



Increased porcine circovirus type 2 replication in porcine leukocytes and by concanavalin A stimulation

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25 Abstract

Previously, it was shown that modulation of the immune system enhances porcine circovirus type 2 (PCV2) replication in pigs. In the present study, the effect of the mitogen concanavalin A (ConA) on PCV2 replication was investigated. Since ConA induces T-lymphocyte activation and initiates the production of interferon-gamma (IFN- γ), a cytokine that enhances PCV2 replication in porcine epithelial and monocytic cell lines *in vitro*, it was examined if the effects observed with ConA were mediated by IFN- γ . In an *in vitro* study, ConA but not IFN- γ enhanced PCV2 replication in peripheral blood mononuclear cells (PBMC). Up to 2.08 % and 0.96 % of PBMC were antigen positive for PCV2 strains 1121 and Stoon-1010 respectively, and a low virus production was observed. PCV2-infected PBMC were identified as CD4⁺ (40 %), CD8⁺ (54 %) and IgM⁺ (11 %). In a subsequent *in vivo* study, caesarean-derived colostrum-deprived piglets were injected with ConA or IFN- γ 12 hours before inoculation and every 3 days for 9 days after inoculation with strain 1121. PCV2 was isolated from inguinal lymph node biopsies from 10 days post inoculation (dpi) in ConA-treated pigs and from 15 dpi in non-treated and IFN- γ -treated pigs. ConA increased PCV2 replication levels, but disease was not observed. Half of the ConA-treated and IFN- γ -treated pigs showed a delayed humoral immune response, but this delay did not result in increased PCV2 replication in these pigs. These experiments demonstrated that ConA enhances PCV2 replication in PBMC *in vitro* and in lymphoid tissues *in vivo*.

Keywords: porcine circovirus type 2; viral replication; concanavalin A; interferon-gamma

1. Introduction

Porcine circovirus type 2 (PCV2) is a small, non-enveloped, single-stranded DNA virus. It is a member of the family of the *Circoviridae* and it is widespread in wild and domestic pigs. In pig dense regions, PCV2 infects all conventional pigs after weaning. The majority of these infections are sub-clinical. Under certain conditions, PCV2 infection causes post-weaning multisystemic wasting syndrome (PMWS), a multifactorial disease that causes severe growth retardation, weight loss and death in weaned piglets (Segales and Domingo, 2002). PMWS is characterized by severe lymphocytic depletion and monocytic infiltration in lymphoid tissues and high amounts of PCV2 are found within these lesions (Segales and Domingo, 2002).

PCV2 lacks its own polymerases and therefore fully depends on cellular enzymes to complete its infectious cycle. Up till now, only 4 different viral proteins have been described in porcine circovirus-infected cells (Mankertz and Hillenbrand, 2002; Liu et al., 2005), which makes PCV2 incapable of actively modifying the immune response of its host in its own favour, as other more complex viruses do (Tortorella et al., 2000). Therefore, specific conditions in the host intensively influence PCV2 replication. Co-infections with other infectious agents such as porcine parvovirus (PPV) (Allan et al., 1999), porcine reproductive and respiratory syndrome virus (PRRSV) (Allan et al., 2000) and *Mycoplasma hyopneumoniae* (Opriessnig et al., 2004) enhance PCV2 replication and exacerbate the clinical outcome of a PCV2 infection. Injection of immune stimulating drugs such as the soluble protein antigen Keyhole limpet hemocyanin (KLH) in incomplete Freund's adjuvant (Krakowka et

75 al., 2001) and vaccination (Opriessnig et al., 2003) enhance PCV2 replication as well, suggesting that the induction of an immune response might be a key factor in the induction of high levels of PCV2 replication. However, strategies involving immune-stimulating drugs or vaccination have not always been successful in enhancing PCV2 replication and PMWS occurrence (Resendes et al., 2004).

80 PCV2 replication in lymphoid tissues occurs in B-lymphocytes, T-lymphocytes, monocytes and macrophages (Sanchez et al., 2004). Recently, it was shown that the plant lectin concanavalin A (ConA) is able to induce PCV2 replication in CD3⁺ peripheral blood mononuclear cells (PBMC) that have been inoculated with PCV2 *in vitro* (Yu et al., 2007). Mitogen stimulation also induces PPV replication in porcine
85 peripheral blood lymphocytes (Paul et al., 1979) and TT virus replication in human PBMC (Mariscal et al., 2002). PPV and TT virus are also non-enveloped, single-stranded DNA viruses. The mitogen ConA causes predominantly T-lymphocyte proliferation and activation (Mosmann et al., 1986), leading to the production of lymphokines. More specifically, ConA induces production of high concentrations of
90 interferon-gamma (IFN- γ) *in vitro* (Verfaillie et al., 2001) and *in vivo* (Miyagi et al., 2004).

Recently, it was shown that interferon-alpha (IFN- α) and especially IFN- γ are able to enhance PCV2 replication in cell lines *in vitro*. When administered after PCV2 inoculation, 1,000 units of IFN- γ / 10⁶ cells increased the number of PCV2-infected
95 cells in epithelial PK-15 and monocytic 3D4/31 cells with 691 % and 423 % respectively (Meerts et al., 2005b). However, an *in vivo* study demonstrated that pigs with high IFN- γ mRNA expression levels in PBMC are less susceptible to PCV2 replication (Meerts et al., 2005c). Up till now, the effects of ConA and IFN- γ on PCV2 replication *in vivo* are not known.

100 It is the aim of the present study to determine the effects of ConA and IFN- γ on PCV2
replication in PBMC *in vitro* and to try to set up a model for the reproduction of
PMWS using ConA and IFN- γ *in vivo*.

105 2. Material and methods

2.1. Cells

PCV negative PK-15 cells were grown in minimal essential medium (MEM)
110 containing Earle's salts (Gibco, Grand Island, USA), supplemented with 5 % foetal
bovine serum (FBS), 0.3 mg ml⁻¹ glutamine, 100 U ml⁻¹ penicillin, 0.1 mg ml⁻¹
streptomycin and 0.1 mg ml⁻¹ kanamycin.

PBMC were separated from blood by density centrifugation at 750 x g on Ficoll-
Paque[®] (Amersham Pharmacia Biotech AB, Uppsala, Sweden). These cells were
115 maintained in RPMI 1640 (Gibco) containing 10 % FBS, 0.3 mg ml⁻¹ glutamine, 100
U ml⁻¹ penicillin, 0.1 mg ml⁻¹ streptomycin, 0.1 mg ml⁻¹ kanamycin, 1 % non-
essential amino acids (100x; Gibco) and 1 % sodium pyruvate (100 mM; Gibco).

Cells were maintained at 37 °C in the presence of 5 % CO₂.

120 2.2. Viruses

PCV2 strains 1121 and Stoon-1010 were used in this study. The replication kinetics
of these 2 strains have been described in PK-15 cells, foetal cardiomyocytes and
alveolar macrophages by Meerts et al. (2005a). For *in vitro* use, 1121 and Stoon-1010

125 were at a 29th and 19th passage level on PK-15 cells respectively. For *in vivo*
inoculation, a 4th passage on PK-15 cells of strain 1121 was used.

