

## Testing the efficacy of medium chain fatty acids against rabbit colibacillosis

Mélanie Gallois, Thierry Gidenne, Juan Orengo, Cécile Caubet, Christian

Tasca, Alain Milon, Séverine Boullier

### ► To cite this version:

Mélanie Gallois, Thierry Gidenne, Juan Orengo, Cécile Caubet, Christian Tasca, et al.. Testing the efficacy of medium chain fatty acids against rabbit colibacillosis. Veterinary Microbiology, 2008, 131 (1-2), pp.192. 10.1016/j.vetmic.2008.03.004 . hal-00532405

## HAL Id: hal-00532405 https://hal.science/hal-00532405

Submitted on 4 Nov 2010

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

### Accepted Manuscript

Title: Testing the efficacy of medium chain fatty acids against rabbit colibacillosis

Authors: Mélanie Gallois, Thierry Gidenne, Juan Orengo, Cécile Caubet, Christian Tasca, Alain Milon, Séverine Boullier

PII:	S0378-1135(08)00101-6
DOI:	doi:10.1016/j.vetmic.2008.03.004
Reference:	VETMIC 3983
To appear in:	VFTMIC
To uppeur in:	V ET MIC
Received date:	3-12-2007
Revised date:	2-3-2008
Accepted date:	4-3-2008



Please cite this article as: Gallois, M., Gidenne, T., Orengo, J., Caubet, C., Tasca, C., Milon, A., Boullier, S., Testing the efficacy of medium chain fatty acids against rabbit colibacillosis, *Veterinary Microbiology* (2007), doi:10.1016/j.vetmic.2008.03.004

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

**Revised manuscript** 

# **ACCEPTED MANUSCRIPT**

1	Testing the efficacy of medium chain fatty acids against rabbit colibacillosis
2	
3	Mélanie Gallois <sup>1,3</sup> , Thierry Gidenne <sup>1</sup> , Juan Orengo <sup>1,4</sup> , Cécile Caubet <sup>2</sup> , Christian Tasca <sup>2</sup> , Alain
4	Milon <sup>2</sup> and Séverine Boullier <sup>2*</sup>
5	
6	<sup>1</sup> : INRA, UMR1289 Tissus Animaux, Nutrition, Digestion, Ecosystème, Métabolisme,
7	Chemin de Borde-Rouge - Auzeville, BP 52627, F-31326 Castanet-Tolosan Cedex, France;
8	INP-ENSAT, F-31326 Castanet-Tolosan Cedex, France; ENVT, F-31076 Toulouse Cedex 3,
9	France
10	<sup>2</sup> : UMR INRA ENVT 1225, Physiopathologie et Immunité des Muqueuses, Ecole Nationale
11	Vétérinaire de Toulouse, 23 chemin des Capelles, BP 87614, 31076 Toulouse, France
12	<sup>3</sup> : present address: Institut National de la Recherche Agronomique, Laboratoire de
13	Pharmacologie-Toxicologie 180 chemin de Tournefeuille, BP3 31931 Toulouse cedex 9,
14	France.
15	<sup>4</sup> : present address: Department of Animal Production, Faculty of Veterinary Science,
16	University of Murcia, Campus de Espinardo, 30100 Murcia, Spain.
17	*: Corresponding author. Tel: 33 5 61 19 32 61, Fax 33 5 61 19 39 75, e-mail:
18	s.boullier@envt.fr
19	
20	Short title: Dietary MCFA against rabbit colibacillosis
21	
22	Abbreviations: MCFA: Medium Chain Fatty Acids; SO: Sunflower Oil; MCT: Medium
23	Chain Triglycerides; LB: L-Broth.
24	

