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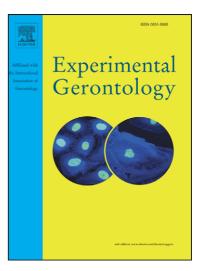
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Biomarkers for Alzheimer's disease and other forms of dementia: clinical needs, limitations and future aspects

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ABSTRACT

An early diagnosis of Alzheimer's disease (AD) and other types of dementia-causing disorders is vital in order to achieve effective treatments. Fortunately, in the recent years the search for specific biomarkers has undergone a rapid evolution. New technologies in proteomics and genomics have permitted great advances in defining biochemical markers in cerebrospinal fluid (CSF) and in blood. Novel imaging techniques are also improving the diagnosis and early detection of brain changes in vivo. Furthermore, combined analysis of different biomolecules, or of biochemical and neuroimaging studies, increase diagnostic sensitivity and specificity. However, the discovery of sensitive and specific biomarkers for neurodegenerative diseases needs to overcome some important challenges. With the available technology, standardization of methods is essential to reducing inconsistency and increasing reliability. Global initiatives, multicenter studies and consensus protocols of analysis are of critical importance. The present review summarizes the results achieved in the search for an early diagnosis of neurodegenerative disorders, and reflects the limitations and the perspectives of the field.

Keywords: Dementia; Alzheimer's disease; proteomics; imaging; biomarkers

INTRODUCTION

A diversity of neurodegenerative disorders are associated with the development of dementia. All of them are characterized by neuronal dysfunctions that eventually lead to neuronal death. All of them are similarly complex, possibly resulting from several neuropathological mechanisms acting together. Alzheimer's Disease (AD) is the most prevalent. It has been calculated that more than half of patients with dementia have AD, and that this number will double every 20 years (Brookmeyer et al., 1998). Lewy Body dementia (LBD) is the second most common dementia after AD (Tarawneh and Galvin, 2007). Also, frontotemporal lobar neurodegenerations such us frontotemporal dementia (FTD), midbrain disorders such as Parkinson's disease (PD) or progressive supranuclear palsy (PSP) and vascular dementia (VaD) are associated with progressive cognitive impairment.

Mild cognitive impairment (MCI) is often, but not always, a transitional state between normal aging and dementia (Winblad et al., 2004). The concept of MCI has been essentially derived from clinical and neuropsychological settings, and its definition is continually being revised. To reduce the heterogeneous nature of MCI, a subdivision of the concept has been further proposed based on the presence of a deficit in the amnestic domain, a single domain other than the amnestic, and multiple domains (Winblad et al., 2004). The estimated annual incidence of MCI in population studies is of about 2% in population studies (Larrieu et al., 2002; Palmer et al., 2008) and about 5-9% in clinical samples (Adak et al., 2004; de Leon et al., 2001). Few longitudinal data are available at the population level on MCI subtypes and their role as a transition between normal aging and dementia. The rates of transition between MCI and AD have been calculated between one-third to one-half for the MCIamnestic subtype (Palmer et al., 2008; Petersen et al., 1999) and two-thirds for MCImultidomains subtype over a period of three years. However, a substantial number of MCI patients are reclassified as normal aging on follow-up (Larrieu et al., 2002) and many MCI do not progress to any sort of dementia (de Jager and Budge, 2005). In

other words, the standard clinical diagnostic evaluations, including neuropsychological tests failed to identify those people with global cognitive deficits who have high risk of progressing to dementia.

As cognitive decline advances, other specific disease-related features appear, such as visual hallucinations and Parkinsonism in LBD (McKeith et al., 1996) or personality changes in FTD (Neary et al., 2005). Although the clinical picture of these diseases is often related to the area of the brain that is primarily affected, mixed pathologies are common and clinical manifestations in early disease phases are often similar. Furthermore, a single type of pathology can produce different cognitive outcomes making the diagnose difficult.

There is evidence that the pathological changes in dementia-causing diseases begin decades before the appearance of the first clinical manifestations (Jack et al., 2009). The challenge of finding effective treatments for AD and other dementias is parallel to the prospect of detecting the preclinical changes at the earliest stage possible, and of accurately identifying the pathology (or pathologies) responsible. To develop and validate biomarkers capable to early detect and discriminate between disorders with similar clinical outcomes is one of the most important goals of neuroscience research.

The quest for biomarkers for AD and other dementias includes research in multiple fields: genomics, proteomics, structural and functional imaging and neurophysiology. The present review summarizes the most recent advances and analyses the current and future directions in the pursuit of biomarkers for AD and other dementia disorders.

BIOCHEMICAL MARKERS

Biomarkers in CSF

CSF is a useful diagnostic window to neurodegenerative diseases for many reasons, mainly because it reflects metabolic processes in the brain in a direct manner, due to the free exchange of molecules between the brain and the CSF (Reiber and Peter, 2001). Amyloid plaques in AD brain are mainly composed of AB-42 and AB-43 (Welander et al., 2009), while in cerebral amyloid angiopathy (CAA) predominantly contain Aβ-40. The CSF biomarkers total tau (tTau), hyperphosphorylated tau (p-Tau), and the 42 amino acid form of A β (A β -42) are now established markers for AD (Zetterberg et al., 2003) and can identify AD in the early, MCI stage of the disease with high accuracy (Hansson et al., 2006). CSF of AD and MCI patients shows decreased values of Aβ-42 and increased total tau (tTau) or phosphorylated tau (pTau) (Ewers et al., 2007). Recent studies have been focused in investigating if the pattern of different length A^β peptides could enhance the diagnostic power. The levels of Aβ (1-37, -38, -39, -40, -42) gave a 91% sensitivity and a 64% specificity in predicting the development of AD form MCI during a follow-up study (Hoglund et al., 2008). While progressive increase of CSF tTau CSF concentrations was found along the progression of AD (Andersson et al., 2007), this is not the case for Aβ-42 levels (Stefani et al., 2006). Both Aβ and tau levels in CSF are influenced by other factors that participate in AD, such age and apoE. Aging and the presence of the APOE $\varepsilon 4$ allele accelerate the deposition of A β -42 in brain and decrease of CSF A β -42 levels (Peskind et al., 2006). Also, CSF ApoE levels in correlate with t-tau and 24Shydroxycholesterol (24S-OHC) in patients with cognitive disorders (Shafaati et al., 2007). The association between A β load in the brain and CSF A β reduction remains puzzling, but it has been suggested that aggregation of this peptide into oligomeric forms could result in concomitant reduction of soluble forms in CSF and in the brain,

and increased A β deposition in plaques. Also, the relationship between CSF biomarkers (A β -42, t-tau and p-Tau) and plaque and tangle burden in the brain is not clear. Although initial studies suggested a direct correlation between that CSF p-Tau and neurofibrillary pathology in AD patients (Buerger et al., 2006), recent reports have shown no association (Buerger et al., 2007; Engelborghs et al., 2007).

Both increased CSF tau and decreased A β is also seen in other neurodegenerative disorders (Jellinger et al., 2008). P-tau181 differentiates AD and LBD, whereas p-tau231 differentiates AD and FTD (Hampel et al., 2004). Recent studies have also shown associations between several A β peptides and specific dementias. For example, decreased A β -38 levels correlated with FTD (Bibl et al., 2007b) and A β -37 (Bibl et al., 2006) with LBD.

Neurofilament light unit has been suggested as a possible CSF biomarkers to distinguish VaD from AD (Petzold et al., 2007). CSF neurofilament protein levels may also help to discriminate between FTD and early onset AD (Petzold et al., 2007).

However, the combination of tau and A β measurements in CSF enhance the accuracy of the diagnosic power. CSF tau/A β -42 ratio predicts cognitive decline in cognitively intact older adults (Fagan et al., 2007) and in individuals with MCI (Hansson et al., 2007; Li et al., 2007). Moreover, grouping these biomarkers is today the only biochemical tool with enough sensitivity and specificity to distinguishing AD from other dementias in the early stages (Finehout et al., 2007; Wiltfang et al., 2005), LBD (Mollenhauer et al., 2006a) and PD patients with dementia (Mollenhauer et al., 2006b).

Several other CSF biomarkers for AD and other dementia disorders have been described (Table 1). Some of these molecules have been shown to play a role in AD pathology, like the β -secretase BACE1 (Ewers et al., 2008; Vassar et al., 1999), cholesterol carriers (Mollenhauer et al., 2006a), cholesterol metabolites (Bjorkhem et

al., 2009; Mateos et al., 2009), or antioxidant molecules (Akterin et al., 2006), while other have an unknown relation to the disease. Comparative proteomics or metabolomics of CSF detecting a large number of molecules may provide a further means to diagnose and assess neurodegenerative diseases. However some considerations need to be taken into account when designing future studies, .d. including the standardization of CSF extraction and the avoidance of factors like

Biomarkers in plasma

An ideal early detection of different types of dementia would require simple, noninvasive and non-expensive diagnostic tests. However, up to date, no validated peripheral diagnostic markers for the early and differential diagnosis of dementiacausing disorders are available. The structure of the blood brain barrier (BBB) limits the presence of potential biomarkers that are closely associated to brain pathogenesis to small molecules, lipophilic molecules, and molecules with specific transporters (Irizarry, 2004). Brain derived proteins and metabolites that pass into the plasma will also become markedly diluted in a biochemically complex medium. It is also possible the presence of high intra- and inter-person variability due to factors that could influence the levels of a potential biomarker. The majority of the efforts in finding plasma biomarkers for neurodegenerative disorders have been made in AD, starting more than a decade ago with the traditional approach of measuring one molecule closely related to the pathology of the disease, in this case beta-amyloid (Aβ) (Mayeux et al., 1999; Tamaoka et al., 1996; Vanderstichele et al., 2000). Plasma total A β or A β -42 are increased in cases of familial AD (Kosaka et al., 1997; Scheuner et al., 1996) and in Downs syndrome with amyloid precursor protein (APP) triplication (Schupf et al., 2001) but the results were not consistent with the diagnosis of sporadic AD (Assini et al., 2004; Fukumoto et al., 2003; Mayeux et al., 1999; Schupf et al., 2001; Tamaoka et al., 1996; Vanderstichele et al., 2000). Peripheral Aß is transported via receptor for advanced glycation end products (RAGE) across the BBB into the brain (Deane et al., 2004) and A β elimination from brain across the BBB by cell surface low-density lipoprotein receptor related protein-1 (LRP) (Sagare et al., 2007). On the other hand, APP is also produced by platelets and is thus an alternative source A β of exists in plasma.

