

Procalcitonin is a reliable marker of severe systemic infection in neutropenic haematological patients with mucositis

Loredana Sarmati, Angela Beltrame, Luca Dori, Gaetano Maffongelli, Laura Cudillo, Gottardo de Angelis, Alessandra Picardi, Licia Ottaviani, Maria G Cefalo, Adriano Venditti, et al.

▶ To cite this version:

Loredana Sarmati, Angela Beltrame, Luca Dori, Gaetano Maffongelli, Laura Cudillo, et al.. Procalcitonin is a reliable marker of severe systemic infection in neutropenic haematological patients with mucositis. American Journal of Hematology, 2010, 10.1002/ajh.21685 . hal-00552309

HAL Id: hal-00552309 https://hal.science/hal-00552309

Submitted on 6 Jan 2011

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



American Journal of Hematology

Procalcitonin is a reliable marker of severe systemic infection in neutropenic haematological patients with mucositis

Journal:	American Journal of Hematology
Manuscript ID:	AJH-09-0833.R2
Wiley - Manuscript type:	Letters
Date Submitted by the Author:	09-Feb-2010
Complete List of Authors:	Sarmati, Loredana; Clinical Infectious Diseases, Tor Vergata University, Rome, Pubblic Health Beltrame, Angela; Clinical Infectious Diseases, Tor Vergata University, Rome, Pubblic Health Dori, Luca; Clinical Infectious Diseases, Tor Vergata University,, Pubblic Health Maffongelli, Gaetano; Clinical Infectious Diseases, Tor Vergata University, Rome, Pubblic Health Cudillo, Laura; Haematology Stem Cell Transplant Unit, Tor Vergata University De Angelis, Gottardo; Haematology Stem Cell Transplant Unit, Tor Vergata University Picardi, Alessandra; Haematology Stem Cell Transplant Unit, Tor Vergata University Ottaviani, Licia; Oncohaematologic Unit, Tor Vergata University Cefalo, Maria; Oncohaematologic Unit, Tor Vergata University Venditti, Adriano; Oncohaematologic Unit, Tor Vergata University Amadori, Sergio; Oncohaematologic Unit, Tor Vergata University Arcese, William; Haematology Stem Cell Transplant Unit, Tor Vergata University
Keywords:	Infections, Infections in imunocompromised hosts, Infectious disease



Procalcitonin is a reliable marker of severe systemic infection in neutropenic haematological

1	
2	
3 4	1
5	-
6 7	2
8	С
9	5
10	4
12	
13	5
14 15	
16	6
17	
18	7
20	
21	8
22	
23 24	9
25	
26	10
27	11
29	11
30 21	12
32	12
33	13
34 25	
35 36	14
37	
38	15
39 40	
41	16
42	
43 44	17
45	
46	18
47 48	4.0
49	19
50	20
51 52	20
53	21
54	
ວວ 56	
57	
58	
59 60	

2	patients with mucositis
3	
1	
5	Sarmati Loredana, Beltrame Angela, Dori Luca, Maffongelli Gaetano, Cudillo Laura°, De Angelis
5	Gottardo°, Picardi Alessandra°, Ottaviani Licia°°, Cefalo Maria Giovanna°°, Venditti Adriano°°,
7	Amadori Sergio°°, Arcese William°, Andreoni Massimo.
3	
9	Clinical Infectious Diseases, Tor Vergata University,
)	° Haematology Stem Cell Transplant Unit, Tor Vergata University,
L	°°Oncohaematologic Unit, Tor Vergata University,
2	Rome, Italy
3	Running head: procalcitonin and mucositis
1	
5	Corresponding author : Loredana Sarmati, MD
5	Clinical Infectious Diseases
7	Tor Vergata University
3	V. Montpellier 1
9	00133 Rome , Italy
)	Phone +390672596873
L	sarmati@med.uniroma2.it

Patients with neutropenia are exposed to a high risk for infections in which fever is often the unique symptom (1). Systemic infections remain the main cause of mortality in these patients therefore, the policy for infection management is to promptly administer empirical antibiotic therapy in order to avoid the increased risk of mortality related to the treatment delay (2). However, microbiological diagnostic tests are not sufficiently rapid, sensitive or specific to indentify the microbial causes of fever, and a considerable number of patients suffer febrile episodes over a prolonged period without a definite microbiological aetiology.

