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Altered TGFbeta signaling and cardiovascular manifestations in patients with autosomal recessive cutis laxa type I caused by fibulin-4 deficiency

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25 Running title: TGFb signaling in patients with *FBLN4* deficiency

26

27 ABSTRACT

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29 Fibulin-4 is a member of the fibulin family, a group of extracellular matrix proteins 30 prominently expressed in medial layers of large veins and arteries. Involvement of the 31 FBLN4 gene in cardiovascular pathology was shown in a murine model and in three 32 patients affected with cutis laxa in association with systemic involvement. In order to elucidate the contribution of FBLN4 in human disease, we investigated two cohorts of 33 34 patients. Direct sequencing of 17 patients with cutis laxa revealed no FBLN4 mutations. In 35 a second group of 22 patients presenting with arterial tortuosity, stenosis and aneurysms, FBLN4 mutations were identified in three patients, two homozygous missense mutations 36 (p.Glu126Lys and p.Ala397Thr) and compound heterozygosity for missense mutation 37 p.Glu126Val and frameshift mutation c.577delC. Immunoblotting analysis showed a 38 decreased amount of fibulin-4 protein in the fibroblast culture media of two patients, a 39 finding sustained by diminished fibulin-4 in the extracellular matrix of the aortic wall on 40 immunohistochemistry. pSmad2 and CTGF immunostaining of aortic and lung tissue 41 42 revealed an increase in TGFbeta signaling. This was confirmed by pSmad2 immunoblotting of fibroblast cultures. In conclusion, patients with recessive FBLN4 mutations are 43 predominantly characterized by aortic aneurysms, arterial tortuosity and stenosis. This 44 45 confirms the important role of fibulin-4 in vascular elastic fiber assembly. Furthermore, we

46 provide the first evidence for the involvement of altered TGFbeta signaling in the47 pathogenesis of *FBLN4* mutations in humans.

48

49 **KEYWORDS**

50 TGFbeta, fibulin-4, cutis laxa, aortic aneurysm, arterial tortuosity

51

52 **INTRODUCTION**

53

Fibulins are a seven-member family of extracellular matrix proteins that play a role 54 in both elastic fiber assembly and function ¹⁻⁴. Fibulin-4 is prominently expressed in the 55 medial layers of large veins and arteries and in small capillaries ⁵. The importance of 56 fibulin-4 to elastogenesis was shown in studies of *fibulin-4^{-/-}* mice which have severe lung 57 and vascular defects including emphysema, aortic/arterial aneurysms or stenosis and 58 arterial tortuosity ⁶⁻⁷. In the aorta of the *fibulin-4^{-/-}* mouse, an increase in TGFbeta signaling 59 was shown⁶, a phenomenon that has previously been described in other human aortic 60 aneurysm syndromes, including Marfan syndrome (MFS, MIM 154700), Loeys-Dietz 61 syndrome (LDS, MIM 610168 and 609192) and Arterial Tortuosity Syndrome (ATS, MIM 62 208050)⁸⁻¹⁰. Recently, it was shown that fibulin-4 deficiency not only leads to altered 63 64 elastic fiber assembly due to its role in elastogenesis, but also results in a defective smooth muscle cell terminal differentiation with downregulation of smooth muscle contractile 65 proteins ¹¹. Both processes can potentially contribute, separately or in concert, to aneurysm 66 67 formation.

So far, FBLN4 (also called EFEMP2) mutations have been described in three 68 patients with autosomal recessive cutis laxa type I (MIM 219100)¹²⁻¹⁴. These patients 69 presented prenatally with fragmented elastic tissues, cutis laxa and variable pulmonary and 70 cardiovascular involvement (Table 1). Cardiorespiratory complications often lead to death 71 72 during childhood. Mutations in FBLN5, the gene encoding the fibulin-5 protein, also cause autosomal recessive and autosomal dominant cutis laxa type I¹⁵⁻¹⁸ and are typically 73 associated with pulmonary emphysema. On the other hand, the X-linked form (MIM 74 75 304150), caused by mutations in the gene encoding a copper transporting ATPase (ATP7A), 76 is now classified within the group of copper deficiency syndromes. Type II autosomal 77 recessive cutis laxa (MIM 219200) presents with joint laxity and developmental delay. 78 Recently, it was shown that mutations in the ATP6V0A2 gene, encoding the α^2 unit of the V-type H^+ ATPase, resulted in this cutis laxa subtype by causing impaired glycosylation ¹⁹. 79 An additional form of autosomal recessive cutis laxa resembling type II is De Barsy 80 syndrome (MIM 219150). Patients have a progeroid-like appearance, typical facial features, 81 growth delay and cutis laxa²⁰⁻²¹. Recently, disease causing mutations were identified in 82 *PYCR1*, the gene encoding pyrroline-5-carboxylate reductase 1^{22-23} . Patients presenting 83 with autosomal dominant cutis laxa (MIM 123700), caused by alterations in the gene 84 encoding the elastin protein, ELN²⁴⁻²⁷, have a normal life span and internal organ 85 86 abnormalities are less prominent.

In this study we screened *FBLN4*, which encodes the fibulin-4 protein, in two phenotypically distinct cohorts, the first with cutis laxa, and the second with arterial tortuosity, stenosis and aneurysms. A potential link with TGFbeta-signaling and the effect of the different mutations on fibulin-4 protein expression were investigated.

