

Horizons and humus forms in beech forests of the Belgian Ardennes

Jean-François Ponge

▶ To cite this version:

Jean-François Ponge. Horizons and humus forms in beech forests of the Belgian Ardennes. Soil Science Society of America Journal, 1999, 63 (6), pp.1888-1901. 10.2136/sssaj1999.6361888x. hal-00504791v2

HAL Id: hal-00504791 https://hal.science/hal-00504791v2

Submitted on 23 Aug 2010

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1	Horizons and humus forms in beech forests of the Belgian Ardennes
2	
3	Jean-François Ponge*
4	
5	*Museum National d'Histoire Naturelle, Laboratoire d'Ecologie Générale, 4 avenue du Petit-Chateau,
6	91800 Brunoy, France.
7	Phone number: +33 1 60479213
8	Fax number: +33 1 60465009
9	E-mail: <u>Jean-Francois.Ponge@wanadoo.fr</u>
10	
11	
12	
13	ACKNOWLEDGEMENTS
14	
15	The author is greatly indebted to Pr. Dr. F. Delecour (Faculté des Sciences Agronomiques
16	Gembloux, Belgium) for the selection of study sites.
17	

Horizons and humus forms in beech forests of the Belgian Ardennes

4 ABSTRACT

Forest soil organic horizons are named on the basis of visual observations made directly in the field, thus this is often subjective. Humus form horizons were compared in thirteen beech stands (*Fagus sylvatica* Ehrh.) of the Belgian Ardennes (western Europe) embracing a wide range of ecological conditions, in order to find more objective bases for their classification. We used a semi-quantitative micromorphological method for the description of horizons, and a multivariate method for data analysis. These methods helped to discern objective discontinuities between Oi, Oe and Oa horizons, adding new criteria for their characterization, such as the root system of trees. Within these horizons, transitions between subhorizons are gradual, thus do not lie on clear-cut criteria. The transition between Oa and A horizons was also gradual. The composition of Oa and A horizons varies according to humus form. The vertical distribution of soil organisms and their vertical movements were considered the origin of discontinuous and continuous processes taking part in the transition from one horizon to another. The observation of horizons under a dissecting microscope may help to find more reliable bases for their nomenclature, even without the use of costly soil sections.

INTRODUCTION

Since the pioneering work of Müller (1889), who described two basic humus forms, mull and torf, beneath Danish beech forests, on the basis of microscopic observations, there have been many attempts to classify humus profiles in forest soils. Different horizons or sub-horizons are generally recognized within the O Horizon (Brady, 1984) or Ao (Duchaufour, 1997) horizon. Although discrepancies concerning the terminology of these strata exist, three main strata are recognized, Oi (entire leaves), Oe (fragmented leaves), and Oa (holorganic faecal pellets). Differences in the development of these horizons, together with structure and chemical properties of the underlying A horizon, allow recognition of three main humus forms, now called mull, moder, and mor (Klinka et al., 1981; Delecour, 1983; Green et al., 1993; Brêthes et al., 1995; Jabiol et al., 1994, 1995). The mor humus form, together with dysmoder (moder with a thick H stratum), has been also called raw humus (Kubiëna, 1953; Delecour, 1980). The recognition of biological processes taking place in the development of humus profiles (Hartmann, 1965; Zachariae, 1965; Bal, 1982) suggested that some features, such as compaction of the soil matrix, deposition of faecal pellets, skeletonization of leaves or tunnelling of needles, are the direct result of soil faunal activity. Thus, as for most biological processes, the transition from one horizon to another should be considered discontinuous, due to tolerance limits and food and habitat preferences of soil organisms.

Characterization of organic horizons and humus forms on the basis of morphological features is common; however, the existence of clear-cut changes between one horizon and another has been poorly demonstrated through quantitative or semi-quantitative morphological data. Federer (1982) pointed out that simple remeasurements of thickness of forest soil horizons may lead to false conclusions if done by two individuals. Discrepancies between field and laboratory observable features of horizons have been reported (Bernier et al., 1993). This may indicate that some morphological criteria used for the definition of organic horizons are inadequate or difficult to employ with accuracy in the field. This is particularly true for the transition from Oa to A horizons and for subdivisions which have been recognized within Oi and Oe horizons (Babel, 1971).

2

3 4

5

6

7

8

9

10

11

12

13

14

15

16 17

18

19

20 21

22

23

horizon.

24

25

26

27

In the present study, our purpose was to determine whether the transition from one organic horizon to another is a continuous or, rather, a step-by-step process, with sharp delineations in horizon properties. In doing so, we addressed the question as to whether horizons exhibit true emergent properties and have not been created by soil scientists for only classification purposes.

MATERIAL AND METHODS

Nomenclature

Comparable organic horizons have several different names depending on the taxonomic system in use. The commonly used nomenclature of Brêthes et al. (1995) corresponds to USDA nomenclature as follows: OL equals Oi, OF equals Oe, and OH equals Oa. These horizons correspond to L, F and H horizons, respectively, recognized by Hesselmann (1926) and later refined by Babel (1971).

The classification of humus forms by Brêthes et al. (1995) was used in this study. It is not based on a strong relationship between the features O and A horizon, as this is the case in other classifications. For instance, a crumby A horizon (typical of mull) may coexist with a thick O horizon (typical of moder). Some humus forms, such as hemimoder and amphimull, were described by other authors as mull-moder (Duchaufour, 1997) or mull-like moder (Kubiëna, 1953; Delecour, 1980, 1983). A hemimoder means a moder humus form (A horizon with organic matter juxtaposed to mineral matter) without any Oa horizon. An amphimull means a mull humus form (A horizon with organo-mineral assemblages) with a distinct Oa

The nomenclature of soil types used in this study followed the FAO-UNESCO classification (Driessen and Dudal, 1991). Cambisols and podzols correspond to Inceptisols and Spodosols,

respectively, in USDA Soil Taxonomy (Brady, 1984). Nomenclature of plant species followed Rameau et al. (1989).

4 Study sites

Thirteen mature beech stands were selected in the Belgian Ardennes, covering a wide range of humus forms (Table 1). Beech (*Fagus sylvatica*) forests were chosen because of their wide distribution and the need for a wide range of environmental conditions, in particular soil and climate, without strong variation in the composition of litter. All studied sites were located on nutrient-poor geological substrates (schists, graywackes, quartzites) ranging from Cambrian to Devonian age. Altitude and related regional factors (climate, mineral richness of parent rock) were found to be the main source of variation of soil animal communities, humus forms and site quality over the studied range (Ponge et al., 1997). Results of litter and soil chemical analyses were also reported in the aforementioned paper.

Categories of the soil matrix in humus horizons

At each site, two humus profiles were sampled for micromorphological description of horizons. These profiles were chosen to represent the range of observed within-site variation of humus forms. Sampling was completed in June 1989. Preparation of the samples (5 x 5 cm section monoliths) was carried out according to the method of Bernier and Ponge (1994), except that only the top 1 cm of the A horizon was sampled. Horizons were separated in the field on the basis of variation visible to the naked eye, without reference to any *a priori* classification. Afterwards, they were classified into Oi, Oe, and Oa horizons according to the abovementioned field criteria, and numbered according to their order from the top to the bottom of a given horizon, i.e. Oi1, Oi2, Oi3, Oe1, Oe2, etc... All 172 horizons were immediately immersed in ethyl alcohol then transported to the laboratory. The composition of each horizon was analyzed by observing the soil matrix in alcohol under a dissecting microscope. No attempt was made to quantify the volume or mass of each category. Rather, we used the following visual semi-quantitative

coding of the abundance of a given category in a given horizon:

3	absent	0
4	present but scarce	1
5	present and common	2
6	present and dominant	3

A total of 185 categories were recognized. Most of them were plant litter, in varying degrees of decomposition or comminution by fauna. Animal faeces were classified according to the corresponding animal group, their degree of comminution by other animals, and their degree of connection with uneaten plant categories. Animals were counted and classified into broad groups.