#### 25 Abstract

26 Enteropathogenic Escherichia coli (EPEC) represents a major cause of lethal diarrhea in 27 young mammals. Although the pathogenicity mechanisms of EPEC are now well understood, 28 the intrinsic and environmental factors that control the expression of EPEC virulence remain 29 largely unknown. In the rabbit, suckling reduces pups' sensitivity to EPEC infection. Hence, 30 we have hypothesized that uncharacterized factors present in doe milk may mediate this 31 protection. Medium chain fatty acids (MCFA), known to possess antimicrobial properties, are 32 highly abundant in doe milk. We demonstrate that caprylic acid exhibits a clear bacteriostatic 33 effect in vitro against the rabbit EPEC strain E22 (O103:H2:K-), in a dose-dependent manner. 34 In vivo, the dietary inclusion of triglycerides of MCFA did not however reduce the sensitivity 35 of young rabbits challenged with this EPEC strain. The mortality and fecal excretion of EPEC 36 were not reduced, and the bacterial adhesion to ileum was not inhibited. Amount of MCFA 37 reaching the ileal level might have been too low and/or their association to other milk 38 antimicrobials may have been required to observe a positive effect on disease evolution in a 39 context of a highly virulent challenge.

40

41 Keywords: Enteropathogenic *E. coli* (EPEC), milk, rabbit, medium chain fatty acids
42 (MCFA), diarrhea

- 43
- 44

#### 45 **1. Introduction**

46 Maternal milk is very efficient in preventing colibacillosis in young rabbits infected with a 47 highly pathogenic strain of enteropathogenic E. coli (EPEC) belonging to the O103 serogroup 48 (Gallois et al., 2007), a dominant serogroup in French fattening rabbit units (Camguilhem and 49 Milon, 1989). Milk Igs are not responsible for this protection (Gallois et al., 2007) and a cross 50 protection with Abs directed towards other strains of E. coli is unlikely, as all trials using heterologous challenges (Donnenberg et al., 1998) or vaccines (Milon et al., 1992) failed to 51 52 demonstrate cross protection between different EPEC strains. Less specific milk components 53 are considered as good candidates for this protection, through antimicrobial or 54 immunomodulating properties, or by interfering with the expression of virulence factors 55 (Baranyi et al., 2003; Jia et al., 2001; Recio and Visser, 1999, Cravioto et al., 1991, Sun et al., 56 2002). Doe milk composition is very unusual among domestic mammals. Triglycerides are 57 mainly composed of medium chain fatty acids (MCFA), with caprylic (C8:0) and capric 58 (C10:0) acids representing up to 65% (molar proportion) of total fatty acids (Demarne et al., 59 1978, Pascual et al., 1999). This content is very high as compared to other species such as 60 cows (5%), ewes (17%), goats (18%) or humans (2.5%) (Freeman et al., 1965). MCFA are 61 very active antimicrobials in vitro, including against coliforms (Sun et al., 2002, Marounek et 62 al., 2002). In vivo, MCFA have been implicated in the control of the stomacal microflora in 63 suckling rabbits (Canas-Rodriguez and Smith, 1966), and provide health benefits during the 64 post-weaning period (Skrivanova et al., 2004). The known antimicrobial properties of MCFA 65 and their particularly high content in doe milk led us to determine whether MCFA could 66 protect pups against colibacillosis during suckling. In this study, the bacteriostatic activity of 67 MCFA against EPEC were demonstrated in vitro. The disease evolution was studied in vivo in 68 rabbits inoculated with a non-pathogenic (BM21) or a pathogenic strain of *E. coli* (strain E22, 69 O103:H2:K-), and fed with a diet supplemented or not with triglycerides of MCFA.

70

#### 71 **2. Material and methods**

#### 72 <u>2.1. Bacterial strains</u>

An enteropathogenic *E. coli* strain belonging to O103:H2:K- serotype (E22 strain), which does not metabolize the rhamnose and is fimbriae AF/R2 positive (Camguilhem and Milon, 1989), and the non-pathogenic BM21 strain of *E. coli* (Boullier et al., 2003a) were used in this study. For the inhibition of adhesion assay, a genetically-modified pEGFP-E22 strain was used (Boullier et al., 2003b).

