

Prevalence of infection in wild and farmed ungulates

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1	Veterinary Microbiology_Short Communication
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3	Prevalence of Coxiella burnetti infection in wild and farmed ungulates
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19 Abstract

20 The aim of this study was to evaluate by serology and PCR analyses the prevalence of C. 21 burnetti infection in ungulates in Spain. Sera were collected from red deer (Cervus elaphus; 22 n=116), roe deer (*Capreolus capreolus*; n=39), fallow deer (*Dama dama*; n=13) and cattle 23 (n=79). Sera were tested for anti-C. burnetii antibody detection by means of an 24 immunofluorescence antibody assay (IFA) and C. burnetii DNA was amplified by PCR in 25 samples from ungulates that had antibodies to phase II antigens. Twenty-nine, fifteen and 26 thirty-nine percent of the red deer, roe deer and cattle had antibodies against C. burnetii, 27 respectively. None of the fallow deer sera tested positive. Seroprevalence was statistically 28 higher in farmed than in wild red deer and higher in northern than in southern populations, 29 whereas an inverse pattern was observed for the roe deer. Most of the seropositive animals had only anti-C. burnetii phase II antibodies, thus showing the acute nature of infections in 30 31 the sampled ungulates. These results show that C. burnetii circulates in wild ungulates in 32 Spain and suggest that they can act as pathogen reservoirs for both domestic animals and 33 humans. 34

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³⁵ Keywords: *Coxiella*; Q fever; tick; vector-borne diseases; wildlife

43 Introduction

44 Q fever is a worldwide zoonosis produced by the gram-negative bacteria, *Coxiella* 45 *burnetii*. Multiple hosts can serve as reservoir of infection, including many wild and 46 domestic mammals, birds and ticks (Willeberg et al., 1980; Marrie et al., 1986; Webster et 47 al., 1995; Dunbar et al., 1998; Maurin and Raoult, 1999; Komiya et al., 2003). However, 48 domestic ruminants represent the most frequent source of *C. burnetii* infection in humans 49 (Maurin and Raoult, 1999).

50 Over 40 tick species are naturally infected with *C. burnetii*, including 51 *Rhipicephalus, Haemaphysalis, Amblyomma, Dermacentor, Ixodes* and *Otobius* species. 52 However, ticks are not considered essential in the natural cycle of *C. burnetii* in livestock 53 because other sources of infection are more important in animals that live in close contact 54 (Maurin and Raoult, 1999). In contrast, ticks may play a significant role in the transmission 55 of coxiellosis among wild vertebrates (Marrie et al., 1986; Maurin and Raoult, 1999).

56 Despite efforts to eradicate coxiellosis from cattle, goat and sheep herds, the disease 57 remains a serious risk for human and animal health in Spain (Tellez et al., 1988: Maurin 58 and Raoult, 1999; Maltezou and Raoult, 2002; Bolaños et al., 2003; Marrie, 2004; 59 Cardenosa et al., 2006; Oporto et al., 2006; Sanz et al., 2006). The disease seems to be 60 more prevalent in the Basque and Navarra provinces in northern Spain than in the central 61 and southern regions of the country (Tellez et al., 1988; Maurin and Raoult, 1999). Recent research efforts have been focused on the characterization of C. burnetii infection in 62 63 domestic ruminants and humans (reviewed by Woldehiwet, 2004; Kazar, 2005). However, 64 little is known about the prevalence of C. burnetii in wild ungulates (Enright et al., 1971; Ejercito et al., 1993; Marrie et al., 1993) and to our knowledge, C. burnetii infection has not 65 66 been characterized in Spanish wild and farmed cervids. Wild and farmed cervid populations

are growing in Spain and in other countries due to their value as game animals, thus
increasing the risk for disease transmission to humans and domestic animals (Gortázar et
al., 2006; Vicente et al., 2005).

The objective of the study reported herein was to evaluate by serology and PCR analyses the prevalence of *C. burnetii* infection in wild and farmed cervids from two regions in northern and southern Spain.

73 Materials and Methods

74 <u>Study sites</u>

In southern Spain, we collected sera of Iberian red deer (wild and farmed), European roe deer and cattle in a 3,000 hectares fenced hunting estate (LO) located in the Natural Park of "Los Alcornocales" in the province of Cádiz, Andalucía. We also collected European roe deer and cattle sera from a neighbouring hunting estate (OJ). In northern Spain, Iberian red deer, European roe deer and fallow deer serum samples were collected in Asturias region. The number of samples through ruminant species and habitat are summarized in table 1.