Several studies have investigated plasma Aβ levels and most groups have found no significant differences between AD and control cases (Fukumoto et al., 2003; Kosaka

et al., 1997; Mayeux et al., 1999; Scheuner et al., 1996; Tamaoka et al., 1996; Vanderstichele et al., 2000). Similarly, other studies showed that elevated levels of A β -42 (van Oijen et al., 2006) low levels of A β -40 (Sundelof et al., 2008) or a reduced A β -42/A β -40 ratio in plasma (Graff-Radford et al., 2007) in aging could indicate the conversion from a normal cognitive status to MCI or AD. In contrast, others reported that reduced A β -42 plasma levels might reflect the evolution from normal aging or MCI to AD (Song et al., 2007), rather than a marker for neuropathological events occurring in the disease.

CSF A β levels do not correlate with plasma A β levels in individual patients (Mehta et al., 2001; Vanderstichele et al., 2000). As previously discussed, other sources of A β , such platelets, are contributing to the total A β present in plasma. Also, it has been shown that the concentration of A β in plasma is considerably influenced by simultaneous medication (Blasko et al., 2005), including insulin treatments (Kulstad et al., 2006).

Decreases in plasma A β have been shown in cerebral CAA, amyotrophic lateral sclerosis, and LBD (Jellinger et al., 2008), and a decrease in the ratio of plasma A β -38/-40 peptides in vascular dementia (VaD) (Bibl et al., 2007a).

Although not useful for diagnosis, plasma A β measurement could be potentially helpful in clinical studies to evaluate the pharmacological effects of modulators of APP processing, such as β - or γ -secretase inhibitors.

Plasma homocysteine has been shown to be directly related to $A\beta$ -40 levels, while the association with $A\beta$ -42 was not significant, suggesting that homocysteine is related to aging but not specifically to AD, but it could interact to affect AD risk and cognition in PD (Irizarry, 2004; Luchsinger et al., 2007). Elevated homocysteine is a risk factor for cardiovascular disease (Boushey et al., 1995) but its relation with AD risk is unknown.

Chronic inflammation with microglia activation is believed to play a central role in AD pathogenesis. Whether the accumulation of inflammatory markers within the brain is also reflected in serum or plasma is unclear because many of these proteins do not easily cross the BBB. Inflammatory molecules including C-reactive protein, interleukin (IL)-1 β , tumor necrosis factor- α , IL-6, IL-6 receptor complex, α 1-antichymotrypsin and transforming growth factor- β show inconsistent changes across studies, while other cytokines such as IL-12, interferon- α , and interferon- β remain unchanged (Teunissen et al., 2002).

Increasing evidence has suggested that cholesterol, or cholesterol metabolism, plays a role in some neurodegenerative disorders including AD. The relation between hypercholesterolemia and AD or MCI has been confirmed repeatedly (Evans et al., 2004; Kivipelto et al., 2002; Launer et al., 2001). The presence of the E4 isoform of the apolipoprotein E (apoE), the main cholesterol transporter in the body, is the major known risk factor for AD and has been associated with either risk or severity of several other dementia-causing disorders (Cedazo-Minguez, 2007). The presence of the APOE ε4 allele has been associated with less apoE protein in plasma (Schiele et al., 2000) However, there was no consistent association of plasma apoE levels and AD. Studies have documented elevated apoE levels (Taddei et al., 1997) no difference, (Panza et al., 2003) or reduced levels relative to controls (Siest et al., 2000). Blood lipids are modifiable by dietary or pharmacologic intervention, and the lipoprotein cholesterol profile is an established marker of the effects of cholesterollowering medications and the associated reduction in cardiac risk. Metabolites of cholesterol such as 24S-hydroxycholesterol (24S-OHC) and 27-hydroxycholesterol (27-OH) can pass the BBB, and a large amount of 27-OH is taken up from the blood, acting as an important link between extra-cerebral and intra-cerebral pools of cholesterol (Bjorkhem et al., 2009). The central origin of 24S-OHC and its relatively constant hepatic metabolism has led several investigators to examine if 24S-OHC

may be a marker of brain cholesterol balance. Patients with advanced AD have significantly reduced levels of 24S-OHC in plasma (Bretillon et al., 2000; Kolsch et al., 2000). In a recent work it was shown that plasma levels of 24S-OHC decrease in parallel with decreases in the volume of the caudate measured with MRI in patients with different stages of Huntington's disease (Teunissen et al., 2007). Very recent studies on patients with PD have reported a markedly decreased level of 24S-OHC in plasma, which is consistent with its possible utility as a marker of CNS neuronal mass (Bretillon et al., 2000). Although 24S-OHC is not a diagnostic marker in itself, it may be a modifiable risk factor. It has been suggested that the determination of 24S-OHC levels may complement MRI morphology as a valuable tool to follow neurodegenerative changes in the early stages of this disease.

Other proteins present in plasma with recently discovered associations to AD pathology have also been proposed as biomarkers, such ALZAS (ALZheimer ASsociated protein). Pilot studies in serum of patients with probable AD have detected an up to tenfold increase of ALZAS in patients with MCI, but only moderately increased titres in autopsy-confirmed AD (Jellinger et al., 2008).

Since the pathological processes in neurodegenerative disorders are multiple, the diagnostic accuracy may be further improved by combining several markers. Recent proteomic discovery of various plasma signaling proteins may allow the development of a simple, cost-effective test for AD (German et al., 2007; Ray et al., 2007). A multifactorial biomarker can be approached in two ways: a "knowledge-based" approach, incorporating "known" putative biomarkers (molecules with known association with the disease); or an unbiased survey of many hundreds or thousands of molecules. A few knowledge-based approaches have been attempted for AD, integrating selected molecules known to be involved in the disease (Teunissen and Scheltens, 2007). In one study a panel of 29 serum biomarkers for inflammation,

homocysteine metabolism, cholesterol metabolism, and brain specific proteins were evaluated. A model incorporating IL-6 receptor, cysteine, protein fraction α1 and cholesterol levels proved to be the best combination to discriminate AD from controls, although specificity to other cognitive disorders and PD was weaker (Teunissen and Scheltens, 2007). In another study, 120 different signaling proteins were evaluated and a model including 18 proteins was generated. This combination showed high accuracy (89%) to differentiate 42 AD from 39 control cases and in predicting those MCI patients that later converted to AD (Ray et al., 2007).

Unbiased approaches have been pursued to evaluate a broad range of proteins (proteomics), small molecule metabolites (metabolomics), or transcripts (transcriptomics) in blood. A proteomic study using 2D-PAGE identified more than 70 proteins that differed between AD and control cases (Ueno et al., 2000). In another proteomic case-control study the combination of several molecules gave a sensitivity of 56% and specificity of 80% (Hye et al., 2006). The complexity of serum and plasma, and difficulties in assay standardization make these approaches challenging, but the rapid evolution in technological and computational facilities will allow greater diagnostic accuracy and broader applicability of these methods.

GENETIC BIOMARKERS

Another approach to seek for biomarkers for neurodegenerative diseases is the transcriptional profiling of genes in peripheral samples. The potential use of bloodbased gene expression profiling in the diagnosis of brain disorders has been described by several independent groups (Burczynski and Dorner, 2006; Sharp et al., 2006). Expression profiling of whole blood RNA offers several advantages in deciphering aberrant patterns of gene regulation in neurodegenerative processes: it might predict much earlier changes than that measured by means of protein

alteration; the uniform chemical nature of RNA make transcriptome studies less of a challenge than both proteome and metabolome studies and it has been shown that peripheral blood cells shares significant gene expression similarities with central nervous system (CNS) tissues (Burczynski and Dorner, 2006; Scherzer et al., 2007). A supportive example of this is a recent study that showed that the Parkinson disease-linked α -synuclein gene was up regulated both in blood and in the substantia nigra of patients with Parkinson disease (Kalman et al., 2005). Some gene expression studies have been done for AD biomarker discovery using blood as the clinical sample. A global gene expression analysis on fibroblasts from 33 individuals (both healthy and demented mutation carriers as well as wild-type siblings) from three families segregating the swedish APP, artic APP and presenilin 1 H163Y mutations showed that mutation carriers share a common gene expression profile significantly different from that of their wild-type siblings (Nagasaka et al., 2005). This suggests that the disease process starts decades before the onset of cognitive decline, and that presymptomatic diagnosis of AD may be feasible in the near future. Another study of AD patients vs. controls identified a group of 20 candidate genes (from 3200 genes investigated) that showed an altered expression in AD (Kalman et al., 2005). Transcriptional profiling of Alzheimer blood mononuclear cells by microarray identified 19 up- and 136 downregulated genes comparing to controls (Maes et al., 2007). In a recent study, the expression pattern of 33 genes (including APP. Insulin degrading enzyme, Histone cluster 1, H3e (HIST1H3E) and Cannabinoid receptor 2 (CNR2)) was investigated in whole blood samples from a population clinically diagnosed with AD and from healthy controls. These 33 genes were chosen according to previous findings of gene expression profiling in postmortem AD brains (Grunblatt et al., 2007; Jacob et al., 2007). The profiling of these genes was carried our four times in a period of one year in correlation to AD and it was found that the expression of HIST1H3E was correlated in a negative

manner to the MMSE scores, while. the expression of CNR2 showed some tendency to positively correlate with the decreased MMSE score (Grunblatt et al., 2009). The promising results point to the fact that gene expression profiling might be a promising method to seek for biomarkers for early dementia. More studies done in a longitudinal large scale manner and including multiple considerations such as diagnosis, MMSE score, and medication are need in order to establish the sensitivity JUSC and the specificity of this diagnostic tool.

