



HAL
open science

Development of specific diagnostic test for small ruminant lentivirus genotype E

Ramses Reina, Elena Grego, Margherita Profiti, Idoia Galaria, Patrizia Robino, Antonio Quasso, Beatriz Amorena, Sergio Rosati

► **To cite this version:**

Ramses Reina, Elena Grego, Margherita Profiti, Idoia Galaria, Patrizia Robino, et al.. Development of specific diagnostic test for small ruminant lentivirus genotype E. *Veterinary Microbiology*, 2009, 138 (3-4), pp.251. 10.1016/j.vetmic.2009.04.005 . hal-00514607

HAL Id: hal-00514607

<https://hal.science/hal-00514607>

Submitted on 3 Sep 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Accepted Manuscript

Title: Development of specific diagnostic test for small ruminant lentivirus genotype E

Authors: Ramses Reina, Elena Grego, Margherita Profiti, Idoia Glaria, Patrizia Robino, Antonio Quasso, Beatriz Amorena, Sergio Rosati



PII: S0378-1135(09)00179-5
DOI: doi:10.1016/j.vetmic.2009.04.005
Reference: VETMIC 4406

To appear in: *VETMIC*

Received date: 19-12-2008
Revised date: 13-3-2009
Accepted date: 3-4-2009

Please cite this article as: Reina, R., Grego, E., Profiti, M., Glaria, I., Robino, P., Quasso, A., Amorena, B., Rosati, S., Development of specific diagnostic test for small ruminant lentivirus genotype E, *Veterinary Microbiology* (2008), doi:10.1016/j.vetmic.2009.04.005

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1 **Development of specific diagnostic test for small ruminant lentivirus genotype E**

2 Ramses Reina ¹, Elena Grego ¹, Margherita Profiti ¹, Idoia Galaria ², Patrizia Robino ¹, Antonio
3 Quasso³, Beatriz Amorena ², Sergio Rosati ^{1*}

4

5 ¹*Dipartimento di Produzioni Animali, Epidemiologia ed Ecologia, Facoltà di Medicina Veterinaria,*
6 *Università degli Studi di Torino, Via Leonardo da Vinci, 44, 10095 Grugliasco (TO), Italy.*

7 ²*Instituto de Agrobiotecnología (CSIC-UPNA-Gobierno de Navarra) Ctra. Mutilva Baja s/n 31192*
8 *Mutilva Baja, Navarra, Spain.*

9 ³*Dipartimento di Prevenzione, Servizio veterinario, Divisione sanità animale ASL19 Asti, Italy*
10

11

12

13 *Corresponding Author

14 Via Leonardo da Vinci 44

15 10095 Grugliasco (TO)

16 tel ++39 011 6709187

17 fax ++39 011 6709196

18 e-mail sergio.rosati@unito.it (S. Rosati)

19

Abstract

Small ruminant lentivirus (SRLV) belonging to the highly divergent genotype E has recently been identified in the Italian goat breed Roccaverano. In this report we have developed a specific serological test based on recombinant matrix/capsid antigen fusion protein. Performance has been evaluated and compared with a similar test based on genotype B antigen. Herds under study were selected according to the infectious status characterized by blood PCR and sequencing. Results clearly showed that B and E based recombinant ELISA only detected homologous infection and an apparent cross-reactivity was recorded in a herd in which co-infection was present. Three commercially available ELISAs showed different abilities in detecting genotype E infection, being the whole virus-based immunoassay the best choice. Genotype E-recombinant antigen was not detected in ELISA by three commercially available Mabs known to be cross-reactive among CAEV and MVV capsid antigens, further supporting the high divergence of the E genotype from others. Finally, a SRLV-free herd according to commercial ELISA testing, was analysed in the same area where genotype E was identified and few animals belonging to Roccaverano breed were found slightly reactive with the E antigens. Our results suggest that the prevalence of genotype E in other small ruminant populations may be conveniently estimated using a comparative assay based on a combination of genotype specific recombinant antigens and may highlight a wider space in which SRLVs evolve.

Keywords

Small ruminant lentivirus, genotype E, recombinant antigen, diagnosis

46

47 **Introduction**

48 Small ruminant lentiviruses (SRLVs) are a heterogeneous group of viruses affecting sheep and goat
49 and are responsible for chronic debilitating diseases known as Maedi Visna (MV) and Caprine
50 Arthritis-Encefalitis (CAE) (Pepin et al., 1998). Viral isolates characterized so far, show different
51 genetic, antigenic and biological properties and are no longer considered species-specific (Pisoni et
52 al., 2005; Shah et al., 2004b). From an antigenic point of view, most SRLVs can be classified as
53 MVV-like or CAEV-like, corresponding to genotype A and B respectively (Shah et al., 2004a).
54 Previous studies have suggested that early serological diagnosis can be achieved using homologous
55 antigen (Lacerenza et al., 2006). Nevertheless, most of the currently available diagnostic tests are
56 produced using a single strain-based antigen preparation which is believed to detect cross-reacting
57 antibodies against epitopes located in structural proteins (Gogolewski et al., 1985). Recently, a
58 novel genotype E has been identified in the local breed Roccaverano in north-west Italy. First
59 sequences were obtained by chance in a caprine herd, using a set of degenerated primers designed
60 to amplify a *gag* fragment from the majority of known genotypes, encompassing major linear
61 capsid antigen epitopes. Following a preliminary sequence screening, it seemed quite clear that this
62 viral cluster might have escaped diagnosis in the field using conventional antigen preparations,
63 likely due to the low similarity found in the major immunodominant regions (Grego et al., 2007). A
64 viral strain was subsequently isolated from an apparently healthy goat highly reactive by ELISA
65 against genotype E-derived major capsid antigen epitopes (Reina et al., 2009). Genetic features of
66 this genotype have been described in three epidemiologically unrelated herds. The complete
67 genome (~8,4Kb) presented two major deletions corresponding to the dUTPase subunit of the *pol*
68 gene and to the *vpr* accessory gene. Based on previous studies in which such subunits were
69 independently deleted from a pathogenic infectious clone, these deletions could explain why the
70 viral cluster is not related to any known clinical signs, representing a natural, well host-adapted, low
71 pathogenic lentivirus (Harmache, 1996; Turelli, 1996; Zhang, 2003). Moreover, preliminary