78

#### 79 <u>2.2. In vitro experiments</u>

80 Antimicrobial properties of MCFA were evaluated in vitro as follows. Solutions of C8:0, 81 C10:0 (Sigma) and of a mix of both, ranging from 0-40 mM, were prepared as previously described (Sun et al., 2002), and adjusted to a pH of 6.5 with NaOH. 10<sup>3</sup> CFU of E22 were 82 83 cultured in 5ml of MCFA + LB media for 18h with rotary shaking or spread on LB agar plate 84 to determine the initial exact amount of bacteria added to each media. CFU were counted for 85 each concentration of MCFA after ON culture on LB agar plates. The inhibition growth rate 86 was calculated as: (number of CFU/ml in LB medium) / (number of CFU/ml in MCFA 87 medium).

Ability of MCFA to inhibit EPEC adhesion to epithelial cells was measured on HeLa cells, cultivated as previously described (Boullier et al., 2003b). 20  $\mu$ l of pEGFP-E22 bacteria grown ON (10<sup>9</sup> CFU.mL<sup>-1</sup>) were pre-incubated with different amounts of C8:0 and C10:0 media for one hour at RT prior to their incubation for 30 min with HeLa cells (Boullier et al., 2003b). After washing, 10<sup>4</sup> HeLa cells per sample were immediately acquired with a Facscalibur (Becton Dickinson). Quantification of adhesion was monitored using Cellquest software (Becton Dickinson). The percentage of inhibition was calculated using the following

- 95 formula: ((Percentage of pEGFP-E22 + HeLa cells without C8:0-C10:0 media) (Percentage
- 96 of pEGFP-E22 + HeLa cells pre-incubated with C8:0-C10:0 media))/(Percentage of pEGFP-
- 97 E22 + HeLa cells without C8:0-C10:0 media)).
- 98

### 99 <u>2.3. In vivo experiment</u>

This experiment was approved by a regional ethical committee. 32 litters of 9 rabbits were 100 101 equally divided in 4 groups, according to their diet and to the E. coli strain inoculated at 28 days of age. Pelleted diets, without antibiotics, were formulated to cover the nutritional 102 103 requirements of the young rabbit (Gidenne, 2000) and were supplied with either 2% of 104 sunflower oil (SO) or 2% of medium chain triglycerides (MCT), a synthesized oil containing 55% of C8:0 and 44% of C10:0 (Radia®7104, Oléon France). Chemical composition of diets 105 106 was determined according to European recommendations (EGRAN, 2001), and fatty acids 107 composition was assayed by the ISTE (In Situ TransEsterification) method described by Park 108 and Goins (Park and Goins, 1994). From 14 to 35 days of age (the day of weaning), does and 109 their pups were reared in separated cages, to allow the feeding of pups with experimental diets 110 provided ad libitum. Suckling was controlled every morning by maintaining does with their pups for about 5 minutes. Rabbits were orally inoculated at 28 days of age with 10<sup>4</sup> CFU of 111 112 BM21 or E22 strains, 3 hours after suckling and were weaned at day 35. Mortality controls 113 were done twice a day and clinical examination every morning. Euthanasia was practiced in dying rabbits (intramuscular injection of Imalgène<sup>®</sup>1000 followed by an intracardiac injection 114 of T61<sup>®</sup>). Necropsy and a bacterial analysis of cecal content were systematically carried out to 115 116 identify rabbits which died from colibacillosis. Moreover, bacterial analyses were performed 117 on 10g of feces per cage twice a week. Cecal and fecal enumeration of E. coli was determined 118 as previously described (Gallois et al., 2007). At 36 days of age, one representative rabbit per 119 litter was sacrificed, in order to analyze composition of stomacal and ileal contents in fatty