92 MATERIALS AND METHODS

93 Patient data

Two patient cohorts were investigated in this study. The first cohort consisted of 17 patients with predominant cutis laxa without major cardiovascular findings (supplementary table S1). The 22 patients in the second cohort had mild skin involvement but significant cardiovascular features, such as arterial tortuosity, stenosis and aneurysms (supplementary table S1). We describe here the clinical details of all three patients in whom we found *FBLN4* mutations, and summarize the clinical data of two previously reported patients from whom we studied fibroblasts or tissues.

101 A currently 20-year-old female (patient 1) presented at the age of 2 months with airway compression, ascending aortic aneurysm, proximal pulmonary arterial stenosis, 102 103 distal pulmonary arterial dilatation and innominate artery dilatation. She underwent cardiovascular surgery at 2.5, 7 and 8 months of age. By age 39 months, she had abdominal 104 arterial tortuosity and dilatation. Clinical findings included a high arched palate, 105 micrognathia, mild joint hypermobility, velvety skin, no cutis laxa and normal scarring. A 106 cerebral angiogram at the age of 5 years showed arterial tortuosity of the internal carotid, 107 108 anterior/middle cerebral, and vertebral arteries. Histology of aortic biopsies showed 109 disrupted elastic fibers and increased deposition of glycosaminoglycans. Detailed clinical description has been published previously 28 . 110

Patient 2 is a 3-year-old boy with a flat face, prominent forehead, hypertelorism, highly arched palate, pectus excavatum and joint hypermobility. He had a wide mediastinum on routine chest radiography. Echocardiography showed a dilated ascending aorta and aortography showed pseudocoarctation and aortic tortuosity. The clinical features
 of this patient have been reported previously ²⁹. To date, the patient is 7 years old and the
 dilatation of his ascending aorta increased to 54 mm. The patient is asymptomatic.

Patient 3 was born at 37 weeks of gestation. Clinical features included a prominent 117 118 forehead, mild hypertelorism, down-slanting palpebral fissures, depressed nasal bridge, 119 low-set and distorted external ears and long fingers. An echocardiogram showed severe 120 dilatation of the ascending aorta and severe tortuosity of the entire aorta with hypoplasia of 121 the transverse aortic arch, proximal descending, thoracic, and abdominal aorta. MRI 122 showed severe tortuosity of the carotid and cerebral arteries. At 5 weeks of age, she 123 suffered a large hemispheric stroke. She died from cardiorespiratory failure at 124 approximately 18 months. Autopsy was declined. Neither parent of patient 3 had phenotypcial characteristics of ARCL type I. 125

Patient 4 was described recently ¹². This female patient presented with mild cutis laxa, arachnodactyly and systemic involvement, including pulmonary hypertension, mild tricuspid valve insufficiency, abdominal aortic tortuosity, and dilatation of the ascending aorta and the main branches of the pulmonary arteries, with dissection. The parents denied consanguinity. She died at 27 days of age from respiratory distress and inoperable systemic vascular abnormalities.

Patient 5 is a child from consanguineous parents. She died shortly after birth from profound bradycardia. Clinical features included facial dysmorphism, cutis laxa, joint contractures and arachnodactyly. She had a thickened myocardium with a minimal pericardial effusion, emphysema and fragile vessels. A detailed clinical description has been reported recently ¹⁴.

138 Molecular analysis

For patients 1-3, genomic DNA was extracted from EDTA blood samples using the Puregene method (Qiagen, Venlo, The Netherlands). All coding exons and flanking introns of *FBLN4* were amplified by PCR on genomic DNA level (primer sequences are available upon request). Products were sequenced using the BigDye terminator cycle sequencing method on the ABI3730 XL automatic sequencer (Applied Biosystems, Halle, Belgium).

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145 Cell culture

146 Skin fibroblasts were available from patients 1 and 5, and age- and sex-matched 147 controls. Fibroblasts were cultured in Dulbecco's modified Eagle's medium (DMEM, 148 Invitrogen, Merelbeke, Belgium) supplemented with 10% fetal calf serum (FCS, 149 Invitrogen) in the presence of antibiotics at 37°C in 5% CO₂.

150

151 **Protein analysis**

Confluent fibroblasts of patient 1 and patient 5 and age- and sex-matched controls 152 were grown in DMEM medium in the absence of FCS for 24 hours. Media were harvested 153 and concentrated (Centriprep-30, Millipore, Brussels, Belgium) in the presence of protease 154 155 inhibitors (Complete Mini tablet, Roche Applied Science, Vilvoorde, Belgium). Protein samples were loaded on gel together with 5x non-reducing lane marker sample buffer 156 (Thermo Fisher Scientific, Aalst, Belgium). SDS-PAGE electrophoresis was performed 157 158 using NuPage 4-12% Bis-Tris precast gels (Invitrogen) followed by electrotransfer of the proteins onto a Hybond-ECL nitrocellulose membrane (GE healthcare) using the iBlot 159

system (Invitrogen). Blocking of the membrane was done in 2% ECL-advantage buffer (GE 160 161 healthcare). Immunoblotting was carried out using a mouse monoclonal anti-human fibulin-4 antibody (clone 226,347,597) (1:500) in combination with a mouse monoclonal anti-162 human fibronectin-1 antibody (clone 84) (1:500), followed by incubation with the 163 164 secondary antibody ECLplex goat-anti-mouse IgG-Cy5 (1/2000) (GE healthcare). 165 Monoclonal antibodies were generated in mice immunized with full-length recombinant human fibulin-4³⁰. Epitopes for all three monoclonal antibodies are located in the N-166 167 terminal half of fibulin-4. Membranes were scanned using the Typhoon 9400 (GE healthcare). Immunoblotting of pSmad2 was performed as previously described ³¹. 168 169 Quantification of the immunoblots was performed using ImageJ software.