72 epidemiological data suggest that infection can persist in the population through familiar lineage,
73 with a low tendency to spread horizontally. To date, no information is available on the prevalence
74 of genotype E in larger goat populations due to the lack of a specific antibody detection system. To
75 address this issue, in this report we have developed genotype E specific recombinant antigens which
76 were tested with a panel of monoclonal antibodies (Mabs) known to be reactive against CAEV and
77 MVV capsid antigens. These genotype E derived antigens were also used to develop an indirect
78 ELISA in order to test a panel of goat sera belonging to herds in which the SRLV infectious status
79 was determined by PCR product sequence analysis. Serological and sequence data were in
80 agreement highlighting the importance of using genotype specific tests when determining SRLV
81 seroprevalence, assessing SRLV-free status and searching for epidemiological information.

82

83 **Material and Methods**

84 *Virus and plasmids*

85 *Roccoverano* strain was originally isolated using mammary gland explants from an adult goat and a
86 complete proviral sequence was obtained (Genbank accession number EU293537). The *gag* gene
87 was amplified by concatenating overlapping PCR fragments and cloned in pCRTopoXL
88 (Invitrogen). The gene fragment coding for Matrix (P16) and major Capsid Antigen (P25) was
89 subsequently amplified and was subcloned between the *BamHI/EcoRI* sites of pGEX6His
90 following site-directed PCR-mediated mutagenesis suppressing an internal *EcoRI* restriction site.
91 This plasmid, derived from pGEX6P prokaryote expression vector (GE Healthcare), was modified
92 by inserting an in frame 6Xhis-tag between *EcoRI* and *Sall* restriction sites thus allowing a double
93 step affinity purification.

94 *Expression and purification of P16-25 recombinant antigens and P25-B3 subunit epitopes*

95 Transformed *E.coli* BL21 bacteria were induced at early log phase for 2 h with 0.5mM IPTG under
96 agitation. Bacterial cells were recovered by centrifugation and lysed by physicochemical methods.
97 Recombinant GST/P16-25/6H fusion protein was recovered in the soluble fraction and the first

98 affinity step was carried out in batch using glutathione Sepharose 4B (GE Healthcare). GST
99 cleavage was achieved in pooled eluted fractions using PreScission Protease (2U/mg) (GE
100 Healthcare). Solution containing GST and P16-25/6H was dialyzed for 24h to remove reducing and
101 chelating agents and loaded into a Hi-Trap chelating HP column (GE Healthcare), positively
102 charged with nickel ions. Following immobilised metal chelate affinity chromatography, purity and
103 yield of recombinant antigen was estimated by SDS-PAGE and DC protein assay (BioRad).

104 Using the same protocol, the same gag antigens derived from strain It-Pi1 (genotype B) were
105 employed to generate the antigenic CAEV-like counterpart.

106 Subunit immunodominant capsid antigen epitopes of MVV and CAEV had previously been
107 characterised. In this study a third version using genotype E (P25-B3) derived sequence was
108 produced, generating a GST fusion protein as previously described (Grego et al., 2002; Rosati et al.,
109 1999).

110 *Blood samples, polymerase chain reaction and sequencing*

111 Twelve caprine herds were selected in this and in a previous study (Grego et al., 2007). Heparinized
112 blood samples were obtained from a number of adult animals, representative of each herd: and
113 DNA was extracted from white blood cells using DNA blood kit (Qiagen). A *gag* nested PCR
114 previously developed (Grego et al., 2007) and known to detect the highest number of SRLV
115 genotypes/subtypes was applied to each sample and all positive results with suitable bands were
116 sequenced. After, this preliminary screening, five herds of Roccaverano breed were selected: a first
117 herd, BL (n=52) in which only genotype E was detected; a second herd, NG (n=40) in which
118 genotype B and E were detected; and a third group of goats, TM (n=20), BM (n=18) and CF (n=6)
119 in which only genotype B was present. In this study, all caprine herds were retested when possible
120 at completion and additional sequences were obtained.

121 To date, the presence of genotype E had been limited to few herds of the Roccaverano breed.
122 Therefore, an additional three-breed long term SRLV negative herd (n=400) was also included for

123 serological testing. The herd consisted of 109 Roccaverano, 107 Saanen and 184 French Alpine
124 goats.

125

126 *Serum samples and ELISAs*

127 For P16-25 recombinant ELISA, microplates (Immunomaxi TPP) were coated with 100 ng of P16-
128 25 derived from B and E genotypes or water as negative antigen (Fig. 1). Plates were allowed to dry
129 overnight at 37°C and then blocked with 2.5% bovine casein for 1h at 37°C. After four washes,
130 serum samples were diluted 1/20 in phosphate-buffered saline containing 1.25% casein and
131 incubated for 1h at 37°C. Subsequently to the washing step, anti-sheep/goat IgG peroxidase labelled
132 Mab diluted in the same buffer was added and the plates incubated as above. After a final washing
133 step, the reaction was developed with ABTS and plates were read at 405 nm. Net absorbances were
134 obtained by subtracting the absorbance of negative antigen from the absorbance of each
135 recombinant antigen and visualized by box plots. Cut off was previously defined for genotype B as
136 having a reactivity of >40% relative to the positive control serum reactivity included in each plate
137 (Lacerenza et al., 2006). The same absorbance value was applied for genotype E, due to the lack of
138 a truly negative flock regarding this genotype.

139 For subunit ELISA (P25-B3 epitope), microplates were coated with 220 ng of GST-B3 derived
140 from genotype B (sequence KLNEEAERWRRNNPPPP), genotype E (sequence
141 KLNKEAETWMRQNPQPP) and an equimolar amount of GST as negative control. Net
142 absorbances were obtained by subtracting the GST antigen absorbance from that of each
143 recombinant subunit.