NEUROPHYSIOLOGICAL BIOMARKERS

In comparison with biochemical or imaging approaches little attention has been paid to the application of electroencephalograms, quantitative electroencephalography (qEEG), event-related potential (ERP), transcranial magnetic stimulation (TMS), and vagus nerve stimulation (VNS) as useful clinical markers of early disease detection or progression. A recent meta-analysis explored the diagnostic accuracy of spontaneous EEG in dementia in articles published between 1980 and 2008, concluding that this method was not sufficient for the initial diagnosis of MCI in routine clinical practice (Jelic and Kowalski, 2009)

Nonetheless, in parallel with the dramatic improvements in technology recent promising findings have been reported. Multiple different qEEG components, as well as ERP activity, are altered during the transitional stage between healthy aging and AD (Jackson and Snyder, 2008; Koenig et al., 2005). For example, abnormalities in P600 and N400, two ERP components, have been associated with the conversion from amnestic MCI to AD (Olichney et al., 2008). Recently, a novel VNS technique, vagus somatosensory-evoked potentials, was developed in an attempt to detect dementia at an early stage (Merrill et al., 2006).

The profiling of EEG, ERP or VNS markers for the diagnostic assessment of early

cognitive changes is important. These are simple and cost-effective methods that could help in the identification of patients with MCI and in the prediction of which individuals will ultimately progress from MCI to AD. This would allow the use of therapeutic interventions targeted to those persons at greatest risk for conversion to

NEUROIMAGING

Along the progression of the neuropathological processes, continuous morphological and functional changes occur in the brain, which can be detected and quantified by several brain imaging techniques. The most apparent aging-related brain change is an atrophy resulting from synaptic and neuronal loss. This atrophy is larger in individuals with dementia and often shows a regional variation depending on the form of dementia that the patient suffers (Small et al., 2008). However, there are also areas in which anatomical changes do not distinguish between these disorders.

Habitually, neuroimaging techniques are classified as either structural or functional, according to the principal information that they give. Structural imaging techniques as computed tomography (CT) or magnetic resonance imaging (MRI) are often done to rule out other causes of dementia, such as normal pressure hydrocephalus or some particular lesions like tumors. Also, they can serve to clarify the diagnosis by detecting areas with atrophy or vascular lesions. While CT provides good spatial resolution (the ability to distinguish two structures from each other as separate), MRI provides comparable resolution with far better contrast resolution (the ability to distinguish the differences between two arbitrarily similar but not identical tissues).

Functional imaging techniques, such PET or SPECT, provide information about brain structure as well, but their spatial resolution is lower than structural techniques. However, their strong side is that they can be used to measure parameters of brain metabolism, like regional cerebral blood flow or regional cerebral glucose metabolism. Importantly, these parameters are often impaired in AD and other dementias before the morphological changes occur. Therefore functional imaging techniques are more suitable for helping the physicians in an earlier diagnosis. The clinical use of available neuroimaging techniques is expanding. At the same time, new technologies are now undergoing rapid evolution and novel applications of imaging techniques are being explored.

Magnetic Resonance Imaging (MRI)

MRI has high-resolution and gives high-quality information on brain structure, including delineation of grey and white matters. Generally, patients with AD show atrophy of the parietal lobes and the hippocampus on MRI compared to controls (Jack et al., 1997) (Figure 1). Several structural MRI studies localize the pattern of the atrophy in early-onset AD to more posterior regions (precuneus and posterior cingulate) (Frisoni et al., 2007; Karas et al., 2007), amygdala (Krasuski et al., 1998), occipital lobes, corpus callosum and prominent posterior cortical involvement (Teipel et al., 2003). Atrophies in hippocampus and entorhinal cortex can predict memory progression (Mungas et al., 2005) and have shown to be associated with increased risk of developing AD (Apostolova et al., 2006). However, it has also been shown that these changes are not specific to AD (Rodrigue and Raz, 2004). In clinical practice, structural MRI can help to support clinical diagnosis, but is not sufficient for establishing a definitive diagnosis, as there is a substantial overlapping between atrophy associated with normal aging and several neurodegenerative disorders, including AD. Non-specific age-related changes in the white matter appear frequently in elderly patients with stroke or MCI, but are also common in healthy elderly individuals.

On the other hand, some studies have shown some discriminative diagnostic power using structural MRI. For example, patients with amnestic MCI who converted to AD showed larger atrophy in several areas including hippocampus and inferior and middle temporal gyri, compared with non-converters (Chetelat et al., 2005). With the progression of AD, the corpus callosum usually appears atrophic anteriorly, differentiating this disease from FTLD, in which the posterior area of the corpus callosum is more affected (Likeman et al., 2005).

It is expected that with the development of higher resolution volumetric MRI and new automated, computerized analytical programs (as voxel-based morphometry), the

capacity to identify subtle specific abnormality patterns in various forms of dementia will be greater, and the diagnostic utility of MRI improved.

Functional MRI (fMRI)

Functional MRI (fMRI) measures brain function over time during rest, or more commonly, in association with a cognitive task or stimulus that require the involvement of related brain areas and networks (Wagner et al., 1998). Blood Oxygen Level Dependent fMRI is the most common functional imaging method and measures alterations in blood flow associated with neuronal activity, which indirectly reflects brain activity. Currently, the high inter- and intra-individual variability of the signals and its inherent dependency of hemodynamics has precluded the use of this technique for differential diagnostic of dementias. However, fMRI could be useful to characterize the functional abnormalities specific to every disease. For example, in comparison to controls, individuals with AD show lowered brain activity in parietal and hippocampal regions and higher activity in primary cortices unaffected by the disease. Some studies found differential fMRI activation responses to a specific paradigms in MCI compared to healthy controls (Teipel et al., 2003). Recent advances in fMRI have helped to define intrinsic functional networks in the human brain (Fox and Raichle, 2007). The study of cognitive-behavioral functions in early phases of neurodegenerative disorders could identify the neuroanatomical networks affected by these diseases (Sperling, 2007) and it is possible that function-critical neural networks could be affected differentially by variant neurodegenerative diseases (Seeley et al., 2009). This novel perspective has opened new exciting possibilities for the use on fMRI in the differential diagnosis of dementia-causing disorders.

Positron Emission Tomography (PET) and Single Photon Emission Computed Tomography (SPECT)

Molecular imaging by PET or SPECT with radiopharmaceuticals permits quantitative evaluation of physiological functions, protein pharmacokinetics and distribution of receptors with high sensitivity. PET and SPECT have been extensively evaluated as diagnostic procedures for dementia, since they have demonstrated diagnostic and prognostic utility for evaluating patients with cognitive impairment and in distinguishing among primary neurodegenerative disorders.

Studies often use the radiolabelled glucose [18F]FDG-PET to measure cerebral glucose metabolism, which indirectly indicates synaptic activity. In AD, FDG-PET studies have shown distinctive symmetric cortical hypometabolism from posterior cingulate and parietal regions, spreading to the temporal and prefrontal cortice (Figure 1) (Pakrasi and O'Brien, 2005; Silverman et al., 2001). These alterations in AD have been shown to be distinctive from controls and from other types of dementia and to correlate with cognitive impairment in MCI patients (Small et al., 2008). In terms of diagnostic accuracy, PET assessment of glucose metabolism has been found to have high sensitivity (94%) but low specificity (73–78%) (Silverman et al., 2001). Similar specificity for diagnosis has been found with SPECT studying regional blood flow with Tc-hexamethylpropyleneamine oxime (O'Brien, 2007).

Several low-molecular-weight probes have been developed to measure $A\beta$ deposits with PET in vivo. The most studied is the Pittsburgh compound B (PIB) that shows significantly greater cortical binding in patients with AD compared with controls (Figure 1) (Klunk et al., 2004). A 2-year longitudinal follow-up showed PIB not significant binding increase in AD patients despite declines in glucose metabolism and cognitive function (Engler et al., 2006). PIB shows retention in patients with CAA, low binding with variable pattern in LBD, and absent binding in FTD (Rowe et al., 2007). PIB binding in MCI patients appeared bimodal, with approximately 50% of MCI patients showing high binding (like AD patients), and 50% low binding (as cognitive normal controls). Amyloid binding studies in vivo may serve also to better

define the role of $A\beta$ in AD. Recently, correlation of PIB binding with brain atrophy (determined by MRI) has been shown. Higher amyloid deposition was generally not associated with more severe gray matter atrophy, except in the medial temporal lobes (amygdala and hippocampus), suggesting that different brain areas have diverse susceptibility to amyloid toxicity, or that amyloid is nonessential for neurodegeneration (Frisoni et al., 2009).

New PET ligands, such as 2-(1-6-[(2-[F-18]fluoroethyl)(methyl)amino]-2naphthyl}ethylidene)malononitrile ([18F]FDDNP), are being developed to measure amyloid plaques and tau inclusions that may facilitate the investigation of this pathologies in living humans (Braskie et al., 2008). When comparing binding potential of PIB and [18F]FDDNP it was found low correlation between them, with differences in regional binding and discrepant findings in MCI, suggesting that they measure related, but different, characteristics of the disease (Tolboom et al., 2009).

CONCLUSION

The current biochemical and neuroimaging markers have very limited predictive value. Combining neuroimaging studies with the analysis of some CSF proteins increase diagnostic sensitivity and specificity, especially in AD, but the predictive significance is still narrowed to the final stages of the disease (Figure 2).