82 <u>Animals and sample preparation</u>

83 Wild cervids from Asturias region were necropsied after being hunter harvested. 84 Blood from shot wild cervids was collected directly from the heart or thoracic cavity into 85 sterile tubes without anticoagulant. Wild cervids from southern Spain were life-captured by gamekeepers with fixed capture boxes and bled by cervical puncture. Cattle were bled 86 87 during the tuberculosis control campaign. All samples were collected in 2004 and 2005. 88 Blood from captured/farmed wild cervids and cattle from southern Spain was collected into 89 sterile tubes with and without anticoagulant (EDTA) and maintained at 4°C until arrival at 90 the laboratory. Plasma and serum were then separated after centrifugation and stored at -

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91 20°C. DNA was extracted from sediments of serum samples using TriReagent (Sigma, St.

92 Louis, MO, USA). Ticks were collected from cervids and cattle and classified as previously

- 93 described (Ruiz-Fons et al., 2006).
- 94 <u>C. burnetii serologic tests</u>

95 Serum antibodies were determined for *C. burnetii* using an indirect 96 immunofluorescence antibody (IFA) test (Fuller Laboratories, Fullerton, CA, USA) and 97 following manufacturer's recommendations. The test for *C. burnetii* uses phase I and II 98 antigens with a cut-off dilution of the test serum of 1:16 (Torina et al., 2007). The IFA test 99 has been previously used to test deer sera for anti-*Coxiella burnetti* antibodies (Yasumoto 90 et al., 1997).

101 <u>C. burnetii htpB polymerase chain reaction (PCR) and sequence analysis</u>

The *C. burnetii htpB* gene, coding for a 62 kDa antigen (Vodkin and Williams, 103 1988) was amplified by nested PCR as previously reported (To et al., 1996). To check the 104 specificity of the PCR, amplified fragments were resin purified (Wizard, Promega) and 105 cloned into the pGEM-T vector (Promega) for sequencing both strands by double-stranded 106 dye-termination cycle sequencing (Secugen SL, Madrid, Spain). At least two independent 107 clones were sequenced for each PCR analyzed.

108 <u>Statistical analysis</u>

109 A chi-square test was used to test for the distribution patterns of *C. burnetii* 110 seroprevalence between sampling area (northern *vs.* southern), ungulate species and habitat 111 (wild *vs.* farmed). The same statistical test was used to evaluate the differences in the anti-112 phase I and II antibodies seroprevalence in cattle from LO farm. Statistical analyses were 113 performed using the SPSS v14.0 (SPSS Inc., Chicago, USA) package. The *p* value was set 114 at 0.05.

115 Results

Evidence of *C. burnetii* infection was found in red deer in northern and southern Spain and in roe deer and cattle in southern Spain. Seroprevalence of *C. burnetti* in the sampled ruminants was 15.4% in roe deer, 5.6% in wild red deer, 40% in farmed red deer and 39% in cattle. None of the fallow deer analyzed showed evidence of contact with *C. burnetii*. Seroprevalence level through species, habitat and location is shown in table 1.

Hyalomma marginatum marginatum and Hy. lusitanicum were found parasitizing
on red deer and cattle. Roe deer had infestations with Hy. m. marginatum. Rhipicephalus
bursa and R. pusillus were collected from roe deer and cattle and red deer and roe deer,
respectively. R. (Boophilus) annulatus was found on cattle only. Ixodes ricinus and
Haemaphysalis concinna were found infesting red deer and roe deer in Asturias.

The observed seroprevalence to *C. burnetii* antigens was higher in the southern red deer population (χ^2 =4.85, df=1, *p*<0.05; Table 1) and it was higher in farmed than in wild animals (χ^2 =9.05, df=1, *p*<0.01; Table 1). Furthermore, the observed seroprevalence of *C. burnetii* infection in wild red deer was higher in northern than in southern populations (Table 1). However, the opposite result was obtained for wild roe deer with higher observed seroprevalence in the southern population (χ^2 =8.27, df=1, *p*<0.01; Table 1).