144 Three commercially available ELISAs, based on whole virus (brand A), double recombinants
145 (brand B) or recombinant and synthetic (brand C) antigens, were used to detect and quantify the
146 infection status in BL herd. Assays were carried out according to the manufacturer's protocols.

147 Three commercially available monoclonal antibodies (Mabs), namely 5A1, 10A1 and 8B1, known
148 to be reactive against CAEV and MVV capsid antigen (McGuire et al., 1987) were obtained from

149 VMRD, Inc. (Pullman WA, USA) and tested in P16-25 recombinant ELISA. Mabs were used at
150 dilution of 1µg/well, using mouse-specific secondary antibody. In the latter experiment, a
151 previously produced recombinant capsid antigen from the MVV strain K1514 (Rosati et al., 1999),
152 was used to confirm Mabs cross-reactivity between genotype A and B.

153

154 *Western blot*

155 The genotype B and E recombinant P16-25 proteins were also tested by Western blot using the
156 same three Mabs (5A1, 10A1 and 8B1), as well as a polyclonal serum from mice immunized with
157 recombinant P16-25 of genotype E (positive control) and serum from the same goat from which the
158 Roccaverano strain (genotype E prototype) had originally been isolated.

159 *Sequence analysis and phylogenetic trees*

160 In order to create phylogenetic trees, a model of molecular evolution was estimated using a
161 hierarchical likelihood ratio test approach and the Akaike information criterion (Akaike, 1973)
162 implemented in the software ModelTest ver. 3.7 (Posada and Crandall, 1998, , 2001). Bayesian
163 methods implemented in the computer program MrBayes ver. 3.1.1 (Huelsenbeck and Ronquist,
164 2001. Ronquist and Huelsenbeck, 2003) were used to create phylogenetic trees and to assess
165 statistical support for clades. A Markov chain Monte Carlo search for 1,000,000 generations using
166 two runs with four chains (temperature = 0.05) was performed and results were represented as a
167 50% majority rule consensus tree. Tree statistics and phylogenetic manipulations were calculated
168 from the computer program PAUP* ver. 4.0b10 (Swofford, 2000). Genetic diversity was expressed
169 as nucleotide diversity (Nei, 1987), or the mean proportion of nucleotide differences among
170 sequences.

171 *Statistical analysis*

172 Maximum expected prevalence of both genotypes in the herds was estimated using WinEpiScope
173 2.0 (<http://www.clive.ed.ac.uk/winepiscope/>). Agreement beyond chance between genotype E

174 specific ELISA and commercial assays was analysed using Fleiss'κ coefficient (Fleiss, 1981).
175 Differences in O.D. values between tests were evaluated using Wilcoxon test.

176 *Nucleotide sequence accession numbers*

177 Nucleotide sequences reported in this paper were deposited in the GenBank database with accession
178 numbers FJ547242-46. A subset of additional sequences (EF675997-6026; EU726488-525) had
179 been generated in previous studies (Grego et al., 2007; Reina et al., 2009).

180 **Results**

181 Recombinant antigen derived from genotype E was successfully expressed and purified. SDS-
182 PAGE of the purified product revealed a single protein band of a molecular weight corresponding to
183 the expected size of matrix and capsid antigen fusion protein (41kDa) (Fig 1).

184 Out of 124 blood DNA samples, 56 were PCR positive and the 39 suitable for sequence analysis,
185 were used to construct a phylogenetic tree using CAEV-CO (M33677), Roccaverano (EU293537)
186 and K1514 (M10608) as reference strains. As shown in Fig. 2, all sequences, representative of herd
187 BL belonged to genotype E (8 sequences out of 12 positive PCR reactions). This sample size
188 allowed us to estimate a maximum expected prevalence of B genotype lower than 25% (CI 95%)
189 within this herd.

190 Conversely, sequences obtained from herds TM, BM and CF were grouped into the B1 subtype (15
191 sequences out of 22 PCR reactions), allowing an estimation of a maximum expected prevalence of
192 E genotype lower than 13.5% (CI 95%). Finally, both infections were detected in herd NG with a
193 high degree of co-infections (B and E genotypes) within the same animal, as a previously described
194 genotype specific PCR had revealed (Grego et al., 2007).

195 Box plots of serum reactivity against B and E recombinant antigens are shown in Fig 3. In herd BL
196 and the TM-BM-CF herd group, only one homologous antigen was clearly detected among infected
197 animals based on infectious status determined by phylogenetic analysis, while an apparent cross-
198 reactivity between the two antigens was present only in herd NG. To further study the serological
199 reactivity against genotype specific gag antigens, sera were also tested by subunit ELISA, using

200 P25-B3 epitope from genotypes B and E. Indeed, several samples reacted in a type specific manner
201 (Fig. 4). Serum samples belonging to herd BL showed a distribution mainly oriented towards the E
202 derived epitope and on the contrary, B1 infected herds (TM-BM-CF) were clearly reactive only
203 with the B derived epitope (Fig 4 a and 4b). Herd NG showed a wider distribution of absorbances
204 reaching high values against both peptides (Fig 4 c).

205 Genotype E derived p16-25 antigen reacted also with few sera from the long term negative herd
206 tested and these sera were from Roccaverano goats, while no reactivity was recorded in Saanen and
207 French Alpine goats cohabiting in the same herd (not shown).

208 Commercial ELISAs showed a different capacity in detecting infection in herd BL, being the whole
209 virus ELISA in perfect agreement with genotype E derived ELISA ($\kappa= 1$), followed by the double
210 recombinant ELISA ($\kappa= 0.83$) and the recombinant/synthetic ELISA ($\kappa= 0.47$).

211 Distribution of absorbance values in infected animals of the same herd revealed that the best signal -
212 to - noise ratio belongs to genotype E antigen and is significantly higher than those obtained from
213 the three commercial ELISAs ($p<0.05$) (Fig.5).