2.3. Concanavalin A and recombinant IFN- γ

130 ConA was purchased from Sigma (Heidelberg, Germany). Recombinant IFN- γ (rIFN-
 γ) was purchased from R&D Systems (Abingdon, UK).

2.4. Pigs

135 Six nine-month-old conventional PCV2 seronegative pigs were used as blood donors
for the isolation of PBMC.

Seventeen caesarean-derived colostrum-deprived (CD/CD) pigs were obtained from 2
Landrace sows and were raised in sterile conditions in individual isolators.

140 2.5. *In vitro* experiments

2.5.1. Inoculation and treatment

After isolation, PBMC were inoculated with a dose of $10^{4.3}$ TCID₅₀ / 10^6 PBMC for 1
145 h at 37 °C. After inoculation, cells were washed twice, resuspended in fresh medium
and seeded in 24-well cell culture plates (Nunc, Roskilde, Denmark) at a
concentration of 10^6 PBMC ml⁻¹. Immediately after seeding, ConA or rIFN- γ was
added at a dose of 5 μ g / 10^6 PBMC or 1,000 units / 10^6 PBMC respectively. A dose
of 5 μ g ConA / 10^6 PBMC is commonly used to induce proliferation in porcine

150 PBMC (Verfaillie et al., 2001). A dose of 1,000 units of rIFN- γ / 10^6 cells was the most efficient dose to increase the number of PCV2-infected cells in PK-15 and 3D4/31 cells (Meerts et al., 2005b). This dose was effective for stimulating PBMC *in vitro*, because 24 hours after treatment, a 5-fold increase in interleukin-12 levels was observed in supernatant fluids of rIFN- γ -treated PBMC when compared to non-treated
 155 PBMC. This was measured with a commercial ELISA for interleukin-12 (R&D Systems).

PBMC inoculated with PK-15 cell culture medium and treated with ConA or rIFN- γ were included as negative controls.

160 2.5.2. Fixation and stainings

At 0, 12, 24, 36, 48, 72 and 96 hours post inoculation (hpi), PBMC and supernatant fluids were collected. Cells were smeared onto glass slides, fixed in methanol at -20 °C for 10 min and stored at -20 °C until use. Supernatant fluids were centrifuged at
 165 2,900 x g for 10 min and stored at -20 °C until virus titration on PK-15 cells.

Expression kinetics of the non-structural Rep protein and the structural capsid protein were determined with double immunofluorescence stainings using the monoclonal antibody (mAb) F210, directed against the PCV2 Rep protein (McNeilly et al., 2001), and biotinylated purified porcine polyclonal antibodies (pAbs), directed against the
 170 PCV2 capsid protein (Meerts et al., 2005a). Briefly, fixed cell smears were air-dried at room temperature for 10 min. Next, they were incubated with an optimal dilution of mAb F210 in phosphate-buffered saline (PBS). Subsequently, a 1 : 500 dilution of fluorescein isothiocyanate (FITC)-labelled goat anti-mouse pAbs (Molecular Probes, Eugene, USA) in PBS was applied. This was followed by an optimal dilution of

175 biotinylated purified porcine pAbs in PBS, followed by incubation with a 1 : 100
 dilution of Texas Red-labelled streptavidin (Molecular Probes) in PBS. Each of these
 incubations were performed for 1 h at 37 °C. Finally, Hoechst 33342 (Molecular
 Probes) was applied at a concentration of 10 µg ml⁻¹ for 10 min at room temperature,
 in order to visualize the nucleus. Cell smears were washed three times with PBS
 180 between the incubations. Stained cell smears were mounted with a glycerol solution
 containing 1,4-diazabicyclo(2.2.2)octane (DABCO) anti-fading agent (Janssen
 Chimica, Beerse, Belgium). A LEICA DM/RBE fluorescence microscope (Leica
 Microsystems GmbH, Heidelberg, Germany) was used for visualization. Expression
 kinetics of the Rep protein and the capsid protein were assessed at 0, 12, 24, 36, 48,
 185 72 and 96 hpi, by determination of nuclear and cytoplasmic localization of PCV2
 antigens in 5,000 PBMC at each time point.

PCV2 positive PBMC (strain 1121) were further characterized at 0, 36 and 72 hpi by
 using mAbs directed against cell markers CD4, CD8, IgM and SWC3, respectively.
 The cell marker CD4 is present on T-helper and memory T-helper lymphocytes,
 190 regulatory T-lymphocytes and natural interferon-alpha producing cells (Pescovitz et
 al., 1984; Charley and Lavenant, 1990). CD8 is present on cytotoxic and gamma-delta
 T-lymphocytes, memory T-lymphocytes and natural killer cells (Pescovitz et al.,
 1984; Saamuller et al., 1994). IgM is present on naïve B-lymphocytes, B-
 lymphoblasts and IgM-producing plasma cells (Van Zaane and Hulst, 1987). SWC3 is
 195 present on monocytes, macrophages, dendritic cells and granulocytes (Pescovitz et al.,
 1984; Paillot et al., 2001). The staining procedure was similar to the technique
 described above. Cell smears were incubated with optimal dilutions of mAbs against
 CD4, CD8, IgM or SWC3, respectively, followed by FITC-labelled goat anti-mouse
 pAbs. Thereafter, cell smears were stained with biotin-conjugated porcine anti-PCV2

200 pAbs, followed by Texas Red-labelled streptavidin. On each cell smear, all PCV2
positive cells were analyzed by fluorescence microscopy.

2.5.3. Virus titration of PBMC culture supernatant

205 Ten-fold dilutions of PBMC culture supernatants were inoculated on semi-confluent
monolayers of PCV negative PK-15 cells. After 72 hours of incubation, cells were
fixed in 4 % paraformaldehyde in PBS. PCV2-infected cells were detected via an
immunoperoxidase monolayer assay (IPMA) as described by Labarque et al. (2000).

210 2.5.4. Statistical analysis

The statistical significance of differences between non-treated PBMC and ConA-
treated PBMC and between non-treated PBMC and rIFN- γ -treated PBMC was tested
by applying two-sided Kruskal-Wallis tests. Differences were considered significant
when $p < 0.05$. Statistical analyses were performed using S-PLUS (S-PLUS 6.1,
215 Insightful Corporation, Seattle, USA, 2002).

2.6. *In vivo experiments*

2.6.1. Inoculation and treatment

220

The set up of the experiment is illustrated in Table 1. The 17 CD/CD pigs were
randomly divided in 5 groups. At 19 days of age, all pigs from groups A (PCV2/-), B
(PCV2/ConA) and C (PCV2/rIFN- γ) were inoculated intraperitoneally and oronasally
with a total dose of $10^{4.3}$ TCID₅₀ of strain 1121. Pigs of groups D (-/ConA) and E (-

225 /rIFN- γ) were mock inoculated with equal volumes of cell culture medium. Treatment regimes with ConA or rIFN- γ were designed based on the results of a preliminary experiment in conventional pigs. In PCV2-inoculated PK-15 cells, addition of plasma of a ConA-treated conventional pig or a rIFN- γ -treated conventional pig increased the number of infected cells with maximum 524 % and 154 % respectively, when compared with plasma samples from the same pigs before ConA or rIFN- γ treatment. As a result from this experiment, pigs from groups B (PCV2/ConA) and D (-/ConA) were injected every 3 days with ConA at a dose of 1.5 mg kg⁻¹ body weight. Half of the dose was injected intramuscularly (IM) in the neck and the other half was injected subcutaneously (SC) in the region of the lower abdomen that is drained by the inguinal lymph nodes. The first injection was administered 12 h before PCV2 inoculation of the pigs and repeated at 2, 5 and 8 days post inoculation (dpi). Pigs from groups C (PCV2/rIFN- γ) and E (-/rIFN- γ) were treated at the same time points with rIFN- γ at a dose of 100,000 units (3 μ g of recombinant protein) per treatment: 50,000 units were injected IM in the neck and 50,000 units SC in the lower abdomen. Pigs were monitored for a period of 21 days after PCV2 inoculation and humanly euthanized afterwards, by intravenous injection of a barbiturate overdose (Natriumpentobarbital 20 %, Kela, Hoogstraten, Belgium).

