

Influence of inoculum size on the selection of resistant mutants of Escherichia coli in relation to mutant prevention concentrations of marbofloxacin.

Aude Ferran, Véronique Dupouy-Guiraute, Pierre-Louis Toutain, Alain

Bousquet-mélou

▶ To cite this version:

Aude Ferran, Véronique Dupouy-Guiraute, Pierre-Louis Toutain, Alain Bousquet-mélou. Influence of inoculum size on the selection of resistant mutants of Escherichia coli in relation to mutant prevention concentrations of marbofloxacin.. Antimicrobial Agents and Chemotherapy, 2007, 51 (11), pp.4163-6. 10.1128/AAC.00156-07. hal-00517559

HAL Id: hal-00517559 https://hal.science/hal-00517559

Submitted on 16 Sep 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Influence of Inoculum Size on the Selection of <i>Escherichia coli</i> Resistant Mutants in
2	Relation to Mutant Preventive Concentrations of Marbofloxacin
3	
4	Aude Ferran, Véronique Dupouy, Pierre-Louis Toutain and Alain Bousquet-Mélou*
5	
6	UMR181 Physiopathologie et Toxicologie Expérimentales, INRA, ENVT, Ecole Nationale
7	Vétérinaire de Toulouse, 23 chemin des Capelles, BP 87 614, 31076 Toulouse Cedex 3,
8	France.
9	
10	* Corresponding author. Mailing address : UMR181 Physiopathologie et Toxicologie
11	Expérimentales, INRA, ENVT, Ecole Nationale Vétérinaire de Toulouse, 23 chemin des
12	Capelles, BP 87 614, 31076 Toulouse Cedex 3, France. Phone : +33 (0) 5 61 19 39 25. Fax :
13	+33 (0) 5 61 19 39 17. E-mail : a.bousquet-melou@envt.fr
14	

15

- 16 We demonstrated using an *in vitro* pharmacodynamic model that the likelihood of selection of
- 17 Escherichia coli resistant mutants to a fluoroquinolone was increased when the initial size of
- 18 the bacterial population exposed to fluoroquinolone concentrations within the mutant
- 19 selection window was increased

20

Resistant bacteria selected under the pressure of fluoroquinolone exposure expand 21 from few spontaneously resistant mutant present before any treatment. When the bacterial load at the infectious site exceeds 10^9 - 10^{10} CFU (ref), it can be presumed if spontaneous 22 mutation rate is about 10⁻⁹-10⁻⁷ (Lindgren, AAC, 2003) that before any antibiotic treatment a 23 24 small resistant subpopulation of first-step resistant mutants already coexists with the larger 25 susceptible wild-type population.

26 Minimal Inhibitory Concentrations (MIC) allows the determination of the 27 susceptibility of the major bacterial population and Mutant Prevention Concentration (MPC) 28 the susceptibility of the small resistant subpopulation (2, 6, 10). MIC and MPC define the Mutant Selection Window (MSW), a range of antibiotic concentrations favouring the 29 selection of first-step mutants (10). Previous studies (5, 11) have indicated that prevention of 30 31 first-step mutants selection was obtained when fluoroquinolone concentrations exceeded the MPC for more than 80% of the dosage interval, i.e. when time within MSW (T_{MSW}) was 32 33 inferior to 20%. However, these studies only tested one inoculum size whereas the bacterial 34 load increases during the time course of infections, and the likelihood of a mutant appearing 35 may increase with inoculum size.

The aim of this study was to determine, using marbofloxacin a fluoroquinolone 36 extensively used in veterinary medicine, an interaction between inoculum sizes $(10^5, 10^7 \text{ or})$ 37 10^9 CFU/mL) and marbofloxacin exposures characterized by different T_{MSW} (0, 25 or 100%) 38 39 on the selection of Escherichia coli resistant mutants.

40

Marbofloxacin MIC for Escherichia coli ATCC 25922 was determined by 41 microdilution technique and MPC by a previously described method (1). MIC and MPC were 42 43 0.008 and 0.256 µg/mL, respectively.