A major limitation in the field is that markers discovered by one group cannot be reproduced by other groups. Most of dementia-causing diseases are complex and heterogeneous, but it is also true that diagnostic criteria and analytical methodology varies among studies.

Standardization of methods seems critical to reducing inconsistency and increasing reliability. It is necessary to implement common protocols for sample preparation, experimental design and generation proteomics data. Global initiatives of standardization are of critical importance. A good example is a recent study coming from the Swedish Brain Power initiative. Mattsson et al (Mattsson et al., 2009) evaluated the utility of the classical CSF biomarkers for AD (A β 42, T-tau and P-tau) by including a large number of samples from several European centers. The study showed that levels A β 42, T-tau and P-tau in CSF could predict to some extent the outcome of individuals with MCI. However, the sensitivity and specificity of these markers were sufficient to be used for screening but not as an early diagnostic test. The inclusion of other molecules could improve the power of diagnostics.

An early detection of different types of dementia using non-invasive diagnostic tests is the ultimate dream. However, the potential use of proteomic analysis of serum or plasma faces increased difficulties in assay standardization, and would require additional multilateral efforts.

The rapid evolution of technological and computational advances will eventually permit the discovery of biomarkers for an early and differential diagnosis of dementia-causing disorders. Identifying the individual risk for having AD or related

disorders will allow the development of preventive / delaying strategies.

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FIGURE LEGENDS.

Figure 1. Representative examples of brain images (MRI, and PET) from age matched cognitive intact individuals and Alzheimer's disease patients. MRI pictures of brains from a control and an AD patient. AD brain shows (symmetric) hippocampus atrophy and enlargement of lateral ventricles. PET pictures of AD brains showed reduced glucose metabolism mainly in parietal

areas (determined with the radiolabelled glucose [18F]FDG) and increased cortical Aβ load (determined with the Pittsburgh compound B (PIB)).

Figure 2. Diagnostic and treatment possibilities along the evolution to Dementia.

REFERENCES

Abdi, F., Quinn, J.F., Jankovic, J., McIntosh, M., Leverenz, J.B., Peskind, E., Nixon, R., Nutt, J., Chung, K., Zabetian, C., Samii, A., Lin, M., Hattan, S., Pan, C., Wang, Y., Jin, J., Zhu, D., Li, G.J., Liu, Y., Waichunas, D., Montine, T.J., Zhang, J., 2006. Detection of biomarkers with a multiplex quantitative proteomic platform in cerebrospinal fluid of patients with neurodegenerative disorders. J Alzheimers Dis 9, 293-348.

Adak, S., Illouz, K., Gorman, W., Tandon, R., Zimmerman, E.A., Guariglia, R., Moore, M.M., Kaye, J.A., 2004. Predicting the rate of cognitive decline in aging and early Alzheimer disease. Neurology 63, 108-14.

Akterin, S., Cowburn, R.F., Miranda-Vizuete, A., Jimenez, A., Bogdanovic, N., Winblad, B., Cedazo-Minguez, A., 2006. Involvement of glutaredoxin-1 and thioredoxin-1 in beta-amyloid toxicity and Alzheimer's disease. Cell Death Differ 13, 1454-65.

Andersson, C., Blennow, K., Johansson, S.E., Almkvist, O., Engfeldt, P., Lindau, M., Eriksdotter-Jonhagen, M., 2007. Differential CSF biomarker levels in APOE-epsilon4-positive and -negative patients with memory impairment. Dement Geriatr Cogn Disord 23, 87-95.

Apostolova, L.G., Dutton, R.A., Dinov, I.D., Hayashi, K.M., Toga, A.W., Cummings, J.L., Thompson, P.M., 2006. Conversion of mild cognitive impairment to Alzheimer disease predicted by hippocampal atrophy maps. Arch Neurol 63, 693-9.

Assini, A., Cammarata, S., Vitali, A., Colucci, M., Giliberto, L., Borghi, R., Inglese, M.L., Volpe, S., Ratto, S., Dagna-Bricarelli, F., Baldo, C., Argusti, A., Odetti, P., Piccini, A., Tabaton, M., 2004. Plasma levels of amyloid beta-protein 42 are increased in women with mild cognitive impairment. Neurology 63, 828-31.

Bibl, M., Esselmann, H., Mollenhauer, B., Weniger, G., Welge, V., Liess, M., Lewczuk, P., Otto, M., Schulz, J.B., Trenkwalder, C., Kornhuber, J., Wiltfang, J., 2007a. Blood-based neurochemical diagnosis of vascular dementia: a pilot study. J Neurochem 103, 467-74.

Bibl, M., Mollenhauer, B., Esselmann, H., Lewczuk, P., Klafki, H.W., Sparbier, K., Smirnov, A., Cepek, L., Trenkwalder, C., Ruther, E., Kornhuber, J., Otto, M., Wiltfang, J., 2006. CSF amyloid-beta-peptides in Alzheimer's disease, dementia with Lewy bodies and Parkinson's disease dementia. Brain 129, 1177-87.

Bibl, M., Mollenhauer, B., Lewczuk, P., Esselmann, H., Wolf, S., Trenkwalder, C., Otto, M., Stiens, G., Ruther, E., Kornhuber, J., Wiltfang, J., 2007b. Validation of amyloid-beta peptides in CSF diagnosis of neurodegenerative dementias. Mol Psychiatry 12, 671-80.

Bjorkhem, I., Cedazo-Minguez, A., Leoni, V., Meaney, S., 2009. Oxysterols and neurodegenerative diseases. Mol Aspects Med 30, 171-179.

Blasko, I., Kemmler, G., Krampla, W., Jungwirth, S., Wichart, I., Jellinger, K., Tragl, K.H., Fischer, P., 2005. Plasma amyloid beta protein 42 in non-demented persons aged 75 years: effects of concomitant medication and medial temporal lobe atrophy. Neurobiol Aging 26, 1135-43.

Boushey, C.J., Beresford, S.A., Omenn, G.S., Motulsky, A.G., 1995. A quantitative assessment of plasma homocysteine as a risk factor for vascular disease. Probable benefits of increasing folic acid intakes. Jama 274, 1049-57.

Braskie, M.N., Klunder, A.D., Hayashi, K.M., Protas, H., Kepe, V., Miller, K.J., Huang, S.C., Barrio, J.R., Ercoli, L.M., Siddarth, P., Satyamurthy, N., Liu, J., Toga, A.W., Bookheimer, S.Y., Small, G.W., Thompson, P.M., 2008. Plaque and tangle imaging and cognition in normal aging and Alzheimer's disease. Neurobiol Aging.

Bretillon, L., Siden, A., Wahlund, L.O., Lutjohann, D., Minthon, L., Crisby, M., Hillert, J., Groth, C.G., Diczfalusy, U., Bjorkhem, I., 2000. Plasma levels of 24Shydroxycholesterol in patients with neurological diseases. Neurosci Lett 293, 87-90.

Brookmeyer, R., Gray, S., Kawas, C., 1998. Projections of Alzheimer's disease in the United States and the public health impact of delaying disease onset. Am J Public Health 88, 1337-42.

Buerger, K., Alafuzoff, I., Ewers, M., Pirttila, T., Zinkowski, R., Hampel, H., 2007. No correlation between CSF tau protein phosphorylated at threonine 181 with neocortical neurofibrillary pathology in Alzheimer's disease. Brain 130, e82.

Buerger, K., Ewers, M., Pirttila, T., Zinkowski, R., Alafuzoff, I., Teipel, S.J., DeBernardis, J., Kerkman, D., McCulloch, C., Soininen, H., Hampel, H., 2006. CSF phosphorylated tau protein correlates with neocortical neurofibrillary pathology in Alzheimer's disease. Brain 129, 3035-41.

Burczynski, M.E., Dorner, A.J., 2006. Transcriptional profiling of peripheral blood cells in clinical pharmacogenomic studies. Pharmacogenomics 7, 187-202.

Carrette, O., Demalte, I., Scherl, A., Yalkinoglu, O., Corthals, G., Burkhard, P., Hochstrasser, D.F., Sanchez, J.C., 2003. A panel of cerebrospinal fluid potential biomarkers for the diagnosis of Alzheimer's disease. Proteomics 3, 1486-94.

Castano, E.M., Roher, A.E., Esh, C.L., Kokjohn, T.A., Beach, T., 2006. Comparative proteomics of cerebrospinal fluid in neuropathologically-confirmed Alzheimer's disease and non-demented elderly subjects. Neurol Res 28, 155-63.

Cedazo-Minguez, A., 2007. Apolipoprotein E and Alzheimer's disease: molecular mechanisms and therapeutic opportunities. J Cell Mol Med 11, 1227-38.

Chetelat, G., Landeau, B., Eustache, F., Mezenge, F., Viader, F., de la Sayette, V., Desgranges, B., Baron, J.C., 2005. Using voxel-based morphometry to map the structural changes associated with rapid conversion in MCI: a longitudinal MRI study. Neuroimage 27, 934-46.

Davidsson, P., Westman-Brinkmalm, A., Nilsson, C.L., Lindbjer, M., Paulson, L., Andreasen, N., Sjogren, M., Blennow, K., 2002. Proteome analysis of cerebrospinal fluid proteins in Alzheimer patients. Neuroreport 13, 611-5.

de Jager, C.A., Budge, M.M., 2005. Stability and predictability of the classification of mild cognitive impairment as assessed by episodic memory test performance over time. Neurocase 11, 72-9.

de Leon, M.J., Convit, A., Wolf, O.T., Tarshish, C.Y., DeSanti, S., Rusinek, H., Tsui, W., Kandil, E., Scherer, A.J., Roche, A., Imossi, A., Thorn, E., Bobinski, M., Caraos, C., Lesbre, P., Schlyer, D., Poirier, J., Reisberg, B., Fowler, J., 2001. Prediction of cognitive decline in normal elderly subjects with 2-[(18)F]fluoro-2-deoxy-D-glucose/poitron-emission tomography (FDG/PET). Proc Natl Acad Sci U S A 98, 10966-71.