2.6.2. Clinical monitoring

245 Pigs were monitored daily for the appearance of clinical signs that are typically associated with PMWS. These clinical signs include cachexia, depression, respiratory distress and jaundice (Segales and Domingo, 2002).

250 2.6.3. PCV2 replication

To follow the degree of PCV2 replication in all individual pigs, inguinal lymph node biopsies were collected at different time points as described before (Meerts et al., 2005c). At 10 dpi, a biopsy was taken from the left inguinal lymph node and at 15 dpi
 255 a biopsy was taken from the right inguinal lymph node. In order to take lymph node biopsies, the pigs were anaesthetized by IM injection of 2.2 mg tiletamine and 2.2 mg zolazepam (Zoletil[®], Virbac, Wavre, Belgium) dissolved in 0.22 ml 2 % xylazine (XYL-M[®] 2 %, VMD, Arendonk, Belgium) kg⁻¹ body weight. An incision was made in the overlaying skin and half the inguinal lymph node was excised. The wound was
 260 closed by mersilene sutures (Mersutures[®], Ethicon, USA). At the time of euthanasia (21 dpi) the remaining parts of both left and right inguinal lymph nodes were collected.

Ten % suspensions were made from these biopsies and from the remaining parts of these lymph nodes, and the level of PCV2 replication in these organs was determined
 265 by PCV2 titration on PK-15 cells. The detection limit of this technique was 10^{2.7} TCID₅₀ / g.

Quantification and immunophenotyping of PCV2 positive cells was performed on inguinal lymph node biopsies that were collected at 15 dpi. Serial cryostat sections were fixed in methanol at -20 °C for 10 min. Double immunofluorescence stainings
 270 for CD4, CD8, IgM or SWC3, respectively, and for PCV2, were performed as described above. Prior to mounting, autofluorescence was quenched by immersion of the sections for 30 min in a 10 mM dilution of CuSO₄ in a 50 mM NH₄-acetate buffer at pH 5.0. PCV2 positive cells were quantified and immunophenotyped using a fluorescence microscope. The total number of PCV2 positive cells and the total

275 number of double positive cells (cell marker and PCV2) were determined over an area
of 10 mm² of tissue.

2.6.4. Humoral immune response against PCV2

280 At 0, 7, 10, 15 and 21 dpi, blood was taken (1/1 in Alsevers solution) from the jugular
vein, in order to monitor total anti-PCV2 Ab titres by an IPMA described previously
(Labarque et al., 2000). PCV2 neutralizing Abs were determined by a sensitive
neutralization assay described previously (Meerts et al., 2005c).

285 The animal experiments described in this study were authorized and supervised by the
Ethical and Animal Welfare Committee of the Faculty of Veterinary Medicine of
Ghent University.

2.6.5 Statistical analysis

290

The statistical significance of differences between non-treated pigs and ConA-treated
pigs and between non-treated pigs and rIFN- γ -treated pigs was tested by applying
two-sided Kruskal-Wallis tests. Differences were considered significant when $p <$
0.05. Statistical analyses were performed using S-PLUS (S-PLUS 6.1, Insightful
295 Corporation, Seattle, USA, 2002).

3. Results

300 3.1. *In vitro* experiments

3.1.1. PCV2 replication kinetics in PBMC

Figure 1 demonstrates mean percentages of PCV2 positive cells and antigen
 305 expression patterns in PCV2-inoculated PBMC of three conventional PCV2
 seronegative pigs (pigs 1, 2 and 3). For strain 1121, maximum numbers of antigen
 positive cells were very low in non-treated PBMC (0.06 % in pig 1 at 48 hpi, 0.14 %
 in pig 2 at 72 hpi and 0.04 % in pig 3 at 48 hpi). Antigen positive cells were observed
 starting from 36 hpi (pigs 2 and 3) or 48 hpi (pig 1). In ConA-treated PBMC,
 310 maximum numbers of antigen positive cells of 2.08 %, 1.02 % and 2.02 % were
 observed at 96 hpi, 72 hpi and 96 hpi for pigs 1, 2 and 3 respectively. Nuclear
 localized viral antigens were detected for the first time at 12 hpi for pig 2 and at 36
 hpi for pigs 1 and 3. Cytoplasmic localized viral antigens were detected starting from
 36 hpi. At this time point, 20 % to 50 % of cells with viral antigens in the nucleus
 315 showed viral antigens in the cytoplasm and all cells with viral antigens in the
 cytoplasm showed viral antigens in the nucleus. From 36 hpi to 96 hpi, the numbers
 of total PCV2 antigen positive cells, positive cells with antigens in the nucleus and
 positive cells with antigens in the cytoplasm were significantly different in ConA-
 treated PBMC when compared to non-treated PBMC. In rIFN- γ -treated PBMC,
 320 maximum numbers of antigen positive cells were very low (0.06 % in pig 1 at 72 hpi,
 0.18 % in pig 2 at 72 hpi and 0.06 % in pig 3 at 36 and 48 hpi). Antigen positive cells
 were observed starting from 24 hpi (pig 2), 36 hpi (pig 3) or 72 hpi (pig 1).
 For Stoon-1010, maximum numbers of antigen positive cells were very low in non-
 treated PBMC (0.02 % in pig 1 at 96 hpi, 0.22 % in pig 2 at 96 hpi and 0.00 % in pig

325 3). Antigen positive cells were observed starting from 12 hpi (pig 2) or 96 hpi (pig 1).
 In ConA-treated PBMC, maximum numbers of antigen positive cells of 0.96 %, 0.84 % and 0.34 % were observed at 48 hpi for pig 1 and at 72 hpi for pigs 2 and 3 respectively. Nuclear localized viral antigens were detected for the first time at 24 hpi for pig 1 and at 12 hpi for pigs 2 and 3. Cytoplasmic localized viral antigens were
 330 detected for the first time at 24 hpi for pigs 1 and 2 and at 72 hpi for pig 3. At these time points, 12 % to 33 % of cells with viral antigens in the nucleus showed viral antigens in the cytoplasm and all cells with viral antigens in the cytoplasm showed viral antigens in the nucleus. From 24 hpi to 72 hpi, the numbers of total PCV2 antigen positive cells and positive cells with antigens in the nucleus were significantly
 335 different in ConA-treated PBMC when compared to non-treated PBMC. At 96 hpi, the numbers of positive cells with antigens in the cytoplasm were significantly different in ConA-treated PBMC when compared to non-treated PBMC. In rIFN- γ -treated PBMC, maximum numbers of antigen positive cells were very low (0.02% in pig 1 at 36 hpi, 0.16 % in pig 2 at 72 hpi and 0.02 % in pig 3 at 72 hpi). Antigen
 340 positive cells were observed starting from 12 hpi (pig 2), 36 hpi (pig 1) or 72 hpi (pig 3).