214 None of the three Mabs showed reactivity against recombinant antigen derived from genotype E in
215 ELISA while antibody cross-reaction was confirmed when using CAEV and MVV capsid antigens
216 (Fig. 6a). Western blot analysis (Fig. 6b) showed a strong reaction when using p16-25 from
217 genotype B and a weak signal against p16-25 of genotype E, likely due to intrinsic higher
218 sensitivity of WB compared to ELISA under the test conditions applied.

219

220 Discussion

221 In this study we demonstrate that the highly divergent SRLV genotype E can be detected in a type
222 specific manner by a comparative assay using *gag* derived antigens from different genotypes. The
223 true prevalence of viral E genotype might have been underestimated so far for two reasons: i)
224 commercial ELISAs may detect infection without yielding specific serotype information; and ii)
225 some animals may escape diagnosis based on immunodominant epitopes highly divergent from E

226 genotype antigens. The development of a first serological tool specific for genotype E has become
227 therefore essential to monitor natural infection by this genotype in the field. We chose *gag* derived
228 structural proteins since it is well documented that anti-*gag* derived antibody response remains
229 detectable for a long time, if not life long (de Andres et al., 2005), and cross-reactivity between
230 MVV and CAEV P16-25 is acceptable except in the early stage of infection when homologous
231 antigen is more quickly recognised (Lacerenza et al., 2006). Lack of sensitivity of CAEV P16-25 in
232 detecting genotype E infection and vice versa was not surprising. All the diagnostically relevant
233 epitopes characterized so far in the *gag* encoded proteins (summarized in Table 1), show
234 divergences between genotype E and CAEV or MVV. Commercial ELISA based on whole virus
235 preparations demonstrate that genotype E can be detected, likely due to the presence of a larger
236 panel of cross reacting epitopes which may compensate the high variability between genotype E and
237 ELISA antigen. The other commercial ELISAs used are based on a more restricted antigen display
238 and we cannot exclude that, for example, the TM epitope of genotype E might elicit cross-reacting
239 antibodies recognised by heterologous antigen preparations.

240 We previously described P25 immunodominant epitope and matrix protein as specifically detecting
241 A or B genotype infections (Rosati et al., 1999; Grego et al., 2005). Here, we show that genotype E
242 derived P25 immunodominant epitope also recognises only homologous infection. Lack of reactivity
243 of three Mabs, known to be reactive against CAEV and MVV, with genotype E P25 protein was
244 surprising and demonstrates that there are at least three different type specific epitopes in P25. This
245 is relevant when aiming for antigenic detection of a wide spectrum of SRLVs. The hypothesis that
246 genotype E infection could have escaped through traditional diagnostic pressure (test and slaughter
247 policy), was evaluated in a SRLV free herd in which few old aged animals of the Roccaverano
248 breed were slightly reactive against genotype E recombinant antigen. We cannot rule out a false
249 positive reaction since an attempt to amplify viral sequences from the same animals gave
250 inconclusive results. It should be taken into account that no positive reaction was recorded in any of
251 the 292 animals belonging to other breeds of the same herd. On the other hand, animals with a low

252 proviral load could be misdiagnosed using PCR. Interestingly, young animals of the three breeds
253 (Roccaverano, Saanen and French Alpine) were negative, confirming that horizontal transmission is
254 probably hampered due to genetic deletions present in this low pathogenic virus. Thus, artificial
255 feeding of kids, a prevention strategy for SRLV control still used in the herd, is an efficient way to
256 co-eradicate both CAEV and genotype E infections.

257 To date we could identify only three herds infected with genotype E and no more than about a
258 hundred E-infected animals, all belonging to the Roccaverano goat breed (total heads 1500) which
259 was at risk of extinction in the seventies. We are therefore aware that the geographical distribution
260 of genotype E may have a very limited impact in other countries. However, this study shows that
261 the lack of E specific serological tests does not allow the determination of the true prevalence of
262 genotype E infection. Molecular tools such as PCR are now available for genotype E as well as a
263 congruous number of GenBank deposited sequences. The low agreement between PCR and ELISA
264 in detecting genotype E infected animals was not surprising, since this has been widely
265 demonstrated by other authors regarding A or B genotypes. This is likely due to low viral load or to
266 high heterogeneity at the primer binding site (de Andres et al., 2005). It should also be noted that
267 not all the recently described PCRs developed for generic detection of SRLVs may be used for
268 genotype E identification. Primers described by Shah (Shah et al., 2004a) designed to amplify 1.2kb
269 of *pol* gene, will result in a product of 0.8kb, resembling unspecific, due to internal dUTPase
270 deletion. In addition, due to the same deletion, PCR primers described by Eltahir (Eltahir et al.,
271 2006) will fail to amplify as a result of the lack of forward primer binding site.

272 In conclusion, our results demonstrate that, despite the limited information on the distribution of
273 genotype E in small ruminant population, specific antigen design is required for accurate diagnosis.
274 Subunit ELISA using a panel of A, B and E immunodominant region of capsid antigens can be used
275 as a rapid preliminary screening test that would allow serotype determination. On the other hand,
276 evidence regarding this highly divergent genotype has widened the space in which SRLVs have
277 evolved with potential implications for control strategies.

278 **Acknowledgement**

279 This work was supported by Regione Piemonte: Ricerca Scientifica Applicata 2008 and Italian

280 MIUR.

281

282

283

284

Accepted Manuscript

285 Fig legends

286 Fig 1

287 Sodium dodecyl sulfate-polyacrylamide gel electrophoresis showing expression and purification of
288 recombinant antigens in *E.coli*. MW, molecular-weight standard; lane 1 *E. coli* lysate; lanes 2 and 4
289 bacterial lysate expressing GST/P16-25 of It-Pi1 (genotype B) and Roccaverano (genotype E)
290 respectively; lanes 3 and 5, affinity purified recombinant P16-25 of the same strains after GST
291 cleavage and IMAC.

292 Fig 2

293 Phylogenetic tree of amplified *gag* region from different herds. The reference strains of genotype A
294 (M10608), B (M33677) and E (EU293537) are shown.