120 acids according to the ISTE method (Park and Goins, 1994). At 63 days of age, all remaining rabbits were sacrificed. Survival rates were submitted to a  $\chi^2$  test (procedure CATMOD, SAS 121 Online<sup>®</sup>). Bacterial shedding and MCFA composition of digestive contents were analyzed 122 with the GLM procedure (SAS Online<sup>®</sup>), depending on diet, the strain of *E. coli* and their 123 124 interaction as main effects. On day 28, two additional litters of nine MCT or SO-fed suckling rabbits were inoculated with 10<sup>4</sup> CFU of E22 strain, and sacrificed 3 days later. Distal ileum 125 126 was fixed in 10% formaldehyde and embedded in paraffin. Bacteria fixed on the epithelia 127 were revealed with an anti-E22 serum prepared in the laboratory (Boullier et al., 2003b). 128 Tissue samples were scored blindly for bacterial adhesion.

129

#### 130 **<u>3. Results</u>**

#### 131 <u>3.1 MCFA inhibit bacterial growth *in vitro*</u>

132 The possible bacteriostatic or bactericidal effects of MCFA were first tested against E22 133 bacteria in vitro. At any tested concentrations, C8:0 did not display any detectable bactericidal or bacteriostatic effect, with an inhibition rate inferior to 1 log at 40 mM (5.76 g.kg<sup>-1</sup>, Figure 134 1). In contrast, C10:0 exhibited a strong dose-dependent bacteriostatic activity. The mean 135 inhibition rate reaches values of up to 91 at 40 mM (6.88 g.L<sup>-1</sup>). Interestingly, no synergistic 136 137 effect was observed between the two MCFA on the inhibition of bacterial growth. The dose-138 response curve obtained was similar to the one obtained when using C10:0 alone. There was 139 no detectable bactericidal activity with any of the MCFA concentrations tested (data not 140 shown). These in vitro findings prompted us to test whether MCFA could modify bacterial 141 growth *in vivo* and protect rabbits against colibacillosis when added in pup's diet.

#### 143 <u>3.2 MCT supplementation is inefficient to protect young rabbits from colibacillosis</u>

144 The chemical composition of diets was similar among groups (CP: 17.2%, NDF: 35.9%, 145 starch: 9.3% on an as-fed basis), with major differences restricted to their composition in 146 MCFA, MCT diet containing 19 percent units more C8:0 and 15 percent units more C10:0 147 than SO diet. In E22 groups, the first deaths due to EPEC occurred on the day of weaning and 148 the mortality rate reached 50% at 48-49 days of age, whatever the oil added in feed (Figure 2). 149 On day 63, 46% and 44% of rabbits had survived to colibacillosis in SO-E22 and MCT-E22 groups, respectively (P = 0.85). None of BM21-inoculated rabbits died from E22 150 colibacillosis, and their mortality rate was unaffected by the diet (P = 0.19). Morbidity, 151 152 characterized by anorexia, weight loss, liquid and sometimes hemorrhagic diarrhea, and 153 dehydration was similar in both E22 groups, and never exceeded 31% on a daily basis 154 whatever the diet (data not shown). Only 12 and 8 rabbits in MCT-E22 and SO-E22 groups, 155 respectively, did not show any signs of diarrhea all the experiment through. In BM21 groups, 156 excretion of E. coli remained stable and no colony belonging to the O103 serogroup was 157 identified (Figure 2). In infected rabbits, fecal excretion of E. coli increased as soon as 31 158 days of age, and most colonies were rhamnose negative. The level of excretion was between  $10^6$  and  $10^8$  CFU per g of feces from 31 to 45 days of age, then decreased to less than  $10^4$ 159 160 CFU per g of feces at the end of the experiment independently from the diet.

