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Title: FSH-stimulated PTEN activity accounts for the lack of FSH mitogenic effect in prepubertal rat Sertoli cells

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ABSTRACT

Follicle-stimulating hormone (FSH) controls the proliferation and differentiation of Sertoli cells of the testis. FSH binds a G protein-coupled receptor (GPCR) to stimulate downstream effectors of the phosphoinositide-3 kinase (PI3K)-dependent pathway, without enhancing PI3K activity. To clarify this paradox, we explored the activity of phosphatase and tensin homolog deleted in chromosome 10 (PTEN), the PI3K major regulator, in primary cultures of rat Sertoli cells. We show that, within minutes, FSH increases PTEN neosynthesis, requiring the proteasomal degradation of an unidentified intermediate, as well as PTEN enzymatic activity. Importantly, introducing an antisense cDNA of PTEN into differentiating Sertoli cells restores FSH-dependent cell proliferation. In conclusion, these results provide a new mechanism of PTEN regulation, which could serve to block entry into S phase of Sertoli cells, while they are proceeding through differentiation in pre-pubertal animals.

INTRODUCTION

G protein-coupled receptors represent the largest class of cell surface receptors, involved in most physiological regulations, including reproduction. Once activated by their cognate ligand, they mediate diverse intracellular signalling events, mainly transduced by their interaction with heterotrimeric G proteins, and with their companion scaffolding proteins, β -arrestins (Reiter and Lefkowitz, 2006). Among those signalling events, some GPCRs stimulate the activity of class IB phosphatidylinositol-3 kinase (Stephens et al., 1993), which converts phosphoinositide(4, 5)P₂ (PIP₂) to phosphoinositide(3, 4, 5)P₃ (PIP₃) in the lipid bilayer. PIP₃ then serves as a second messenger which recruits signalling proteins containing a pleckstrin homology domain (Klarlund et al., 1997).

The biological activity of class I PI3K is antagonized by a key regulator of phosphoinositide steady-state, PTEN, which dephosphorylates PIP₃ in position 3 (Vazquez and Devreotes, 2006). As such, PTEN has been largely acknowledged as a tumor suppressor, inactivated by loss of heterozygosity on chromosome 10q23, by mutation or by promoter methylation silencing (Carnero et al., 2008). While those properties of PTEN have been widely documented, relatively little is known about PTEN regulation. *Pten* gene transcription has been shown to be up-regulated through DNA binding sequences for early growth regulated transcription factor 1 (EGR1) (Virolle et al., 2001) for example, and down-regulated through mitogen-activation protein kinase kinase-4 (MKK4)-dependent NF- κ B binding sites (Xia et al., 2007). The PTEN protein is stabilized by direct interaction with casein kinase II that limits its caspase-mediated degradation (Torres et al., 2003). PTEN is also regulated by post-translational modifications, such as mono/polyubiquitination, phosphorylation, acetylation or oxidation, which control its activity, stability or sub-cellular localization (Tamguney and Stokoe, 2007).

The FSH-R is a GPCR mainly expressed at the cell surface of Sertoli cells of the seminiferous tubules, where it transduces activation of diverse signalling pathways, which greatly vary according to the differentiation stage, leading to appropriate cellular responses (Crépieux et al., 2001; Lecureuil et al., 2007; Meroni et al., 2002; Meroni et al., 2004). For example, 5 day-old rats Sertoli cells proliferate in response to FSH. At this stage, FSH enhances the phosphorylation of MAP kinases ERK1 and ERK2, thus increasing cyclin D1 expression. In contrast, FSH totally switches off the ERK pathway in 19 day-old rat differentiating Sertoli cells, and *in vitro*, cell proliferation becomes insensitive to FSH (Crépieux et al., 2001). Likewise, we observed that the signaling pathways activated by FSH upstream of the p70 S6 kinase (p70S6K1) are different at each of those developmental stages, resulting in differential phosphorylation patterns of the enzyme (Lécureuil et al., 2005) (Musnier *et al.*, submitted). In particular, in differentiating Sertoli cells, FSH stimulates downstream targets of PI3K, such as mammalian Target Of Rapamycin (mTOR). However, we provided evidence for a tonic PI3K activation in these cells (Lécureuil et al., 2005). To gain a better insight onto PI3K regulation in differentiating Sertoli cells, here we analyzed expression, activity, and functionality of PTEN, the key regulator of PI3K biological function.

MATERIALS AND METHODS

Purification of Sertoli cells

Animals were treated following the current ethical guidelines of the European Community. Sertoli cells were isolated from testes of 5 or 19 day-old Wistar rat, as published (Guillou et al., 1986), and fifty thousand cells /cm² were plated on CellBind plates (Corning) in Dulbecco's minimum essential medium (DMEM) supplemented with penicillin (100 IU/ ml), streptomycin (100 µg/ml), glutamine (2 mM), retinol (50 ng/ml), vitamin E (200 ng/ml) and human transferrin (5 µg/ml). The medium was replaced after 24 hours to remove unattached cells, and Sertoli cells were stimulated by 100 ng/ ml porcine FSH (bioactivity of 160 x NIH-FSH-P1) purified by Dr J. Closset Université de Liège, Belgium (Closset and Hennen, 1989), or by 10 ng/ml tumour-necrosis factor α (TNF α). MG132 and lactacystin were from Calbiochem (Merck Chemical Ltd). All other reagents were from Sigma Chemical Co (St-Louis, MO).