In the study reported herein, anti-phase I antibodies were found in three cattle only in the farm LO in southern Spain (Table 1), which statistically differed from the number of anti-phase II antibodies positive cattle from the same farm ($\chi^2=22.6$, df=1, p<0.001). Interestingly, the observed seroprevalence of *C. burnetii* in cattle was higher in farm LO than in farm OJ in Andalucía, southern Spain ($\chi^2=21.5$, df=1, p<0.001; Table 1).

C. burnetii DNA was amplified by PCR in samples from wild and farmed red deerand cattle that had antibodies to phase II antigens (Table 1). As expected, the sequence of

- the amplicons corresponded to the *C. burnetii htpB* gene. Sequence analysis evidenced that
 all sequences were identical to the sequence reported for the Nine Mile phase I isolate
 (Genbank accession number M20482; Vodkin and Williams, 1988).
- 142 **Discussion**

This is to our knowledge the first report on *C. burnetti* infection in Spanish wild ungulates. As abovementioned, there is a need of research on pathogens circulating in wild ungulate populations due to its increasing demographic trend and especially due to their possible role as pathogen reservoirs for domestic animals and humans (see for example Ruiz-Fons et al., 2007).

The seroprevalence of C. burnetii found in wild roe (15.4%) and red deer (5.6%) 148 149 populations in Spain is within the range reported for other wild cervids. Recent 150 seroepidemiological studies in wild cervids have shown anti-C. burnetii antibody 151 prevalence of 16.5% and 1.5% in moose (Alces alces) and white-tailed deer (Odocoileus 152 virginianus) in Nova Scotia, USA, respectively (Marrie et al., 1993) and 69% and 56% in 153 Hokkaido deer (Cervus nippon vesoensis) and Japanese deer (Cervus nippon centralis) in 154 Hokkaido, Japan, respectively (Ejercito et al., 1993). We found no fallow deer to show 155 evidence of contact with C. burnetii, but the number of samples tested was smaller than in 156 other host species included in the study. Alternatively, this species may have innate 157 resistance to infection.

Our results suggest that in farmed red deer, animal-to-animal contact may increase the risk of *C. burnetii* transmission. Additionally, differences in the observed prevalence of *C. burnetii* infection in wild cervid populations may depend upon living environments (Komiya et al., 2003), which are different between northern and southern Spain. Management widely differs between the areas sampled in northern and southern Spain.

While sampled wild cervids in southern Spain live in fenced estates, northern wild cervids live in natural conditions. Feeders for both red and roe deer are present in the sampled southern estates and cervids use to concentrate on them, thus increasing the risk of *C*. *burnetii* and other pathogens transmission between individuals (Acevedo et al., 2007).

167 Anti-C. burnetii phase II antibodies are generally associated with acute infections 168 whereas in chronic infections, the antibody level to phase I is usually higher than that to 169 phase II (Maurin and Raoult, 1999). Our results suggest that most of the animals analyzed 170 had acute C. burnetii infections, which may correlate with heavy tick infestations in these 171 animals. Tick species found are common on wild ungulates in Spain (de la Fuente et al., 172 2004; Ruiz-Fons et al., 2006) and have been suspected to play an important role as vectors 173 of C. burnetii among wild vertebrates (Marrie et al., 1986; Maurin and Raoult, 1999). Nevertheless, in order to definitively determine the real role of wild ungulates as C. burnetti 174 175 reservoirs for domestic animals and humans, an experimental approach is needed.