161

#### 162 <u>3.3. MCFA does not inhibit bacterial adhesion *in vivo* nor *in vitro*</u>

The addition of MCT in pup's diet could have modified the level of ileal bacterial attachment and the severity of tissue lesions induced by E22. *In vivo*, the profile of E22 adhesion was similar among MCT and SO rabbits, with very rare adhesion spots of bacteria on the ileum (a mean of 2 to 3 colonies per slide) without any intestinal inflammation lesion (Figure 3). *In* 

- 167 *vitro*, we showed that incubation of E22 bacteria with C8:0 and C10:0 (10 to 40 mM), alone
- 168 or combined, did not reduce their capacity to adhere on HeLa cells (Figure 3).
- 169

#### 170 <u>3.4. Transit of fatty acids along the stomach and the ileum</u>

171 As MCT showed no in vivo beneficial effect against colibacillosis, we wondered whether the 172 available amount of MCFA along the digestive tract was sufficient. As expected, 173 concentrations of C8:0 and C10:0 were higher in the stomach of MCT-fed rabbits compared 174 to SO rabbits, both in infected and in control groups (5.5-times higher for C8:0, P < 0.01, and 175 4.1-times higher for C10:0, P < 0.05). These values reached 18-19 mM for both fatty acids in 176 MCT-E22 group. In the ileum, the concentrations of C8:0 and C10:0 also remained higher in 177 MCT compared to SO rabbits, both in infected and control groups (P < 0.001). However, these values were low and did not exceed 0.1 g.kg<sup>-1</sup> (<0.7 mM) of fresh ileal content both for 178 179 C8:0 and C10:0 and were much lower than the active concentrations tested *in vitro*.

180

#### 181 **<u>4. Discussion</u>**

In the rabbit, the disease caused by EPEC is delayed by suckling, which is in agreement with our previous study (Gallois et al., 2007). Indeed, after inoculation of 32 to 42-days-old rabbits with similar doses of EPEC strains belonging to the O103 serogroup, deaths are classically observed 3 to 14 days post-infection (d.p.i.) (Camguilhem and Milon, 1989, Boullier et al., 2003a). In our study, rabbits mainly died in a period ranging from 13 to 17 d.p.i., which corresponds to 6 to 10 days after weaning. This protection during the suckling period is not conferred by specific milk Igs, as all does were seronegative towards the E22 strain.

Antimicrobial properties of MCFA against bacteria including coliforms were shown *in vitro*(Canas-Rodriguez and Smith, 1966, Marounek et al., 2002, Sun et al., 2002). In this study, we

191 show that only C10:0 inhibits the growth of E22 strain at neutral pH. Despite this clear

192 bacteriostatic effect *in vitro*, the dietary inclusion of triglycerides of MCFA does not reduce 193 the sensitivity of young rabbits to colibacillosis. Rates of morbidity and mortality, as well as 194 fecal E. coli excretion, were similar in all groups of rabbits regardless of their diet. This could 195 be partly due to an insufficient MCFA intake. The MCT incorporation level in diet (2%) was 196 chosen according to the MCFA content in doe milk and in reference to ingestion curves of 197 milk/solid feed (Gidenne and Lebas, 2006), to reach dietary MCFA concentrations similar to 198 the ones found in milk at the time of weaning. In rabbits fed with MCT, intake of C8:0 and 199 C10:0 (via feed) was similar to the intake before weaning (thus via the milk) (data not 200 shown). Alternatively, intestinal concentrations of MCFA were probably too low to produce a 201 bacteriostatic effect. For MCFA not to be absorbed too quickly and thus remain available in 202 EPEC sensitive tissues (ileum and cecum), they had to be included in the diet in the form of 203 MCT. In addition, because medium chain monoglycerides exert a higher antimicrobial activity than their corresponding free fatty acids (Isaacs et al., 1995, Petschow et al., 1996), 204 205 MCFA absorption needed to be delayed. Feeding rabbits with MCT should have allowed a 206 slower absorption of MCFA (Perret, 1980). Their monoesterified derivates would have thus 207 reached ileal and cecal segments. Even if the stomacal and ileal MCFA contents were higher in MCT-fed rabbits, they remained lower than 0.2 mg.g<sup>-1</sup> of fresh matter for both C8:0 and 208 209 C10:0 in the ileum. It was shown that C10:0 can inhibit the growth of E. coli at concentrations lower than 0.3 g.L<sup>-1</sup> (2mM) at a pH of 4.6 (Sun et al., 2002), but this minimal inhibiting 210 211 concentration increases quickly when the pH nears neutrality (Hsiao and Siebert, 1999). 212 Accordingly, in the present study, no bacteriostatic activity of C8:0 even at a dose of 5.76 g.L<sup>-</sup> <sup>1</sup> (40mM) was observed at neutral pH (close to conditions found in rabbit ileum and cecum), 213 214 whereas a 2log bacteriostatic effect was obtained with C10:0, but at a concentration of 6.88 g.L<sup>-1</sup> (40mM) which is much higher than the concentrations of MCFA found in the ileum of 215 infected MCT-fed rabbits (0.16 g.kg<sup>-1</sup>). MCT are in part digested and absorbed at stomacal 216