Cell transfections and plasmids

Once purified from testes, Sertoli cells were seeded at a density of 3, 750 cells / mm² in 24-well CellBind plates. On the next day, the JetPEI reagent from Polyplus was used, to transfect cells in each well with 100 ng of E2F1-luciferase reporter plasmid, 25 ng of Renilla luciferase plasmid, and 900 ng of pBK-CMV plasmid or pBK-CMV plasmid encoding antisense PTEN. Forty-eight hours after transfection, cells were stimulated with FSH for 8 hours, and then were lysed in 100 µl/well of passive lysis buffer, according to the manufacturer's instructions (Dual luciferase kit, Promega). Cell lysates were then transferred to blind 96-well plates and the chemiluminescence produced by the firefly and renilla luciferases was detected on an Ascent luminiscan (Thermo Labsystems). The E2F1-driven luciferase plasmid (E2F1-luc) contains 3 E2F1- and one Sp1-binding sites linked to the thymidine kinase promoter, and was donated by Dr A. Harel-Bellan (Villejuif, France). The mammalian expression vector pBK-CMV, driven by the cytomegalovirus immediate-early promoter and containing the neomycin resistance gene, was purchased from Stratagene Cloning Systems (La Jolla, CA). The PTEN cDNA (1218 pb, a gift from Dr. Kaz Matsumoto, NIDCR, NIH, Bethesda, MD) was subcloned into the EcoRI and XbaI sites of PBK-CMV in the antisense orientation, as described in (Wu et al., 2004). The orientation of the construct was verified by PCR and DNA sequencing. The plasmid encoding Renilla luciferase was provided in the Dual luciferase kit.

Western blot analysis

Sertoli cells were lysed in TNET lysis buffer (20 mM Tris pH 7.8, 50 mM NaCl, 5 mM EGTA, 1 % vol/vol Triton X-100, 1 mM phenylmethylsulfonyl fluoride, 4 mM Na₃VO₄, 5 µg/ml leupeptin, 5 µg/ml pepstatin, and 5 µg/ml aprotinin, and 40 µg of proteins were separated by SDS-PAGE and transferred to nitrocellulose membrane (Whatman, Dassel, Germany). Membranes were saturated with TBST-BSA (Tris Buffer Saline with 0.1 % Tween 20, 3 % Bovine Serum Albumin) and were probed with the following antibodies, all diluted 1: 1000 in Tris Buffer Saline / 0.1 % Tween 20 (TBS/T) with 3 % BSA: rabbit anti-phospho-I κ B α , anti-PTEN (Cell Signalling Technology), anti-phospho-Akt [Ser473] (Abcam), anti-rabbit secondary antibody (Eurobio). Membranes were also probed with anti-vinculin or anti-actin (Sigma Aldrich) antibodies to monitor gel loading. Western blots were revealed with enhanced

chemiluminescence (NEN Life Science Products). Films were scanned and the optical density of the signals was measured with ImageMaster 1D Elite version 4 software (Amersham Biosciences).

In vitro PTEN enzymatic activity

After stimulation, Sertoli cells were disrupted on ice for 30 min in lysis buffer (137 mM NaCl, 20 mM Tris HCl pH 7.5, 1 mM MgCl₂, 1 mM CaCl₂, 150 mM Na₃VO₄, 1 % Nonidet P-40, 10 % glycerol, 2 mM phenylmethylsulfonyl fluoride (PMSF), 10 µg/ml aprotinin). Briefly, Sertoli cell extracts were immunoprecipitated with anti-PTEN antibody. PTEN phosphatase activity correlates with the amount of phosphate released from diC₈-PIP₃, visualized by a colorimetric assay (Biomol Green™ reagents) read at 620 nm.

Rat transferrin quantification

Secreted transferrin was measured in 100 µl of Sertoli cell culture media by radioimmunoassay, as described (Suire et al., 1997). All standards and samples were assayed in triplicate.

PIP3 assay by protein-lipid overlay

To estimate PI3K activity, PIP3 was quantified from 10⁷ Sertoli cells as described recently (Guillou et al., 2007). Briefly, PtdIns-rich fractions were extracted with chloroform/ methanol, selectively isolated on neomycin beads, eluted with methanol-chloroform-HCl. Purified PtdIns were spotted onto a nitrocellulose membrane, blotted overnight at 4°C with the PH domain of the General Receptor for Phosphoinositide 1 (GRP1) fused to green fluorescent protein (GFP). Membranes were then probed with a rabbit anti-GFP polyclonal antibody coupled to HRP, the chemiluminescence was detected with the ECL reagent (Amersham) and acquired with the Image Reader LAS-1000 charge-coupled device (CCD) camera (Fujifilm). Densitometry was measured with the AIDA software with local background subtraction.