In ConA-treated PBMC, all cells with nuclear localized capsid protein also contained Rep protein in their nucleus. In 89 % (1121) and 81 % (Stoon-1010) of the cells with nuclear localized Rep protein, nuclear localized capsid protein was found. Cells with
 345 cytoplasmic staining for capsid protein, without nuclear staining for Rep or capsid proteins, were observed starting from 48 hpi (pig 2 and 3) or 72 hpi (pig 1) for strain 1121 and starting from 48 hpi (pig 1), 72 hpi (pig 2) or 96 hpi (pig 3) for Stoon-1010. This staining pattern was accompanied with morphological changes typical for cellular degeneration. Rep protein was never detected in the cytoplasm of PCV2

350 positive cells, except in a small fraction of cells with typical morphological signs of degeneration.

Positive reactions were not observed when staining non-inoculated PBMC for PCV2 antigens or when staining PCV2-inoculated PBMC with an irrelevant isotype-matched mAb and with porcine PCV2-negative control pAbs.

355

3.1.2. Identification of PCV2 positive PBMC

PBMC were obtained from three conventional PCV2 seronegative pigs (pigs 4, 5 and 6). The proportions of ConA-treated PCV2 positive PBMC per cell smear, identified as CD4⁺, CD8⁺, IgM⁺ or SWC3⁺ after inoculation with strain 1121, were determined 360 at 0, 36 and 72 hpi. At 0 hpi, PCV2 positive cells were not observed in ConA-treated PBMC. At 36 hpi, PCV2 positive cells were identified as CD4⁺ (48 % ± 5 %), CD8⁺ (56 % ± 4 %) and IgM⁺ (12 % ± 3 %). At 72 hpi, 40 % ± 5 % of PCV2 positive cells were CD4⁺, 54 % ± 11 % were CD8⁺ and 11 % ± 4 % were IgM⁺. Viral antigen 365 positive SWC3⁺ PBMC were not found. The numbers of non-treated and rIFN-γ-treated PCV2 positive PBMC per cell smear were too low for a correct interpretation of results.

3.1.3. Virus titration of PBMC culture supernatant

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Figure 2 demonstrates the evolution of mean virus titres in culture supernatants of PCV2-inoculated PBMC of three conventional PCV2 seronegative pigs (pigs 1, 2 and 3). Time-dependent rises in titres were seen for supernatants of ConA-treated PBMC, starting from 72 hpi for strain 1121 and from 48 hpi for Stoon-1010. Maximum virus

375 titres of $10^{3.2}$ TCID₅₀ / ml supernatant and $10^{4.1}$ TCID₅₀ / ml supernatant were
 observed for strain 1121 and Stoon-1010 at 96 hpi. Significant differences between
 virus titres in supernatants of non-treated PBMC and ConA-treated PBMC were
 observed at 72 and 96 hpi for strain 1121 and at 36, 48 and 96 hpi for Stoon-1010. A
 small but statistically significant difference between virus titres in supernatants of
 380 non-treated PBMC and rIFN- γ -treated PBMC was observed for strain 1121 at 96 hpi.

3.2. *In vivo* experiments

3.2.1. Clinical monitoring

385 Clinical signs suggestive of PMWS were not observed. Differences in general
 conditions were not observed between any of the pigs. Inguinal lymph nodes of all
 pigs of groups B (PCV2/ConA), C (PCV2/rIFN- γ), D (-/ConA) and E (-/rIFN- γ) were
 noticeably enlarged and pale. This was observed from the first time a biopsy was
 390 taken, up to the end of the study. Enlarged, pale inguinal lymph nodes were not
 observed in any of the pigs of group A (PCV2-inoculated, non-treated pigs). Pig 6
 (group B, PCV2/ConA) died during blood sampling at 15 dpi.

3.2.2. PCV2 replication

395 The evolution in PCV2 titres in the inguinal lymph nodes of individual pigs is shown
 in Figure 3. From all pigs of groups A, B and C (PCV2-inoculated pigs), PCV2 was
 isolated at at least one time point. PCV2 could be detected for the first time at 15 dpi
 in pigs of groups A (PCV2/-) and C (PCV2/rIFN- γ). In pigs of group B

(PCV2/ConA), PCV2 was already isolated from the lymph node biopsies taken at 10 dpi. Titres at this time point ranged between $10^{3.0}$ and $10^{4.3}$ TCID₅₀ / g. At 15 and 21 dpi, PCV2 titres in pigs of group B (PCV2/ConA) were generally higher than in pigs of groups A (PCV2/-) and C (PCV2/rIFN- γ), but due to the high variation in PCV2 titres between pigs in the same groups and due to the limited number of pigs per group, a significant difference between groups A (PCV2/-) and B (PCV2/ConA) could only be observed at 10 dpi and not at 15 and 21 dpi.

The general evolution in PCV2 titres in time was similar in all pigs from groups A, B and C (PCV2-inoculated pigs), except in one pig (pig 7) from group B (PCV2/ConA). Eleven out of twelve pigs of groups A, B and C (PCV2-inoculated pigs) showed a peak of PCV2 replication at 15 dpi. Afterwards, the PCV2 titres decreased in all these pigs. Pig 7 (group B, PCV2/ConA) showed a gradual increase in PCV2 titre until the end of the study.

In pigs of groups D and E (non-inoculated pigs), PCV2 was never isolated during the study.

Table 2 shows the results of the quantification and immunophenotyping of PCV2 positive cells in inguinal lymph node biopsies at 15 dpi. In biopsies from pigs of groups A (PCV2/-) and C (PCV2/rIFN- γ), low numbers of PCV2 positive cells were observed (less than 20 PCV2 positive cells per 10 mm²). In these pigs, PCV2 was mainly localized in SWC3⁺ cells, but in some pigs small proportions of PCV2 positive cells were CD4⁺, CD8⁺ or IgM⁺. PCV2 positive cells were mainly observed as single cells in follicular and parafollicular regions.

The number of PCV2 positive cells was significantly higher in biopsies from group B (PCV2/ConA) pigs 5, 6 and 7 (389 - 664 PCV2 positive cells per 10 mm²), when compared to pigs from group A (PCV2/-). In pigs 5, 6 and 7, PCV2 was found in

425 CD4⁺, CD8⁺, IgM⁺ and SWC3⁺ cells. In pigs 6 and 7, SWC3⁺ cells were the
predominantly PCV2 positive cell type (75 % and 66 % respectively). In pig 5, the
percentages of CD4⁺, CD8⁺, IgM⁺ and SWC3⁺ PCV2 positive cells ranged from 14 %
to 33 %. In pigs 5, 6 and 7, the vast majority of PCV2 positive cells were located in
the peripheral areas of the follicular regions and in the parafollicular regions. In these
430 pigs, PCV2 antigens were occasionally (< 5 %) observed in the nucleus of CD4⁺,
CD8⁺, IgM⁺ and SWC3⁺ cells. For pig 8 (group B, PCV2/ConA), similar observations
were made as for pigs from groups A (PCV2/-) and C (PCV2/rIFN- γ).
PCV2 positive cells were not observed in any of the pigs of groups D and E (non-
inoculated pigs).