Multivariate analysis

Correspondence analysis (Greenacre, 1984) was used to give an overall picture of affinities and differences between horizons and/or categories. This method, using the chi-square distance, allows horizons and categories to be simultaneously projected on factorial axes, thus groups of samples could be directly associated with groups of categories. As was theoretically developed by Benzécri (1973), this method has been refined for particular purposes such as community gradient analysis [detrended correspondence analysis (Hill and Gauch, 1980)] or the study of community-environment relationships [canonical correspondence analysis (Ter Braak, 1987)]. Here, our purpose was to analyze the structure of a data matrix (the composition of horizons) without any a priori hypotheses concerning their relationships with environmental factors and with more than one single factor suspected to result from the analysis. Additional (passive) variables were used to help interpretation of the factorial axes and not to quantify causal relationships. Thus, we used the approach of original correspondence analysis, that was devised to evaluate global patterns underlying complex data matrices, and not that of the aforementioned derived methods.

2

3

4

5

6

7

8 9 10

12 13

11

15 16

14

17

18 19

20

21

22

23

24

25

26

27

We analyzed first the total 172 horizons and 185 categories. Only axes 1 and 3 were considered for the global description of humus horizons. Axis 2 was neglected, because it isolated only one horizon (Oe1 in the second profile studied in site 16), which was characterized by the development of the root system of hair-grass (Deschampsia flexuosa) at the inside of decaying plant fragments. This development

Matrices were analyzed so that horizons were observations (rows) and categories were active variables (columns). Animal groups, horizon names, humus form names, phytosociological types, and altitude were added as passive variables. They had no influence on the results, but they were projected on factorial axes as the active variables.

Categories were coded and counted as indicated above. Animal groups were recorded as total

counts. Horizons, humus forms and phytosociological types were coded as 1 when a given horizon (row)

belonged to a given category (column), 0 when not. Altitude was recorded in meters. All variables (active

and passive) were standardized with unit variance and a mean of 20. This allowed contributions to

factorial axes to be proportional to factorial coordinates and data of a different nature, such as counts,

semi-guantitative coding and measurements, to be included in the same analysis (Ponge and Delhaye

1995). In the case of altitude, high elevation and low elevation were distinguished. Standardized altitude

values (high elevation) were complemented to 40 in order to create a new variable, varying in an opposite

sense (low elevation). The new variable had similar mean (20) and standard deviation (1). Following this

procedure, already used by Ponge and Delhaye (1995), the range of elevation values was described by

two symmetrical points.

RESULTS

Composition of the humus profile

was not apparent in other humus profiles, due to scarcity of ground vegetation, and we preferred to discard the corresponding axis rather than to discard this horizon from the analysis.

The projection of horizons in the plane of axes 1 and 3 (Fig. 1) indicated that they could be classified into three groups, corresponding to Oi, Oe, and Oa+A horizons, respectively. All horizons sampled did not fall exactly in each of the three branches depicted by the plane of axes 1 and 3. This does not indicate that Oi, Oe, and Oa+A horizons were poorly differentiated on the basis of their composition. Rather, this phenomenon indicates that a few horizons had been badly classified in the field; correspondence analysis revealed that they had more points in common with another group than with their own group. Differences between Oa and A horizons were negligible when compared with differences between Oi and Oe horizons. Oa and A horizons did not exhibit distinct branches. In addition, differences within each of the Oi, Oe or Oa horizons were negligible compared to differences between these horizons.

The projection of categories in the plane of axes 1 and 3 (Appendix 1) allowed us to characterize each horizon or group of horizons by its composition (Table 2). The Oi horizon was characterized by entire leaves of beech at varying stages of fungal conditioning and small recently fallen tree categories such as bud scales, male flowers, and twigs. Smears of faecal material, holorganic or organo-mineral, were observed at the surface of some beech leaves. A laminated mixing of leaf fragments and faeces of litter-consuming enchytraeids, which we called "sandwich material", together with plant categories such as male flowers of beech, grass stems, twigs, at some stage of decomposition, was characteristic of the Oe horizon. The presence of beech petioles and nerves, mycelial strands and living bases of wood-rush (*Luzula forsteri*) was also noticeable. Both Oa and A horizons were characterized by dead or senescent fine roots, living and dead woody roots, old (compacted) holorganic enchytraeid faeces, and fragments of parent rock (both intact or weathered). Charcoal was also present in this group of horizons. Sclerotia of the mycorrhizal ascomycete *Cenococcum geophilum* also occurred.

Living fine roots of beech and faeces of litter-consuming enchytraeids were placed in an intermediate position between the Oe horizon and the Oa+A horizons. This indicated that they were present in all of these horizons and, thus, they did not help in differentiating among horizons. Maple (*Acer pseudoplatanus*) and ash (*Fraxinus excelsior*) plant materials were different. These categories only occurred at one site (100) which was characterized by an intense earthworm activity, with abundant cast deposition within the Oi horizon (Fig. 1). The projection of corresponding samples was influenced by the presence of organo-mineral material (typical of the A horizon of mull humus) together with categories typical of the Oi horizon, which influenced in turn the position of the abovementioned ash and maple categories.

Oa and A horizons, which were considered as diagnostic horizons for the separation of mull versus moder humus forms (Delecour, 1983; Brêthes et al., 1995), were not clearly separable on the basis of composition as revealed by the use of a dissecting microscope. In order to reveal possible variation in the composition of these horizons, we analyzed separately Oa and A horizons (partial analysis). The projection of horizons in the plane of axes 1 and 2 (Fig. 2) separated Oa and A (classified by field observation), but with a high degree of overlapping. This indicated that the distinction between Oa and A horizons that had been made in the field (Oa = holorganic, A = organo-mineral) did not reflect true composition. The projection of some passive variables such as humus form, altitude and phytosociological type (Fig. 3) helped to establish a link between Axis 2 of partial analysis and ecological conditions prevailing in the studied sites. Mull humus forms were on the negative side of Axis 2; moder humus forms were placed on the positive side of Axis 2.

The composition of Oa and A horizons, as indicated by the projection of categories in the plane of axes 1 and 2 of partial analysis (Appendix 2), depended on the humus form. The Oa horizon of moder humus forms (eumoder, hemimoder and dysmoder) was mostly characterized by compacted holorganic faeces of enchytraeids, different plant organs hard to decay, bundles of skeletonized beech leaf fragments, living woody roots of beech, and dead black mycorrhizae of beech produced by *Cenococcum*

geophilum. The upper part of the A horizon of mull humus forms (oligomull, amphimull, dysmull) was mostly characterized by organo-mineral earthworm faeces, milliped faeces, intact stones, beech leaves skeletonized by macrofauna or mesofauna and categories belonging to ground vegetation such as the root system of grass species or bleached leaves of wood-sorrel (*Oxalis acetosella*). The Oa horizon of amphimull shared many features with the A horizon of the mull group (indicated by its position on the negative side of Axis 2). The A horizon of the moder group was placed in an intermediary position (not far from the origin, on both sides of Axis 2), thus without characteristic features distinguishing both the horizon and the humus form.