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171 Histological analysis

172 Paraffin-embedded lung and aortic tissue were available from patient 4. For immunohistochemical staining of fibulin-4 (clone 347), connective tissue growth factor 173 (CTGF) and pSmad2 (the phosphorylated form of Smad2) in lung and aortic tissue, we 174 selected representative specimens from three healthy control individuals. From these 175 formalin-fixed, paraffin-embedded specimens, 5µm thick sections were cut, deparaffinized 176 and rehydrated. Antigens were unmasked using 10mM sodium citrate buffer and auto-177 178 peroxidase activity was inhibited. Sections were blocked with normal goat serum (pSmad2 and CTGF) or normal horse serum (fibulin-4) (Vectastain, Burlingame, CA, USA). 179 Antibodies directed against CTGF (Abcam, Cambridge, UK), pSmad2 (Ser465/467) (Cell 180 181 Signaling Technology, Boston, MA, USA), and fibulin-4 (see protein analysis section) were used. Subsequently, sections were incubated with a secondary antibody, goat-anti-182

rabbit IgG (pSmad2 and CTGF) or horse-anti-mouse (fibulin-4) (Vectastain), ABC (Avidin:
Biotinylated enzyme Complex) reagent (Vectastain) and DAB (3,3'-Diaminobenzidine)
peroxidase (Vectastain). Sections were dehydrated in xylene and mounted. Light
microscopy was performed on a Zeiss Axio Imager A1 microscope.

187

188 **RESULTS**

189 FBLN4 mutation screening

190 We sequenced FBLN4 in two cohorts of patients. The 17 patients of the first cohort 191 presented predominantly with cutis laxa and had screened negative previously for ELN and 192 FBLN5 mutations. The second cohort included 22 patients who were diagnosed with 193 arterial tortuosity, stenosis and aneurysms. This cohort screened negative previously for SLC2A10, FBN1, TGFBR1 and TGFBR2 mutations. Molecular study of the first cohort did 194 195 not identify any FBLN4 mutations. In the second cohort FBLN4 mutations were identified in three patients. Patients 1 and 2 have homozygous missense mutations, c.376G>A196 (p.Glu126Lys) and c.1189G>A (p.Ala397Thr) (Figure 2A and B). The first mutation 197 replaces the same glutamic acid residue from the EGF consensus sequence as the 198 previously reported p.Glu57Lys¹³ but in a different cbEGF domain, whereas the second 199 200 mutation affects a highly conserved amino acid in the fibulin-type module. The parents of 201 patient 2 were heterozygous for the *p.Ala397Thr* mutation. The third patient (Figure 1) harbors a missense mutation, c.377A > T (p.Glu126Val), in combination with a frameshift 202 mutation, c.577delC leading to a premature termination codon (p.Gln193Ser fs X12) 203 204 (Figure 2C). Parental studies confirmed that these mutations occurred in trans. The missense mutation in patient 3 affects the same glutamic acid residue (at position 126) as 205

identified in patient 1, but leads to a different amino acid substitution. Parental samples
were not available for molecular analysis. None of the mutations were identified in 200
control chromosomes. An overview of the molecular and clinical data of these three
patients and the *FBLN4*-mutation positive patients described previously in the literature
(patients 4-6) is given in Table 1.

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212 Fibulin-4 expression in human skin fibroblasts and aortic tissue

213 To investigate the effect of the FBLN4 mutations on the expression of fibulin-4 214 protein, an immunoblotting experiment was performed on the culture medium of dermal 215 fibroblasts of patient 1 (homozygous c.376G > A (p.Glu126Lys) missense mutation), patient 216 5 (homozygous c.800G>A (p.Cys267Tyr) missense mutation), and two age- and sexmatched controls. Immunoblotting showed a distinct protein band of approximately 49 kDa 217 218 in the culture medium of the control samples. In patient 5 a complete absence and in patient 1 a slightly diminished amount of fibulin-4 in the culture medium was observed (Figure 219 3B). In addition, we performed immunohistochemical staining of fibulin-4 on aortic tissue 220 of a control individual and patient 4, which confirmed the near-absence of extracellular 221 fibulin-4 in the aortic wall of patient 4 in comparison to its control. Similar intracellular 222 223 fibulin-4 staining was observed in both individuals (Figure 3A). These results suggest that 224 the mutant proteins are either not secreted, or are secreted into the extracellular space to a lesser extent. The impaired secretion of mutant fibulin-4 leads to a decrease of available 225 fibulin-4 protein in the extracellular matrix for elastogenesis, and disturbed elastic fiber 226 227 formation.