Procalcitonin (PCT) (3) has become increasingly popular as a novel marker of infection. Several studies have underscored its value in clinical conditions by identifying infectious processes (4, 5, 6). The use of this marker in haematological patients has provided controversial results and no agreement exists about the capacity of PTC to differentiate fever by other inflammatory processes such as mucositis and Graft versus Host diseases (GVHD) (7-11).

In order to evaluate the usefulness of PCT in diagnostic and therapeutic approaches to fever in haematological patients, we evaluated the values of PCT, C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) in the course of febrile neutropenia by assessing the influence of mucositis and GVDH on these three parameters.

The demographics and the clinical characteristics of the study population are shown in Table 1. One hundred and sixteen infectious events were observed, 50% of them occurring in patients with mucositis. Nine (30%) subjects out of 30 transplanted patients developed GVHD. FUO, pneumonia, sepsis and infection of CVC were the most common clinical features associated with fever.

In order to assess the power of the investigated inflammatory parameters in discriminating the clinical severity of infection episodes, the median values of PCT, CRP and ESR as measured during the major events were compared with those recorded during the minor events. During major infectious events, the PCT levels were significantly higher at any considered time-point (PCT1: 0.55 vs 0.13 ng/ml, p<0.001; PCT2: 1.04 vs 0.21 ng/ml, p<0.001; PCT3: 0.54 vs 0.16 ng/ml, p<0.001), CRP was significantly higher only at times one and two (CRP1: 112 vs 56 mg/dl, p=0.001; CRP2: 150 vs 104 mg/dl, p=0.02) and the ESR values were not significantly different at any time.

Figure 1 shows the results of the PCT and CRP determinations during febrile major and minor events in patients with and without mucositis. The analysis showed that the PCT values were significantly higher at all three time-points during major events compared to minor events, regardless of the presence of mucositis. On the contrary, the values of CRP were significantly higher at the time-point one in patients with major events and mucositis, and at time-point one and two in those with major events without mucositis, compared to patients with minor events.

The sensitivity and specificity of PCT and CRP to identify major infectious events in patients with mucositis was evaluated through ROC curves (Fig. 2). The area under the curve accounts for the statistical significance for PCT1 (68%; 95% CI: 54 to 82; p=0.016), PCT2 (68%; 95% CI: 53.9 to 81.7; p = 0.02) and PCT3 (65%; 95% CI: 51 to 80; p = 0.035). Otherwise, the area under the ROC

American Journal of Hematology

curve was statistical significant for CRP1 (68%, 95% CI: 54 to 81; p=0,019), but not for CRP2 (54%, 95% CI: 38.8 to 68.9; p = 0.6) and CRP3 (48%, 95% CI: 33 to 63; p = 0.8). Moreover, in patients with mucositis, PCT2 values of \leq 0.5 ng/ml and \leq 0.2 ng/ml discriminates major infectious events with sensitivities of 48% and 79% and specificities of 70% and 52%, respectively.

A bacterial isolate was obtained in 50/59 (85%) major infectious events and in 22/57 (38%)
minor events. Gram-negative isolates were significantly more frequent during major events
compared to minor events (44/59 vs. 4/ 57, p <0.001).

The PCT values, but not the CRP values, were significantly higher at all three time-points in patients with a Gram-negative isolate relative to patients with an isolate other than a Gramnegative (PCT1: 1.9 vs 0.2 ng/ml, p=0.001; PCT2: 3.5 vs 0.4 ng/ml, p<0.001; PCT3: 0.7 vs 0.4 ng/ml, p=0.005). Moreover, a PCT2 value of \geq 0.98 ng/ml was found to discriminate between the presence or absence of a gram-negative bacterial isolation during fever with a sensitivity of 80% and a specificity of 78%.

The study of PCT and CRP was conducted separately on the group of transplant patients with the occurrence of GVHD. To this end, only patients at risk of GVHD (allogeneic, haploidentical and umbilical cord blood transplants) have been considered. GVHD occurred in 12 out of 30 patients, and it was skin-localised in 11/12 cases and bowel-localised in 1 case. Major infectious events occurred in 9/12 cases and in all of these patients high levels of PCT (>1.3 ng/ml) were detected at all three time points. During minor infectious events, the values of PCT1, 2 and 3 were ≤ 0.2 ng/ml regardless of GVHD.