295 Fig 3

296 Box plots of net absorbances in ELISA testing of sera from BL herd (panel A), NG (panel B) and
297 TM-BM-CF group (panel C) against recombinant P16-25 of genotypes B (left box) and E (right
298 box). Bottom and top of each box represent the first and the third quartiles of the distributions. The
299 median is represented by the line inside the box.

300 Fig4

301 Net absorbance against P25-B3 subunit ELISA: each serum sample was tested against the B derived
302 epitope (X axis) and E derived epitope (Y axis). Samples are from BL herd (panel A), NG (panel
303 B) and TM-BM-CF group (panel C).

304 Fig 5

305 Distribution of O.D. values in BL herd using different serological tests. Sera which were positive to
306 at least two ELISA tests were considered as true positive. Brand A: whole virus based ELISA (O.D.
307 450nm); brand B: double recombinant based ELISA (O.D. 450nm-595nm); brand C recombinant
308 and synthetic based ELISA (O.D. 450nm-595nm); genotype E specific recombinant P16-P25
309 ELISA (O.D. 405nm). Bottom and top of each box represent the first and the third quartiles of the
310 distributions respectively. The median is represented by the line inside the box.

311 Fig.6

312 a) Monoclonal antibody reactivity in ELISA (O.D. 405 nm) against capsid antigen of CAEV isolate
313 (McGuire et al. 1987). Net absorbances against recombinant P16-25 of genotypes E and B and
314 recombinant P25 of genotype A are shown.

315 b) Western blot analysis of the same Mabs (5A1, 10A1 and 8B1) against P16-25 of genotypes B
316 (lanes 1, 2 and 3) and E (lanes 4, 5 and 6). Lanes 7 and 8 were developed using a polyclonal mouse
317 antiserum raised against E derived fusion protein, and a serum from an E infected goat respectively.

318 MW, molecular weight marker.

319

320 Table: Immunodominant linear epitopes of SRLV identified so far along the structural proteins in
321 genotypes A, B and E. Genbank accession numbers, source references and amino acid sequences
322 from each genotype are indicated. Dashes represent the same amino acid as that of genotype B.

323

324

325

326

327

328

329

330

331

332

333

334

335

336

Genotype	GenBank	Matrix (Grego et al., 2005)	Capsid antigen B (Grego et al., 2002)	Capsid antigen D (Rosati et al., 1999)	Nucleoprotein (Lacerenza et al., 2008)	Transmembrane (Bertoni et al., 1994)
B	M33677	KLLTPPEESNKKDFMSL	LNEEAERWRRNNPPPPA	VQQASVEEKMQACRDVGSE	GNGRRGIRVVPSAPPME	ELDCWHYHQYCITS
A	M10608	-N-----TS-RE-A--	--D-----V-Q---G-N	----T-----	--N---P-----L	-----QH--V--
E	EU293537	RSM----ESR---V--	--K---T-M-Q-Q--GP	A-TST----L----E-E-S	--SQ--P-----Q	-I-----G--V--

340
341

Accepted Manuscript

342 REFERENCES

- 343
344 Akaike, H., 1973. Information theory and an extension of an extension of the maximum likelihood
345 principle. In: Proc. 2 Inter. Symposium on Information Theory, Budapest, pp. 267-281.
- 346 Bertoni, G., Zahno, M.L., Zaroni, R., Vogt, H.R., Peterhans, E., Ruff, G., Cheevers, W.P., Sonigo,
347 P., Pancino, G., 1994, Antibody reactivity to the immunodominant epitopes of the caprine
348 arthritis-encephalitis virus gp38 transmembrane protein associates with the development of
349 arthritis. *J Virol* 68, 7139-7147.
- 350 de Andres, D., Klein, D., Watt, N.J., Berriatua, E., Torsteinsdottir, S., Blacklaws, B.A., Harkiss,
351 G.D., 2005, Diagnostic tests for small ruminant lentiviruses. *Veterinary Microbiology* 107,
352 49-62.
- 353 Eltahir, Y.M., Dovas, C.I., Papanastassopoulou, M., Koumbati, M., Giadinis, N., Verghese-
354 Nikolakaki, S., Koptopoulos, G., 2006, Development of a semi-nested PCR using
355 degenerate primers for the generic detection of small ruminant lentivirus proviral DNA. *J*
356 *Virol Methods* 135, 240-246.
- 357 Fleiss, J.L., 1981, *Statistical methods for rates and proportions*. 2nd edn, (J Wiley, New York), 212-
358 236.
- 359 Gogolewski, R.P., Adams, D.S., McGuire, T.C., Banks, K.L., Cheevers, W.P., 1985, Antigenic
360 cross-reactivity between caprine arthritis-encephalitis, visna and progressive pneumonia
361 viruses involves all virion-associated proteins and glycoproteins. *J Gen Virol* 66 (Pt 6),
362 1233-1240.
- 363 Grego, E., Bertolotti, L., Carrozza, M.L., Profiti, M., Mazzei, M., Tolari, F., Rosati, S., 2005,
364 Genetic and antigenic characterization of the matrix protein of two genetically distinct ovine
365 lentiviruses. *Vet Microbiol* 106, 179-185.
- 366 Grego, E., Bertolotti, L., Quasso, A., Profiti, M., Lacerenza, D., Muz, D., Rosati, S., 2007, Genetic
367 characterization of small ruminant lentivirus in Italian mixed flocks: evidence for a novel
368 genotype circulating in a local goat population. *J Gen Virol* 88, 3423-3427.
- 369 Grego, E., Profiti, M., Giammarioli, M., Giannino, L., Rutili, D., Woodall, C., Rosati, S., 2002,
370 Genetic heterogeneity of small ruminant lentiviruses involves immunodominant epitope of
371 capsid antigen and affects sensitivity of single-strain-based immunoassay. *Clin Diagn Lab*
372 *Immunol* 9, 828-832.
- 373 Harmache, A., Russo, P., Vitu, C., Guiguen, F., Mornex, J.F., Pepin, M., Vigne, R., Suzan, M.,
374 1996, Replication in goats in vivo of caprine arthritis encephalitis virus deleted in vif or tat
375 genes: Possible use of this deletion mutants as live vaccines. *AIDS Res Hum Retroviruses*
376 12, 409-411.
- 377 Huelsenbeck, J.P., Ronquist, F., 2001, *MRBAYES: Bayesian inference of phylogenetic trees*.
378 *Bioinformatics (Oxford, England)* 17, 754-755.
- 379 Lacerenza, D., Giammarioli, M., Grego, E., Marini, C., Profiti, M., Rutili, D., Rosati, S., 2006,
380 Antibody response in sheep experimentally infected with different small ruminant lentivirus
381 genotypes. *Vet Immunol Immunopathol* 112, 264-271.
- 382 Lacerenza, D., Genovese, F., Profiti, M., Nucera, D., Rosati, S., 2008, Characterization of an
383 immunodominant epitope of small ruminant lentivirus (SRLV) nucleoprotein. *Vet Immunol*
384 *Immunopathol* 125, 361-367.
- 385 McGuire, T.C., Brassfield, A.L., Davis, W.C., Cheevers, W.P., 1987, Antigenic and structural
386 variation of the p28 core polypeptide of goat and sheep retroviruses. *J Gen Virol* 68 (Pt 8),
387 2259-2263.
- 388 Nei, M., 1987, *Molecular Evolutionary Genetics*. Columbia University Press, New York, NY.
- 389 Pepin, M., Vitu, C., Russo, P., Mornex, J.F., Peterhans, E., 1998, Maedi-visna virus infection in
390 sheep: a review. *Vet Res* 29, 341-367.