Sertoli cell metabolic labeling

Twenty million Sertoli cells were pre-incubated with actinomycin D (5 µg/ml), then labelled with 200 µCi (0.1 nmole) of ³⁵S-Met/³⁵S-Cys, stimulated by 100 ng/ml FSH for one hour, and lysed in 300 µl RIPA buffer (150 mM NaCl, 50 mM Tris pH 8.0, mM EDTA, 1 % Triton X-100). PTEN was immunoprecipitated in the cell lysates with rabbit anti-PTEN antibody (1: 100), overnight at 4°C, and the immunoprecipitates were analyzed by SDS-PAGE. Then the gel was vacuum-dried and exposed to a phosphorImager screen.

RNA extraction and Q-RT-PCR

Total RNA were extracted from 3 x 10⁶ cells per condition, by using the RNABle reagent (Eurobio, les Ulis, France), according to the manufacturer's instructions. For each time point, three independent sets of total RNA from FSH-stimulated cells were isolated and used as a template for probe generation. RNA integrity and quantity was assessed using RNA 6000 nanochips with an Agilent 2100 bioanalyser (Agilent Technologies, Inc., Palo Alto, CA). One µg of total RNAs was reverse transcribed with the Superscript II Reverse Transcriptase from Invitrogen (Life Technologies, Gaithersburg, MD, USA) according to the manufacturer's instructions,

and 1/40th of the cDNA was used as template for PCR amplification as previously described (Cederroth et al., 2007). A mean quantity was calculated from triplicate PCR reactions for each sample, and this quantity was normalized to two similarly measured quantities of normalization genes (*GAPDH*, *rps29* and *tubb*). The statistical significance of fold-changes was determined by a paired Student's *t*-test. Rat PTEN primers used for qRT-PCR are:

forward 5' AATGTTTCAGTGGCGGAACTTG 3' and
reverse 5' TCCTGAGTTGGAGGAGTAGATCTTC 3'.

Softwares

The Cell Designer graphic interface (<http://celldesigner.org>) has used to describe the signalling pathways explored herein.

Data analysis

Measurements were performed in duplicates or triplicates and the mean values \pm s.e.m of at least 3 independent experiments are presented. Results were generally expressed as percent of the maximum values. Statistical analysis of the data was performed using one-way ANOVA (Bonferroni's multiple comparison tests) to compare samples (GraphPad PRISM Software, San Diego, CA). Values with $p < 0.05$ were considered as significantly different.

RESULTS

Phosphoinositide level in Sertoli cells

To estimate PI3K activity, we analyzed the PIP3 content of rat Sertoli cells by protein-lipid overlay (Guillou et al., 2007) and expressed the results as % of the response to insulin, taken as a positive control. As expected from our previous report (Lécureuil et al., 2005), in differentiating Sertoli cells isolated from prepubertal 19-day old rats, the level of PIP3 was basally elevated, when compared to the level of PIP3 measured in proliferating Sertoli cells from 5-day old rat neonates (Figure 1A). Furthermore, in differentiating Sertoli cells, the PIP3 level was not significantly increased upon FSH stimulation (Figure 1B) nor by Insulin stimulation (data not shown). Accordingly, phosphorylation of the Akt Ser/Thr kinase on Ser473 was not significantly enhanced upon FSH stimulation, whereas it was clearly increased by Insulin stimulation for 60 min, as expected (Figure 1C). Neither was phosphorylation of Akt Thr308 visible in FSH-stimulated cells (data not shown). Accordingly, phosphorylation of the PtdIns-dependent transcription factor Foxo1 was not enhanced upon FSH stimulation of these cells (Musnier et al., 2009).

FSH enhances PTEN protein level and activity

Since the PIP3 level was basally high in differentiating Sertoli cells, we anticipated that the expression of PTEN could be basally low. This was actually the case, but surprisingly, time-course stimulations led to a gradual increase in PTEN protein, as soon as 15 min of FSH stimulation (Figures 2A, 2B). This increase peaked around 60 min and remained high thereafter. It required the continuous presence of the hormone in the cell medium, since when the FSH-containing medium was replaced by fresh medium for the last 120 minutes of incubation, PTEN protein expression returned close to its basal level (Figure 2C). Importantly, PTEN activity paralleled PTEN upregulation in response to FSH (Figure 2D), unless the hormone was withdrawn for the last 120 minutes as above (Figure 2E). Insulin stimulation also led to an increase in PTEN activity (Figure 2F). Therefore, enhanced PTEN activity by FSH could counterbalance PI3K activity by maintaining a constant level of PIP3, as shown in Figure 1B.

FSH enhances PTEN neo-synthesis

As expected from such an early cellular process, preventing transcription with actinomycin D did not significantly impair FSH-induced PTEN up-regulation (Figure 3A), suggesting that FSH enhanced PTEN expression by a post-transcriptional mechanism. The efficacy of actinomycin D was assessed by quantifying extracellular transferrin, whose gene has been reported to be regulated by FSH mainly transcriptionally (Suire et al., 1995) (Figure 3B). To conclusively support a post-transcriptional regulation of PTEN, we assessed by quantitative real time RT-PCR the expression levels of PTEN upon gradual increase of FSH stimulation. We found that PTEN transcripts decreased over time (Figure 3C), thus ruling out the possibility that the accumulation of PTEN proteins could be due to an up-regulation of PTEN transcription. Therefore, one possibility was that FSH would enhance PTEN proteo-synthesis. When newly synthesized PTEN was labelled with $^{35}\text{S-Met}/^{35}\text{S-Cys}$, we observed that FSH was able to increase PTEN protein level (160 % when normalized with a non-specific band), even when transcription was blocked (Figure 3D).