The present study shows that PCT is a reliable marker of infection in patients with haematological neoplasms and febrile neutropenia. A fair number of publications (7-11, 12-16) have evaluated the utility of PCT as a marker of infection in neutropenic patients with solid-organ cancer or haematological malignancy. Overall, these studies showed a correlation between an elevation of PCT and the presence of infection. However, in a recent review (9) that explored studies published between January 1990 and October 2006, the authors were unable to reach definitive conclusions about the usefulness of PCT in febrile neutropenia because of the heterogeneity of the populations evaluated and the variability of the results obtained in the different articles.

In our study, the significant increase of PCT during infectious events was not affected by inflammatory conditions such as mucositis and GVHD. To date, only two articles have addressed the role of PCT in haematological patients with neutropenic fever and mucositis and / or GVHD, and they came to conflicting conclusions. Both these studies used the LUMI Test B.R.A.H.M.S. to assess the levels of PCT. Blijlevens et al. (7) demonstrated the inability of PCT to discriminate between infection and other inflammatory conditions in 12 haematological transplanted patients. In particular, the authors found out that GVHD rather than infection accounted for the increase of PCT. Fleischhack et al. (11), in a study including paediatric patients with haematological or solid tumours without infections, concluded that chemotherapy-induced tissue damage (such as severe mucositis) did not cause considerable increases of the PCT plasma levels. Some considerations can be made to justify these different results. In the first study, the limited number of patients impacted the statistical evaluation of the results; in the second study, none of the patients was transplanted and half of the febrile events occurred in children with solid tumours.

American Journal of Hematology

3 4	103
5 6 7	104
, 8 9	105
10 11 12	106
13 14	107
15 16	108
18 19	109
20 21	110
22 23 24	111
25 26	112
28 29	113
30 31	114
33 34	115
35 36 27	116
38 39	117
40 41 42	118
43 44	119
45 46 47	120
48 49	121
50 51 52	122
53 54	123
55 56 57	124
58 59	125
60	

In our study, the levels of PCT at all three time-points detected during the course of major infectious events in patients with and without mucositis were significantly higher than in patients with minor infections. Regarding the influence of GVHD on the PCT increase in our study population, although the sample size does not allow statistical evaluation, it is clear that the PCT was not affected by GVHD and values greater than 1 ng / ml are present only in patients with severe infections regardless of the simultaneous occurrence of GVHD.

In some studies (12, 17, 18) it was reported that PCT elevation could be significantly higher in patients with Gram-negative bacteraemia than in those with Gram-positive bacteraemia. Kocazeybek and coworkers (18) in a study on patients with infectious endocarditis found that the median serum PCT level of Gram-positive-related endocarditis was 2.92 ng/ml, whereas this level was 8.62 ng/ml in cases of endocarditis due to gram-negative bacteria. Charles and colleagues (17), in a review on bacteraemia in critically ill patients, observed that a high PCT value was found to be independently associated with Gram-negative bacteraemia and that a PCT level of 16.0 ng/ml yielded an 83% positive predictive value and a 74% negative predictive value. In our study, values of PCT of \geq 0.98 ng/ml were able to discriminate between the presence and absence of Gram-negative bacteraemia with a sensitivity of 80% and a specificity of 78%. This is of great importance in relation to the clinical severity of Gram-negative bacteraemia in patients with neutropenia and suggest the use of PCT in guiding the choice of empirical antibiotic therapy during febrile neutropenia.

In conclusion, our experience indicates that the determination of PCT at the given timein conclusion, our experience indicates that the determination of PCT at the given timepoints might represent a valuable tool to predict the occurrence of major infectious episodes in patients with febrile neutropenia, even in presence of confounding associated conditions such as mucositis and/or GVHD. This would allow an early and appropriate antibiotic therapy to be