Distribution of soil animals

Animal groups found during the dissection of humus horizons (Table 3) were projected as passive variables in the plane of axes 1 and 3 of total analysis (Fig. 4). Comparison with the position of horizons (Fig. 1) revealed that the Oi horizon was very poor in fauna at the time of sampling (June), no animal group being placed far from the origin in the direction of the Oi Ihorizon (see also Table 3). The Oe horizon was characterized by mites other than *Platynothrus peltifer* (MIT, ORI, PHT) and springtails (SPR), the latter being present at a greater depth than the former as ascertained by the respective position of corresponding points along Axis 1. The only group that characterized the Oa+A horizons was enchytraeids (ENC). Enchytraeids mainly characterized the Oa horizon, as it appeared from the comparison of Oa and A horizons (Fig. 5, Table 3). Most fauna (here mostly mesofauna) were concentrated in the Oe and Oa horizons, the most abundant group being enchytraeids, followed at a far lower level of abundance by mites and springtails, with an increasing preference for deeper horizons at the time of sampling in the order mites < springtails < enchytraeids.

DISCUSSION AND CONCLUSION

The existence of a transitional horizon between typical Oa and A horizons, due to a progressive enrichment in mineral particles of the Oa horizon, has been suggested by Delecour (1980, 1983). He even considered it as a characteristic feature of the humus forms belonging to the moder group, rather than the accumulation of holorganic faeces in Oe and Oa horizons which is commonly used to define moder humus (Klinka et al., 1981; Green et al., 1993, Duchaufour, 1997). This feature is also used in the classification of forest humus forms by Brêthes et al. (1995) where the A horizon of moder humus forms is considered to be made of holorganic faeces juxtaposed to mineral particles, without true incorporation of organic matter to mineral matter. This composition can be achieved by the vertical movements of small animals carrying mineral particles onto their tegument and depositing them, together with faeces, in the course of their wandering between food sources (Oe horizon) and refuges (Oa and A horizons). In moder humus forms, where no other agent mixes organic matter with mineral matter, this behaviour is typical of animals with a sticky tegument such as enchytraeid worms (Ponge, 1991), whose daily vertical movements of several centimeters are well-known (Springett et al., 1970). In this case the gradual passage from the Oa to the A horizon can be interpreted as an active (biological) diffusion process, the source of organic matter being decaying litter, which is actively consumed by these animals and humified (Ponge, 1991), and the source of mineral matter being underlying mineral horizons. As has been observed in other studies (Zachariae, 1965; Ponge, 1991) enchytraeids also consume faeces of other animals such as oribatid mites. As they live deeper (on average) than other litter-dwelling animals (Table 3) they progressively incorporate faeces of litter-consuming animals into their own faecal material. This may explain why the Oa horizon of moder humus forms appears mainly made of compacted enchytraeid faeces and unconsumed material rather than remains of the activity of all litter-feeding animals. They also ingest silt particles (unpublished data), and carry them onto their tegument (Ponge, 1988). This pattern may lead to the observed soft transition from Oa to A horizons.

24

25

26

27

1

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

The presence of clear discontinuities between Oi and Oe horizons and between Oe and Oa+A horizon, and the absence of clearcut subdivisions within them, may be explained by threshold levels in the vertical distribution of soil animals and other organisms. We cannot speculate from our data about the

depth levels at which animals consume litter and defecate at any time of the year, and about all the factors which govern these patterns. Nevertheless, it can be inferred from existing literature that the vertical distribution of soil fauna is determined, among others, by i) genetically fixed behavioural features (Stevenson and Dindal, 1982; Kretzschmar, 1984), ii) ecophysiological requirements of animals (Atalla and Hobart, 1964; Haukka, 1987), iii) the stage of decomposition of litter (Hayes, 1963; Soma and Saitô, 1983; David, 1986). For instance, if we take into account only leaf litter as a food, the depth level at which animals will consume it can be interpreted as a compromise between the need for food of a high nutritional value such as freshly fallen litter (Soma and Saitô, 1983), the toxicity or repellence of some compounds present in undecayed litter (Satchell and Lowe, 1967), and the search for better micro-climate conditions (Joosse, 1971). If we examine diagnostic features of Oi and Oe horizons as ascertained in our analysis (Table 2) it appears that the passage from Oi to Oe horizon is characterized by the deposition of faeces of small litter-consuming animals, such as oribatid mites and enchytraeids, these categories being near absent in the Oi horizon.

The passage from the Oe to the Oa+A horizon is marked by the appearance of the root system of beech (Table 2). This creates a new discontinuity along the humus profile. According to the humus form, the feeder root system of beech will develop throughout accumulated organo-mineral earthworm faeces (A horizon of mull humus) or holorganic enchytraeid faeces (Oa horizon of moder humus). This common feature is probably the main reason for the absence of a clear distinction between Oa and A horizons in the global analysis (Fig. 1). From existing literature, it can be concluded that the vertical distribution of root tips is strongly influenced by humus form horizons and the humus form itself (Meyer and Göttsche, 1971; Persson, 1983; Harvey et al., 1986; Ponge, 1988; Bernier and Ponge, 1994). These results point out that the upper part of the Oa horizon together with the bottom of the Oe horizon is the main micro-site for mycorrhizal root development in moder humus forms, mull humus forms being characterized by a lower number of root tips which are widely distributed throughout the A horizon. Unfortunately mechanisms which could explain these patterns are poorly understood, although the need for available water and nutrients has been advocated as a reason for the preferential development of feeder roots in

carbon-rich substrates (Persson, 1983).

2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

1

As this is now well-admitted the genesis of humus form horizons, and thus of humus forms, is placed under the influence of microflora, fauna, and plant subterranean parts, whose activity is determined by climate, parent rock and quantity and quality of vegetation (Kubiëna, 1955; Bal, 1982; Toutain, 1987; Green et al., 1993; Brêthes et al., 1995). The existence of horizons, not for the purpose of classification, but as a discernable entity, with clear emergent properties, can be understood if we take into account the accumulation through time of key categories with a high longevity. This is the case for dead leaves, enchytraeid faeces, earthworm faeces, or fine roots of trees. If a category present in a given horizon has a low longevity, i.e. if it is rapidly transformed into another category or if it disappears from the soil matrix through mineralization or leaching, then it cannot directly participate in the building of this horizon. Rather, it may, i) participate in the building of underlying horizons (for instance colloidal organic compounds precipitating in B horizons), ii) be taken-up by organisms, iii) diffuse in the atmosphere or in the water flow. This is the case, among others, of oribatid faeces which rapidly disappears from the moder profile when consumed by enchytraeids (Ponge, 1991). This is also the case for enchytraeid faeces when ingested in turn by earthworms living in mull humus (Bal, 1982). The passage from one horizon to another should thus be considered as a check in a continuous process (the slow maturation of individual categories in the absence of disturbances), and a start for another continuous process of maturation. The fact that at a given depth level the bulk of categories of a given type are transformed into categories of another type (due to the activity of vertically distributed organisms) will create discontinuities visible to the naked eye and thus will help us to distinguish horizons and humus forms on the field. In the absence of these discontinuities (as between Oa and A horizons of moder humus forms) this distinction may be more difficult and field observation may lead to false conclusions, as has been always suggested by Bernier et al. (1993). In this case the identification of horizon components under a dissecting microscope and the use of counting methods (Bernier et al., 1993; Bernier and Ponge, 1994) may help a better characterization of these horizons.