229 Increased TGFbeta signaling in patients with *FBLN4* mutations

230 To determine whether mutations in the human FBLN4 gene have an effect on the TGFbeta signaling pathway, immunohistochemical staining for phosphorylated Smad2 231 (pSmad2, an effector of TGFbeta signaling) and CTGF (a TGFbeta-driven gene product) in 232 233 the aorta and lungs of a normal individual and patient 4 was performed. We demonstrated a 234 more intense and increased nuclear pSmad2 staining in the aorta and lung of the patient that 235 was not present in the control tissues (Figure 4). Following TGFbeta stimulation, 236 immunoblotting indicated a significantly increased pSmad2 signal in fibroblast cultures of 237 patient 1 and patient 5 in comparison with their respective controls (Figure 5). These 238 findings suggest that mutations in FBLN4 have an effect on the TGFbeta signaling 239 pathway. This hypothesis is supported by increased intensity of staining for CTGF in the aortic and lung tissue of patient 4. 240

241

242 **DISCUSSION**

243

244 We identified FBLN4 mutations in three unrelated patients clinically characterized by major cardiovascular defects, including aortic aneurysms, arterial tortuosity and 245 stenosis, and minor skin involvement. This phenotype is different from that seen in patients 246 247 with mutations in the genes encoding the elastin protein, ELN, and the fibulin-5 protein, FBLN5, in which skin involvement is the most prominent clinical finding. Although the 248 patients described in this paper share clinical characteristics with previously reported 249 FBLN4 mutation positive patients ¹²⁻¹⁴, there is considerable clinical overlap with Loeys-250 Dietz syndrome (LDS) and Arterial Tortuosity syndrome (ATS). LDS is an autosomal 251

dominant disorder characterized by a triad of arterial tortuosity and aneurysms, 252 253 hypertelorism and bifid uvula or cleft palate. The disorder is caused by mutations in the genes encoding transforming growth factor beta receptor 1 and 2 (TGFBR1 and TGFBR2, 254 respectively)^{9,31}. The autosomal recessive disorder, ATS, is characterized by tortuosity, 255 elongation, stenosis and aneurysm formation of the major arteries ³². Coucke et al.⁸ 256 257 identified SLC2A10, the gene encoding a facilitative glucose transporter GLUT10, as the 258 causative gene. The patients investigated in this study were referred because of clinical 259 features reminiscent of one or more of these disorders. This report doubles the number of known patients with FBLN4 mutations ¹²⁻¹⁴. Our findings underscore the prominent 260 261 vascular involvement and the rather mild cutaneous findings associated with FBLN4 262 mutations. As such, screening of FBLN4 should be considered particularly in patients with findings suggestive of LDS or ATS, in whom TGFBR1, TGFBR2 or SLC2A10 mutation 263 264 screening is negative. We report for the first time long-term survival in patient 1 and patient 2, 20 and 7 years old, respectively. This is an interesting observation in view of the early 265 266 lethality previously reported in FBLN4-mutation positive patients and Fbln4-deficient mice 7. 267

We report four novel *FBLN4* mutations. Several lines of evidence suggest the disease causing nature of these mutations. First, the three missense mutations cause either a change of charge (p.Glu126Lys and p.Glu126Val), a change of hydrophilic side chain to hydrophobic side chain (p.Glu126Val), or the addition of a hydroxyl group (p.Ala397Thr). Secondly, these missense mutations modify highly conserved residues within the fibulin-4 sequence. The glutamic acid residue at position 126 is located in the DINE consensus sequence of the second cbEFG-like module, which is essential for Ca²⁺ binding. Ca²⁺ binding of EGF modules is crucial for module-module interactions, and hence, for the structural and binding properties of the intact protein ³³. Third, *in silico* prediction programs, Polyphen and SIFT, classify the missense mutations as probably (*p.Glu126Lys* and *p.Glu126Val*) or possibly (*p.Ala397Thr*) damaging and not tolerated, respectively. Finally, the alanine residue at position 397 is highly conserved among species and between the homologous fibulin-3, -4 and -5 proteins (Figure 2B).

281 Immunoblotting analysis showed the absence (patient 5) or a slightly decreased 282 amount (patient 1) of the mutant fibulin-4 protein in the culture media of the patients in 283 comparison with the controls. This result mirrors the difference in disease severity 284 manifested clinically between these two patients. A possible explanation for the difference 285 in secretion between these two patients is that the homozygous alteration of a cysteine residue in patient 5 (p.Cys267Tyr) has a more dramatic effect on protein folding and 286 287 trafficking by the altered disulfide bridge formation compared to the glutamic acid to lysine alteration in patient 1. However, two previously reported FBLN4 mutations, p.Glu57Lys¹³ 288 and *p.Arg279Cys* in combination with *c.1070_1073dupCCGC*¹² show complete absence of 289 the mutant protein in the extracellular matrix, the latter was confirmed in this report by 290 immunohistochemical staining of fibulin-4 on aortic tissue. This indicates that the 291 diminished amount of mutant fibulin-4 present in the extracellular matrix of patient 1 is the 292 293 first example of a patient with FBLN4 mutation with the production and secretion of the mutant protein. This might be in accordance with the longer survival of the patient. Given 294 these observations we suggest that mutations in *FBLN4* impair the stability and/or secretion 295 of fibulin-4, similarly to prior findings for fibulin-5 mutations³⁴. The resulting decrease of 296 protein in the extracellular matrix leads to altered interactions with fibulin-4 binding 297