PTEN up-regulation requires FSH-induced proteolysis

We next sought to explore the mechanism whereby PTEN protein level was enhanced by FSH. Since PTEN degradation has been shown in other systems to be mediated by the proteasome (Torres and Pulido, 2001), we tested whether FSH would inhibit a constitutive proteasome-mediated degradation of PTEN, in addition to increasing PTEN proteosynthesis. In time-course experiments, the MG 132 proteasome inhibitor hampered FSH-induced PTEN up-regulation (Figure 4A, 4B). Efficacy and specificity of MG 132 were further confirmed by stabilization of the I κ B α protein (Figure 4C) in TNF α -treated cells (Traenckner et al., 1994). Similar results were obtained with an unrelated proteasome inhibitor, lactacystin (Figure 4D), indicating that proteolysis of an inhibitory protein was a prerequisite to PTEN upregulation by FSH.

PTEN depletion restores FSH mitogenic effect in Sertoli cell from prepubertal rats

Owing to its well-known biological function as a tumor suppressor, PTEN could be involved in triggering Sertoli cells in prepubertal animals towards their differentiation program, by limiting their mitogenic ability. To address this question, a pBK-CMV plasmid encoding antisense PTEN or wild-type pBK-CMV plasmid was introduced in differentiating Sertoli cells, together with a reporter plasmid encoding the luciferase cDNA under the control of E2F1-responsive elements. E2F1 binding sites can be used as sensors of the G1/S transition of the cell cycle. In Sertoli cells transfected with the pBK-CMV empty vector, FSH could not enhance luciferase activity (Figure 5A), consistently with previous data indicating that FSH does not enhance proliferation of Sertoli cells isolated from prepubertal rats (Crépieux et al., 2001; Orth, 1982). In contrast, in FSH-stimulated cells transfected with antisense PTEN, luciferase activity was significantly enhanced, suggesting that Sertoli cells have gained sensitivity to FSH to undergo proliferation. Therefore, upregulation of PTEN could serve to limit Sertoli cell proliferation when undergoing differentiation. In line with this view, FSH stimulation could not enhance PTEN expression in Sertoli cells isolated from neonates (Figure 5B).

DISCUSSION

In this paper, we have identified a novel regulation of PTEN by FSH. It is shown here that FSH enhances the protein level of the PI3K regulator PTEN, within minutes, while PI3K activity remains apparently constant, as seen by the absence of Akt phosphorylation. We propose that FSH-induced PI3K activity could not be observed precisely because PTEN expression is upregulated by FSH stimulation at the same time. In contrast, insulin enhances PTEN activity, probably as a negative feedback mechanism, as previously reported (Lackey et al., 2007; Moorehead et al., 2003). Hence, we favour a different mechanism for PTEN activation by insulin since insulin, unlike FSH, activates Akt in our cell model.

FSH-induced upregulation of PTEN was due to an enhanced PTEN neosynthesis rate. We postulate that in the absence of hormone, an undefined inhibitor X degrades or keeps PTEN from accumulating, which explains that PI3K activity is constitutively elevated. Obviously, this does not imply that the effect is direct, and this inhibitor could destabilize the PTEN protein or mRNA, although a direct input on the translational machinery cannot be ruled out. FSH stimulation would then target this inhibitor to the

proteasome for degradation, ultimately leading to PTEN accumulation and subsequent conversion of PIP3 to PIP2, as shown here.

What could be the physiological role of an FSH- induced rapid stimulation of PTEN in Sertoli cells? Sertoli cells isolated from 19 day-old rats were analyzed in this study. At this developmental stage, cells are proceeding through their differentiation program, and the mitosis rate slows down and becomes insensitive to FSH (Crépieux et al., 2001; Orth, 1982). Since PTEN level is not altered by FSH exposure of proliferating Sertoli cells recovered from neonate rats, it was likely that FSH-stimulated PTEN expression could be linked to Sertoli cell terminal differentiation. Clearly, this view is supported by the data presented herein, since the introduction of a plasmid encoding antisense PTEN in differentiating Sertoli cells knocks-down the endogenous PTEN mRNA and restores the responsiveness of these cells to FSH to undergo proliferation. This is consistent with the well admitted role of PTEN as a negative regulator of cell proliferation and parallels the previously proposed role of PTEN in the transition from proliferation to differentiation of granulosa cells, within the ovarian follicle (Froment et al., 2005).