1	REFERENCES
2	
3	Atalla, E.A.R., and J. Hobart. 1964. The survival of some soil mites at different humidities and their
4	reaction to humidity gradients. Ent. Exp. Appl. 7: 215-228.
5	
6	Babel, U. 1971. Gliederung und Beschreibung des Humusprofils in mitteleuropäischen Wäldern.
7	Geoderma 5: 297-324.
8	
9	Bal, L. 1982. Zoological ripening of soils. PUDOC, Wageningen.
10	
11	Benzécri, J.P. 1973. L'analyse des données. II. L'analyse des correspondances. Dunod, Paris.
12	
13	Bernier, N., and J.F. Ponge. 1994. Humus form dynamics during the sylvogenetic cycle in a mountain
14	spruce forest. Soil Biol. Biochem. 26: 183-220.
15	
16	Bernier, N., J.F. Ponge, and J. André. 1993. Comparative study of soil organic layers in two bilberry-
17	spruce forest stands (Vaccinio-Piceetea). Relation to forest dynamics. Geoderma 59: 89-108.
18	
19	Brady, N.C. 1984. The nature and properties of soils. Macmillan, New York.
20	
21	Brêthes, A., J.J. Brun, B. Jabiol, J.F. Ponge, and F. Toutain. 1995. Classification of forest humus forms: a
22	French proposal. Ann. Sci. For. 52: 535-546.
23	
24	David, J.F. 1986. Influence de la durée du séjour dans la litière des feuilles mortes de chêne (Quercus
25	petraea Liebl.) sur leur consommation par le Diplopode Cylindroiulus nitidus (Verhoeff, 1891).
26	C.R. Acad. Sc. Paris, Sér. III 302: 379-381.
27	

1	Delecour, F. 1980. Essai de classification pratique des humus. Pédologie 30: 225-241.
2	
3	Delecour, F. 1983. Les formes d'humus: identification et description. Natural. Belg. 64: 75-86.
4	
5	Driessen, P.M., and R. Dudal. 1991. The major soils of the world. Agricultural University of Wageningen,
6	Wageningen, and Catholic University of Leuven, Leuven.
7	
8	Duchaufour, P. 1997. Pédologie. Sol, végétation, environnement. 5 th ed. Masson, Paris.
9	
10	Federer, C.A. 1982. Subjectivity in the separation of organic horizons of the forest floor. Soil Sci. Soc.
11	Am. J. 46: 1090-1093.
12	
13	Green, R.N., R.L. Trowbridge, and K. Klinka. 1993. Towards a taxonomic classification of humus forms.
14	For. Sci. Monogr. N° 29.
15	
16	Greenacre, M.J. 1984. Theory and applications of correspondence analysis. Academic Press, London.
17	
18	Hartmann, F. 1965. Waldhumusdiagnose auf biomorphologischer Grundlage. Springer-Verlag, Wien.
19	
20	Harvey, A.E., M.F. Jurgensen, M.J. Larsen, and J.A. Schlieter. 1986. Distribution of active
21	ectomycorrhizal short roots in forest soils of the inland Northwest: effects of site and disturbance.
22	USDA Forest Service, Intermountain Research Station, Ogden, Research Paper INT-374.
23	
24	Haukka, J.K. 1987. Growth and survival of Eisenia fetida (Sav.) (Oligochaeta: Lumbricidae) in relation to
25	temperature, moisture and presence of Enchytraeus albidus (Henle) (Enchytraeidae). Biol. Fertil.
26	Soils 3: 99-102.

1	Hayes, A.J. 1963. Studies on the feeding preferences of some phthiracarid mites (Acari: Oribatidae). Ent.
2	Exp. Appl. 6: 241-256.
3	
4	Hesselmann, H. 1926. Studier över barrskogens humustäcke. Meddel. Stat. SkogsförsOksanst. 22: 169-
5	552.
6	
7	Hill, M.O., and H.G. Gauch Jr. 1980. Detrended correspondence analysis: an improved ordination
8	technique. Vegetatio 42: 47-58.
9	
10	Jabiol, B., A. Brêthes, J.J. Brun, J.F. Ponge, and F. Toutain. 1994. Une classification morphologique et
11	fonctionnelle des formes d'humus. Propositions du référentiel pédologique 1992. Rev. For. Fr. 46:
12	152-166.
13	
14	Jabiol, B., A. Brêthes, J.F. Ponge, F. Toutain, and J.J. Brun. 1995. L'humus sous toutes ses formes.
15	ENGREF, Nancy.
16	
17	Joosse, E.N.G., 1971. Ecological aspects of aggregation in Collembola. Rev. Ecol. Biol. Sol 8: 91-97.
18	
19	Klinka, K., R.N.Green, R.L. Trowbridge, and L.E. Lowe. 1981. Taxonomic classification of humus forms in
20	ecosystems of British Columbia. First approximation. Land Manag. Rept. N° 8.
21	
22	Kretzschmar, A. 1984. Besoins biologiques des vers de terre et porosité du sol. Bulletin GFHN 15: 96-
23	102.
24	
25	Kubiëna, W.L. 1953. The soils of Europe. Illustrated diagnosis and systematics. CSIC, Madrid, Spain, and
26	Thomas Murby, London.

1	Kubiëna, W.L. 1955. Animal activity in soils as a decisive factor in establishment of humus forms. p. 73-82
2	+ 2 inlet plates. In D.K. McE. Kevan (ed.) Soil zoology. Butterworths, London.
3	
4	Meyer, F.H., and D. Göttsche. 1971.Distribution of root tips and tender roots of beech. p. 47-52. In H.
5	Ellenberg (ed.) Ecological studies. Analysis and synthesis, vol. 2. Springer-Verlag, Berlin.
6	
7	Müller, P.E. 1887. Recherches sur les formes naturelles de l'humus et leur influence sur la végétation et
8	le sol. Ann. Sci. Agron. Fr. Etrang. 6: 85-423 + 7 inlet plates.
9	
10	Persson, H. 1983. The distribution and productivity of fine roots in boreal forests. Plant Soil 71: 87-101.
11	
12	Ponge, J.F. 1988. Etude écologique d'un humus forestier par l'observation d'un petit volume, premiers
13	résultats. III. La couche F1 d'un moder sous Pinus sylvestris. Pedobiologia 31: 1-64.
14	
15	Ponge, J.F. 1991. Succession of fungi and fauna during decomposition of needles in a small area of
16	Scots pine litter. Plant Soil 138: 99-113.
17	
18	Ponge, J.F., P. Arpin, F. Sondag, and F. Delecour. 1997. Soil fauna and site assessment in beech stands
19	of the Belgian Ardennes. Can. J. For. Res. 27: 2053-2064.
20	
21	Ponge, J.F., and L. Delhaye. 1995. The hetrogeneity of humus profiles and earthworm communities in a
22	virgin beech forest. Biol. Fertil. Soils 20: 24-32.
23	
24	Rameau, J.C., D. Mansion, and G. Dumé. 1989. Flore forestière française. Guide écologique illustré. I.
25	Plaines et collines. Institut pour le Développement Forestier, Paris.
26	
27	Satchell, J.E., and D.G. Lowe. 1967. Selection of leaf litter by Lumbricus terrestris. p. 102-119. In O. Graff