217 level (Perret, 1980). Thus, MCFA and their derivates are likely to be active mainly in the 218 stomach where high MCFA concentrations are observed, and limit the crossing of pylorus by 219 pathogenic *E. coli*. Moreover, acidic pH conditions in the stomach would be more adapted to 220 the expression of the antimicrobial activity of MCFA. If this hypothesis is true, our 221 experimental conditions (infectious dose/unit of time, highly pathogenic strain of EPEC) may 222 have been too drastic to allow the visualization of such phenomenon.

223 The protection conferred by suckling (Gallois et al., 2007) is probably the result of a 224 combination of different milk antimicrobial factors that may act to limit the virulence of 225 pathogens along the digestive tract through additive and synergistic effects (Isaacs et al., 226 1995). Some components inhibit bacterial growth while others are more involved in the 227 control of the colonization of epithelial cells by microorganisms. It was shown that MCFA 228 can limit the invasion of intestinal epithelial cells by Salmonella enterica in chicken (Van 229 Immerseel et al., 2004). However, no inhibition of adhesion of E22 to HeLa cells in presence 230 of MCFA was observed although an inhibiting adhesion capacity of milk both in vivo and in 231 vitro was previously demonstrated (Gallois et al., 2007).

In conclusion, our results show that, despite an *in vitro* effect, MCFA alone do not protect rabbits against colibacillosis. Since the pathogenicity of EPEC is related to their ability to colonize epithelial cells, it would be interesting to characterize milk factors able to inhibit bacterial adhesion. Once characterized, their use in association with bacteriostatic substances like MCFA may lead to synergistic effects aiming at protecting rabbits against colibacillosis.

237

#### 238 <u>5. Acknowledgements</u>

The authors wish to thank the staff of UMR 1289 TANDEM (INRA, Toulouse, France) for
their technical collaboration, as well as M.C. Nicot and F. Enjalbert (Nutrition de la Vache
Laitière, ENV, Toulouse, France). We also tank B. Severac for his constructive comments on

- the manuscript. This work was partly supported by grants from Glon group (France) and
- 243 Région Midi-Pyrénées (no.A2639).
- 244