435

3.2.3. Humoral immune response against PCV2

The evolution in PCV2-specific Abs, as demonstrated by IPMA, is shown in Figure 4.
Anti-PCV2 Abs were demonstrated in all pigs from groups A, B and C (PCV2-
440 inoculated pigs), except in the pig that died at 15 dpi (pig 6, group B, PCV2/ConA).
Differences were observed in the time points at which anti-PCV2 Abs were detected
for the first time. Pigs from group A (PCV2/-) seroconverted at 10 or 15 dpi. In
groups B (PCV2/ConA) and C (PCV2/rIFN- γ), only 2 out of 4 pigs seroconverted at
15 dpi. In the other pigs of groups B (PCV2/ConA) and C (PCV2/rIFN- γ),
445 seroconversion against the virus was delayed until 21 dpi. Statistically significant
differences were not detected at any of the time points. PCV2 neutralizing Abs
evolved in a similar way as PCV2-specific Abs detected by IPMA (data not shown).
None of the pigs from groups D or E (non-inoculated pigs) seroconverted against
PCV2.

450

4. Discussion

In the present study, it was shown that the mitogen ConA increases PCV2 replication
455 in porcine leukocytes both *in vitro* and *in vivo*. An effect of rIFN- γ on PCV2
replication in leukocytes was not observed under the conditions of the present study.
Expression of PCV2 antigens in ConA-stimulated PBMC was observed starting from
12 hpi to 36 hpi, depending on the PCV2 strain and the blood donor that were used.
This is in agreement with another study, in which PCV2 replication was observed in
460 ConA-stimulated PBMC from 18 hpi onwards (Yu et al., 2007). In the present study,
the first detectable viral antigens were localized in the nucleus. These antigens were
identified as Rep proteins and capsid proteins. This suggests that in the early stages of
infection, newly synthesized Rep and capsid proteins are efficiently transported to the
nucleus where genome replication and encapsidation take place. As infection
465 progressed, the number of cells with capsid proteins in the cytoplasm increased,
suggesting the translocation of assembled virions from the nucleus to the cytoplasm,
as suggested previously by Finsterbusch et al. (2005). At the late stages of infection,
cells with intense cytoplasmic staining but without nuclear staining were observed.
This was accompanied with morphological changes typical for cellular degeneration,
470 suggesting that PCV2 infection may lead to cell death. Only in these degenerated
cells, the Rep protein was occasionally found in the cytoplasm, indicating a very
strong affinity of the Rep protein for the nucleus in viable cells. The results further
suggest that the rise in PCV2 titres in culture supernatants coincided with the
appearance of these degenerated cells. The fact that PCV2 antigens were most

475 frequently found in the nucleus of PBMC early in replication contradicts with earlier
 findings in PK-15 cells. The majority of PCV2-infected PK-15 cells have capsid
 antigens in the cytoplasm without having Rep or capsid antigens in the nucleus during
 the early stages of the replication cycle (Meerts et al., 2005a). Since it is known that
 cellular transcription factors of lymphoid and epithelial cell types can have different
 480 specificities for promoters, enhancers and co-activators (Foster et al., 1985; Forsberg
 and Westin, 1991), it can be speculated that in PK-15 cells the RNA polymerase II
 transcription-initiation complex might have a higher affinity for the promoter region
 of the capsid gene than for the promoter region of the Rep gene, leading to expression
 of the capsid protein in a large number of cells without expression of the Rep protein.
 485 In ConA-treated PBMC, the affinities of this complex for both promoters may be
 equal, leading to a simultaneous expression of the Rep protein and the capsid protein.
 In the present study, it was demonstrated that PCV2 replicates in CD4⁺, CD8⁺ and
 IgM⁺ PBMC, suggesting that not only circulating T-lymphocytes, which was
 demonstrated previously by Yu et al. (2007), but also circulating B-lymphocytes may
 490 be susceptible to PCV2 replication *in vitro*. Since CD4 is also present on e.g. natural
 IFN- α producing cells (Charley and Lavenant, 1990) and since CD8 is also present on
 e.g. natural killer cells (Saalmuller et al., 1994), we cannot exclude the possibility that
in vitro stimulation with ConA makes these cell types susceptible to PCV2 replication
 as well. Further research should clarify this issue. As in other *in vitro* studies (Yu et
 495 al., 2007), we failed to demonstrate PCV2 replication in PBMC-derived monocytes.
 Meerts et al. (2005b) have previously shown that IFN- γ enhances PCV2 replication *in*
vitro in porcine epithelial and monocytic cell lines. This enhancing effect by IFN- γ
 was caused by an increased internalization of bound virions into the cell. The
 influence of IFN- γ on the internalization of PCV2 in PBMC was not investigated in

the present study. In cell lines, active cellular polymerases allow PCV2 to complete its infectious cycle. Therefore, we assume that the enhancing effect of ConA on PCV2 replication in PBMC *in vitro* was merely due to its high mitotic activity. We assume that the lack of high mitotic activity of rIFN- γ , under the conditions of the present study, was the main reason why rIFN- γ did not have an effect on PCV2 replication in PBMC *in vitro*.

In preliminary experiments, PCV2 replication was also observed in PBMC stimulated with phytohaemagglutinin and a combination of ionomycin and phorbol dibutyrate but not with LPS (data not shown). The best results were obtained with ConA and therefore this mitogen was selected for the present study. The technique that has been described is inexpensive, easy to perform and therefore it is potentially suitable for detailed *in vitro* studies of the PCV2 replication cycle in lymphocytes, studies that are nowadays merely performed in cell lines.

In the present study, it was shown that treatment of CD/CD pigs with ConA renders them more susceptible to an increased PCV2 replication. Injecting the pigs with rIFN- γ , following the protocol that was used in the present study, could not induce this increased PCV2 replication. The different outcomes of the ConA and rIFN- γ treatments could be due to various mechanisms. ConA is known to cause proliferation and activation of mainly T-lymphocytes (Mosmann et al., 1986), but it also affects other immune cell types such as B-lymphocytes (Andersson et al., 1972) and monocytes/macrophages (Smith and Goldman, 1972). ConA also induces various cytokines and IFN- γ is just one of them. ConA thus has a much wider range of biological activities than IFN- γ . ConA also induces much higher concentrations and a more prolonged production of IFN- γ in animals compared to the IFN- γ concentration that can be reached by injecting rIFN- γ . Injected rIFN- γ is very quickly removed from

the body (Lortat-Jacob et al., 1996), while ConA induces a continuous production of IFN- γ by the T-lymphocytes of the animal. Unfortunately, we were not able to demonstrate the presence of IFN- γ in plasma samples of the CD/CD pigs of the present study, nor with a bio-assay on PK-15 cells, nor with a commercial ELISA for IFN- γ (Biosource Europe, Nivelles, Belgium), indicating that IFN- γ blood levels were very low at the times of blood sampling (data not shown). Nevertheless, the treatments of pigs with ConA or rIFN- γ were effective since inguinal lymph nodes of treated pigs were clearly enlarged, whether these pigs were inoculated with PCV2 or not. Therefore, results from the present study are indicative that the enhancing effect of ConA treatment on PCV2 replication was due to the global effect of ConA on the pig's immune system and not to the effect of IFN- γ alone. Obviously, more research needs to be performed to address this issue.