1	and J.E. Satchell (ed.) Progress in soil biology. Friedr. Vieweg, Braunschweig, and North-Holland
2	Publishing Company, Amsterdam.
3	
4	Soma, K. and T. Saitô. 1983. Ecological studies of soil organisms with reference to the decomposition o
5	pine needles. II. Litter feeding and breakdown by the woodlouse, Porcellio scaber. Plant Soil 75
6	139-151.
7	
8	Springett, J.A., J.E.Brittain, and B.P. Springett. 1970. Vertical movement of Enchytraeidae (Oligochaeta
9	in moorland soils. Oikos 21: 16-21.
10	
11	Stevenson, B.G., and D.L. Dindal. 1982. Effect of leaf shape on forest litter spiders: community
12	organization and microhabitat selection of immature Enoplognatha ovata (Clerck) (Theridiidae). J
13	Arachnol. 10: 165-178.
14	
15	Ter Braak, C.J.F. 1987. The analysis of vegetation-environment relationships by canonical
16	correspondence analysis. Vegetatio 69: 69-77.
17	
18	Thill, A., M. Dethioux, and F. Delecour. 1988. Typologie et potentialités forestières des hêtraies naturelles
19	de l'Ardenne Centrale. IRSIA, Brussels.
20	
21	Toutain, F. 1987. Activité biologique des sols, modalités et lithodépendance. Biol. Fertil. Soils 3: 31-38.
22	
23	Zachariae, G. 1965. Spuren tierischer Tätigkeit im Boden des Buchenwaldes. Forstwiss. Forsch. N° 20.
24	

1	LEGENDS OF FIGURES
2	
3	Fig. 1. Results from total correspondence analysis of organic horizons with faunal characteristics in beech
4	forests of the Belgian Ardennes. Projection of horizons (rows) in the plane of axes 1 and 3.
5	Indicated names of horizons were based on field observation and were not included as main
6	variables.
7	
8	Fig. 2. Partial correspondence analysis on Oa and A horizons. Projection of horizons (rows) in the plane
9	of axes 1 and 2. Otherwise as for Fig. 1.
10	
11	Fig. 3. Partial correspondence analysis on Oa and A horizons. Projection of passive variables in the
12	plane of axes 1 and 2. (festuc = Luzulo-Fagetum festucetosum; melic = Melico-Fagetum
13	festucetosum; typic = Luzulo-Fagetum typicum; vaccin = Luzulo-Fagetum vaccinietosum).
14	
15	Fig. 4. Total correspondence analysis. Projection of animal groups (passive variables) in the plane of
16	axes 1 and 3. Coding of animal groups as in Table 3.
17	
18	Fig. 5. Partial correspondence analysis on Oa and A horizons. Projection of animal groups (passive
19	variables) in the plane of axes 1 and 2. Otherwise as for Fig. 4.
20	

Table 1. Main features of the 13 studied sites. Phytosociological types according to Thill et al. (1988). Soil types according to FAO-UNESCO classification (Driessen and Dudal 1991). Humus forms according to Brêthes et al. (1995).

Site	Altitude	Phytosociological type	Soil type	Humus form
1	370 m	Luzulo-Fagetum festucetosum	Dystric cambisol	Dysmull
3	465 m	Luzulo-Fagetum festucetosum	Dystric cambisol	Eumoder
4	500 m	Luzulo-Fagetum typicum	Dystric cambisol	Dysmoder
5	505 m	Luzulo-Fagetum vaccinietesosum	Dystric cambisol	Eumoder to dysmoder
16	445 m	Luzulo-Fagetum vaccinietesosum	Dystric cambisol	Eumoder
17	430 m	Luzulo-Fagetum typicum	Dystric cambisol	Hemimoder to eumoder
22	400 m	Luzulo-Fagetum typicum	Gleyic cambisol	Eumoder to dysmoder
24	390 m	Luzulo-Fagetum festucetosum	Dystric cambisol	Dysmull to dysmoder
26	430 m	Luzulo-Fagetum vaccinietesosum	Leptic podzol	Dysmoder
28	375 m	Luzulo-Fagetum festucetosum	Dystric cambisol	Amphimull to eumoder
40	385 m	Luzulo-Fagetum vaccinietesosum	Ferric podzol	Dysmoder
100	350 m	Melico-Fagetum festucetosum	Dystric cambisol	Oligomull to dysmull
307	380 m	Luzulo-Fagetum vaccinietesosum	Leptic podzol	Amphimull

Table 2. Most typical categories in the three main groups of horizons depicted by correspondence analysis

Oi

Entire brown leaves of beech Entire bleached leaves of beech Intact bud scales of beech Intact male inflorescences of beech

Entire variegated leaves of beech

Intact twigs

Holorganic faecal material smearing beech leaves

Organo-mineral material smearing beech leaves

Oe

Sandwich material made of beech leaf fragments and holorganic enchytraeid faeces

Brown decaying male inflorescences of beech

Fragments of grass stems browsed by fauna

Pollen

Woodlice shells

Petioles and nerves of beech filled with faeces of phthiracarid oribatid mites

Bud scales of beech, entire but brown and soft

Intact unidentified fragments of seed wings

Intact seed coats of beech

Twigs filled with enchytraeid holorganic faeces

Twigs filled with oribatid holorganic faeces

Living leaf bases of Luzula albida

White rhizomorphs

Oa+A

Dead pale yellow creamy mycorrhizae of beech

Sclerotia of Cenoccum geophilum

Intact dead fine long roots of beech

Compacted holorganic enchytraeid faeces

Intact living woody roots of beech

Strongly decayed bud scales of beech

Well-decayed bark fragments

Intact stones

Decaying woody roots of beech

Weathering stones

Charcoal

Dead fine long roots of beech tunnelled by fauna

Decaying grass roots

Dead black mycorrhizae of beech

Compacted organo-mineral material

Table 3. Mean number of animals which were found during the dissection of humus horizons (L \times I \times h = 5 \times 5 \times 1 cm)

		Oi	Oe	Oa	Α
		(n=24)	(n=59)	(n=44)	(n=24)
ADB	Adult beetles	0.02	0	0.02	0
ADF	Adult flies	0.01	0.02	0	0
ANT	Ants	0.01	0	0	0
BEE	Beetle larvae (miscellaneous)	0.14	0.3	0.09	0.04
ВОО	Booklice	0.01	0.02	0	0
CEC	Cecidomyid fly larvae	0.27	0.32	0.22	0.17
CEN	Centipedes	0	0.06	0.09	0
CER	Ceratopogonid fly larvae	0.05	0.02	0	0
CHI	Chironomid fly larvae	0.25	0.3	0.64	0.25
CLI	Click-beetle larvae	0.06	0.03	0.19	0.04
COC	Cochineals	0.01	0.11	0	0
COP	Copepods	0.04	0.01	0	0
DIP	Diplura	0.14	0.21	0.15	0.08
DOL	Dolichopodid-empidid fly larvae	0.02	0.21	0.28	0.33
EAR	Earthworms	0.07	0.01	0.19	0
ENC	Enchytraeids	12	113	231	50
FLY	Fly larvae (miscellaneous)	0.02	0.11	0.06	0.04
MIL	Millipeds	0	0	0	0.04
MIT	Mites (miscellaneous)	10	19	10	2
ORI	Oribatid mites (miscellaneous)	10	20	14	2
PAU	Pauropods	0.37	0.48	0.64	0.08
PHT	Phthiracarid oribatid mites	2	24	8	1
PLA	Platynothrus peltifer (oribatid mite)	6.9	4.9	3.5	0.5
PRO	Protura	0	0.01	0.2	0.17
PSE	Pseudoscorpions	0.06	0	0	0.04
SCI	Sciarid fly larvae	1.5	4.8	4.4	0.5
SLU	Slugs	0.07	0	0	0
SPI	Spiders	0.17	0.25	0.06	0
SPR	Springtails	14	64	52	17
SYM	Symphiles	0.1	0.17	0.34	0.13
THR	Thrips	0.02	0.02	0	0
TIP	Tipulid fly larvae	0.02	0.02	0	0
woo	Woodlice	0.05	0.01	0	0