#### 245 <u>6. References</u>

- Baranyi, M., Thomas, U., Pellegrini, A., 2003, Antibacterial activity of casein-derived
   peptides isolated from rabbit (Oryctolagus cuniculus) milk. J Dairy Res 70, 189-197.
- Boullier, S., Nougayrède, J., Marches, O., Tasca, C., Boury, M., Oswald, E., De Rycke, J.,
  Milon, A., 2003a, Genetically engineered Enteropathogenic E.coli strain elicits a specific
  immune response and protects against a virulent challenge. Microbes and Infection 5, 857867.
- Boullier, S., Tasca, C., Milon, A., 2003b, A new flow cytometric method to quantify the
  inhibition of enteropathogenic *Escherichia coli* adhesion by anti-adhesin antibodies.
  Cytometry 53, 79-87.
- Camguilhem, R., Milon, A., 1989, Biotypes and O serogroups of Escherichia coli involved in intestinal infections of weaned rabbits: clues to diagnosis of pathogenic strains. J Clin Microbiol 27, 743-747.
- Canas-Rodriguez, A., Smith, W.H., 1966, The identification of the antimicrobial factors of the
   stomach of suckling rabbits. Biochem J. 100, 79-82.
- Cravioto, A., Tello, A., Villafan, H., Ruiz, J., del Vedovo, S., Neeser, J.R., 1991, Inhibition of
  localized adhesion of enteropathogenic Escherichia coli to HEp-2 cells by immunoglobulin
  and oligosaccharide fractions of human colostrum and breast milk. J Infect Dis 163, 12471255.
- Demarne, Y., Lhuillery, C., Pihet, J., Martinet, L., Flanzy, J., 1978, Comparative study of
   triacylglycerol fatty acids in milk fat from two leporidae species: rabbit (oryctolagus
   cuniculus) and hare (lepus europaeus). Comp Biochem Physiol B 61, 223-226.
- Donnenberg, M.S., Tacket, C.O., Losonsky, G., Frankel, G., Nataro, J.P., Dougan, G., Levine,
   M.M., 1998, Effect of prior experimental human enteropathogenic Escherichia coli
   infection on illness following homologous and heterologous rechallenge. Infect Immun 66,
   52-58.
- Freeman, C.P., Jack, E.L., Smith, L.M., 1965, Intramolecular Fatty Acid Distribution in the
   Milk Fat Triglycerides of Several Species. J Dairy Sci 48, 853-858.
- Gallois, M., Gidenne, T., Tasca, C., Caubet, C., Coudert, C., Milon, A., Boullier, S., 2007,
  Maternal milk contains antimicrobial factors that protect young rabbits from
  enteropathogenic Escherichia coli infection. Clin Vaccine Immunol 14, 585-592.
- Gidenne, T., 2000, Recent advances and perspectives in rabbit nutrition. Emphasis on fiber
   requirements. World Rabbit Science 8, 23-32.
- Gidenne, T., Lebas, F., 2006, Feeding behaviour in rabbits., In: Bels, V. (Ed.) Feeding in
   domestic vertebrates. From structure to behaviour. CABI, Wallingford, UK, pp. 179-209.
- Hsiao, C.P., Siebert, K.J., 1999, Modeling the inhibitory effects of organic acids on bacteria.
  Int J Food Microbiol 47, 189-201.
- Isaacs, C.E., Litov, R.E., Thormar, H., 1995, Antimicrobial activity of lipids added to human
   milk, infant formula, and bovine milk. J Nutr Biochem 6, 362-366.
- Jia, H.P., Starner, T., Ackermann, M., Kirby, P., Tack, B.F., McCray, P.B., Jr., 2001,
  Abundant human beta-defensin-1 expression in milk and mammary gland epithelium. J
  Pediatr 138, 109-112.