Deane, R., Wu, Z., Sagare, A., Davis, J., Du Yan, S., Hamm, K., Xu, F., Parisi, M., LaRue, B., Hu, H.W., Spijkers, P., Guo, H., Song, X., Lenting, P.J., Van Nostrand, W.E., Zlokovic, B.V., 2004. LRP/amyloid beta-peptide interaction mediates differential brain efflux of Abeta isoforms. Neuron 43, 333-44.

Engelborghs, S., Sleegers, K., Cras, P., Brouwers, N., Serneels, S., De Leenheir, E., Martin, J.J., Vanmechelen, E., Van Broeckhoven, C., De Deyn, P.P., 2007. No association of CSF biomarkers with APOEepsilon4, plaque and tangle burden in definite Alzheimer's disease. Brain 130, 2320-6.

Engler, H., Forsberg, A., Almkvist, O., Blomquist, G., Larsson, E., Savitcheva, I., Wall, A., Ringheim, A., Langstrom, B., Nordberg, A., 2006. Two-year follow-up of amyloid deposition in patients with Alzheimer's disease. Brain 129, 2856-66.

Evans, R.M., Hui, S., Perkins, A., Lahiri, D.K., Poirier, J., Farlow, M.R., 2004. Cholesterol and APOE genotype interact to influence Alzheimer disease progression. Neurology 62, 1869-71.

Ewers, M., Buerger, K., Teipel, S.J., Scheltens, P., Schroder, J., Zinkowski, R.P., Bouwman, F.H., Schonknecht, P., Schoonenboom, N.S., Andreasen, N., Wallin, A., DeBernardis, J.F., Kerkman, D.J., Heindl, B., Blennow, K., Hampel, H., 2007. Multicenter assessment of CSF-phosphorylated tau for the prediction of conversion of MCI. Neurology 69, 2205-12.

Ewers, M., Zhong, Z., Burger, K., Wallin, A., Blennow, K., Teipel, S.J., Shen, Y., Hampel, H., 2008. Increased CSF-BACE 1 activity is associated with ApoE-epsilon 4 genotype in subjects with mild cognitive impairment and Alzheimer's disease. Brain 131, 1252-8.

Fagan, A.M., Roe, C.M., Xiong, C., Mintun, M.A., Morris, J.C., Holtzman, D.M., 2007. Cerebrospinal fluid tau/beta-amyloid(42) ratio as a prediction of cognitive decline in nondemented older adults. Arch Neurol 64, 343-9.

Finehout, E.J., Franck, Z., Choe, L.H., Relkin, N., Lee, K.H., 2007. Cerebrospinal fluid proteomic biomarkers for Alzheimer's disease. Ann Neurol 61, 120-9.

Fox, M.D., Raichle, M.E., 2007. Spontaneous fluctuations in brain activity observed with functional magnetic resonance imaging. Nat Rev Neurosci 8, 700-11.

Frisoni, G.B., Lorenzi, M., Caroli, A., Kemppainen, N., Nagren, K., Rinne, J.O., 2009. In vivo mapping of amyloid toxicity in Alzheimer disease. Neurology 72, 1504-11.

Frisoni, G.B., Pievani, M., Testa, C., Sabattoli, F., Bresciani, L., Bonetti, M., Beltramello, A., Hayashi, K.M., Toga, A.W., Thompson, P.M., 2007. The topography of grey matter involvement in early and late onset Alzheimer's disease. Brain 130, 720-30.

Fukumoto, H., Tennis, M., Locascio, J.J., Hyman, B.T., Growdon, J.H., Irizarry, M.C., 2003. Age but not diagnosis is the main predictor of plasma amyloid beta-protein levels. Arch Neurol 60, 958-64.

German, D.C., Gurnani, P., Nandi, A., Garner, H.R., Fisher, W., Diaz-Arrastia, R., O'Suilleabhain, P., Rosenblatt, K.P., 2007. Serum biomarkers for Alzheimer's disease: proteomic discovery. Biomed Pharmacother 61, 383-9.

Graff-Radford, N.R., Crook, J.E., Lucas, J., Boeve, B.F., Knopman, D.S., Ivnik, R.J., Smith, G.E., Younkin, L.H., Petersen, R.C., Younkin, S.G., 2007. Association of low plasma Abeta42/Abeta40 ratios with increased imminent risk for mild cognitive impairment and Alzheimer disease. Arch Neurol 64, 354-62.

Grunblatt, E., Bartl, J., Zehetmayer, S., Ringel, T.M., Bauer, P., Riederer, P., Jacob, C.P., 2009. Gene expression as peripheral biomarkers for sporadic Alzheimer's disease. J Alzheimers Dis 16, 627-34.

Grunblatt, E., Zander, N., Bartl, J., Jie, L., Monoranu, C.M., Arzberger, T., Ravid, R., Roggendorf, W., Gerlach, M., Riederer, P., 2007. Comparison analysis of gene expression patterns between sporadic Alzheimer's and Parkinson's disease. J Alzheimers Dis 12, 291-311.

Hampel, H., Mitchell, A., Blennow, K., Frank, R.A., Brettschneider, S., Weller, L., Moller, H.J., 2004. Core biological marker candidates of Alzheimer's disease - perspectives for diagnosis, prediction of outcome and reflection of biological activity. J Neural Transm 111, 247-72.

Hansson, O., Zetterberg, H., Buchhave, P., Andreasson, U., Londos, E., Minthon, L., Blennow, K., 2007. Prediction of Alzheimer's disease using the CSF Abeta42/Abeta40 ratio in patients with mild cognitive impairment. Dement Geriatr Cogn Disord 23, 316-20.

Hansson, O., Zetterberg, H., Buchhave, P., Londos, E., Blennow, K., Minthon, L., 2006. Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: a follow-up study. Lancet Neurol 5, 228-34.

Hoglund, K., Hansson, O., Buchhave, P., Zetterberg, H., Lewczuk, P., Londos, E., Blennow, K., Minthon, L., Wiltfang, J., 2008. Prediction of Alzheimer's disease using

a cerebrospinal fluid pattern of C-terminally truncated beta-amyloid peptides. Neurodegener Dis 5, 268-76.

Hu, Y., Malone, J.P., Fagan, A.M., Townsend, R.R., Holtzman, D.M., 2005. Comparative proteomic analysis of intra- and interindividual variation in human cerebrospinal fluid. Mol Cell Proteomics 4, 2000-9.

Hye, A., Lynham, S., Thambisetty, M., Causevic, M., Campbell, J., Byers, H.L., Hooper, C., Rijsdijk, F., Tabrizi, S.J., Banner, S., Shaw, C.E., Foy, C., Poppe, M., Archer, N., Hamilton, G., Powell, J., Brown, R.G., Sham, P., Ward, M., Lovestone, S., 2006. Proteome-based plasma biomarkers for Alzheimer's disease. Brain 129, 3042-50.

Irizarry, M.C., 2004. Biomarkers of Alzheimer disease in plasma. NeuroRx 1, 226-34.

Jack, C.R., Jr., Lowe, V.J., Weigand, S.D., Wiste, H.J., Senjem, M.L., Knopman, D.S., Shiung, M.M., Gunter, J.L., Boeve, B.F., Kemp, B.J., Weiner, M., Petersen, R.C., 2009. Serial PIB and MRI in normal, mild cognitive impairment and Alzheimer's disease: implications for sequence of pathological events in Alzheimer's disease. Brain 132, 1355-65.

Jack, C.R., Jr., Petersen, R.C., Xu, Y.C., Waring, S.C., O'Brien, P.C., Tangalos, E.G., Smith, G.E., Ivnik, R.J., Kokmen, E., 1997. Medial temporal atrophy on MRI in normal aging and very mild Alzheimer's disease. Neurology 49, 786-94.

Jackson, C.E., Snyder, P.J., 2008. Electroencephalography and event-related potentials as biomarkers of mild cognitive impairment and mild Alzheimer's disease. Alzheimers Dement 4, \$137-43.

Jacob, C.P., Koutsilieri, E., Bartl, J., Neuen-Jacob, E., Arzberger, T., Zander, N., Ravid, R., Roggendorf, W., Riederer, P., Grunblatt, E., 2007. Alterations in expression of glutamatergic transporters and receptors in sporadic Alzheimer's disease. J Alzheimers Dis 11, 97-116.

Jelic, V., Kowalski, J.E., 2009. Evidence-based evaluation of diagnostic accuracy of resting EEG in dementia and mild cognitive impairment. Clin EEG and Neurosci 40, 1-14.

Jellinger, K.A., Janetzky, B., Attems, J., Kienzl, E., 2008. Biomarkers for early diagnosis of Alzheimer disease: 'ALZheimer ASsociated gene'--a new blood biomarker? J Cell Mol Med 12, 1094-117.

Jung, S.M., Lee, K., Lee, J.W., Namkoong, H., Kim, H.K., Kim, S., Na, H.R., Ha, S.A., Kim, J.R., Ko, J., Kim, J.W., 2008. Both plasma retinol-binding protein and haptoglobin precursor allele 1 in CSF: candidate biomarkers for the progression of normal to mild cognitive impairment to Alzheimer's disease. Neurosci Lett 436, 153-7.

Kalman, J., Kitajka, K., Pakaski, M., Zvara, A., Juhasz, A., Vincze, G., Janka, Z., Puskas, L.G., 2005. Gene expression profile analysis of lymphocytes from Alzheimer's patients. Psychiatr Genet 15, 1-6.

Karas, G., Scheltens, P., Rombouts, S., van Schijndel, R., Klein, M., Jones, B., van der Flier, W., Vrenken, H., Barkhof, F., 2007. Precuneus atrophy in early-onset Alzheimer's disease: a morphometric structural MRI study. Neuroradiology 49, 967-76.