Bacteria suspended in Mueller-Hinton (MH) broth were exposed in an *in vitro* pharmacodynamic model to three monoexponential kinetic profiles of marbofloxacin to ensure T_{MSW} of 0, 25 and 100% corresponding to times above MPC ($T_{>MPC}$) of 100, 75 and 0%. Bacteria exposures to marbofloxacin were determined from serial samples by HPLC and killing and regrowth of bacterial population were assessed by counting viable bacteria.

49 For all initial inoculum sizes, bacterial counts without antibiotic revealed similar exponential growth rate until the carrying capacity of the *in vitro* system was reached (about 50 10⁹ CFU/mL). Figures 1A and 1B represent the bacterial counts obtained from inoculum sizes 51 of 10^5 , 10^7 and 10^9 CFU/mL exposed to marbofloxacin with T_{MSW} of 0 and 25% respectively. 52 The bacterial counts for experiments carried out with a T_{MSW} of 100% are shown in Fig. 1C or 53 Fig. 1D depending on the susceptibility of surviving bacteria at the end of experiments. 54 55 Whatever the initial inoculum size, all marbofloxacin regimens showed bactericidal activity 56 during the first hours of exposure. Killing rates then declined with time until regrowth 57 occurred, whatever the T_{MSW} and inoculum size. The minimal counts of surviving bacteria in the central flask seemed to increase with inoculum size, although the limit of detection of 100 58 CFU/mL prevented comparison of the 10^5 and 10^7 CFU/mL inocula (Table 1). Bacteria 59 counts after 32 hours ranged from 10^4 to 2.10^6 CFU/mL when most of the surviving bacteria 60 61 were susceptible to 0.128 µg/mL, i.e. when they were not first-step mutants and ranged from 5.10^7 to 6.10^8 CFU/mL when most of the surviving bacteria were resistant to this 62 concentration, i.e. when they have the same phenotype as first-step mutants. The higher 63 64 regrowth associated with resistant bacteria selection may be explained by a higher growth rate or a slower rate of killing of resistant bacteria in the presence of marbofloxacin. A previously 65 described integrated parameter, called ABBC (3), was used to assess marbofloxacin 66 67 antimicrobial effect during the initial hours of exposure. It describes the ratio of areas from 0 to 10 hours delimited by time-kill curves in the absence and the presence of marbofloxacin 68

69 with the same inoculum sizes. Inoculum size had no net effect on ABBC (Table 1). However, slightly lower ABBC values and higher minimal counts were obtained with 10⁹ CFU/mL 70 71 inocula exposed to T_{MSW} of 100%, i.e. when resistant bacteria to 0.128 µg/mL emerged 72 compared to all other inoculum size/T_{MSW} combinations, suggesting that ABBC decrease 73 might be related to resistant mutants selection. The detection limit might explain that no relation between ABBC and resistance selection was observed for 10⁵ and 10⁷ CFU/mL 74 inocula. The relatively weak effect of inoculum size on fluoroquinolone antimicrobial activity 75 76 observed in the present study is in agreement with a previous report on Escherichia coli 77 exposure to ciprofloxacin or trovafloxacin in an *in vitro* pharmacodynamic model (4).

78 Bacteria growing in the presence of 0.016 (2xMIC), 0.128 (one dilution before MPC) 79 and 0.256 (MPC) µg/mL marbofloxacin were counted before and 32 hours after exposure to 80 marbofloxacin. The frequencies of resistant bacteria were determined by the ratio of bacteria 81 counts growing in the presence and the absence of marbofloxacin. Before exposure to 82 marbofloxacin, very few bacteria were resistant to 0.128 µg/mL and resistance to 0.256 µg/mL was detected in only one initial inoculum of 10⁹ CFU/mL (Fig. 2). At the end of 83 84 control experiments without antibiotic, no mutant resistant to 0.128 µg/mL marbofloxacin was observed whatever the inoculum size. As shown in Fig. 2, bacteria exposed to T_{MSW} of 85 86 100% became mostly resistant to 0.128 µg/mL in five experiments among nine. Most of these 87 resistant bacteria were still susceptible to the MPC of marbofloxacin for Escherichia coli 88 ATCC 25922 (0.256 µg/mL) suggesting that these resistant populations corresponded to first-89 step mutants. The detection of first-step mutants when concentrations were maintained within 90 the MSW is in agreement with previous studies (5, 11). However, resistant mutants emerged systematically in the three experiments carried out with 10^9 CFU/mL, but only in one among 91 three for 10^5 and 10^7 CFU/mL inocula. We calculated AUC/MPC ratios by dividing the 92 93 AUC from 0 to 24 hours by the MPC. The observed AUC/MPC values associated with