- 391 Pisoni, G., Quasso, A., Moroni, P., 2005, Phylogenetic analysis of small-ruminant lentivirus
392 subtype B1 in mixed flocks: evidence for natural transmission from goats to sheep. *Virology*
393 339, 147-152.
- 394 Posada, D., Crandall, K.A., 1998, MODELTEST: testing the model of DNA substitution.
395 *Bioinformatics (Oxford, England)* 14, 817-818.
- 396 Posada, D., Crandall, K.A., 2001, Selecting the best-fit model of nucleotide substitution. *Syst Biol*
397 50, 580-601.
- 398 Reina, R., Grego, E., Bertolotti, L., De Meneghi, D., Rosati, S., 2009, Genome analysis of small
399 ruminant lentivirus genotype E: a caprine lentivirus naturally deleted for dUTPase subunit,
400 vpr-like accessory gene and the 70 bp repeat of U3 region. *J Virol* 83, 1152-1555.
- 401 Ronquist, F., Huelsenbeck, J.P., 2003, MrBayes 3: Bayesian phylogenetic inference under mixed
402 models. *Bioinformatics (Oxford, England)* 19, 1572-1574.
- 403 Rosati, S., Mannelli, A., Merlo, T., Ponti, N., 1999, Characterization of the immunodominant cross-
404 reacting epitope of visna maedi virus and caprine arthritis-encephalitis virus capsid antigen.
405 *Virus Res* 61, 177-183.
- 406 Rosati, S., Profiti, M., Lorenzetti, R., Bandecchi, P., Mannelli, A., Ortoffi, M., Tolari, F., Ciabatti,
407 I.M., 2004, Development of recombinant capsid antigen/transmembrane epitope fusion
408 proteins for serological diagnosis of animal lentivirus infections. *J Virol Methods* 121, 73-
409 78.
- 410 Shah, C., Boni, J., Huder, J.B., Vogt, H.R., Muhlherr, J., Zanoni, R., Miserez, R., Lutz, H.,
411 Schupbach, J., 2004a, Phylogenetic analysis and reclassification of caprine and ovine
412 lentiviruses based on 104 new isolates: evidence for regular sheep-to-goat transmission and
413 worldwide propagation through livestock trade. *Virology* 319, 12-26.
- 414 Shah, C., Huder, J.B., Boni, J., Schonmann, M., Muhlherr, J., Lutz, H., Schupbach, J., 2004b,
415 Direct evidence for natural transmission of small-ruminant lentiviruses of subtype A4 from
416 goats to sheep and vice versa. *J Virol* 78, 7518-7522.
- 417 Swofford, D.L., 2000, PAUP. Phylogenetic Analysis Using Parsimony (and other methods). *In*
418 *Sinauer Associates* Sunderland, Massachusetts.
- 419 Turelli, P., Petursson, G., Guiguen, F., Mornex, J.F., Vigne, R., Querat, G., 1996, Replication
420 properties of dUTPase-deficient mutants of caprine and ovine lentiviruses. *J Virol* 70,
421 1213-1217.
- 422 Zhang, Z., Guo, J., Ni, Y., Bazer, F.W., Giavedoni, L., de la Concha-Bermejillo, A., 2003,
423 Construction and characterization of a recombinant ovine lentivirus carrying the optimized
424 green fluorescent protein gene at the dUTPase locus. *Arch Virol* 148, 1485-1506.
- 425