partners and subsequently, to impaired elastogenesis. Fibulin-4 is known to associate with 298 tropoelastin, possibly connecting elastin to the microfibrils to form the elastic fibers ³⁰. 299 Secondly, fibulin-4 also strongly binds to the N-terminal region of fibrillin-1 in the 300 presence of Ca^{2+ 35-36}. As suggested by Ono and colleagues ³⁶, fibulin-4 competes with 301 302 latent TGFbeta binding proteins (LTBPs) for binding to fibrillin-1. As such, this fibulin 303 protein may play a role in the modulation of LTBP sequestration by the extracellular 304 matrix, an event important in the regulation of the bioavailability of TGFbeta for activation 305 and signaling. As a result of impaired mutant fibulin-4 secretion and/or stability and the 306 resulting impaired elastogenesis, we believe that LTBP is no longer able to bind the 307 fragmented elastic fibers. The TGFbeta-LAP (latency associated peptide) complex is 308 released and TGFbeta is subsequently more prone to activation. An analogous mechanism has previously been suggested to occur in MFS and LDS^{10,31,37}. Additionally, the resulting 309 310 decrease of mutant fibulin-4 protein in the extracellular matrix may lead to defective 311 smooth muscle terminal differentiation as recently suggested by Huang and colleagues. They hypothesized that fibulin-4 deficiency not only results in aneurysm formation through 312 defective elastic fiber formation, but also through extracellular regulation of smooth muscle 313 cell differentiation genes¹¹. 314

To investigate whether increased TGFbeta signaling is involved in the pathogenesis of autosomal recessive cutis laxa type I in humans, as predicted based on the observations of impaired TGFbeta signaling in *Fbln4-/-* mice by Hanada and colleagues ⁶, we assessed the level of pSmad2 and CTGF expression in aorta and lung and subsequently performed pSmad2 immunoblotting on fibroblast cultures before and after TGFbeta stimulation. TGFbeta-stimulated fibroblast cultures of patients with *FBLN4* mutations in this study

showed an increased and more intense level of phosphorylated Smad2 compared to 321 controls. This finding is in line with an "in vivo" increase in TGFbeta signaling 322 demonstrated in the aorta and lung of the fibulin-4 deficient patient 4. The increased 323 TGFbeta signaling in aortic tissue of the FBLN4-mutation positive patients confirms the 324 325 key role of this signaling pathway in the pathogenesis of aortic and arterial aneurysms and 326 tortuosity, as demonstrated previously in other human diseases such as MFS, LDS and ATS ^{8-10,32}. As already shown in Marfan mouse models, this finding offers the potential for 327 328 treatment by blocking TGFbeta signaling through the use of angiotensin receptor type 1 blockers such as losartan³⁸. 329

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344 CONFLICT OF INTEREST

345 The authors declare no conflict of interest.

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- 347 Supplementary information is available at European Journal of Human Genetics' website.
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349 **REFERENCES**

- Argraves WS, Greene LM, Cooley MA, Gallagher WM: Fibulins: physiological and disease
 perspectives. *EMBO Rep* 2003; 4: 1127-1131.
- Nakamura T, Lozano PR, Ikeda Y *et al*: Fibulin-5/DANCE is essential for elastogenesis in vivo.
 Nature 2002; **415**: 171-175.
- 3563.Sasaki T, Gohring W, Miosge N, Abrams WR, Rosenbloom J, Timpl R: Tropoelastin binding357to fibulins, nidogen-2 and other extracellular matrix proteins. *FEBS Lett* 1999; **460:** 280-358284.
- Yanagisawa H, Davis EC, Starcher BC *et al*: Fibulin-5 is an elastin-binding protein essential
 for elastic fibre development in vivo. *Nature* 2002; **415**: 168-171.
- 3635.Giltay R, Timpl R, Kostka G: Sequence, recombinant expression and tissue localization of364two novel extracellular matrix proteins, fibulin-3 and fibulin-4. *Matrix Biol* 1999; **18**: 469-365480.
- 367 6. Hanada K, Vermeij M, Garinis GA *et al*: Perturbations of vascular homeostasis and aortic
 368 valve abnormalities in fibulin-4 deficient mice. *Circ Res* 2007; **100**: 738-746.
- 370 7. McLaughlin PJ, Chen Q, Horiguchi M *et al*: Targeted disruption of fibulin-4 abolishes
 371 elastogenesis and causes perinatal lethality in mice. *Mol Cell Biol* 2006; **26**: 1700-1709.
- Coucke PJ, Willaert A, Wessels MW *et al*: Mutations in the facilitative glucose transporter
 GLUT10 alter angiogenesis and cause arterial tortuosity syndrome. *Nat Genet* 2006; 38:
 452-457.
- 377 9. Loeys BL, Schwarze U, Holm T *et al*: Aneurysm syndromes caused by mutations in the TGF378 beta receptor. *N Engl J Med* 2006; **355**: 788-798.
- Neptune ER, Frischmeyer PA, Arking DE *et al*: Dysregulation of TGF-beta activation
 contributes to pathogenesis in Marfan syndrome. *Nat Genet* 2003; **33**: 407-411.