94 prevention of mutant selection irrespective of inoculum size were 44-54 hours. A value of 22 95 hours was previously reported as sufficient to prevent the emergence of mutants resistant to ciprofloxacin in large inocula (10^{10} CFU) of susceptible *Escherichia coli* strains (9). 96 However, in two thirds of our experiments with inoculum sizes of 10^5 and 10^7 CFU/mL, an 97 AUC/MPC of 9-12 hours was sufficient to prevent the emergence of resistant mutants. These 98 99 results support the hypothesis that breakpoint values of PK/PD parameters associated with the 100 MPC and MSW concepts for preventing the emergence of resistant mutants may depend on 101 the size of exposed bacterial population present at the infection site.

In summary, our results confirmed that maintaining concentrations above the MPC prevents the emergence of resistance. However, the process of mutant selection within the MSW was not evenly linked to underexposure to antibiotics but also influenced by the presence of mutant before antibiotic treatment which is directly linked to the bacterial population size. The *in vivo* relevance of these *in vitro* results merits investigation in animal models of infection.

108

109 We thank Nathalie Arpaillange for technical assistance in bacteriology, Sylvie Puel110 and Charles-Adrien Richard for performing analytical assays.

111

REFERENCES

113 LEGENDS

114 Figure 1:

Observed viable counts of Escherichia coli ATCC 25922 following exposures of 115 initial inoculum sizes of 10^5 (\bullet), 10^7 (\Box) or 10^9 (\blacktriangle) CFU/mL to concentrations of 116 117 marbofloxacin inside the mutant selection window for 0 % (A), 25% (B) or 100% (C-D) of 118 the time. For T_{MSW} of 100%, experiments in which surviving bacteria were mainly susceptible 119 to 0.128 µg/mL are represented in (C) and those in which surviving bacteria were mainly 120 resistant to 0.128 µg/mL are represented in (D). In A, B and C, each symbol represents the 121 mean of 2 experiments. In D, symbols (\bullet) and (\Box) represent results of one experiment and 122 symbols (\blacktriangle) represent the means of 3 experiments. Error bars show standard deviation. Dotted line indicates the lower limit of detection (2 log₁₀ CFU/mL) used for bacterial 123 124 quantification.

125

Figure 2:

Frequencies of bacteria resistant to 0.016 µg/ml (white bars), 0.128 µg/mL (dotted 126 bars) and 0.256 μ g/mL (black bars) before and after exposure of initial inoculum sizes of 10⁵, 127 10^7 or 10^9 CFU/mL to control (one experiment per inoculum size) or to marbofloxacin 128 129 concentrations within the mutant selection window for 100% (3 experiments per inoculum size), 25% (2 experiments per inoculum size) and 0 % (2 experiments per inoculum size) of 130 131 the time. ○, ◎, and ■ indicate that no bacteria resistant to 0.016, 0.128 or 0.256 µg/mL respectively were detected. 132

|--|

Time in the MSW (%)	100				25			0			
Inoculum size (CFU/mL)	1	0 ⁵	10	0^{7}	10^{9}	10 ⁵	107	10 ⁹	10 ⁵	107	10 ⁹
Susceptibility ^a	-	+	-	+	+	-	-	-	-	-	-
Minimal counts (CFU/mL)	<100	<100	<100	<100	2.10^{4}	<100	150	10^{3}	<100	<100	600
Final counts (CFU/mL)	2.10^{6}	1.10^{8}	7.10^{4}	6.10^{8}	5.10^{7}	6.10^{4}	8.10^{4}	3.10^{4}	7.10^{3}	2.10^{4}	1.10^{4}
ABBC (log CFU/mL.h	53	51	60	56	44	54	56	53	54	57	51
^a Suscept	ibility	is asse	essed at	t the	end of the	e exper	iments.	Experin	nents in	which t	the

population is mainly resistant to 0.128 μ g/mL are noted +, those mainly susceptible to 0.128

136 μ g/mL are noted -.