A remarkable difference between ConA-treated pigs and non-treated and rIFN- γ -treated pigs was observed in inguinal lymph node biopsies collected at 15 dpi. High numbers of PCV2-infected cells were observed in 3 out of 4 ConA-treated pigs. PCV2-infected cells were identified as CD4⁺, CD8⁺, IgM⁺ and SWC3⁺, as demonstrated previously by Sanchez et al. (2004). Although PCV2 replicated to high levels in cells of the immune system, only 2 of these 3 pigs showed a delayed onset of the humoral immune response against PCV2. High levels of PCV2 replication in cells of the immune system were not observed in non-treated and rIFN- γ -treated pigs. Still, a delayed onset of the humoral immune response was observed in 2 out of 4 rIFN- γ -injected pigs. This suggests that the delayed humoral immune response observed in 2 out of 4 ConA-treated and in 2 out of 4 rIFN- γ -treated pigs was not caused by high levels of PCV2 replication in cells of the immune system, but might have been caused by a common effect of ConA and rIFN- γ treatments in these specific pigs. Possible

550 responsible mechanisms include the inhibitory effects of ConA-activated suppressor
T-cells on B-cell proliferation and antibody production (Redelman et al., 1976), and
the inhibitory effect of IFN- γ on the early stages of B-cell activation (Abed et al.,
1994). The present results further suggest that an impaired humoral immune response
against PCV2 is not sufficient to cause high levels of PCV2 replication. Further
555 research should deal with these issues.

In the present study, it was again demonstrated that it can be difficult to reproduce
PCV2-associated disease in CD/CD pigs. Even in the ConA-treated pigs that
experienced an increased PCV2 replication, clinical signs were not observed. Still, the
ConA treatment might provide a valuable tool to investigate the link between high
560 PCV2 replication and PMWS. Several treatments of CD/CD pigs that influence the
pig's immune system have been described until now. It has been demonstrated that
immune stimulation (Krakowka et al., 2001), immune suppression by Cyclosporin A
(Meerts et al., 2005c) and mitogen treatment (the present study) are able to increase
PCV2 replication in pigs. This observation may indicate that there is an unstable
565 equilibrium between a specific immune response that is able to neutralize PCV2 and
to eliminate PCV2-infected cells, and an immune response that facilitates the
replication of the virus in the pig. This fragile balance might be the underlying
mechanism responsible for the high variation that is observed in PCV2 replication and
clinical outcome between individual pigs and the even higher variation between
570 different studies.

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Table 1. Distribution of caesarean-derived colostrum-deprived pigs in different
735 groups with specific manipulations per group.

Table 2. Quantification and immunophenotyping of PCV2 positive cells at 15 days
post inoculation in inguinal lymph node biopsies of PCV2-inoculated caesarean-
derived colostrum-deprived pigs of groups A (non-treated), B (treated with
740 concanavalin A, ConA) and C (treated with recombinant interferon-gamma, rIFN- γ).
Pigs from groups D and E (non-inoculated pigs) did not have PCV2 positive cells.

Fig. 1. Mean percentage ($\pm$ SD) of PCV2 antigen positive cells and antigen expression patterns in non-treated, concanavalin A-treated or recombinant interferon-gamma-treated, PCV2-inoculated PBMC of three conventional PCV2 seronegative pigs at different time points after *in vitro* inoculation of PBMC with strains 1121 and Stoon-1010 (pigs 1, 2 and 3).

Fig. 2. Evolution of mean ($\pm$ SD) PCV2 titres in culture supernatants of non-treated, concanavalin A-treated or recombinant interferon-gamma-treated, PCV2-inoculated PBMC of three conventional PCV2 seronegative pigs (pigs 1, 2 and 3).

Fig. 3. Evolution of PCV2 titres in inguinal lymph nodes of PCV2-inoculated caesarean-derived colostrum-deprived pigs of groups A (non-treated), B (treated with concanavalin A, ConA) and C (treated with recombinant interferon-gamma, rIFN- γ), and of non-inoculated pigs of groups D (treated with concanavalin A, ConA) and E (treated with recombinant interferon-gamma, rIFN- γ). The dashed line represents the detection limit of the assay ($10^{2.7}$ TCID₅₀ / g).

Fig. 4. Evolution of anti-PCV2 antibody titres (IPMA) in PCV2-inoculated caesarean-derived colostrum-deprived pigs of groups A (non-treated), B (treated with concanavalin A, ConA) and C (treated with recombinant interferon-gamma, rIFN- γ), and in non-inoculated pigs of groups D (treated with concanavalin A, ConA) and E (treated with recombinant interferon-gamma, rIFN- γ).