- Marounek, M., Skrivanova, V., Savka, O., 2002, Effect of caprylic, capric and oleic acid on
  growth of rumen and rabbit caecal bacteria. J Anim Feed Sci 11, 507-516.
- Milon, A., Esslinger, J., Camguilhem, R., 1992, Oral vaccination of weaned rabbits against
  enteropathogenic Escherichia coli-like E. coli O103 infection: use of heterologous strains
  harboring lipopolysaccharide or adhesin of pathogenic strains. Infect Immun 60, 27022709.
- Park, P.W., Goins, R.E., 1994, In situ preparation of fatty acid methyl esters for analysis of
   fatty acid composition in foods. J Food Sci 59, 1262-1266.
- Pascual, J.J., Cervera, C., Blas, E., Fernandez Carmona, J., 1999, Effect of high fat diets on
  the performance, milk yield and milk composition of multiparous rabbit does. Anim Sci
  68, 151-162.
- Perret, J.P., 1980, Lipolyse gastrique des triglycérides du lait maternel et absorption gastrique
   des acides gras à chaîne moyenne chez le lapereau. J Physiol 76, 159-166.
- Petschow, B.W., Batema, R.P., Ford, L.L., 1996, Susceptibility of Helicobacter pylori to
   bactericidal properties of medium-chain monoglycerides and free fatty acids. Antimicrob
   Agents Chemother 40, 302-306.
- Recio, I., Visser, S., 1999, Two ion-exchange chromatographic methods for the isolation of
   antibacterial peptides from lactoferrin. In situ enzymatic hydrolysis on an ion-exchange
   membrane. J Chromatogr A 831, 191-201.
- 306 Skrivanova, V., Volek, Z., Zita, L., Marounek, M., 2004. Effect of triacylglycerols of caprylic
  307 and capric acid on performance, mortality and digestibility of nutrients in growing rabbits.
  308 In: 8th World Rabbit Congress, Puebla city, Mexico, September 7-10, pp. 991-995.
- 309 Sun, C.Q., O'Connor, C.J., Roberton, A.M., 2002, The antimicrobial properties of milkfat 310 after partial hydrolysis by calf pregastric lipase. Chem Biol Interact 140, 185-198.
- Van Immerseel, F., De Buck, J., Boyen, F., Bohez, L., Pasmans, F., Volf, J., Sevcik, M.,
  Rychlik, I., Haesebrouck, F., Ducatelle, R., 2004, Medium-chain fatty acids decrease
  colonization and invasion through hilA suppression shortly after infection of chickens with
- 314 Salmonella enterica serovar Enteritidis. Appl Environ Microbiol 70, 3582-3587.

Cox

#### 316 Legends

317 **Figure 1.** C10:0 has a bacteriostatic effect on E22 *in vitro* 

318 After 18 hours of E22 culture, mean CFU of each type of culture was determined by plating

319 on LB agar plates in duplicate. Results are expressed as the rate of growth inhibition for each

- 320 fatty acid concentration calculated from the LB control medium. Results represent the mean
- $(\pm SD)$  of 6 independent experiments.

322

**Figure 2.** MCT diet does not confer protection against lethal challenge with O103 EPEC.

324 A: MCT diet does not limit mortality induced by E22 strain. 28-days-old rabbits were orally

inoculated with  $10^4$  CFU of a non-pathogenic strain BM21 or a pathogenic strain E22 of E.

*coli*, and fed either with a diet containing 2% of SO or 2% of MCT. In E22 groups, only rabbits that died from colibacillosis are represented in the graph. In SO group, none of the

rabbits died from E22 colibacillosis. Final survival rates were 45.5%, 43.9%, 87% and 78.6%

in SO-E22, MCT-E22, SO-BM21 and MCT-BM21 groups, respectively.

B: MCT diet does not limit fecal excretion of E22 strain by infected rabbits. Total excretion
of *E. coli* is represented by triangles in BM21 group, and by the height of bars in E22 group.
Full bars indicate the proportion of Rh- strains (E22) among *E. coli* excreted, hatched bars
represent Rh+. SO-fed rabbits are represented in black and MCT-fed rabbits in grey. Data
show arithmetic means ± SD.

335

**Figure 3.** C8:0 and C10:0 do not prevent bacterial adhesion *in vivo* and *in vitro*.

A: MCT diet does not modify ileal bacterial attachment and intestinal inflammation. Ileal
tissue sections of MCT-fed (a) or SO-fed (b) rabbits infected at day 28 and sacrificed at day

339 31 were stained with anti-E22 sera and visualized at X400 magnification. Rabbits from each

- group presented only scarce bacterial adhesion (arrow). Villi were intact and no sign ofinflammation was visible.
- 342 B: MCFA do not inhibit bacterial adhesion on HeLa cells. Different concentrations of C8:0
- 343 and C10:0 were tested for their ability to inhibit bacterial adhesion on HeLa cells. Percentage
- of pEGFP-E22 positive HeLa cells was determined by flow cytometry, by detecting the GFP
- 345 expression of pEGFP-E22. Data are means  $\pm$  SD of 3 independent experiments.





#### ACCEPTED <u>NUSCRIPT</u> M. Δ.

A