Kivipelto, M., Helkala, E.L., Laakso, M.P., Hanninen, T., Hallikainen, M., Alhainen, K., Iivonen, S., Mannermaa, A., Tuomilehto, J., Nissinen, A., Soininen, H., 2002. Apolipoprotein E epsilon4 allele, elevated midlife total cholesterol level, and high midlife systolic blood pressure are independent risk factors for late-life Alzheimer disease. Ann Intern Med 137, 149-55.

Klunk, W.E., Engler, H., Nordberg, A., Wang, Y., Blomqvist, G., Holt, D.P., Bergstrom, M., Savitcheva, I., Huang, G.F., Estrada, S., Ausen, B., Debnath, M.L., Barletta, J., Price, J.C., Sandell, J., Lopresti, B.J., Wall, A., Koivisto, P., Antoni, G., Mathis, C.A., Langstrom, B., 2004. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. Ann Neurol 55, 306-19.

Koenig, T., Prichep, L., Dierks, T., Hubl, D., Wahlund, L.O., John, E.R., Jelic, V., 2005. Decreased EEG synchronization in Alzheimer's disease and mild cognitive impairment. Neurobiol Aging 26, 165-71.

Kolsch, H., Ludwig, M., Lutjohann, D., Prange, W., Rao, M.L., 2000. 7alpha-Hydroperoxycholesterol causes CNS neuronal cell death. Neurochem Int 36, 507-12.

Korolainen, M.A., Nyman, T.A., Nyyssonen, P., Hartikainen, E.S., Pirttila, T., 2007. Multiplexed proteomic analysis of oxidation and concentrations of cerebrospinal fluid proteins in Alzheimer disease. Clin Chem 53, 657-65.

Kosaka, T., Imagawa, M., Seki, K., Arai, H., Sasaki, H., Tsuji, S., Asami-Odaka, A., Fukushima, T., Imai, K., Iwatsubo, T., 1997. The beta APP717 Alzheimer mutation increases the percentage of plasma amyloid-beta protein ending at A beta42(43). Neurology 48, 741-5.

Krasuski, J.S., Alexander, G.E., Horwitz, B., Daly, E.M., Murphy, D.G., Rapoport, S.I., Schapiro, M.B., 1998. Volumes of medial temporal lobe structures in patients with Alzheimer's disease and mild cognitive impairment (and in healthy controls). Biol Psychiatry 43, 60-8.

Kulstad, J.J., Green, P.S., Cook, D.G., Watson, G.S., Reger, M.A., Baker, L.D., Plymate, S.R., Asthana, S., Rhoads, K., Mehta, P.D., Craft, S., 2006. Differential modulation of plasma beta-amyloid by insulin in patients with Alzheimer disease. Neurology 66, 1506-10.

Larrieu, S., Letenneur, L., Orgogozo, J.M., Fabrigoule, C., Amieva, H., Le Carret, N., Barberger-Gateau, P., Dartigues, J.F., 2002. Incidence and outcome of mild

cognitive impairment in a population-based prospective cohort. Neurology 59, 1594-9.

Launer, L.J., White, L.R., Petrovitch, H., Ross, G.W., Curb, J.D., 2001. Cholesterol and neuropathologic markers of AD: a population-based autopsy study. Neurology 57, 1447-52.

Leoni, V., Shafaati, M., Salomon, A., Kivipelto, M., Bjorkhem, I., Wahlund, L.O., 2006. Are the CSF levels of 24S-hydroxycholesterol a sensitive biomarker for mild cognitive impairment? Neurosci Lett 397, 83-7.

Li, G., Sokal, I., Quinn, J.F., Leverenz, J.B., Brodey, M., Schellenberg, G.D., Kaye, J.A., Raskind, M.A., Zhang, J., Peskind, E.R., Montine, T.J., 2007. CSF tau/Abeta42 ratio for increased risk of mild cognitive impairment: a follow-up study. Neurology 69, 631-9.

Likeman, M., Anderson, V.M., Stevens, J.M., Waldman, A.D., Godbolt, A.K., Frost, C., Rossor, M.N., Fox, N.C., 2005. Visual assessment of atrophy on magnetic resonance imaging in the diagnosis of pathologically confirmed young-onset dementias. Arch Neurol 62, 1410-5.

Luchsinger, J.A., Tang, M.X., Miller, J., Green, R., Mehta, P.D., Mayeux, R., 2007. Relation of plasma homocysteine to plasma amyloid beta levels. Neurochem Res 32, 775-81.

Maes, O.C., Xu, S., Yu, B., Chertkow, H.M., Wang, E., Schipper, H.M., 2007. Transcriptional profiling of Alzheimer blood mononuclear cells by microarray. Neurobiol Aging 28, 1795-809.

Mateos, L., Akterin, S., Gil-Bea, F.J., Spulber, S., Rahman, A., Bjorkhem, I., Schultzberg, M., Flores-Morales, A., Cedazo-Minguez, A., 2009. Activity-regulated cytoskeleton-associated protein in rodent brain is down-regulated by high fat diet in vivo and by 27-hydroxycholesterol in vitro. Brain Pathol 19, 69-80.

Mattsson, N., Zetterberg, H., Hansson, O., Andreasen, N., Parnetti, L., Jonsson, M., Herukka, S.K., van der Flier, W.M., Blankenstein, M.A., Ewers, M., Rich, K., Kaiser, E., Verbeek, M., Tsolaki, M., Mulugeta, E., Rosen, E., Aarsland, D., Visser, P.J., Schroder, J., Marcusson, J., de Leon, M., Hampel, H., Scheltens, P., Pirttila, T., Wallin, A., Jonhagen, M.E., Minthon, L., Winblad, B., Blennow, K., 2009. CSF biomarkers and incipient Alzheimer disease in patients with mild cognitive impairment. Jama 302, 385-93.

Mayeux, R., Tang, M.X., Jacobs, D.M., Manly, J., Bell, K., Merchant, C., Small, S.A., Stern, Y., Wisniewski, H.M., Mehta, P.D., 1999. Plasma amyloid beta-peptide 1-42 and incipient Alzheimer's disease. Ann Neurol 46, 412-6.

McKeith, I.G., Galasko, D., Kosaka, K., Perry, E.K., Dickson, D.W., Hansen, L.A., Salmon, D.P., Lowe, J., Mirra, S.S., Byrne, E.J., Lennox, G., Quinn, N.P., Edwardson, J.A., Ince, P.G., Bergeron, C., Burns, A., Miller, B.L., Lovestone, S., Collerton, D., Jansen, E.N., Ballard, C., de Vos, R.A., Wilcock, G.K., Jellinger, K.A.,

Perry, R.H., 1996. Consensus guidelines for the clinical and pathologic diagnosis of dementia with Lewy bodies (DLB): report of the consortium on DLB international workshop. Neurology 47, 1113-24.

Mehta, P.D., Pirttila, T., Patrick, B.A., Barshatzky, M., Mehta, S.P., 2001. Amyloid beta protein 1-40 and 1-42 levels in matched cerebrospinal fluid and plasma from patients with Alzheimer disease. Neurosci Lett 304, 102-6.

Merrill, C.A., Jonsson, M.A., Minthon, L., Ejnell, H., H, C.s.S., Blennow, K., Karlsson, M., Nordlund, A., Rolstad, S., Warkentin, S., Ben-Menachem, E., Sjogren, M.J., 2006. Vagus nerve stimulation in patients with Alzheimer's disease: Additional followup results of a pilot study through 1 year. J Clin Psychiatry 67, 1171-8.

Mollenhauer, B., Bibl, M., Wiltfang, J., Steinacker, P., Ciesielczyk, B., Neubert, K., Trenkwalder, C., Otto, M., 2006a. Total tau protein, phosphorylated tau (181p) protein, beta-amyloid(1-42), and beta-amyloid(1-40) in cerebrospinal fluid of patients with dementia with Lewy bodies. Clin Chem Lab Med 44, 192-5.

Mollenhauer, B., Trenkwalder, C., von Ahsen, N., Bibl, M., Steinacker, P., Brechlin, P., Schindehuette, J., Poser, S., Wiltfang, J., Otto, M., 2006b. Beta-amlyoid 1-42 and tau-protein in cerebrospinal fluid of patients with Parkinson's disease dementia. Dement Geriatr Cogn Disord 22, 200-8.

Mungas, D., Harvey, D., Reed, B.R., Jagust, W.J., DeCarli, C., Beckett, L., Mack, W.J., Kramer, J.H., Weiner, M.W., Schuff, N., Chui, H.C., 2005. Longitudinal volumetric MRI change and rate of cognitive decline. Neurology 65, 565-71.

Nagasaka, Y., Dillner, K., Ebise, H., Teramoto, R., Nakagawa, H., Lilius, L., Axelman, K., Forsell, C., Ito, A., Winblad, B., Kimura, T., Graff, C., 2005. A unique gene expression signature discriminates familial Alzheimer's disease mutation carriers from their wild-type siblings. Proc Natl Acad Sci U S A 102, 14854-9.

Neary, D., Snowden, J., Mann, D., 2005. Frontotemporal dementia. Lancet Neurol 4, 771-80.

O'Brien, J.T., 2007. Role of imaging techniques in the diagnosis of dementia. Br J Radiol 80 Spec No 2, S71-7.

Olichney, J.M., Taylor, J.R., Hillert, D.G., Chan, S.H., Salmon, D.P., Gatherwright, J., Iragui, V.J., Kutas, M., 2008. fMRI congruous word repetition effects reflect memory variability in normal elderly. Neurobiol Aging.

Pakrasi, S., O'Brien, J.T., 2005. Emission tomography in dementia. Nucl Med Commun 26, 189-96.

Palmer, K., Backman, L., Winblad, B., Fratiglioni, L., 2008. Mild cognitive impairment in the general population: occurrence and progression to Alzheimer disease. Am J Geriatr Psychiatry 16, 603-11.