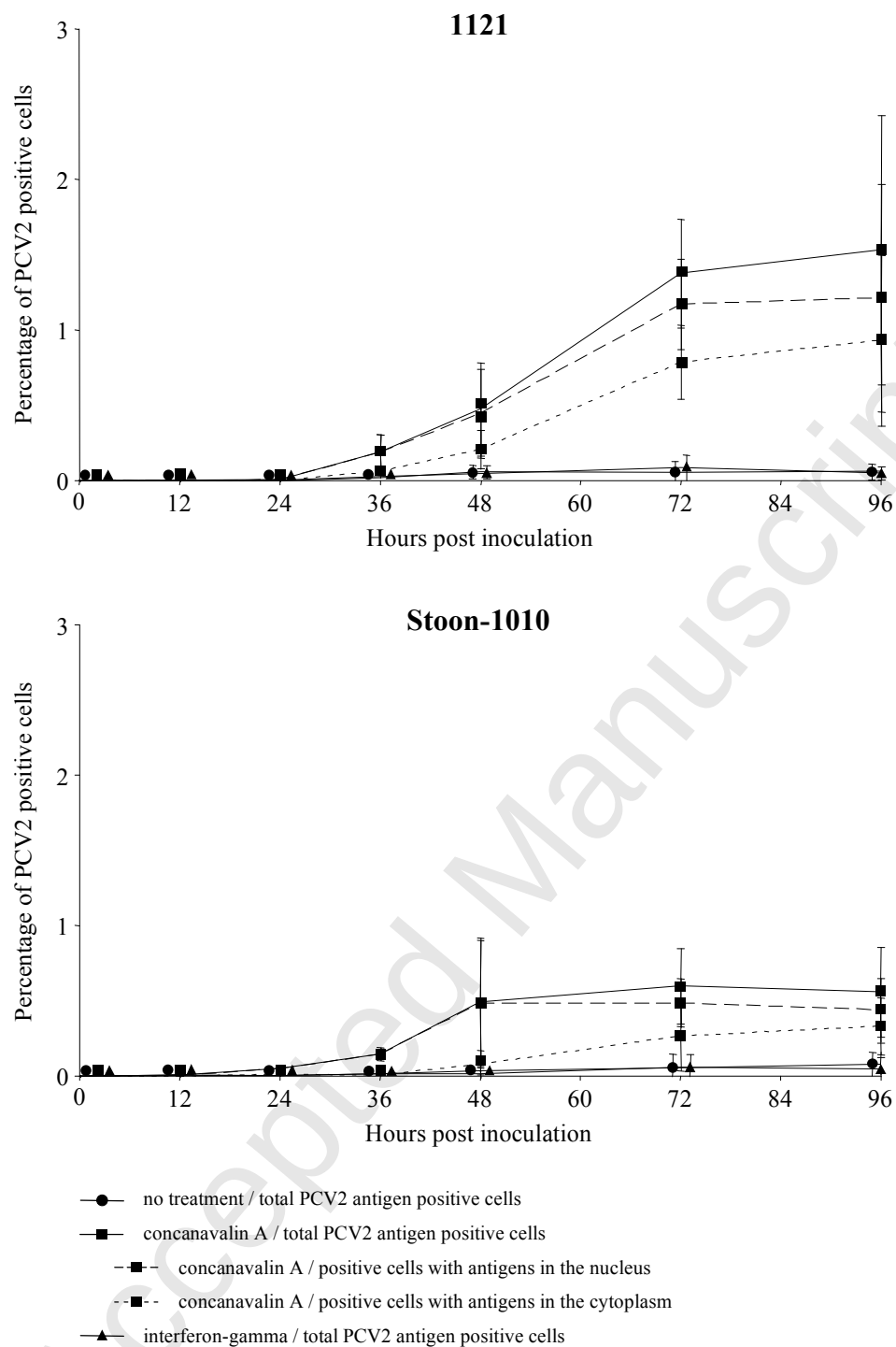
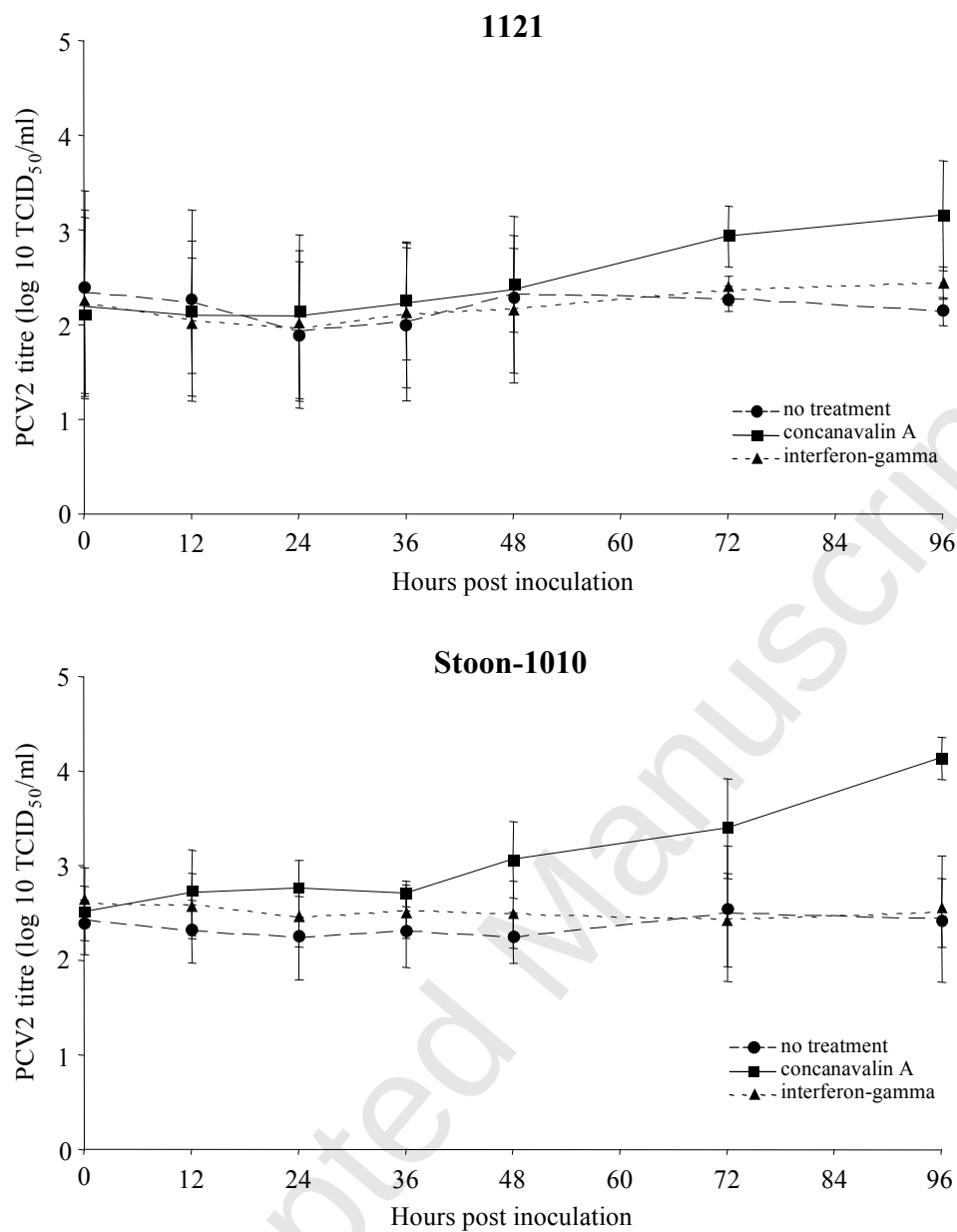
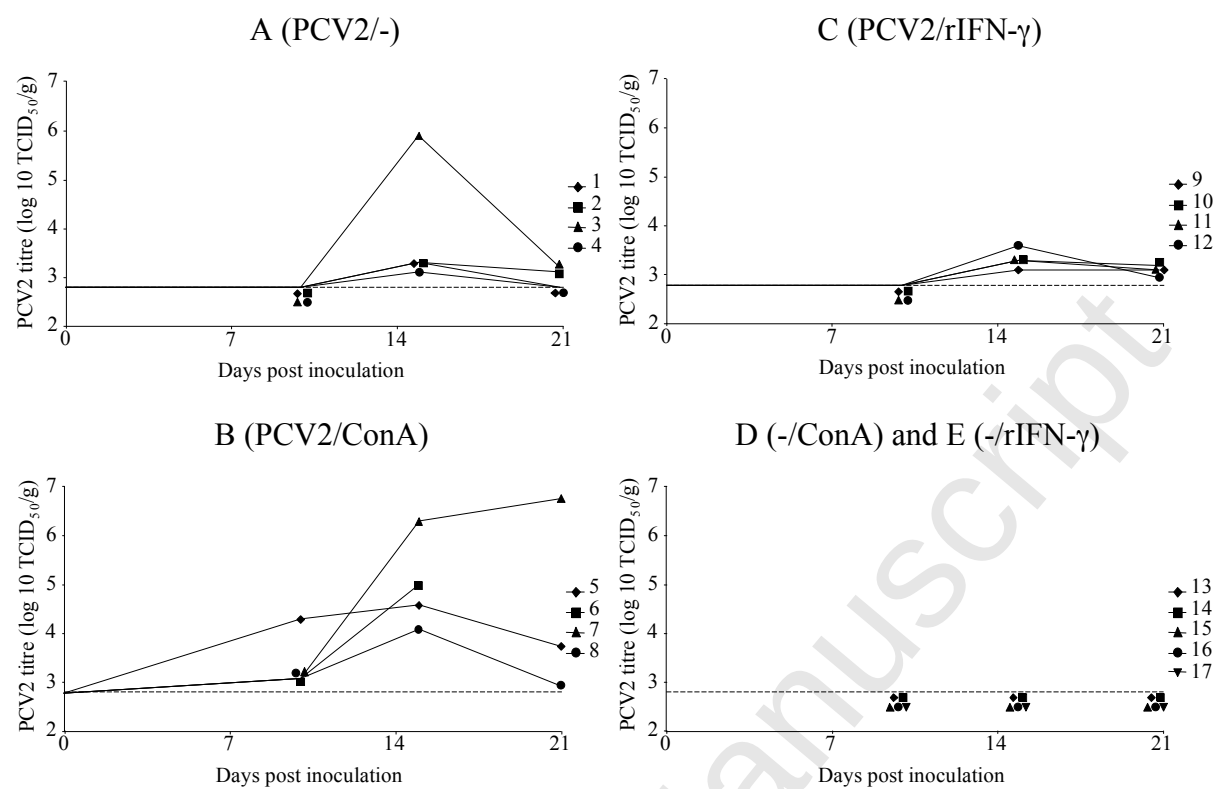