- Huang J, Davis EC, Chapman SL *et al*: Fibulin-4 Deficiency Results in Ascending Aortic
 Aneurysms. A Potential Link Between Abnormal Smooth Muscle Cell Phenotype and
 Aneurysm Progression. *Circ Res* 2009.
- 12. Dasouki M, Markova D, Garola R *et al*: Compound heterozygous mutations in fibulin-4
 causing neonatal lethal pulmonary artery occlusion, aortic aneurysm, arachnodactyly, and
 mild cutis laxa. *Am J Med Genet A* 2007; **143**: 2635-2641.
- Hucthagowder V, Sausgruber N, Kim KH, Angle B, Marmorstein LY, Urban Z: Fibulin-4: a
 novel gene for an autosomal recessive cutis laxa syndrome. *Am J Hum Genet* 2006; **78**:
 1075-1080.
- Hoyer J, Kraus C, Hammersen G, Geppert JP, Rauch A: Lethal cutis laxa with contractural
 arachnodactyly, overgrowth and soft tissue bleeding due to a novel homozygous fibulin-4
 gene mutation. *Clin Genet* 2009.
- 39915.Claus S, Fischer J, Megarbane H *et al*: A p.C217R mutation in fibulin-5 from cutis laxa400patients is associated with incomplete extracellular matrix formation in a skin equivalent401model. J Invest Dermatol 2008; **128**: 1442-1450.
- 40316.Elahi E, Kalhor R, Banihosseini SS *et al*: Homozygous missense mutation in fibulin-5 in an404Iranian autosomal recessive cutis laxa pedigree and associated haplotype. J Invest405Dermatol 2006; **126**: 1506-1509.
- 40717.Loeys B, Van Maldergem L, Mortier G et al: Homozygosity for a missense mutation in408fibulin-5 (FBLN5) results in a severe form of cutis laxa. Hum Mol Genet 2002; 11: 2113-4092118.
- 411 18. Markova D, Zou Y, Ringpfeil F *et al*: Genetic heterogeneity of cutis laxa: a heterozygous tandem duplication within the fibulin-5 (FBLN5) gene. *Am J Hum Genet* 2003; **72**: 998-1004.
- 414 19. Kornak U, Reynders E, Dimopoulou A *et al*: Impaired glycosylation and cutis laxa caused by
 415 mutations in the vesicular H+-ATPase subunit ATP6V0A2. *Nat Genet* 2008; **40**: 32-34.
- 417 20. Kivuva EC, Parker MJ, Cohen MC, Wagner BE, Sobey G: De Barsy syndrome: a review of the
 418 phenotype. *Clin Dysmorphol* 2008; **17**: 99-107.
- 420 21. Morava E, Guillard M, Lefeber DJ, Wevers RA: Autosomal recessive cutis laxa syndrome
 421 revisited. *Eur J Hum Genet* 2009.
- 423 22. Guernsey DL, Jiang H, Evans SC *et al*: Mutation in pyrroline-5-carboxylate reductase 1 gene
 424 in families with cutis laxa type 2. *Am J Hum Genet* 2009; **85**: 120-129.
- 426 23. Reversade B, Escande-Beillard N, Dimopoulou A *et al*: Mutations in PYCR1 cause cutis laxa
 427 with progeroid features. *Nat Genet* 2009; **41**: 1016-1021.
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- Rodriguez-Revenga L, Iranzo P, Badenas C, Puig S, Carrio A, Mila M: A novel elastin gene
 mutation resulting in an autosomal dominant form of cutis laxa. *Arch Dermatol* 2004; **140**:
 1135-1139.
- 433 25. Szabo Z, Crepeau MW, Mitchell AL *et al*: Aortic aneurysmal disease and cutis laxa caused
 434 by defects in the elastin gene. *J Med Genet* 2006; **43**: 255-258.
- 436 26. Tassabehji M, Metcalfe K, Hurst J *et al*: An elastin gene mutation producing abnormal
 437 tropoelastin and abnormal elastic fibres in a patient with autosomal dominant cutis laxa.
 438 *Hum Mol Genet* 1998; **7**: 1021-1028.
- Zhang MC, He L, Giro M, Yong SL, Tiller GE, Davidson JM: Cutis laxa arising from frameshift
 mutations in exon 30 of the elastin gene (ELN). *J Biol Chem* 1999; **274:** 981-986.
- 44328.Ades LC, Knight WB, Byard RW *et al*: Clinicopathologic findings in congenital aneurysms of444the great vessels. *Am J Med Genet* 1996; **66:** 289-299.
- 446 29. Baspinar O, Kilinc M, Balat A, Celkan MA, Coskun Y: Long tortuous aorta in a child with
 447 Larsen syndrome. *Can J Cardiol* 2005; **21:** 299-301.
- Kobayashi N, Kostka G, Garbe JH *et al*: A comparative analysis of the fibulin protein family.
 Biochemical characterization, binding interactions, and tissue localization. *J Biol Chem*2007; **282**: 11805-11816.
- 453 31. Loeys BL, Chen J, Neptune ER *et al*: A syndrome of altered cardiovascular, craniofacial,
 454 neurocognitive and skeletal development caused by mutations in TGFBR1 or TGFBR2. *Nat*455 *Genet* 2005; **37**: 275-281.
- 45732.Callewaert BL, Willaert A, Kerstjens-Frederikse WS *et al*: Arterial tortuosity syndrome:458clinical and molecular findings in 12 newly identified families. *Hum Mutat* 2008; **29:** 150-459158.
- 461 33. Rao Z, Handford P, Mayhew M, Knott V, Brownlee GG, Stuart D: The structure of a Ca(2+)462 binding epidermal growth factor-like domain: its role in protein-protein interactions. *Cell*463 1995; 82: 131-141.
- 465 34. Hu Q, Reymond JL, Pinel N, Zabot MT, Urban Z: Inflammatory destruction of elastic fibers
 466 in acquired cutis laxa is associated with missense alleles in the elastin and fibulin-5 genes. J
 467 Invest Dermatol 2006; **126**: 283-290.
- 469 35. Freeman LJ, Lomas A, Hodson N *et al*: Fibulin-5 interacts with fibrillin-1 molecules and
 470 microfibrils. *Biochem J* 2005; **388**: 1-5.
- 47236.Ono RN, Sengle G, Charbonneau NL *et al*: LTBPS and fibulins compete for fibrillin-1 and473exhibit exquisite specificities in binding sites. *J Biol Chem* 2009.
- 474

