosystems), the copy quantity equation as described in [24] was used, considering a *P. pastoris* genome size of 9.7 Mbp [25] since at the time when this work was performed the genome sequence was not public available. Resulting in 94,000 copies of the genome present in 1 ng of haploid *P. pastoris* genomic DNA. Meanwhile the published sequence (GenBank: FN392319.1, FN392320.1, FN392321.1, FN392322.1, FN392323.1, FN392324.1 and FN392325.1) which still contains a few gaps shows a similar size of about 9.4 Mbp [26].

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The mean Ct values were plotted against the $\log^{10}$ of their initial template copy quantity and standard curves were generated by a linear regression of the plotted points. The PCR efficiency was calculated from the slope of each standard curve by the following equation [24]:

$$E = 10^{-1/slope} - 1$$

Samples displaying Ct values lower than 26 and standard deviations lower than 0.3 were used for copy number determinations according to absolute and relative calculations.

Absolute and relative quantification

For correct determination of the starting copy quantity regardless of precise amounts and qualities of input genomic DNAs, the reference gene *ARG4* was also quantified in parallel. The normalized copy number calculated by absolute quantification is given by the following equation [11]:

$$Copy\ number_{TARGET\ GENE} = \frac{Copy\ quantity_{TARGET\ GENE}}{Copy\ quantity_{ARG4GENE}}$$

Relative quantification of the copy number was performed according to the $2^{-\Delta\Delta Ct}$ method [27], where ΔΔCt = ΔCt of target – ΔCt of calibrator; ΔCt = Ct of target or calibrator – Ct of reference (*ARG4*); E = PCR efficiency.

$$Copy\ number_{TARGET\ GENE} = (1+E)^{-\Delta\Delta Ct}$$

A strain with known copy number (in our case *P. pastoris* X-33 pAOX D2), ideally with one copy of the target gene served as a calibrator strain.

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RESULTS AND DISCUSSION

The quality of the DNA sample is one key step for successful quantification via RT-PCR [13]. Therefore the search of a gDNA preparation method which delivers suitable gDNA with a fast isolation protocol was an important step. *P. pastoris* standard fast isolation protocols using commercially available chemicals delivered high yields of gDNA, but also caused DNA degradation (Figure 1). Excellent results were obtained using glass beads and phenol/chloroform extraction as described by Hoffmann and Winston [23].

In total, six *P. pastoris* strains carrying the GFP-Zeocin expression construct were chosen as models for the establishment of quantitative RT-PCR in *P. pastoris*. gDNA of these *P. pastoris* strains, two under the control of the wild type P_{AOX1} (named X-33 pAOX D2 and E2) and four under the control of the deletion variant P_{AOX1}d6* (named X-33 pAOXd6* A5, D10, F9 and F2) were analyzed (Table 1). Assuming that GFP expression and therefore the measured fluorescence intensity increases with the number of integrated copies we expected that *P. pastoris* X-33 pAOX D2 was a single copy strain. This strain was used for the generation of the standard curve required for the real time PCR experiments.

gDNA standard curves were generated in triplicate determinations and showed to be linear over a range from 1.2×10^4 to 3.1×10^6 copies (based on an estimated 9.7 Mbp full genome size, see material and methods) in the case of SYBR Green detection. A tighter linear range was detected with the TaqMan method in our hands. The standard curves showed high efficiency (>0.92) and were approximately equal for the target and the reference gene, which is a prerequisite for copy number calculations using the relative $2^{-\Delta\Delta Ct}$ method [27]. As at the time when we started these studies no reports for real time PCR performed in *P. pastoris* were available,

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we decided to use the *ARG4* gene as an endogenous control [28] as this is a single copy gene of the haploid host. In this comparative technique, the amount of the target gene *GFP* is normalized by using the reference gene *ARG4* and set relative to the calibrator, which was in our case the single copy X-33 pAOX D2 strain (Figure 2). Hence, the *GFP/ARG4* ratio of a sample was normalized by the *GFP/ARG4* ratio of the calibrator.