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184 **References**

185	Acevedo, P., Vicente, J., Höfle, U., Cassinello, J., Ruiz-Fons, F., Gortázar, C.,
186	2007. Estimation of European wild boar relative abundance and aggregation: a novel
187	method in epidemiological risk assessment. Epidemiol. Infect. 135, 519-527.
188	Bolaños, M., Santana, O.E., Angel-Moreno, A., Perez-Arellano, J.L., Limiñana,
189	J.M., Serra-Majem, L., Martin-Sánchez, A.M., 2003. Seroprevalence of infection by
190	Coxiella burnetii in Canary Islands (Spain). Eur. J. Epidemiol. 18, 259-262.
191	Cardenosa, N., Sanfeliu, I., Font, B., Munoz, T., Nogueras, M.M., Segura, F., 2006.
192	Short report: seroprevalence of human infection by Coxiella burnetii in Barcelona
193	(northeast of Spain). Am. J. Trop. Med. Hyg. 75, 33-35.
194	de la Fuente, J., Naranjo, V., Ruiz-Fons, F., Vicente, J., Estrada-Peña, A., Almazán,
195	C., Kocan, K.M., Martín, M.P., Gortázar, C., 2004. Prevalence of tick-borne pathogens in
196	ixodid ticks (Acari: Ixodidae) collected from European wild boar (Sus scrofa) and Iberian
197	red deer (Cervus elaphus hispanicus) in central Spain. Eur. J. Wild. Res. 50, 187-196.
198	Dunbar, M.R., Cunningham, M.W., Roof, J.C., 1998. Seroprevalence of selected
199	disease agents from free-ranging black bears in Florida. J. Wildl. Dis. 34, 612-619.
200	Ejercito, C.L., Cai, L., Htwe, K.K., Taki, M., Inoshima, Y., Kondo, T., Kano, C.,
201	Abe, S., Shirota, K., Sugimoto, T., Yamaguchi, T., Fukushi, H., Minamoto, N., Kinjo, T.,
202	Isogai, E., Hirai, K., 1993. Serological evidence of Coxiella burnetii infection in wild
203	animals in Japan. J. Wildl. Dis. 29, 481-484.
204	Enright, J.B., Franti, C.E., Behymer, D.E., Longhurst, W.M., Dutson, V.J., Wright,
205	M.E., 1971. Coxiella burneti in a wildlife-livestock environment. Distribution of Q fever in
206	wild mammals. Am. J. Epidemiol. 94, 79-90.
207	Gortázar, C., Acevedo, P., Ruiz-Fons, F., Vicente, J., 2006. Disease risks and
208	overabundance of game species. Eur. J. Wildl. Res. 52, 81-87.

209	Kazar, J., 2005. Coxiella burnetii infection. Ann. N. Y. Acad. Sci. 1063, 105-114.
210	Komiya, T., Sadamasu, K., Kang, M.I., Tsuboshima, S., Fukushi, H., Hirai, K.,
211	2003. Seroprevalence of Coxiella burnetii infections among cats in different living
212	environments. J. Vet. Med. Sci. 65, 1047-1048.
213	Maltezou, H.C., Raoult, D., 2002. Q fever in children. Lancet Infect. Dis. 2, 686-
214	691.
215	Marrie, T.J., Schlech, W.F., Williams, J.C., Yates, L., 1986. Q fever pneumonia
216	associated with exposure to wild rabbits. Lancet 1, 427–429.
217	Marrie, T.J., Embil, J., Yates, L., 1993. Seroepidemiology of Coxiella burnetii
218	among wildlife in Nova Scotia. Am. J. Trop. Med. Hyg. 49, 613-615.
219	Marrie, T.J., 2004. Q fever pneumonia. Curr. Opin. Infect. Dis. 17, 137-142.
220	Maurin, M., Raoult, D., 1999. Q fever. Clin. Microbiol. Rev. 12, 518-553.
221	Oporto, B., Barandika, J.F., Hurtado, A., Moreno, B., Aduriz, G., García-Pérez,
222	A.L., 2006. Incidence of ovine abortion by Coxiella burnetii in Northern Spain. Ann. N. Y.
223	Acad. Sci. 1078, 498-501.
224	Ruiz-Fons, F., Fernández de Mera, I.G., Acevedo, P., Höfle, U., Vicente, J., de la
225	Fuente, J., Gortázar, C., 2006. Ticks (Acari: Ixodidae) parasitizing Iberian red deer (Cervus
226	elaphus hispanicus) and European wild boar (Sus scrofa) from Spain: geographical and
227	temporal distribution. Vet. Parasitol. 140, 133-142.
228	Ruiz-Fons, F., Segalés, J., Gortázar, C., 2007. A review of viral diseases of the
229	European wild boar: Effects of population dynamics and reservoir role. Vet. J., in press.
230	Sanz, J.C., de los Rios, R., Martin, F., Tebar, M.A., Jado, I., Anda, P., 2006.
231	Aplicación de cuatro métodos de ELISA (dos para IgM y dos para IgG) para el diagnóstico
232	serológico de un brote de fiebre Q. Enferm. Infecc. Microbiol. Clin. 24, 178-181.