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445

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452

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475 476 477	37. Ng CM, Cheng A, Myers LA <i>et al</i> : TGF-beta-dependent pathogenesis of prolapse in a mouse model of Marfan syndrome. <i>J Clin Invest</i> 2004; 114: 158				
477 478 479 480 481 482	38.	Habashi JP, Judge DP, Holm TM <i>et al</i> : Losartan, an AT1 antagonist, prevents aortic aneurysm in a mouse model of Marfan syndrome. <i>Science</i> 2006; 312 : 117-121.			
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501 TITLES AND LEGENDS TO FIGURES

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Figure 1. A. Clinical photograph of patient 3 showing a prominent forehead with depressed nasal bridge, mild hypertelorism and downslanting palpebral fissures. The patient has a trachea canulae. **B.** MRI shows severe tortuosity of the carotid arteries (anterolateral oblique view). **C.** A posterior view of a 3D reconstruction of an MR angio showing successively, the dilated aorta ascendens (indicated with an arrow), hypoplasia of the transverse aorta (indicated with an arrowhead) and tortuous descending aorta.

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510 Figure 2. DNA sequence of FBLN4 in A. patient 1, showing the missense c.376G>A511 mutation, leading to the Glu126Lys substitution in the protein. The glutamic acid residue at position 126 is evolutionarily conserved and located in the DINE consensus sequence of the 512 513 second cbEGF-like domain, in **B**. patient 2, showing the c.1189G>A mutation, leading to 514 the Ala397Thr substitution. The alanine residue is located in the C-terminal fibulin-like 515 module and is conserved between fibulin homologues and highly conserved among species, and in C. patient 3, showing the compound heterozygous c.377A > T missense (leading to 516 the Glu126Val substitution) and c.577delC mutations. The missense mutation affects the 517 518 same conserved glutamic acid residue as identified in patient 1 (see panel A for alignment 519 of the conserved sequence).

Figure 3. A. Immunohistochemical staining of aortic tissue of patient 4 and its age- and sex-matched control. Intracellular fibulin-4 staining is seen in both individuals (examples of intracellular staining are indicated with a black arrow). No extracellular fibulin-4 is seen in

the aorta of the patient as compared to its control (extracellular staining is indicated with 524 525 white arrows). Scale bars = $100 \mu m$. B. Immunoblot analysis of the cultured media using fibroblasts of age- and sex-matched controls (C1 and C5) and two patients (P1 and P5). 526 Top panel: anti-fibulin-4 monoclonal antibody (clone 347) demonstrating impaired fibulin-527 528 4 secretion in patient 1 and slightly impaired fibulin-4 secretion in patient 5 in contrast to 529 their matched control. Bottom panel: anti-fibronectin-1 monoclonal antibody indicating the 530 amounts of samples loaded on the gel. The fibulin-4 secretion to the extracellular space as 531 seen on the gel images was quantified using ImageJ software.