1	
2 3 4 126	administered in a very high-risk population such as the one including patients with haematological
5 6 127 7	malignancies and chemotherapy-related febrile neutropenia.
, 8 9 128	
10 11 129 12	Methods
¹³ 14 14	Patients
15 16 131 17	From January 2008 to December 2008, 116 febrile episodes occurring in 88 neutropenic
¹⁸ 132 19	patients were investigated. All the patients were affected with haematological neoplasms and 30
20 21 133 22	of them underwent stem cell transplantation (SCT).
23 24 134	Fever was defined as a single measurement of a temperature of 38.5°C or two or more
25 26 135 27	measurements of 38°C. Neutropenia was defined as an absolute neutrophils count <1000/mm ³ .
28 29 136	Infection events included all episodes of fever associated with evidence of microbiological
30 31 137 32	isolations. Sepsis, septic shock, bacteraemia (BSI) and pneumonia were defined as major infectious
³³ 34 ¹³⁸	events; a fever of unknown origin (FUO), infections of the site of insertion of the central venous
35 36 139 37	catheter (CVC), sinusitis, Herpes simplex virus manifestations and folliculitis were defined as minor
38 39 140	infectious events.
40 41 141 42	BSI was defined as a microbial growth in one blood culture bottle; however, at least two
43 44 142	positive blood cultures bottles collected from different sites were required for microorganisms
45 46 143 47	usually considered as potential contaminants (i.e., coagulase-negative Staphylococcus species).
48 49 144 50	Haemoculture samples were collected upon the first appearance of fever and then on every
50 51 145 52	occasion in which fever was \geq 38°C. Sepsis was defined by the presence of fever or hypothermia,
53 54 146 55	tachypnea and tachycardia. Instances of sepsis associated with hypotension not corrected by
56 147 57 58	proper fluid therapy were scored as septic shock.

American Journal of Hematology

Pneumonia was assumed in cases in which typical clinical and radiological findings were seen on the chest radiograph obtained upon fever onset. CVC infection was suspected in presence of skin inflammation or vein thrombosis at the CVC insertion site. Episodes of fever not associated with microbiological or clinical evidence of infection were regarded as FUO.

Oral and gastrointestinal mucositis was defined and graded according to the Common Terminology Criteria for Adverse Events (CTCAE) version three (19). Acute GVHD was evaluated according to the standard criteria (20)

5 PCT, ESR and CPR evaluations

PCT was determined by a commercially available immunoluminetric assay, LUMI Test PCT kit (B.R.A.H.M.S. Diagnostica, Berlin Germany). Values greater than or equal to 0.5 ng/ml were considered to be pathological in accordance with the manufacturer's instructions (Al-Nawas & Shah, 1996). The precision of the test was estimated in accordance with CLSI guidelines EP5-A (evaluation of precision performance of clinical chemistry devices). The inter-assay coefficient variation at PCT value of 0.1 ng/ml was 15%, 0.2 ng/ml was 10% and >0.3ng/ml was <6%.

ESR was determined with the VES Matic Cube 200 (Diesse Inc., FL 33012, USA), an automatic system using 1 ml of undiluted blood sample with K2-EDTA as an anticoagulant. Blood collected in the EDTA tube and accurately mixed was left to settle. By analogical sensors, the instrument automatically reads the erythrocyte sedimentation levels. The results obtained were compared with the Westergren method and then printed.

The CRP measurement was determined with the use of a Tina-quant immunoturbimetric kit for CRP (latex) highly sensitive assay (Roche). The measuring range is from 0.01 to 2 mg/dl. The turbidity resulting from the immunoprecipitation of CRP in the serum with a specific antibody, in the presence of polyethylene glycol, is measured as an increase in absorbance at 340 nm and is

proportional to the concentration of CRP in the sample. The samples were read with the Hitachi704/705 instruments (Roche).

Blood samples to evaluate PCT, ESR and CRP were collected upon the first appearance of fever and then every day after until its resolution. For the statistical analysis, the values of three specific time-points were considered: time 1 (PCT1, ESR1 and CRP1) on the first day of fever, time (PCT2, ESR2 and CRP2) on the day with the highest values during the febrile episode and time 3 (PCT3, ESR3 and CR 3) on the last day of fever.

.78 Statistical analysis

The statistical analysis was carried out using the Mann-Whitney U test for the medians of non-parametric data and Fischer's Exact Test for the comparison of proportions. The sensitivity, specificity and area under the ROC (Receiving Operator Characteristic) curve were calculated for both PCT and CRP. For the statistical analysis, the data were stored by Excel 2003 and later processed using the software SPSS version 16.