233	Tellez, A., Sainz, C., Echevarria, C., De Carlos, S., Fernandez, M.V., Leon, P.,
234	Rezina, R., 1988. Q fever in Spain: acute and chronic cases 1981–1985. Rev. Infect. Dis.
235	10, 198–202.
236	To, H., Kako, N., Zhang, G.Q., Otsuka, H., Ogawa, M., Ochiai, O., Nguyen, S.V.,
237	Yamaguchi, T., Fukushi, H., Nagaoka, N., Akiyama, M., Amano, K., Hirai, K., 1996. Q
238	fever pneumonia in children in Japan. J. Clin. Microbiol. 34, 647-651.
239	Torina, A., Vicente, J., Alongi, A., Scimeca, S., Turlá, R., Nicosia, S., Di Marco,
240	V., Caracappa, S., de la Fuente, J., 2007. Observed prevalence of tick-borne pathogens in
241	domestic animals in Sicily, Italy during 2003-2005. Zoonoses Public Health, in press.
242	Vicente, J., Ruiz-Fons, F., Vidal, D., Höfle, U., Acevedo, P., Villanua, D.,
243	Fernández-de-Mera, I.G., Martín, M.P., Gortázar, C., 2005. Serosurvey of Aujeszky's
244	disease virus infection in European wild boar in Spain. Vet. Rec. 156, 408-412.
245	Vodkin, M.H., Williams. J.C., 1988. A heat shock operon in Coxiella burnetii
246	produces a major antigen homologous to a protein in both mycobacteria and Escherichia
247	<i>coli</i> . J. Bacteriol. 170, 1227–1234.
248	Webster, J.P., Lloyd, G., Macdonald, D.W., 1995. Q fever (Coxiella burnetii)
249	reservoir in wild brown rat (Rattus norvegicus) populations in the UK. Parasitol. 110, 31-
250	35.
251	Willeberg, P., Ruppanner, R., Behymer, D.E., Haghighi, S., Kaneko, J.J., Franti,
252	C.E., 1980. Environmental exposure to Coxiella burnetii: a sero-epidemiologic survey
253	among domestic animals. Am. J. Epidemiol. 111, 437-443.
254	Woldehiwet, Z., 2004. Q fever (coxiellosis): epidemiology and pathogenesis. Res.
255	Vet. Sci. 77, 93-100.

- 256 Yasumoto, M., Yanase, T., Muramatsu, Y., Morita, C., Ueno, H., 1997.
- 257 Seroepidemiological study of *Coxiella burnetti* in *Cervus nippon* in northern Japan. Jpn. J.
- 258 Zoo Widl. Med. 2, 101-106.

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Table 1. Results of IFA test to phase I and II C. burnetii antigens and htpB gene nested

PCR in wild and domesticated cervids and cattle.

Host	Habitat Locatio	Location	N° of samples	Seropositive (% ± SE at 95% CI)		Nested PCR positive/total
				Phase I	Phase II	tested (%) ^a
	Wild	Northern Spain	21	$0 (0 \pm 0)$	2 (9.5 ± 12.9)	1/2 (50)
Red deer	Wild	Southern Spain	15	$0 (0 \pm 0)$	$0 (0 \pm 0)$	None tested
	Farm LO	Southern Spain	80	$0 (0 \pm 0)$	32 (40 ± 10.8)	4/32 (12)
Subtotal red deer			116	0 (0 ± 0)	<i>34 (29.3</i> ± <i>8.2)</i>	5/34 (14.7)
Roe deer	Wild	Northern Spain	21	$0\ (0\pm0)$	$0 (0 \pm 0)$	None tested
Köe deel	Wild	Southern Spain	18	$0~(0\pm0)$	6 (33 ± 22.4)	0/6 (0)
Subtotal roe deer			39	0 (0 ± 0)	6 (15.4 ± 11.6)	0/6 (0)
Fallow deer	Wild	Northern Spain	13	$0 (0 \pm 0)$	$0 (0 \pm 0)$	None tested
	Farm LO	Southern Spain	49	3 (6±4.9)	26 (53 ± 10.6)	3/26 (12) ^b
Cattle	Farm OJ	Southern Spain	30	$0 (0 \pm 0)$	2 (7 ± 5.1)	0/2 (0)
Subtotal cattle			79	3 (3.8 ± 4.3)	28 (35.4 ± 10.6)	3/28 (10.7)

^a Only samples from seropositive animals were tested by C. burnetii htpB gene nested

PCR.

^b Samples from animals seropositive for phase I *C. burnetii* antibodies were negative for the *htpB* gene nested PCR assay.