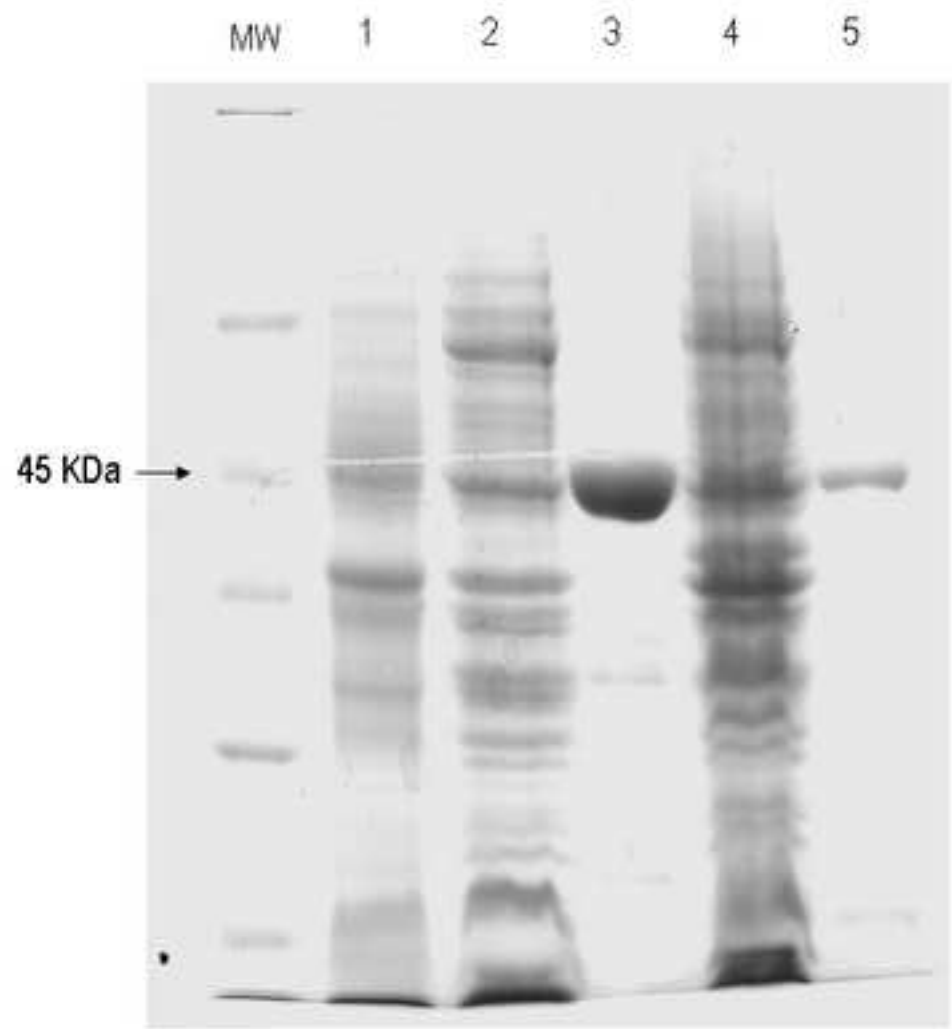


Figure 2

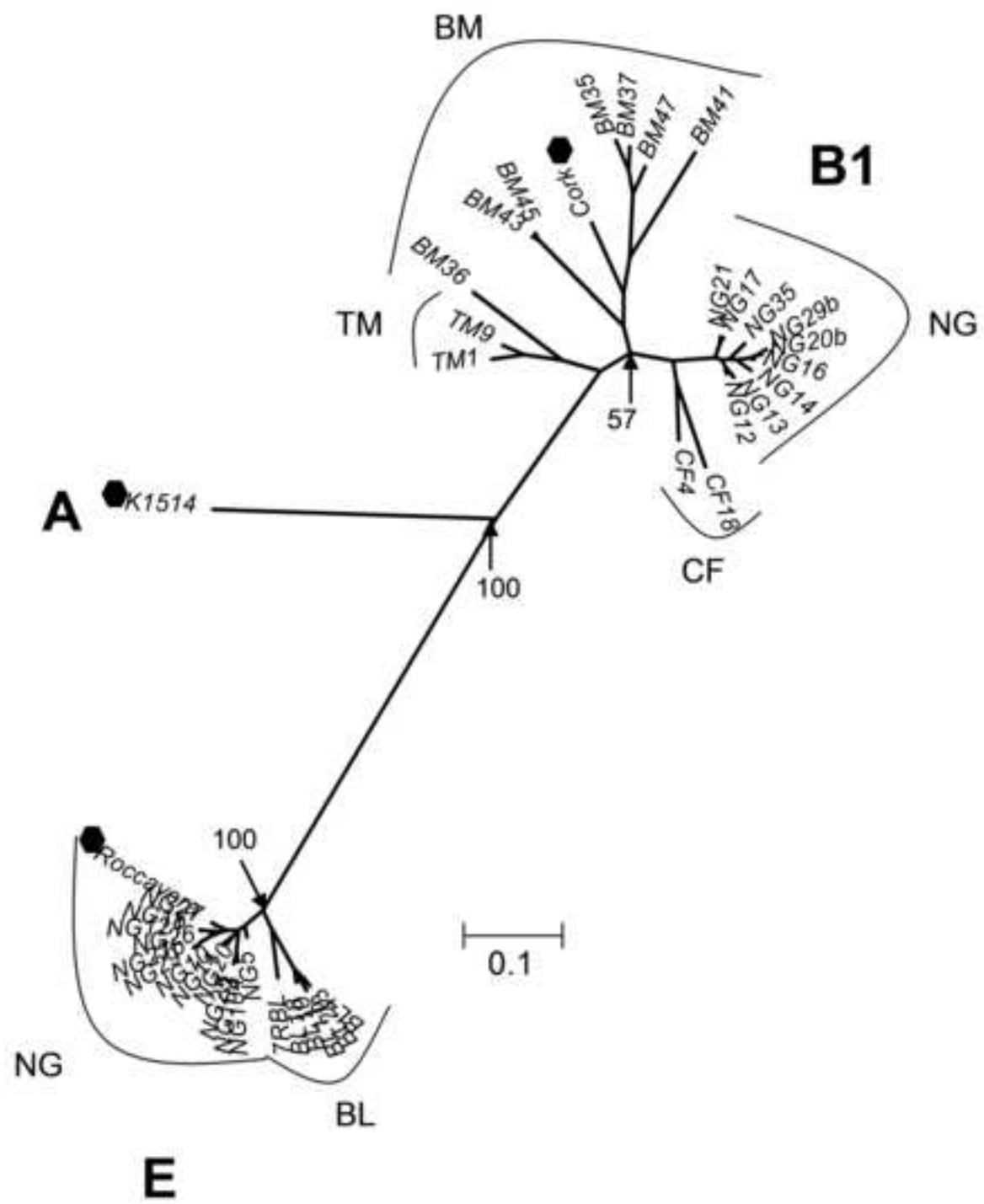
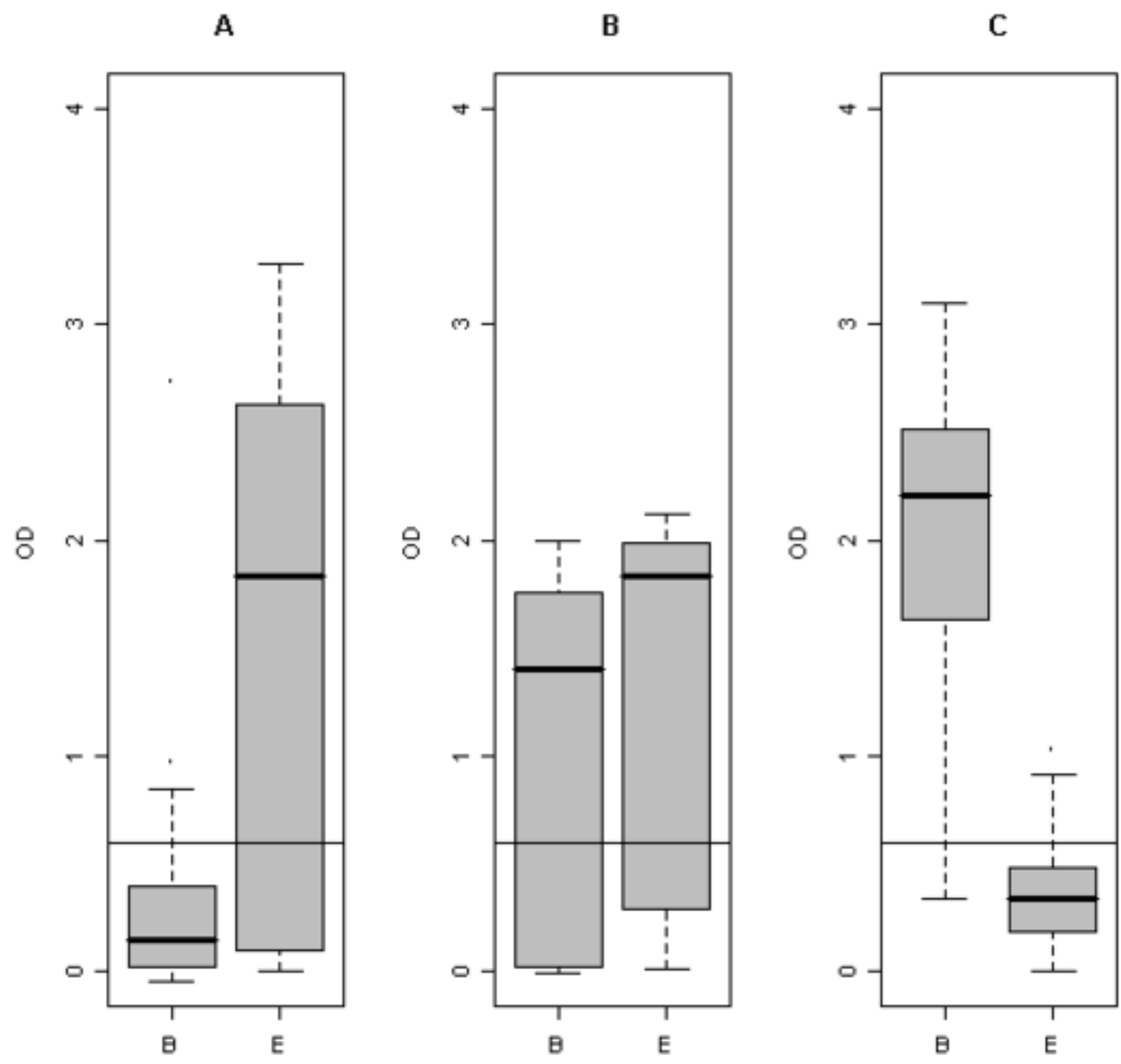