1 2	
3 184 4	References
5 6 185 7	1. Link H, Bohme A, Cornely OA, et al. Antimicrobial therapy of unexplained fever in
8 9 186	neutropenic patients—guidelines of the Infectious Diseases Working Party (AGIHO) of the
10 11 187 12	German Society of Hematology and Oncology (DGHO), Study Group Interventional
13 ₁₈₈ 14	Therapy of Unexplained Fever, Arbeitsgemeinschaft Supportivmassnahmen in der
15 16 189 17	Onkologie (ASO) of the Deutsche Krebsgesellschaft (DKG-German Cancer Society). Ann
¹⁸ 190 19	Hematol 2003; 82(Suppl 2):105–11
20 21 191 22	2. Bodey GP, Buckley M, Sathe YS, Freireich EJ. Quantitative relationships between circulating
²³ 24 ¹⁹²	leukocytes and infection in patients with acute leukemia. Ann Intern Med 1966; 64: 328–
25 26 193 27	340.
28 29 ¹⁹⁴	3. Maruna P, Nedelnikova R, Gurlich R. Physiology and genetics of procalcitonin. Physiol Res
30 31 195 32	2000; 49:S57-S61.
³³ 34 196	4. Assicot M, Gendrel D, Carsin H, et al. High serum procalcitonin concentrations in patients
35 36 197 37	with sepsis and infection. Lancet 1993; 341:515-518.
³⁸ 39 198	5. SimonP, Milbrandt EB, Emlet L. Procalcitonin-guided antibiotics in severe sepsis. Critical
40 41 <u>199</u> 42	Care 2008; 12:309, 1-3
43 44 200	6. Charles PE, Ladoire S, Snauwaert A, et al. Impact of previous sepsis on the accuracy of
45 46 201 47	procalcitonin for the early diagnosis of blood stream infection in critically ill patients. BMC
48 49 202	Infectious Diseases 2008; 8:163, 1-10.
50 51 203 52	7. Blijlevens NMA., Donnelly JP, Meis JFG, et al. Procalcitonin Does Not Discriminate Infection
53 54 204	from Inflammation after Allogeneic Bone Marrow transplantation. Clinical and Diagnostic
55 56 205 57 58 59 60	Laboratory Immunology, 2000; 7(6) p. 889–892.

1	
3 4 206	8. Schuttrumpf S, Binder L, Hagemann T, et al. Procalcitonin: a useful discriminator between
5 6 207 7	febrile conditions of different origin in hemato-oncological patients? Ann Hematol 2003;
8 9 208	82:98–103
10 11 209 12	9. Sakr Y, Sponholz C, Tuche F, et al. The role of procalcitonin in Febrile Neutropenic Patients:
¹³ 210 14 15	review of the Literature. Infection 2008; 36: 396–407
16 211 17	10. Svaldi M, Hirber J, Lanthaler A., et al. Procalcitonin-reduced sensitivity and specificity in
¹⁸ 212 19 20	heavily leucopenic and immunosuppressed patients. British Journal of Haematology,
21 213 22	2001; 115, 53-57
23 24 214 25	11. Fleischhack G, Kambeck I, Cipic D, et al. Procalcitonin in paediatric cancer patients: its
26 215 27	diagnostic relevance is superior to that of C-reactive protein, interleukin 6, interleukin 8,
²⁸ 29 216 30	soluble interleukin 2 receptor and soluble tumour necrosis factor receptor II. British
31 217 32	Journal of Haematology, 2000; 111, 1093-1102
³³ 34 218 35	12. Svaldi M., Hirber J., Lanthaler AI, et al.: Procalcitonin-reduced sensitivity and specificity in
36 219 37	heavily leucopenic and immunosuppressed patients. Br J Haematol 2001; 115: 53–57.
³⁸ 39 220 40	13. Carnino L, Betteto S, Loiacono M, et al. Procalcitonin as a predictive marker of infections
41 221 42	in chemoinduced neutropenia. J Cancer Res Clin Oncol. 2009 Oct 30. [Epub ahead of print]
⁴³ 44 222 45	14. Semeraro M, Thomée C, Rolland E, et al. A predictor of unfavourable outcome in
46 223 47	neutropenic paediatric patients presenting with fever of unknown origin. Pediatr Blood
48 49 224 50	Cancer. 2009 Nov 19. [Epub ahead of print]
51 225 52	15. Hatzistilianou M, Rekliti A, Athanassiadou F, Catriu D. Procalcitonin as an early marker of
53 54 226 55	bacterial infection in neutropenic febrile children with acute lymphoblastic leukemia.
56 227 57	Inflamm Res. 2009 Oct 6. [Epub ahead of print]
58 59 60	