Panza, F., Solfrizzi, V., Colacicco, A.M., Basile, A.M., D'Introno, A., Capurso, C., Sabba, M., Capurso, S., Capurso, A., 2003. Apolipoprotein E (APOE) polymorphism influences serum APOE levels in Alzheimer's disease patients and centenarians. Neuroreport 14, 605-8.

Peskind, E.R., Li, G., Shofer, J., Quinn, J.F., Kaye, J.A., Clark, C.M., Farlow, M.R., DeCarli, C., Raskind, M.A., Schellenberg, G.D., Lee, V.M., Galasko, D.R., 2006. Age and apolipoprotein E*4 allele effects on cerebrospinal fluid beta-amyloid 42 in adults with normal cognition. Arch Neurol 63, 936-9.

Petersen, R.C., Smith, G.E., Waring, S.C., Ivnik, R.J., Tangalos, E.G., Kokmen, E., 1999. Mild cognitive impairment: clinical characterization and outcome. Arch Neurol 56, 303-8.

Petzold, A., Keir, G., Warren, J., Fox, N., Rossor, M.N., 2007. A systematic review and meta-analysis of CSF neurofilament protein levels as biomarkers in dementia. Neurodegener Dis 4, 185-94.

Puchades, M., Hansson, S.F., Nilsson, C.L., Andreasen, N., Blennow, K., Davidsson, P., 2003. Proteomic studies of potential cerebrospinal fluid protein markers for Alzheimer's disease. Brain Res Mol Brain Res 118, 140-6.

Ray, S., Britschgi, M., Herbert, C., Takeda-Uchimura, Y., Boxer, A., Blennow, K., Friedman, L.F., Galasko, D.R., Jutel, M., Karydas, A., Kaye, J.A., Leszek, J., Miller, B.L., Minthon, L., Quinn, J.F., Rabinovici, G.D., Robinson, W.H., Sabbagh, M.N., So, Y.T., Sparks, D.L., Tabaton, M., Tinklenberg, J., Yesavage, J.A., Tibshirani, R., Wyss-Coray, T., 2007. Classification and prediction of clinical Alzheimer's diagnosis based on plasma signaling proteins. Nat Med 13, 1359-62.

Reiber, H., Peter, J.B., 2001. Cerebrospinal fluid analysis: disease-related data patterns and evaluation programs. J Neurol Sci 184, 101-22.

Rodrigue, K.M., Raz, N., 2004. Shrinkage of the entorhinal cortex over five years predicts memory performance in healthy adults. J Neurosci 24, 956-63.

Rowe, C.C., Ng, S., Ackermann, U., Gong, S.J., Pike, K., Savage, G., Cowie, T.F., Dickinson, K.L., Maruff, P., Darby, D., Smith, C., Woodward, M., Merory, J., Tochon-Danguy, H., O'Keefe, G., Klunk, W.E., Mathis, C.A., Price, J.C., Masters, C.L., Villemagne, V.L., 2007. Imaging beta-amyloid burden in aging and dementia. Neurology 68, 1718-25.

Sagare, A., Deane, R., Bell, R.D., Johnson, B., Hamm, K., Pendu, R., Marky, A., Lenting, P.J., Wu, Z., Zarcone, T., Goate, A., Mayo, K., Perlmutter, D., Coma, M., Zhong, Z., Zlokovic, B.V., 2007. Clearance of amyloid-beta by circulating lipoprotein receptors. Nat Med 13, 1029-31.

Scherzer, C.R., Eklund, A.C., Morse, L.J., Liao, Z., Locascio, J.J., Fefer, D., Schwarzschild, M.A., Schlossmacher, M.G., Hauser, M.A., Vance, J.M., Sudarsky, L.R., Standaert, D.G., Growdon, J.H., Jensen, R.V., Gullans, S.R., 2007. Molecular

markers of early Parkinson's disease based on gene expression in blood. Proc Natl Acad Sci U S A 104, 955-60.

Scheuner, D., Eckman, C., Jensen, M., Song, X., Citron, M., Suzuki, N., Bird, T.D., Hardy, J., Hutton, M., Kukull, W., Larson, E., Levy-Lahad, E., Viitanen, M., Peskind, E., Poorkaj, P., Schellenberg, G., Tanzi, R., Wasco, W., Lannfelt, L., Selkoe, D., Younkin, S., 1996. Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. Nat Med 2, 864-70.

Schiele, F., De Bacquer, D., Vincent-Viry, M., Beisiegel, U., Ehnholm, C., Evans, A., Kafatos, A., Martins, M.C., Sans, S., Sass, C., Visvikis, S., De Backer, G., Siest, G., 2000. Apolipoprotein E serum concentration and polymorphism in six European countries: the ApoEurope Project. Atherosclerosis 152, 475-88.

Schupf, N., Patel, B., Silverman, W., Zigman, W.B., Zhong, N., Tycko, B., Mehta, P.D., Mayeux, R., 2001. Elevated plasma amyloid beta-peptide 1-42 and onset of dementia in adults with Down syndrome. Neurosci Lett 301, 199-203.

Seeley, W.W., Crawford, R.K., Zhou, J., Miller, B.L., Greicius, M.D., 2009. Neurodegenerative diseases target large-scale human brain networks. Neuron 62, 42-52.

Shafaati, M., Solomon, A., Kivipelto, M., Bjorkhem, I., Leoni, V., 2007. Levels of ApoE in cerebrospinal fluid are correlated with Tau and 24S-hydroxycholesterol in patients with cognitive disorders. Neurosci Lett 425, 78-82.

Sharp, F.R., Xu, H., Lit, L., Walker, W., Apperson, M., Gilbert, D.L., Glauser, T.A., Wong, B., Hershey, A., Liu, D.Z., Pinter, J., Zhan, X., Liu, X., Ran, R., 2006. The future of genomic profiling of neurological diseases using blood. Arch Neurol 63, 1529-36.

Siest, G., Bertrand, P., Qin, B., Herbeth, B., Serot, J.M., Masana, L., Ribalta, J., Passmore, A.P., Evans, A., Ferrari, M., Franceschi, M., Shepherd, J., Cuchel, M., Beisiegel, U., Zuchowsky, K., Rukavina, A.S., Sertic, J., Stojanov, M., Kostic, V., Mitrevski, A., Petrova, V., Sass, C., Merched, A., Salonen, J.T., Tiret, L., Visvikis, S., 2000. Apolipoprotein E polymorphism and serum concentration in Alzheimer's disease in nine European centres: the ApoEurope study. ApoEurope group. Clin Chem Lab Med 38, 721-30.

Silverman, D.H., Small, G.W., Chang, C.Y., Lu, C.S., Kung De Aburto, M.A., Chen, W., Czernin, J., Rapoport, S.I., Pietrini, P., Alexander, G.E., Schapiro, M.B., Jagust, W.J., Hoffman, J.M., Welsh-Bohmer, K.A., Alavi, A., Clark, C.M., Salmon, E., de Leon, M.J., Mielke, R., Cummings, J.L., Kowell, A.P., Gambhir, S.S., Hoh, C.K., Phelps, M.E., 2001. Positron emission tomography in evaluation of dementia: Regional brain metabolism and long-term outcome. Jama 286, 2120-7.

Simonsen, A.H., McGuire, J., Hansson, O., Zetterberg, H., Podust, V.N., Davies, H.A., Waldemar, G., Minthon, L., Blennow, K., 2007. Novel panel of cerebrospinal fluid biomarkers for the prediction of progression to Alzheimer dementia in patients with mild cognitive impairment. Arch Neurol 64, 366-70.

Simonsen, A.H., McGuire, J., Podust, V.N., Davies, H., Minthon, L., Skoog, I., Andreasen, N., Wallin, A., Waldemar, G., Blennow, K., 2008. Identification of a novel panel of cerebrospinal fluid biomarkers for Alzheimer's disease. Neurobiol Aging 29, 961-8.

Small, G.W., Bookheimer, S.Y., Thompson, P.M., Cole, G.M., Huang, S.C., Kepe, V., Barrio, J.R., 2008. Current and future uses of neuroimaging for cognitively impaired patients. Lancet Neurol 7, 161-72.

Song, M.S., Mook-Jung, I., Lee, H.J., Min, J.Y., Park, M.H., 2007. Serum antiamyloid-beta antibodies and Alzheimer's disease in elderly Korean patients. J Int Med Res 35, 301-6.

Sperling, R., 2007. Functional MRI studies of associative encoding in normal aging, mild cognitive impairment, and Alzheimer's disease. Ann N Y Acad Sci 1097, 146-55.

Stefani, A., Martorana, A., Bernardini, S., Panella, M., Mercati, F., Orlacchio, A., Pierantozzi, M., 2006. CSF markers in Alzheimer disease patients are not related to the different degree of cognitive impairment. J Neurol Sci 251, 124-8.

Sundelof, J., Giedraitis, V., Irizarry, M.C., Sundstrom, J., Ingelsson, E., Ronnemaa, E., Arnlov, J., Gunnarsson, M.D., Hyman, B.T., Basun, H., Ingelsson, M., Lannfelt, L., Kilander, L., 2008. Plasma beta amyloid and the risk of Alzheimer disease and dementia in elderly men: a prospective, population-based cohort study. Arch Neurol 65, 256-63.

Taddei, K., Clarnette, R., Gandy, S.E., Martins, R.N., 1997. Increased plasma apolipoprotein E (apoE) levels in Alzheimer's disease. Neurosci Lett 223, 29-32.

Tamaoka, A., Fukushima, T., Sawamura, N., Ishikawa, K., Oguni, E., Komatsuzaki, Y., Shoji, S., 1996. Amyloid beta protein in plasma from patients with sporadic Alzheimer's disease. J Neurol Sci 141, 65-8.

Tarawneh, R., Galvin, J.E., 2007. Distinguishing Lewy body dementias from Alzheimer's disease. Expert Rev Neurother 7, 1499-516.