Figure 3



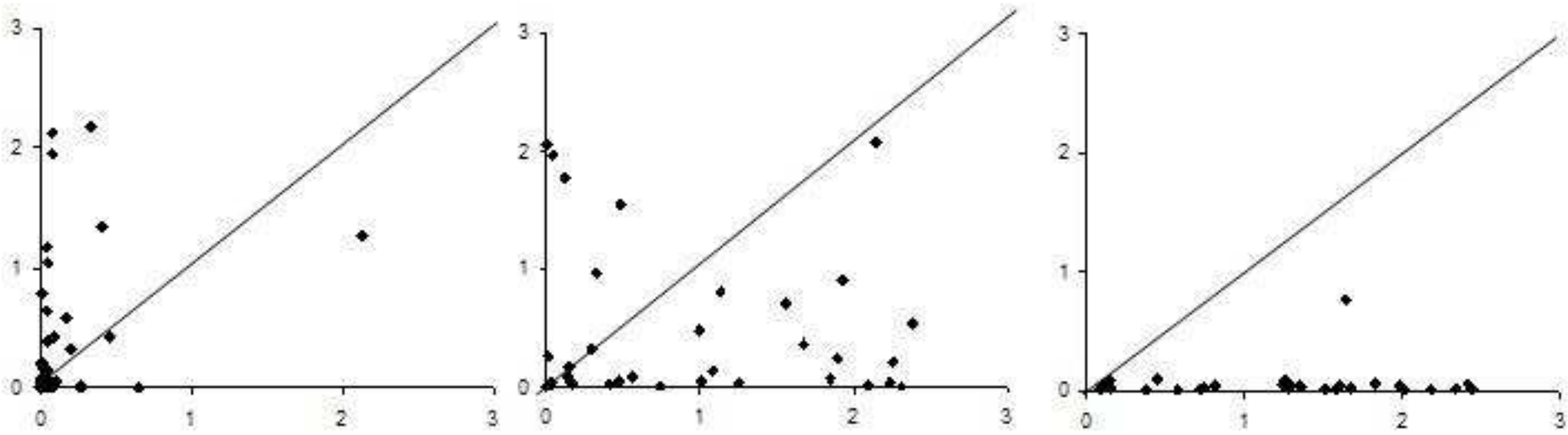


Figure 5

IScrip