Two detection strategies were followed: 1) sequence-specific TaqMan probes and 2) SYBR Green. It was tested if the use of SYBR Green leads to similar results as the more expensive TaqMan probe based detection system, which is described to be the more sensitive detection method [29] (Table 1). In the range of one to five copies of the expression cassette the results were similar, no matter if SYBR Green or TaqMan probe was used. Important to mention are the higher Ct values obtained with the TaqMan detection method which is in line with the already mentioned narrow linear range compared with the SYBR Green method. In any case absolute or relative quantification delivered consistent results, although a modest deviation for the multi copy strain was detected with the TaqMan method.

The quantitation of integrated expression cassettes showed that pAOX D2 and pAOXd6* A5, although displaying different GFP intensities, are single copy transformants (or have identically integrated expression cassettes). These results confirm that the deletion d6* leads to a decrease of promoter activity to about 60% [3] under the described cultivation conditions.

Sustaining the aim of a universal tool for copy number determination, which fits for numbers of different expression constructs, good results have been obtained by using common sequences (e.g. P_{AOX1}), which are present in the integrated expression cassette and in the *P. pastoris* haploid genome. This setting resolves the identification of one single copy strain (calibrator), since the wild type strain already

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contains one copy of the sequence, and designed primers and probes can be used as universal tools.

To show the importance of this method in a laboratory routine, a real case scenario has been followed. The already mentioned benefits of helper protein co-expression have been focused in a so called non-rational approach. Therefore, a reporter strain was transformed with a *P. pastoris* genomic DNA library cloned under the control of the P_{GAP}. The *P. pastoris* reporter strain harbored 7-8 copies of a *Cal*B expression cassette under the control of the P_{AOX1} (named *P. pastoris* pPICK-*Cal*B H5), and displayed rather low levels of active enzyme in the culture supernatant compared to single copy strains. Previous experiments showed that certain bottlenecks during the secretion pathway led to a limited growth of the *P. pastoris Cal*B strains and low expression levels of the target gene, which could be at least in part be removed by co-expression of *Pp*PDI (S. Abad unpublished data). Following this outcome, we aimed at identifying additional putative helper proteins by a second transformation and individual co-overexpression of *P. pastoris* genomic DNA library clones. Therefore genomic *P. pastoris* DNA fragments had been ligated behind the GAP promoter of the pGAPZB expression plasmid. Putative new helper proteins from this library were expected to lead to an increased *Cal*B expression which can be determined by measuring the lipase activity in the culture supernatant of the reporter strain.

In total 3250 transformants were screened, which seemed to be an acceptable number for our initial purpose since four candidates (*Cal*B H5/2-L18, H5/9-H21, H5/10-H22, H5/10-F22) displayed higher *Cal*B expression levels as the reporter strain. These improved transformants were chosen for a re-screen and the expression improvement was confirmed for all candidates. Interestingly, none of the chosen co-expression strains delivered higher expression values than single copy

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*Cal*B strains, and no reproducibility of the results was possible after transformation of the recovered construct into the reporter strain. The copy number determination performed with the SYBR Green method using primers designed for P_{AOX1} sequence and the *P. pastoris* KM71H strain as calibrator explained the described results. The chosen "improved" transformants showed lower numbers of integrated *Cal*B expression cassettes. The transformants H5/2-L18 and H5/10-F22 harbored four and two copies, respectively while H5/9-H21 and H5/10-H22 displayed only one *Cal*B expression cassette (Table1).

CONCLUDING REMARKS

Quantitative PCR allows the determination of the number of integrated expression cassettes of *P. pastoris*. Strains harboring up to five copies were analyzed and delivered consistent results with two different detection methods. However, the narrow linear range of the TaqMan method should be taken into account for further experiments. This PCR based procedure provides a reliable but faster and simple alternative to standard southern blot analysis. It also allows correlations between integration events into the genome and the respective expression levels which is highly relevant for expression studies employing different promoters, redesigned genes or alternative signal sequences. Due to its simplicity it can be applied routinely..