So far, we have never been able to visualize phosphorylation of Akt, an indirect target of PI3K activity, before two hours of FSH stimulation (our unpublished results). Our results contradict those previously published which show that FSH enhances Akt phosphorylation in Sertoli cells from 20 day-old rats (Meroni et al., 2004). Differences in cell culture conditions could be amenable for this discrepancy, which otherwise remains obscure. However, it is clearly reminiscent of the observation that TSH, a glycoprotein hormone structurally related to FSH, stimulates p70S6K1 in a PI3K-dependent manner, without activating Akt (Suh et al., 2003). Furthermore, the muscarinic M3 receptor mediates p70S6K1 activation and enhances its phosphorylation on the T389 residue through an mTOR-dependent mechanism, in an astrocytoma cell line. Only a permissive, low level of PI3K activity is required (Tang et al., 2003) similarly to our observations in differentiating Sertoli cells stimulated by FSH (Lécureuil et al., 2005). While Akt is not activated in astrocytoma cells when stimulated by the M3 cognate agonist carbachol, PDK1 could be the permissive signal. Noteworthy, GPCRs have developed strategies of their own to attenuate PI3K signalling, by virtue of their ability to recruit the adapter proteins β -arrestins. For example, in response to the ligand-activated PAR2, PI3K is sequestered from the lipid components of the plasma membrane by β -arrestins (Wang and DeFea, 2006). Whether this is also the case in Sertoli cells stimulated with FSH remains to be investigated, since, the FSH-bound receptor also recruits β -arrestins to regulate its desensitization (Nakamura et al., 1998; Troispoux et al., 1999) and cell signalling (Kara et al., 2006).

In conclusion, in primary cultures of Sertoli cells from prepubertal rats, in which the molar ratio of the signalling components are balanced, we have identified a novel regulation of tumor suppressor PTEN by FSH. This mechanism appears to be critically involved in the loss of mitogenic potential of Sertoli cells, leading to their terminal differentiation.

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FIGURE LEGENDS

Figure 1: PI3K activity in rat Sertoli cells. A. Basal PIP3 content in 10^7 Sertoli cells isolated from 5- and 19-day old rats. B. PIP3 production in 19 day old rat Sertoli cells stimulated with FSH (100 ng/ml) for 60 min. C = unstimulated cell control. Results are expressed as mean % of the PIP3 response to insulin +/- SEM, in 4 independent overlays, each performed in triplicates. C. Representative Western blot of Akt phosphorylated on Ser473 as a function of FSH stimulation time (n= 3 independent experiments). An actin probe was used to monitor gel loading. Shared superscripts indicate no significant difference, while different superscripts indicate significant differences, at the $P < 0.05$ level.

Figure 2: PTEN expression and activity is rapidly enhanced by FSH signalling in differentiating Sertoli cells.

Time-course stimulation of Sertoli cells isolated from 19-day old rats by FSH (100 ng/ml). A. Representative Western blot of a time-course of FSH-induced PTEN expression. An actin probe was used to monitor gel loading. B. Quantification of the PTEN signal normalized with the actin signal, in 3 independent experiments. Results are expressed as % of the maximum PTEN induction +/- SEM C. and E. After 120 min, cells were maintained in the presence of FSH, or the hormone was withdrawn (chase). D and E. PTEN activity in response to FSH. A representative experiment (n = 3), with each sample assayed in triplicate, is shown. Crude values in nmoles of phosphate released have been normalized to the % of PTEN activity in response to insulin, taken as a positive control. F. PTEN activity in response to insulin. A representative experiment (n = 3), with each sample assayed in triplicate, is shown. Results are expressed as nmoles of phosphate released. Shared superscripts indicate no significant difference, while different superscripts indicate significant differences, at the $P < 0.05$ level.

Figure 3: FSH increases PTEN neosynthesis in differentiating Sertoli cells. A. Representative Western blot of FSH-induced PTEN expression in the presence of Actinomycin D (5 μ g/ml) (n = 3 independent experiments). An actin probe was used to monitor gel loading. B. Radioimmunoassay of transferrin secreted in the culture medium to monitor the efficiency of Actinomycin D, in Sertoli cells stimulated for 24 hours with FSH. C = unstimulated cell control. C. Detection of PTEN messenger RNA expression levels by real-time RT-PCR, in Sertoli cells stimulated with FSH for the indicated period of time. Results are expressed as mean \pm SEM (n = 3 per time point). REL = Relative Expression Level. D. Sertoli cells were stimulated with FSH for one hour in the presence of actinomycin D and 35 S-Met/ 35 S-Cys, or were left untreated. Cell lysates were incubated with anti-PTEN antibody or with a pre-immune rabbit serum (PI). Here is shown a representative autoradiogram out of 3 independent experiments. The fold-stimulation *versus* control is indicated beneath the autoradiogram. n.s. = non specific band. Shared superscripts indicate no significant difference, while different superscripts indicate significant differences, at the $P < 0.05$ level.

Figure 4: FSH signaling launches a proteasome-mediated proteolysis step to enhance PTEN protein level. A. Differentiating Sertoli cells were incubated with the MG132 proteasome inhibitor (10 μ M) for 2 hours prior stimulation with FSH for the indicated

period of time. Representative Western blot of a time-course of FSH-induced PTEN expression in the presence of MG132. B. Quantification of the normalized PTEN signal obtained in the presence of MG132 (n = 3 independent experiments). Results are expressed as the % of the maximum PTEN induction +/- SEM. C. Control Western blot hybridized with an anti-phospho-I κ B α to monitor MG132 efficiency, in Sertoli cells stimulated for 10 min with TNF α . D. Differentiating Sertoli cells were incubated with the lactacystin proteasome inhibitors (10 μ M) for 2 hours prior stimulation with FSH for the indicated period of time. Representative Western blot of a time-course of FSH-induced PTEN expression in the presence of lactacystin (n = 2 independent experiments). A vinculin probe was used to monitor gel loading. Shared superscripts indicate no significant difference, while different superscripts indicate significant differences, at the P < 0.05 level.