Appendix 1. Categories used for the description of humus horizons. Coordinates on factorial axes 1 and 3 of total correspondence analysis

Categories	Axis 1	Axis 3
Entire brown leaves of beech	-0.0280	-0.0142
Bundles of entire brown leaves of beech	-0.0114	-0.0058
Brown leaves of beech skeletonized by macrofauna	-0.0140	0.0020
Bundles of brown leaves of beech skeletonized by macrofauna Brown leaves of beech skeletonized by mesofauna	-0.0124	0.0011
Bundles of brown leaves of beech skeletonized by mesofauna	-0.0085 -0.0065	-0.0056 -0.0035
Entire variegated leaves of beech	-0.0219	-0.0035
Bundles of entire variegated leaves of beech	-0.0146	-0.0078
Entire variegated leaves of beech skeletonized by macrofauna	-0.0050	-0.0013
Entire variegated leaves of beech skeletonized by mesofauna	-0.0077	0.0018
Bundles of variegated leaves of beech skeletonized by macrofauna	-0.0045	0.0030
Entire bleached leaves of beech	-0.0265	-0.0108
Bundles of entire bleached leaves of beech	-0.0143	-0.0066
Bleached leaves of beech skeletonized by macrofauna	-0.0160	0.0090
Bundles of bleached leaves of beech skeletonized by macrofauna Bleached leaves of beech skeletonized by mesofauna	-0.0131 -0.0149	0.0048 0.0028
Bundles of bleached leaves of beech skeletonized by mesofauna	-0.0149	-0.0028
Pits done by caterpillars in beech leaves	-0.0115	-0.0017
Nests done by foliage-consuming insects	-0.0085	-0.0073
Organo-mineral material smearing beech leaves	-0.0184	-0.0128
Holorganic faecal material smearing beech leaves	-0.0216	-0.0035
Intact petioles and nerves of beech	-0.0022	0.0027
Petioles and nerves of beech tunnelled by fauna	0.0001	0.0092
Petioles and nerves of beech filled with enchytraeid faeces	-0.0084	0.0045
Petioles and nerves of beech filled with faeces of Adoristes ovatus (oribatid mite)	-0.0045	0.0030
Petioles and nerves of beech filled with faeces of phthiracarid oribatid mites	-0.0025	0.0188
Petioles and nerves of beech filled with faeces of sciarid dipteran larvae	-0.0014	0.0091
Petioles and nerves of beech filled with grass roots Petioles and nerves of beech brown and tough	-0.0045 -0.0112	0.0030 -0.0027
Petioles and nerves of beech bloached	-0.0018	0.0027
Sandwich material made of beech leaf fragments and holorganic enchytraeid faeces	0.0039	0.0250
Sandwich material made of beech leaf fragments and holorganic earthw orm faeces	-0.0045	0.0034
Sandwich material made of beech leaf fragments and holorganic oribatid faeces	0.0033	0.0106
Sandwich material made of beech leaf fragments and organo-mineral earthworm faeces	-0.0003	0.0024
Sandwich material made of beech leaf fragments and organo-mineral enchytraeid faeces	0.0037	-0.0025
Sandwich material made of beech leaf fragments and holorganic sciarid faeces	0.0001	-0.0007
Skeletonized beech leaf fragments	0.0098	-0.0046
Bundles of skeletonized beech leaf fragments	0.0055	0.0060
Brown beech leaf fragments untouched by fauna	0.0141	-0.0019
Intact bud scales of beech Bud scales of beech, entire but brown and soft	-0.0299 0.0008	-0.0037 0.0191
Strongly decayed bud scales of beech	0.0217	-0.0043
Intact male inflorescences of beech	-0.0232	-0.0120
Brown decaying male inflorescences of beech	-0.0057	0.0236
Pollen mass	-0.0022	0.0204
Intact seed coats of beech	0.0038	0.0175
Seed coats of beech tunnelled by phthiracarid mites	0.0006	0.0077
Seed coats of beech tunnelled by enchytraeids	0.0051	0.0037
Seed coats of beech tunnelled by sciarid larvae	-0.0009	0.0025
Seed coats of beech penetrated by roots	0.0028	0.0004
Intact fragments of beech burn	-0.0098	0.0019
Soft fragments of beech burr Soft fragments of beech burr tunnelled by oribatid mites	0.0099	0.0028
Soft fragments of beech burr tunnelled by enchytraeids	0.0025 0.0068	0.0108 0.0069
Soft fragments of beech burr tunnelled by sciarid larvae	0.0029	0.0069
Soft fragments of beech burr tunnelled by springtails	0.0012	0.0043
Soft fragments of beech burr penetrated by grass roots	0.0033	-0.0048
Beech cupules tunnelled by fauna	0.0006	-0.0024
Intact beech gallnuts	0.0051	0.0010
Intact twigs	-0.0225	-0.0033
Twigs decayed by white-rot	-0.0087	-0.0031
Twig fragments tunnelled by fauna	-0.0074	0.0106
Bark remnants of twigs	0.0079	0.0072
Twigs filled with enchytraeid holorganic faeces	0.0115	0.0205
Twigs filled with enchytraeid organo-mineral faeces	0.0102	-0.0024
Twigs filled with sciarid holorganic faeces Twigs filled with oribatid holorganic faeces	0.0015 0.0085	0.0101 0.0138
Twigs penetrated by beech roots	0.0129	0.0138