Absolute and relative quantification showed to have no influence in the final outcome of the experiment, whereas the latter demands more precise parameters. The house-keeping gene *ARG4* showed to be a reliable endogenous control. However, caution should be taken when an Arg^- auxotroph strain is used as platform for expression experiments.

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As demonstrated by application of the described method multiple transformation steps can lead tto the potential loss of previously integrated expression cassettes, especially if the same locus is targeted for the genomic integration. Gene dosage control of previously integrated expression cassettes after every further transformation step is essential.

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CONFLICT OF INTEREST STATEMENT

The authors have declared no conflict of interest.

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FIGURE LEGENDS

Figure 1: Comparison of *P. pastoris* gDNA quality of diverse isolation methods.

G: GeneRuler 1kb (Fermentas), λ: λ-HindIII Standard (Fermentas), 1: 5 µL of gDNA isolated using Y-PER (Novagen) and chloroform/phenol extraction followed by ethanol precipitation, 2: 5 µL of gDNA isolated according to Easy DNA™-Kit (Invitrogen), 3: 5 µL of gDNA isolated using a E.Z.N.A.® Yeast DNA Kit, 4: 5 µL of gDNA isolated using glass beads and phenol/chloroform extraction as described by Hoffmann and Winston [23].

Figure 2: RT-PCR determination using SYBR Green and TaqMan methods. Target and reference gene amplification of *P. pastoris* GFP-Zeocin strains and standard curve. A: SYBR Green detection, B: TaqMan detection.

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Table 1: Copy number calculation according to absolute (Abs. Q) and relative (Rel. Q) quantification.

Pichia pastoris strain	SYBR Green[a)]		TaqMan[b)]		Resulting copy number	Measured Expression GFP: [RFU], *Cal*B: [mU/L]	Expected copy number[e)]
	Abs. Q	Rel. Q	Abs Q.	Rel Q.			
X-33 pAOX D2	1.0	1.0	0.8	1.0	1[c)]	987 ± 81	1
X-33 pAOX E2	1.7	1.8	1.6	2.0	2[c)]	2068 ± 219	2
X-33 pAOXΔ6* A5	1.1	1.1	1.0	1.2	1[c)]	422 ± 22	1
X-33 pAOXΔ6* D10	1.9	2.1	2.0	2.4	2[c)]	1063 ± 189	2
X-33 pAOXΔ6* F9	2.0	2.1	1.8	2.2	2[c)]	925 ± 58	2
X-33 pAOXΔ6* F2	4.6	4.8	4.0	5.0	4-5[c)]	1678 ± 437	4-5
KM71H pPICK-*Cal*B H5	8.9	8.6	-	-	7-8[d)]	0.033 ± 0.005	-
KM71H *Cal*B H5/2-L18	5.7	5.8	-	-	4-5[d)]	0.390 ± 0.051	-
KM71H *Cal*B H5/9-H21	2.2	2.3	-	-	1[d)]	0.853 ± 0.086	-
KM71H *Cal*B H5/10-H22	2.6	2.7	-	-	1-2[d)]	0.876 ± 0.054	-
KM71H *Cal*B H5/10-F22	3.1	3.1	-	-	2[d)]	0.955 ± 0.075	-

[a)] ARG(sense/antisense): 5'-TCCTCCGGTGGCAGTTCTT-3'/5'-TCCATTGACTCCCGTTTTGAG-3' and

GFP(sense/antisense): 5'-AAATTTAAGGGTAAGCTTTCCGTATG-3'/5'-ATGGTGATGTTAATGGGCACAA-3' or

P_{AOX1}(sense/antisense): 5´-GAAGCTGCCCTGTCTTAAACCTT-3´/5´-CAAAAGCTTGTCAATTGGAACCA-3´

[b)] GFP-probe: 5'-ATCACCTTCACCCTCTCCACTGACAGAAAA-3' , ARG-probe: 5'-TTCAACACTGGCTTCAAAGTCAAACGTTGAA-3'

[c)] based on absolute and relative quantifications results with SYBR Green and TaqMan methods.