Figure 5: PTEN depletion restores FSH-dependent proliferation of Sertoli cells from prepubertal rats. A. Cells were transfected with the E2F1-luciferase reporter plasmid, and either the empty pBK-CMV plasmid or the pBK-CMV plasmid encoding antisense PTEN, as indicated. After 48 hours of expression, cells were stimulated with FSH for 8 hours before lysis. Results are expressed as relative light units +/- SEM normalized for transfection efficiency. Here are pooled the data from two experiments each done in six replicates. B. Representative Western blot of proliferating Sertoli cells from neonate rats stimulated by FSH for 60 min. The membrane was hybridized with an anti-PTEN antibody, then reprobbed with an actin probe to monitor gel loading. Shared superscripts indicate no significant difference, while different superscripts indicate significant differences, at the P < 0.05 level.

Figure 1

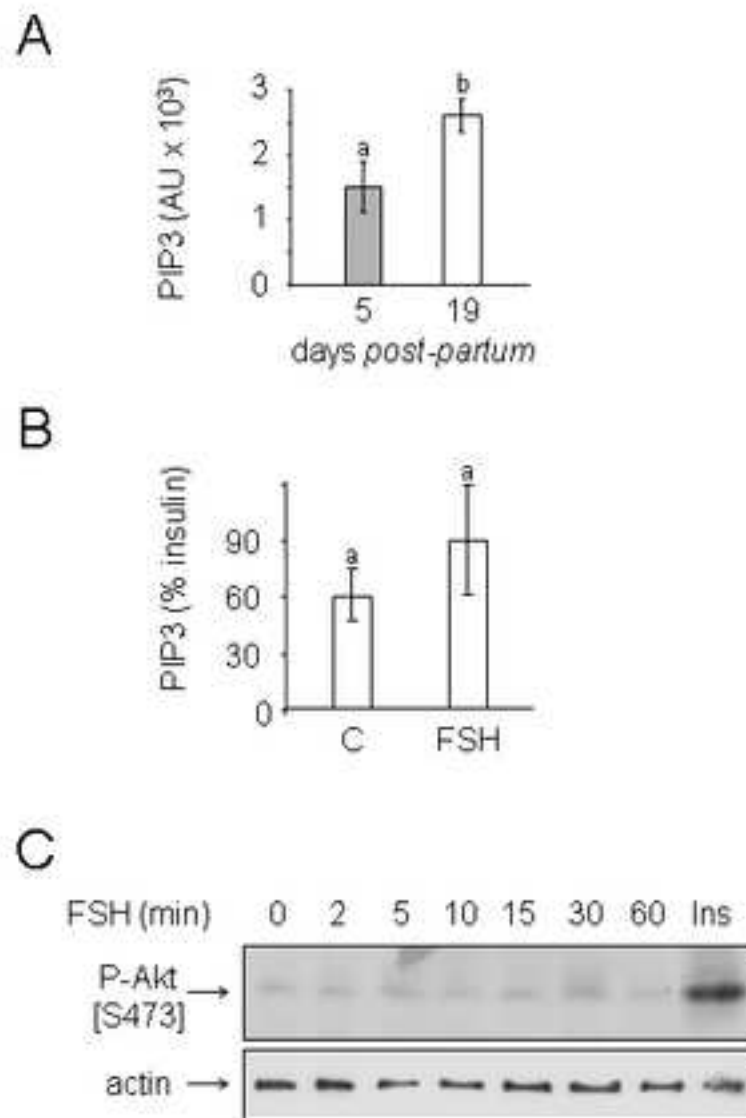


Figure 2

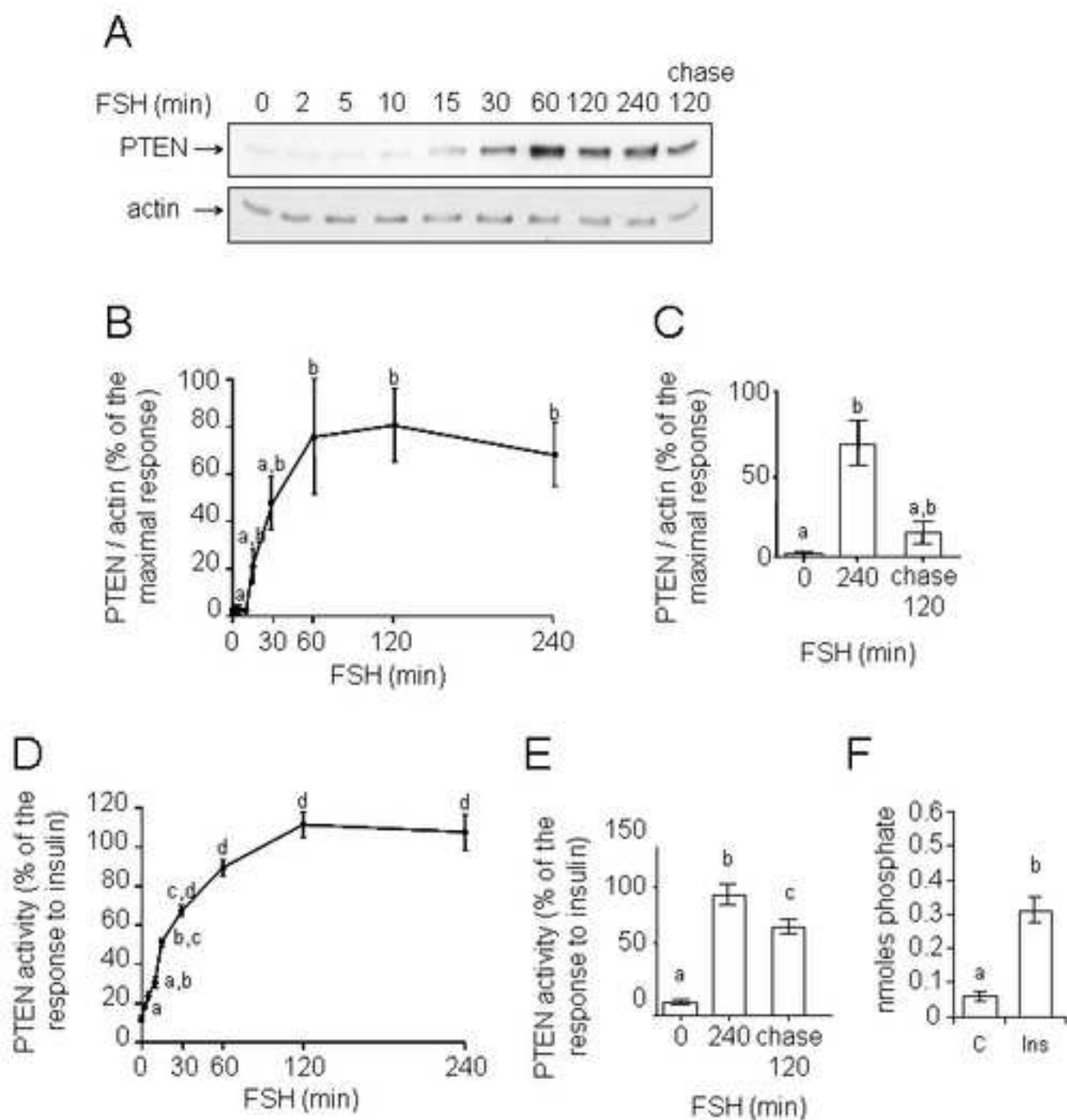


Figure 3

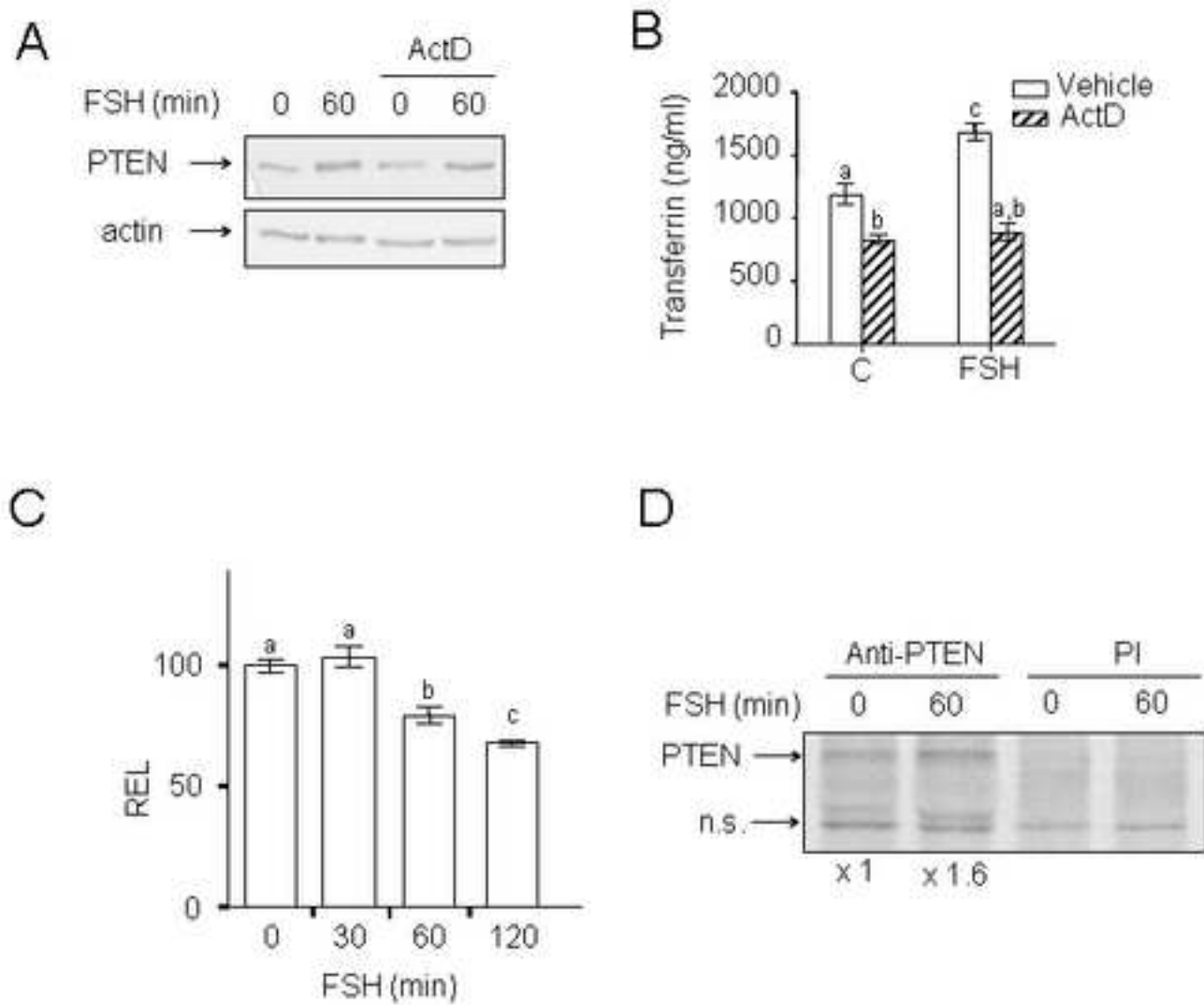


Figure 4

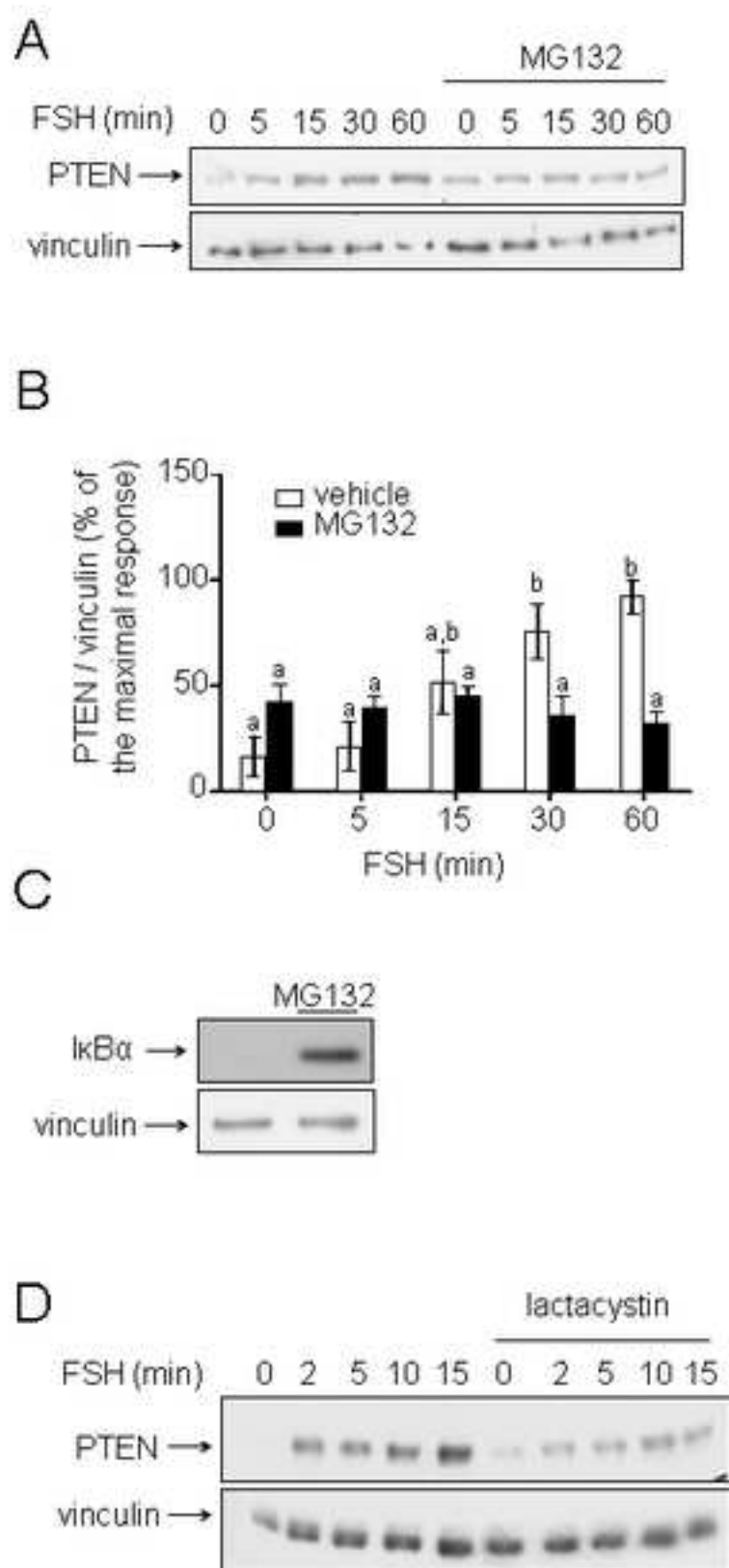


Figure 5