Appendix 1. Continued

Categories	Axis 1	Axis 3
Intact w ood fragments	-0.0048	-0.0029
Decayed wood fragments	0.0016	-0.0093
Wood fragments tunnelled by fauna	0.0096	0.0036
Wood fragments penetrated by grass roots	-0.0045	0.0030
Wood fragments penetrated by beech fine roots	0.0025 0.0072	-0.0063
Intact bank fragments	0.0072	-0.0060 -0.0147
Well-decayed bark fragments Bark fragments tunnelled by enchytraeids	-0.0018	0.0094
Bark fragments tunnelled by phthiracarid mites	0.0001	0.0075
Bark fragments tunnelled by sciarid larvae	-0.0022	0.0046
Bark fragments penetrated by grass roots	-0.0045	0.0030
Intact living fine long roots of beech	0.0231	0.0184
Living fine long roots of beech brow sed by fauna	0.0123	-0.0061
Intact dead fine long roots of beech	0.0238	-0.0178
Dead fine long roots of beech tunnelled by fauna	0.0145	-0.0119
Dead fine long roots of beech penetrated by grass roots	0.0031	-0.0069
Dead fine long roots of beech, voided	0.0046	-0.0075
Living w oody roots of beech	0.0237	-0.0031
Living w oody roots of beech brow sed by fauna	0.0069	-0.0058
Decaying w oody roots of beech	0.0160	-0.0133
Living pale yellow creamy mycorrhizae of beech	0.0221	0.0136
Pale yellow creamy mycorrhizae of beech browsed by fauna	0.0181	0.0003 -0.0139
Dead pale yellow creamy mycorrhizae of beech Living orange brown mycorrhizae of beech with woolly mycelium	0.0260 0.0106	0.0078
Orange brown mycorrhizae of beech with woolly mycelium brow sed by fauna	0.0016	0.0078
Dead orange brown mycorrhizae of beech with woolly mycellum	0.0128	-0.0068
Living black mycorrhizae of beech (produced by Cenoccum geophilum)	0.0120	0.0104
Living black mycorrhizae of beech browsed by fauna	0.0019	0.0043
Dead black mycorrhizae of beech	0.0141	-0.0087
Living yellow mycorrhizae of beech with woolly mycelium	0.0028	0.0074
Living shoots of Polytrichum formosum	-0.0067	0.0087
Fragments of stems of Polytrichum formosum, red and tough	0.0032	0.0044
Fragments of stems of Polytrichum formosum, voided	0.0051	0.0095
Dead stem bases of Polytrichum formosum	0.0024	-0.0001
Decaying stem bases of Polytrichum formosum	0.0047	-0.0024
Living shoots of Scleropodium purum	-0.0073	0.0111
Dead shoots of Scleropodium purum	0.0042	0.0043
Living shoots of Leucobryum glaucum	-0.0064	-0.0054
Dead shoots of Leucobryum glaucum Dead moss, undetermined	0.0003	-0.0012
Intact leaves of Luzula forsteri	-0.0057 -0.0060	-0.0035 -0.0057
Bleached leaves of Luzula forsteri	-0.0034	-0.0037
Living leaf bases of Luzula forsteri	0.0000	0.0136
Decaying leaf bases of Luzula forsteri	0.0047	-0.0024
Intact leaves of Deschampsia flexuosa	-0.0074	-0.0030
Decaying leaves of Deschampsia flexuosa	-0.0135	0.0017
Living leaf bases of Deschampsia flexuosa	-0.0037	-0.0003
Decaying leaf bases of Deschampsia flexuosa	0.0120	-0.0012
Intact inflorescences of Deschampsia flexuosa	-0.0058	-0.0045
Decaying inflorescences of Deschampsia flexuosa	-0.0098	-0.0003
Living grass roots	0.0090	-0.0022
Decaying grass roots	0.0144	-0.0083
Intact grass stems	0.0008	-0.0136
Fragments of grass stems brow sed by fauna	-0.0022	0.0204
Fragments of decaying grass roots	-0.0062	-0.0026
Intact leaves of Vaccinium myrtillus	-0.0075	-0.0063
Skeletonized leaves of Vaccinium myrtillus	-0.0101	-0.0034
Roots of Vaccinium myrtillus	0.0053	0.0013
Living rhizomes of Vaccinium myrtillus	0.0016	0.0018
Decaying rhizomes of Vaccinium myrtillus Bleached leaves of Oxalis acetosella	0.0072 0.0013	-0.0001 -0.0039
Brown entire leaves of Acer pseudoplatanus	-0.0013	-0.0039
Brown leaves of Acer pseudoplatanus skeletonized by macrofauna	-0.0051	-0.0198
Bleached leaves of Acer pseudoplatanus	-0.0051	-0.0198
Bleached leaves of Acer pseudoplatanus skeletonized by macrofauna	-0.0051	-0.0198
Leaves of Acer pseudoplatanus skeletonized by mesofauna	-0.0018	-0.0002

Appendix 1. Continued

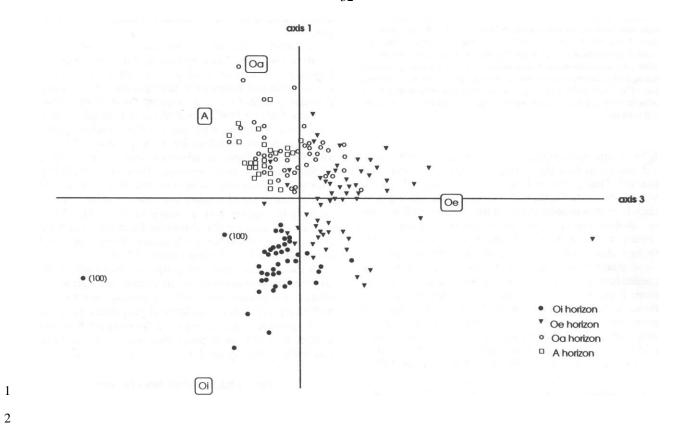
Categories	Axis 1	Axis 3
Winged seed of Acer pseudoplatanus with intact wing	0.0001	-0.0007
Winged seed of Acer pseudoplatanus with skeletonized wing	-0.0062	0.0030
Wingless seed of Acer pseudoplatanus	-0.0012	0.0051
Winged seed of Fraxinus excelsior with intact wing	-0.0053	-0.0192
Brown entire leaves of Quercus petraea	-0.0076	-0.0067
Leaves of Quercus petraea skeletonized by mesofauna	-0.0049	-0.0013
Intact unidentified fragments of seed wings	-0.0050	0.0176
Skeletonized unidentified fragments of seed wings	-0.0027	0.0036
Brown entire needles of Picea abies	-0.0010	-0.0002
Bleached entire needles of Picea abies	-0.0108	-0.0034
Needles of Picea abies brow sed by fauna	-0.0002	0.0086
Seed wings of Picea abies	-0.0053	-0.0032
Brown rhizomorphs	-0.0047	0.0075
White rhizomorphs	-0.0023	0.0130
Yellow rhizomorphs	0.0028	0.0074
Dead rhizomorphs of Armillaria	0.0100	-0.0074
Dead rhizomorphs of Armillaria tunnelled by fauna	0.0100	-0.0074
Sclerotia of Cenoccum geophilum	0.0245	-0.0136
Lichens	-0.0090	-0.0053
Intact caterpillar faeces	-0.0137	0.0099
Caterpillar faeces tunnelled by phthiracarid mites	0.0056	0.0078
Intact slug faeces	-0.0142	-0.0045
Slug faeces tunnelled by enchytraeids	-0.0089	-0.0049
Slug faeces tunnelled by sciarid larvae	-0.0089	-0.0049
Intact holorganic earthw orm faeces	-0.0124	0.0118
Holorganic earthworm faeces tunnelled by enchytraeids	0.0007	0.0057
Unidentified holorganic faeces	-0.0056	-0.0037
Intact organo-mineral earthw orm faeces	-0.0025	0.0036
Compacted organo-mineral earthw orm faeces	0.0088	-0.0016
Organo-mineral earthworm faeces tunnelled by enchytraeids	0.0042	0.0034
Holorganic w oodlice faeces	-0.0120	0.0033
Holorganic milliped faeces	0.0048	0.0049
Holorganic milliped faeces tunnelled by enchytraeids	0.0036	0.0010
Holorganic milliped faeces tunnelled by phthiracarid mites	0.0036	0.0010
Holorganic cranefly faeces	0.0004	0.0048
Intact holorganic sciarid faeces	-0.0044	-0.0007
Compacted holorganic sciarid faeces	0.0074	-0.0069
Intact holorganic enchytraeid faeces	0.0141	0.0081
Compacted holorganic enchytraeid faeces	0.0219	-0.0120
Organo-mineral enchytraeid faeces	0.0078	-0.0044
Compacted organo-mineral enchytraeid faeces	0.0070	-0.0117
Compacted organic-dominant organo-mineral material	0.0066	-0.0064
Compacted organo-mineral material	0.0114	-0.0120
Compacted mineral-dominant organo-mineral material	0.0040	-0.0085
Unidentified mineral assemblages	-0.0016	-0.0011
Charcoal	0.0132	-0.0153
Snail shells	-0.0028	0.0098
Woodlice shells	-0.0022	0.0204
Intact stones	0.0161	-0.0103
Weathering stones	0.0147	-0.0128
Weathering stones impregnated with organic matter	0.0048	-0.0096