[d)] based on results of absolute and relative quantifications with SYBR Green method. The endogenous P_{AOX1} sequence has been subtracted.

[e)] based on fluorescence measurement assuming a linear correlation between GFP intensity and the number of integrated copies.

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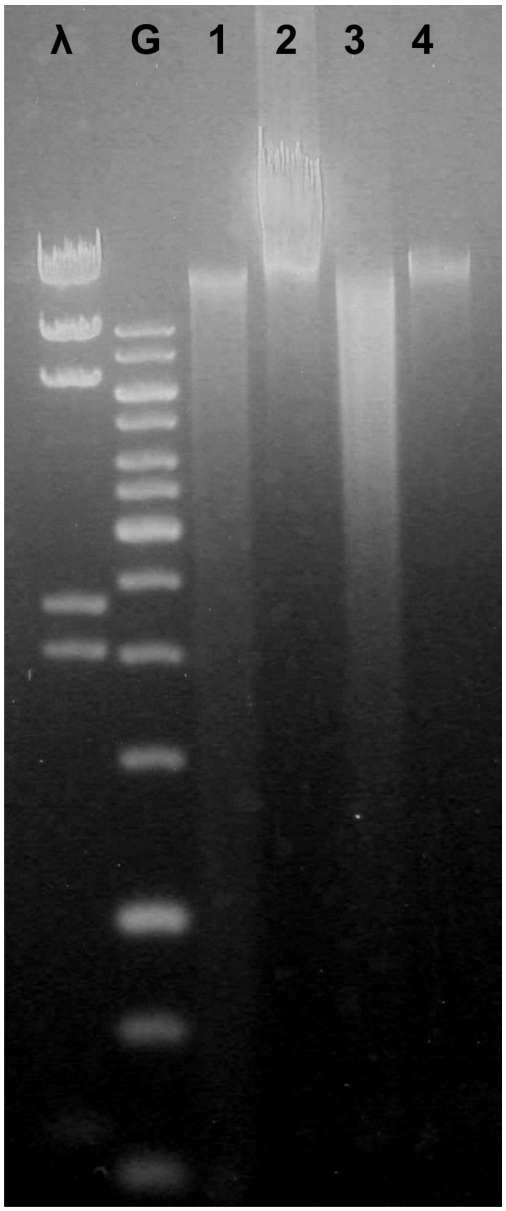

Figure 1: Comparison of P. pastoris gDNA quality of diverse isolation methods.
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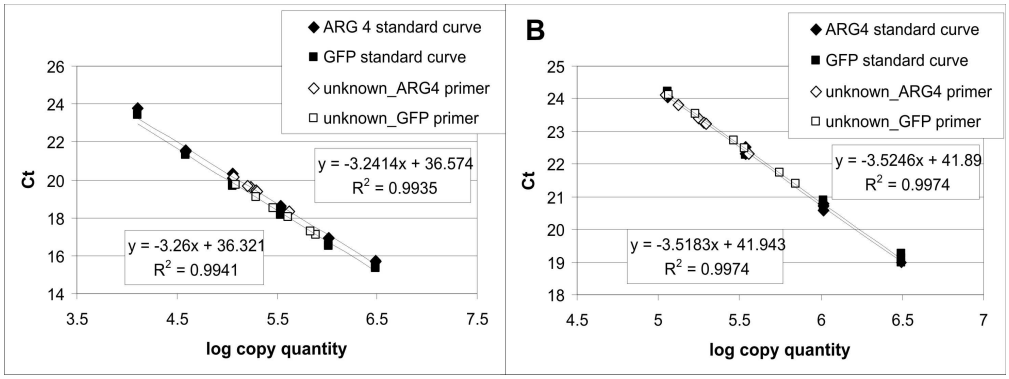

Figure 2: RT-PCR determination using SYBR Green and TaqMan methods. Target and reference gene amplification of P. pastoris GFP-Zeocin strains and standard curve. A: SYBR Green detection, B: TaqMan detection.
249x92mm (300 x 300 DPI)

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