Fig. 1.

**Fig. 2.**

**Fig. 3.**

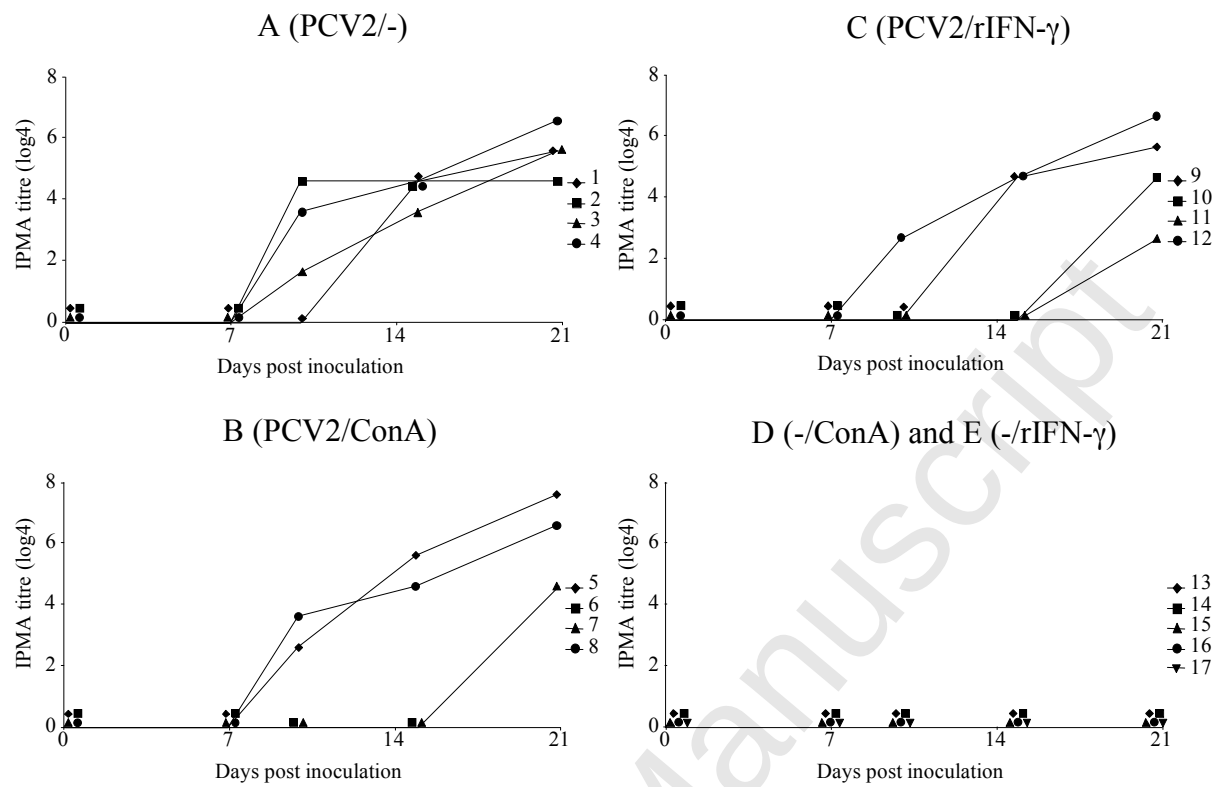


Fig. 4.

Table 1.

Group	n	PCV2 inoculation	Treatment	Pig numbers
A	4	yes	-	1 to 4
B	4	yes	ConA	5 to 8
C	4	yes	rIFN- γ	9 to 12
D	3	no	ConA	13 to 15
E	2	no	rIFN- γ	16 to 17

Table 2.

Group	Pig no.	Treatment	Virus titre (log ₁₀ TCID ₅₀ /g)	Identification of PCV2 positive cells as*			
				CD4 ⁺	CD8 ⁺	IgM ⁺	SWC3 ⁺
A	1	-	3.3	2 / 9 (22%)	1 / 5 (20%)	1 / 5 (20%)	5 / 7 (71%)
	2	-	3.3	0 / 4 (0%)	0 / 10 (0%)	0 / 2 (0%)	1 / 1 (100%)
	3	-	5.8	4 / 18 (22%)	0 / 13 (0%)	3 / 10 (30%)	7 / 11 (64%)
	4	-	3.0	0 / 2 (0%)	0 / 2 (0%)	0 / 4 (0%)	1 / 1 (100%)
B	5	ConA	4.5	140 / 434 (32%)	60 / 432 (14%)	89 / 389 (23%)	136 / 418 (33%)
	6	ConA	5.0	77 / 635 (12%)	69 / 664 (10%)	70 / 555 (13%)	339 / 454 (75%)
	7	ConA	6.3	72 / 570 (13%)	84 / 555 (15%)	44 / 537 (8%)	394 / 601 (66%)
	8	ConA	4.0	0 / 4 (0%)	0 / 5 (0%)	0 / 1 (0%)	4 / 4 (100%)
C	9	rIFN- γ	3.0	1 / 8 (13%)	0 / 4 (0%)	0 / 7 (0%)	12 / 16 (75%)
	10	rIFN- γ	3.3	0 / 2 (0%)	0 / 1 (0%)	0 / 2 (0%)	1 / 2 (50%)
	11	rIFN- γ	3.3	0 / 1 (0%)	0 / 2 (0%)	0 / 1 (0%)	2 / 2 (100%)
	12	rIFN- γ	3.5	1 / 10 (10%)	1 / 2 (50%)	0 / 6 (0%)	10 / 15 (67%)

*Number of both PCV2 and cell marker positive cells / total number of PCV2 positive cells per 10 mm² tissue.