Teipel, S.J., Schapiro, M.B., Alexander, G.E., Krasuski, J.S., Horwitz, B., Hoehne, C., Moller, H.J., Rapoport, S.I., Hampel, H., 2003. Relation of corpus callosum and hippocampal size to age in nondemented adults with Down's syndrome. Am J Psychiatry 160, 1870-8.

Teunissen, C.E., de Vente, J., Steinbusch, H.W., De Bruijn, C., 2002. Biochemical markers related to Alzheimer's dementia in serum and cerebrospinal fluid. Neurobiol Aging 23, 485-508.

Teunissen, C.E., Floris, S., Sonke, M., Dijkstra, C.D., De Vries, H.E., Lutjohann, D., 2007. 24S-hydroxycholesterol in relation to disease manifestations of acute experimental autoimmune encephalomyelitis. J Neurosci Res 85, 1499-505.

Teunissen, C.E., Scheltens, P., 2007. Use of proteomic approaches to identify disease biomarkers. Lancet Neurol 6, 1036-7.

Tolboom, N., Yaqub, M., van der Flier, W.M., Boellaard, R., Luurtsema, G., Windhorst, A.D., Barkhof, F., Scheltens, P., Lammertsma, A.A., van Berckel, B.N., 2009. Detection of Alzheimer pathology in vivo using both 11C-PIB and 18F-FDDNP PET. J Nucl Med 50, 191-7.

Ueno, I., Sakai, T., Yamaoka, M., Yoshida, R., Tsugita, A., 2000. Analysis of blood plasma proteins in patients with Alzheimer's disease by two-dimensional electrophoresis, sequence homology and immunodetection. Electrophoresis 21, 1832-45.

van Oijen, M., Hofman, A., Soares, H.D., Koudstaal, P.J., Breteler, M.M., 2006. Plasma Abeta(1-40) and Abeta(1-42) and the risk of dementia: a prospective case-cohort study. Lancet Neurol 5, 655-60.

Vanderstichele, H., Van Kerschaver, E., Hesse, C., Davidsson, P., Buyse, M.A., Andreasen, N., Minthon, L., Wallin, A., Blennow, K., Vanmechelen, E., 2000. Standardization of measurement of beta-amyloid(1-42) in cerebrospinal fluid and plasma. Amyloid 7, 245-58.

Vassar, R., Bennett, B.D., Babu-Khan, S., Kahn, S., Mendiaz, E.A., Denis, P., Teplow, D.B., Ross, S., Amarante, P., Loeloff, R., Luo, Y., Fisher, S., Fuller, J., Edenson, S., Lile, J., Jarosinski, M.A., Biere, A.L., Curran, E., Burgess, T., Louis, J.C., Collins, F., Treanor, J., Rogers, G., Citron, M., 1999. Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. Science 286, 735-41.

Wagner, A.D., Schacter, D.L., Rotte, M., Koutstaal, W., Maril, A., Dale, A.M., Rosen, B.R., Buckner, R.L., 1998. Building memories: remembering and forgetting of verbal experiences as predicted by brain activity. Science 281, 1188-91.

Welander, H., Franberg, J., Graff, C., Sundstrom, E., Winblad, B., Tjernberg, L.O., 2009. Abeta43 is more frequent than Abeta40 in amyloid plaque cores from Alzheimer disease brains. J Neurochem.

Wiltfang, J., Lewczuk, P., Riederer, P., Grunblatt, E., Hock, C., Scheltens, P., Hampel, H., Vanderstichele, H., Iqbal, K., Galasko, D., Lannfelt, L., Otto, M., Esselmann, H., Henkel, A.W., Kornhuber, J., Blennow, K., 2005. Consensus paper of the WFSBP Task Force on Biological Markers of Dementia: the role of CSF and blood analysis in the early and differential diagnosis of dementia. World J Biol Psychiatry 6, 69-84.

Winblad, B., Palmer, K., Kivipelto, M., Jelic, V., Fratiglioni, L., Wahlund, L.O., Nordberg, A., Backman, L., Albert, M., Almkvist, O., Arai, H., Basun, H., Blennow, K., de Leon, M., DeCarli, C., Erkinjuntti, T., Giacobini, E., Graff, C., Hardy, J., Jack, C., Jorm, A., Ritchie, K., van Duijn, C., Visser, P., Petersen, R.C., 2004. Mild cognitive impairment--beyond controversies, towards a consensus: report of the International Working Group on Mild Cognitive Impairment. J Intern Med 256, 240-6.

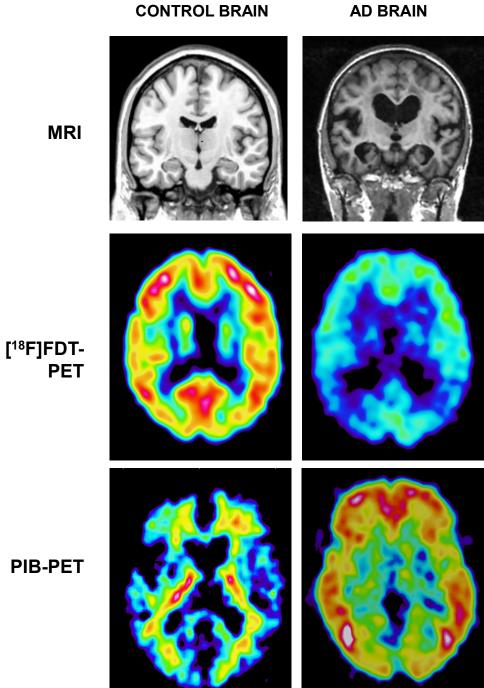
Zetterberg, H., Wahlund, L.O., Blennow, K., 2003. Cerebrospinal fluid markers for prediction of Alzheimer's disease. Neurosci Lett 352, 67-9.

Zhang, J., Goodlett, D.R., Montine, T.J., 2005. Proteomic biomarker discovery in cerebrospinal fluid for neurodegenerative diseases. J Alzheimers Dis 8, 377-86.

v, h stase (E ,airment, / Zhong, Z., Ewers, M., Teipel, S., Burger, K., Wallin, A., Blennow, K., He, P., McAllister, C., Hampel, H., Shen, Y., 2007. Levels of beta-secretase (BACE1) in cerebrospinal fluid as a predictor of risk in mild cognitive impairment. Arch Gen

PROTEIN	Up / Downregulated and references
Albumin	(Abdi et al., 2006; Zhang et al., 2005)
Amyloid β A4 protein	(Abdi et al., 2006; Simonsen et al., 2007; Zhang et al., 2005)
Angiotensinogen	↑(Zhang et al., 2005)
Apolipoprotein Al	(Castano et al., 2006; Davidsson et al., 2002;
	Puchades et al., 2003)↓
Apolipoprotein All	↑(Abdi et al., 2006; Zhang et al., 2005)
Apolipoprotein E	(Davidsson et al., 2002; Puchades et al.,
	2003)↓
	↑(Finehout et al., 2007)
BACE 1	(Ewers et al., 2008; Zhong et al., 2007)
24S-Hydroxycholesterol	\uparrow (Leoni et al., 2006) \uparrow (Simonage et al., 2007; Simonage et al., 2008)
C3a C4a	↑(Simonsen et al., 2007; Simonsen et al., 2008) ↑(Abdi et al., 2006; Simonsen et al., 2007;
048	Zhang et al., 2005)
Cystatin C	↑(Carrette et al., 2003; Hu et al., 2005)
Cystatin C, 8 amino acid N-terminal	\uparrow (Simonsen et al., 2007; Simonsen et al., 2008)
truncation	
Immunoglobulin heavy chain	↓(Abdi et al., 2006; Zhang et al., 2005)
Leucine-rich repeat-containing protein 4B	↑(Abdi et al., 2006; Zhang et al., 2005)
Leucine-rich repeat-containg protein 4B	↓(Abdi et al., 2006; Zhang et al., 2005)
N-acetyllactosamine	↓(Abdi et al., 2006; Zhang et al., 2005)
Neuronal pentraxin-1	(Abdi et al., 2006; Finehout et al., 2007;
	Zhang et al., 2005)
Prostaglandin-H2 D-isomerase	\downarrow (Korolainen et al., 2007; Puchades et al., 2002)
	2003) *(Hu at al. 2005)
Retinol-binding protein	↑(Hu et al., 2005) ↑(Abdi et al., 2006; Davidsson et al., 2002;
realition billianing protein	Zhang et al., 2005)
	\downarrow (Jung et al., 2008; Puchades et al., 2003)
Thioredoxin	(Abdi et al., 2006; Hu et al., 2005)
Transthyretin	↓ (Castano et al., 2006; Korolainen et al., 2007;
	Puchades et al., 2003)
	↑(Abdi et al., 2006; Davidsson et al., 2002;
	Finehout et al., 2007; Zhang et al., 2005)
VGF	\downarrow (Carrette et al., 2003; Simonsen et al., 2007;
a 1 Antitrumain	Simonsen et al., 2008)
α-1-Antitrypsin	(Abdi et al., 2006; Finehout et al., 2007;
	Zhang et al., 2005) ↓(Puchades et al., 2003)
α-1β Glycoprotein	(Fuchades et al., 2003) ↑(Hu et al., 2005; Puchades et al., 2003)
α-2HS Glycoprotein	(Thang et al., 2005)
	\downarrow (Puchades et al., 2003)
β Fibrinogen	(Abdi et al., 2006; Finehout et al., 2007)
β-2-Microglobulin	(Abdi et al., 2006; Carrette et al., 2003;
	Davidsson et al., 2002; Hu et al., 2005;
	Simonsen et al., 2007)
	\downarrow (Puchades et al., 2003; Simonsen et al.,
	2008)

TABLE 1: Molecules that showed differential levels in CSF from Alzheimer's Disease patients as compared to controls.



PIB-PET