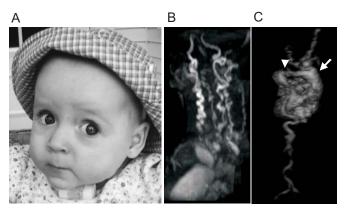
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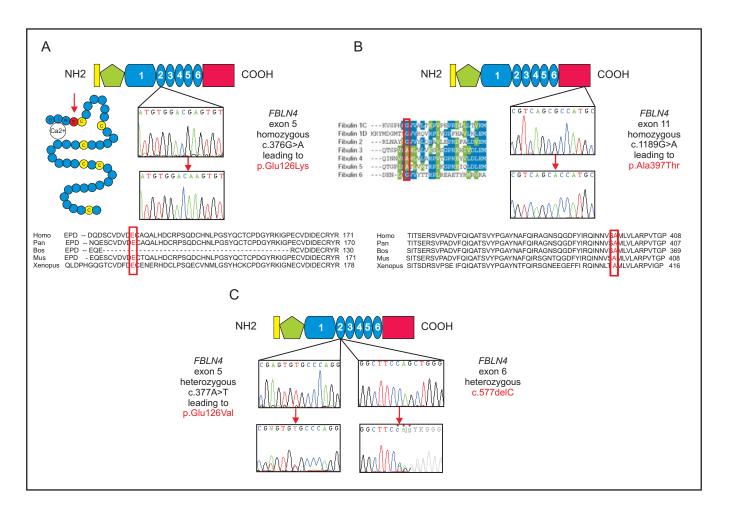
533 Figure 4. Immunohistochemical staining of aorta (A-D and I-L) and lung (E-H and M-P) 534 tissue in FBLN4-mutation positive patient 4 (B,D,F,H,J,L,N,P) and in control (A,C,E,G,I,K,M,O). Increased nuclear accumulation of phosphorylated Smad2 is present in 535 536 both the aorta (B,D) (some of the nuclei are indicated with an arrow) and the lung (F,H) of 537 the patient, as compared with the control aorta (A,C) and lung (E,G). Similarly, increased 538 levels of CTGF expression are noted in the aorta (J,L) and lung (N,P) of the patient, compared to control aorta (I,K) and lung (M,O). These results are indicative of increased 539 TGF beta signaling in the *FBLN4*-mutation positive patient. Scale bars = $100 \mu m$. 540

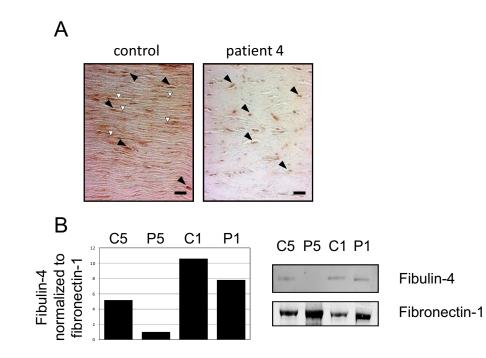
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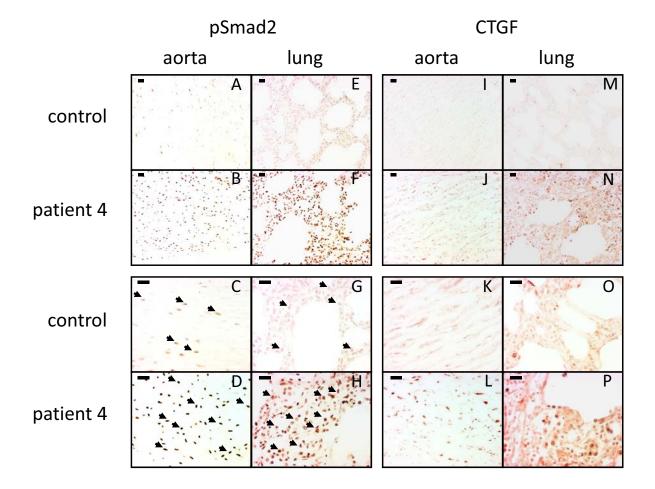
Figure 5. Immunoblot analysis of the fibroblast cell extracts of age- and sex-matched controls (C1 and C5) and patients 1 (P1) and 5 (P5). The graph and the pSmad2 immunoblotting panel show an increase in pSmad2 level in patient 1 and patient 5 when cells were stimulated with TGFbeta protein in contrast to their controls. When cells are not stimulated basal expression of pSmad2 is seen. Bottom panel: anti-vinculin antibody,

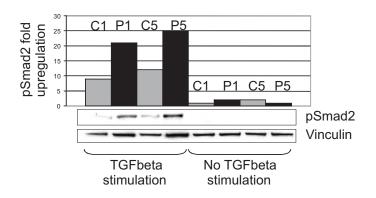
- 547 indicating the amounts of samples loaded on the gel. This immunoblotting analysis is
- representative of three independent experiments (see supplemental figure S1).











TABLES

	patient 1	patient 2	patient 3	patient 4	patient 5	patient 6
	current report	current report	current report	Dasouki et al, 2006 (12)	Hoyer et al, 2009 (14)	Hucthagowder et al, 2006 (13)
mutation	p.Glu126Lys homozygous	p.Ala397Thr homozygous	p.Glu126Val c.577delC compound heterozygous	<i>p.Arg279Cys</i> <i>c.1070_1073dupCCGC</i> compound heterozygous	p.Cys267Tyr homozygous	p.Glu57Lys homozygous
sex	female	male	female	female	female	female
current age	20 years	7 years	died at 18 months	died at 27 days	died at birth	2 years
skin	velvety skin normal scars	-	-	mild cutis laxa	generalized cutis laxa hyperextensibility	cutis laxa
arteries	tortuosity aneurysm stenosis	tortuosity aneurysm	tortuosity aneurysm	tortuosity aneurysm dissection	-	tortuosity aneurysm
emphysema	-	-	-	-	++	++
joint laxity	-	+	-	-	-	++
other	arched palate retrognathia	hypertelorism flat facies	hypertelorism long fingers hemispheric stroke	arachnodactyly	hypertelorism retrognathia cardiorespiratory insufficiency	fractures hernia diaphragmatica

Table 1. Overview of the clinical and molecular data of the three patients reported in

 this article compared to previously described patients ¹¹⁻¹³.