_	Targeted time inside the MSW (%)	T _{>MIC} (%)	$T_{>MPC}$ (%)	AUC/MPC
	100	100	0	9-12
	25	100	75	44-54
	0	100	100	176-210
38				
139				
40				
41				

137 TABLE 2. Marbofloxacin pharmacokinetic parameters in relation to MIC, MPC and MSW.

142 FIGURE 4



143



145

146

147 1. Blondeau, J. M., X. Zhao, G. Hansen, and K. Drlica. 2001. Mutant prevention 148 concentrations of fluoroquinolones for clinical isolates of Streptococcus 149 pneumoniae. Antimicrob Agents Chemother 45:433-8.

- Dong, Y., X. Zhao, B. N. Kreiswirth, and K. Drlica. 2000. Mutant prevention 150 2. 151 concentration as a measure of antibiotic potency: studies with clinical isolates of 152 Mycobacterium tuberculosis. Antimicrob Agents Chemother 44:2581-4.
- 153 3. Firsov, A. A., D. Saverino, M. Ruble, D. Gilbert, B. Manzano, A. A. Medeiros, and S. H. Zinner. 1996. Predictors of effect of ampicillin-sulbactam against TEM-154 155 1 beta-lactamase-producing Escherichia coli in an in vitro dynamic model: 156 enzyme activity versus MIC. Antimicrob Agents Chemother 40:734-8.
- 157 Firsov, A. A., S. N. Vostrov, O. V. Kononenko, S. H. Zinner, and Y. A. Portnoy. 4. 1999. Prediction of the effects of inoculum size on the antimicrobial action of 158 159 trovafloxacin and ciprofloxacin against Staphylococcus aureus and Escherichia coli in an in vitro dynamic model. Antimicrob Agents Chemother 43:498-502. 160
- 161 5. Firsov, A. A., S. N. Vostrov, I. Y. Lubenko, K. Drlica, Y. A. Portnoy, and S. H. 162 Zinner. 2003. In vitro pharmacodynamic evaluation of the mutant selection window hypothesis using four fluoroquinolones against Staphylococcus aureus. 163 Antimicrob Agents Chemother 47:1604-13. 164
- 165 6. Marcusson, L. L., S. K. Olofsson, P. Komp Lindgren, O. Cars, and D. Hughes. 2005. Mutant prevention concentrations of ciprofloxacin for urinary tract 166 infection isolates of Escherichia coli. J Antimicrob Chemother 55:938-43. 167
- 168 7. Mizunaga, S., T. Kamiyama, Y. Fukuda, M. Takahata, and J. Mitsuyama. 2005. 169 Influence of inoculum size of Staphylococcus aureus and Pseudomonas

- 170aeruginosa on in vitro activities and in vivo efficacy of fluoroquinolones and171carbapenems. J Antimicrob Chemother 56:91-6.
- 172 8. Morrissey, I., and J. T. George. 1999. The effect of the inoculum size on bactericidal activity. J Antimicrob Chemother 43:423-5.
- Olofsson, S. K., L. L. Marcusson, P. Komp Lindgren, D. Hughes, and O. Cars.
 2006. Selection of ciprofloxacin resistance in Escherichia coli in an in vitro kinetic model: relation between drug exposure and mutant prevention concentration. J
 Antimicrob Chemother 57:1116-21.
- 178 10. Zhao, X., and K. Drlica. 2001. Restricting the selection of antibiotic-resistant
 179 mutants: a general strategy derived from fluoroquinolone studies. Clin Infect Dis
 180 33 Suppl 3:S147-56.
- 181 11. Zinner, S. H., I. Y. Lubenko, D. Gilbert, K. Simmons, X. Zhao, K. Drlica, and A.
 182 A. Firsov. 2003. Emergence of resistant Streptococcus pneumoniae in an in vitro 183 dynamic model that simulates moxifloxacin concentrations inside and outside the 184 mutant selection window: related changes in susceptibility, resistance frequency 185 and bacterial killing. J Antimicrob Chemother 52:616-22.
- 186