1 2			
3 4 2	28	16.	Martinez-Albarran M, Perez-Molina Jde J, Gallegos-Castorena S, et al. Procalcitonin and
5 6 22 7	29		C-reactive protein serum levels as markers of infection in a pediatric population with
8 9 23	30		febrile neutropenia and cancer. Pediatr Hematol Oncol. 2009; Sep;26(6):414-25
10 11 23 12	31	17.	Charles PE, Ladoire S, Aho S, et al. Serum procalcitonin elevation in critically ill patients at
13 14 14	32		the onset of bacteraemia caused by either gram negative or gram positive bacteria BMC
15 16 23 17	33		Infectious Diseases 2008; 8:38 ,2334-8
18 19 23	34	18.	Kocazeybek B, Küçükoglu [°] S, Öner YA Procalcitonin and C-Reactive Protein in Infective
20 21 23 22	35		Endocarditis: Correlation with Etiology and Prognosis. Chemotherapy 2003; 49:76–84.
²³ 24 ²³	36	19.	Cancer Therapy Evaluation Program, Common Terminology Criteria for Adverse Events,
25 26 23 27	37		Version 3.0, DCTD, NCI, NIH, DHHS, March 31, 2003 (http://ctep.cancer.gov), Publish Date:
28 29 23	38		August 9, 2006
30 31 23 32	39	20.	Przepiorka D, Weisdorf D, Martin P et al. Consensus Conference on Acute GVHD Grading.
33 24 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60	40		Bone Marrow Transplant 1995; 15:825-828
			12

1	
³ 241	
4 5	
6 242 7	Legends
8 9 243	
10	
11 244 12	Figure 1
13 14 15	PCT (a and c) and CRP (b and d) determinations during febrile major and minor events in patients
16 246 17	with and without mucositis.
18 19 247	
20	
21 248	
22	
23 24 25	Figure 2.
26 250 27	Comparison between the PCT and CRP ROC curves in febrile events in the presence of mucositis
²⁸ 29 ²⁵¹	according to major infectious events. A . PTC1, 68% (95% CI: 54 to 82; p=0.016); CRP1, 68% (95% CI:
30 31 252 32	54 to 81; p=0,019). B PCT2, 68% (95% CI: 53.9 to 81.7; p = 0.02); CRP2, 54% (95% CI: 38.8 to 68.9;
³³ 253 34	p = 0.6). C. PCT3, 65% (95% Cl: 51 to 80; p = 0.035); CRP3, 48% (95% Cl: 33 to 63; p = 0.8).
35 36 254	
37	
38	
39	
40	
42	
43	
44	
45 46	
47	
48	
49	
50 51	
52	
53	
54	
55 56	
57	
58	
59	
60	

TABLE 1. Patients and febrile episode characteristics

Characteristic	N° (%)
Total no. of patients	88
No. of non-transplanted patients	58 (66)
LMA°	40 (69)
LLA°°	9 (16)
LMC°°°	1 (2)
LNH^	3 (5)
LT^^	1 (2)
LB^^^	2 (3
No. of transplanted patients	30 (34)
Allogeneic	7 (23)
Autologous	14 (47)
Haploidentical	3 (10)
Umbilical cord Blood	6 (20)
Age (years)	
Median (range)	47 (16-70)
Gender	
Male	52 (59)
Female	36 (41)
Total no. of febrile episodes	116
Sepsis	21 (18)
Septic shock	9 (8)
Pneumonia	22 (19)
Bacteraemia	9 (8)
CVC*	20 (17)
FUO§	29 (25)
Other+	6 (5)
Febrile episodes associated with mucositis (in	58 (50)
total)	
Febrile episodes with GVHD (only in transplanted)	9 (30)
° LMA Acute myeloid leukaemia	
*Acute Lymphatic leukaemia	

°°°Chronic myeloid leukaemia

^Lymphoma non Hodgkin

^^^Burkitt Lymphoma

*Central venous catheter infection

§ Fever of unknown origin

+ Folliculitis, H. simplex, sinusitis

Figure 1



Figure 2