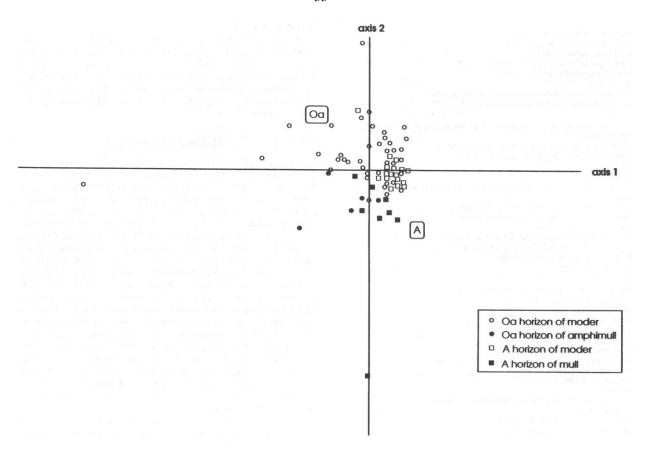
Appendix 2. Categories used for the description of humus horizons. Coordinates on factorail axes 1 and 2 of partial correspondence analysis (OH and A).

Categories	Axis 1	Axis 2
Brown leaves of beech skeletonized by macrofauna	-0.0002	-0.0319
Bleached leaves of beech skeletonized by macrofauna	-0.0002	-0.0319
Bleached leaves of beech skeletonized by mesofauna	-0.0002	-0.0319
Intact petioles and nerves of beech	-0.0130	-0.0013
Petioles and nerves of beech tunnelled by fauna	-0.0100	0.0135
Petioles and nerves of beech filled with enchytraeid faeces	-0.0051	0.0069
Petioles and nerves of beech filled with faeces of phthiracarid oribatid mites Sandwich material made of beech leaf fragments and holorganic enchytraeid faeces	-0.0094 -0.0343	-0.0090 0.0089
Sandwich material made of beech leaf fragments and holorganic earthworm faeces	-0.0343	-0.0001
Sandwich material made of beech leaf fragments and holorganic oribatid faeces	-0.0383	-0.0025
Sandwich material made of beech leaf fragments and organo-mineral earthworm faeces	-0.0031	-0.0038
Sandwich material made of beech leaf fragments and organo-mineral enchytraeid faeces	-0.0002	-0.0008
Skeletonized beech leaf fragments	0.0047	0.0114
Bundles of skeletonized beech leaf fragments	0.0002	0.0201
Brown beech leaf fragments untouched by fauna	-0.0051	-0.0081
Intact bud scales of beech	-0.0214	-0.0201
Bud scales of beech, entire but brown and soft	-0.0292	-0.0056
Strongly decayed bud scales of beech	0.0016	0.0205
Brown decaying male inflorescences of beech	-0.0246	0.0038
Intact seed coats of beech	-0.0105	0.0065
Seed coats of beech tunnelled by phthiracarid mites	0.0020	0.0054
Seed coats of beech tunnelled by enchytraeids	-0.0015	0.0051
Intact fragments of beech burr	-0.0152	-0.0048
Soft fragments of beech burr	0.0041	0.0022
Soft fragments of beech burr tunnelled by oribatid mites	-0.0106	0.0136
Soft fragments of beech burr tunnelled by enchytraeids	0.0012	0.0197
Beech cupules tunnelled by fauna	0.0051	0.0050
Twigs decayed by white-rot	0.0000	-0.0066
Twig fragments tunnelled by fauna	-0.0013	0.0229
Bark remnants of twigs	0.0008	-0.0122
Twigs filled with enchytraeid holorganic faeces	-0.0119 -0.0289	0.0159 -0.0002
Twigs filled with enchytraeid organo-mineral faeces Twigs filled with sciarid holorganic faeces	0.0002	0.0068
Twigs filled with oribatid holorganic faeces	-0.0287	0.0008
Twigs penetrated by beech roots	-0.0207	0.0076
Decayed wood fragments	0.0010	-0.0055
Wood fragments tunnelled by fauna	-0.0168	-0.0104
Wood fragments penetrated by beech fine roots	0.0024	-0.0066
Intact bark fragments	-0.0140	0.0038
Well-decayed bark fragments	0.0091	0.0086
Bark fragments tunnelled by enchytraeids	-0.0090	0.0079
Bark fragments tunnelled by phthiracarid mites	-0.0022	-0.0101
Intact living fine long roots of beech	-0.0212	-0.0033
Living fine long roots of beech browsed by fauna	-0.0021	0.0103
Intact dead fine long roots of beech	0.0061	-0.0014
Dead fine long roots of beech tunnelled by fauna	0.0045	0.0073
Dead fine long roots of beech penetrated by grass roots	0.0049	-0.0025
Dead fine long roots of beech, voided	0.0047	-0.0033
Living woody roots of beech	0.0028	0.0182
Living woody roots of beech browsed by fauna	0.0040	0.0103
Decaying woody roots of beech	0.0113	0.0113
Living pale yellow creamy mycorrhizae of beech	-0.0172	-0.0035
Pale yellow creamy mycorrhizae of beech browsed by fauna	-0.0016	0.0125
Dead pale yellow creamy mycorrhizae of beech	0.0040	0.0047
Living orange brown mycorrhizae of beech with woolly mycelium	-0.0052	0.0069
Dead orange brown mycorrhizae of beech with woolly mycelium	0.0058	0.0065
Living black mycorrhizae of beech (produced by Cenoccum geophilum)	-0.0079	0.0027
Dead black mycorrhizae of beech	0.0057	0.0152
Living shoots of Polytrichum formosum Fragments of stems of Polytrichum formosum, voided	-0.0054	-0.0088
Fragments of stems of Polytrichum formosum, voided Dead stem bases of Polytrichum formosum	0.0010 0.0037	-0.0105 -0.0076
Dead shorts of Leucobryum glaucum	-0.0037	-0.0076
Bleached leaves of Luzula forsteri	-0.0012	-0.0043
Living leaf bases of Luzula forsteri	0.0012	-0.0002

Appendix 2. Continued

Categories	Axis 1	Axis 2
Decaying leaf bases of Luzula forsteri	0.0037	-0.0076
Decaying leaves of Deschampsia flexuosa	-0.0268	0.0051
Living leaf bases of Deschampsia flexuosa	0.0045	-0.0002
Decaying leaf bases of Deschampsia flexuosa	0.0066	0.0079
Living grass roots	0.0065	-0.0133
Decaying grass roots	0.0085	-0.0002
Intact grass stems	0.0026	-0.0070
Roots of Vaccinium myrtillus	-0.0053	0.0095
Decaying rhizomes of Vaccinium myrtillus	-0.0014	0.0092
Bleached leaves of Oxalis acetosella	-0.0002	-0.0319
Brown entire needles of Picea abies	-0.0145	0.0016
Brown rhizomorphs	0.0010	-0.0071
White rhizomorphs	-0.0140	0.0071
Dead rhizomorphs of Armillaria	-0.0012	0.0015
Dead rhizomorphs of Armillaria tunnelled by fauna	-0.0012	0.0015
Sclerotia of Cenoccum geophilum	0.0012	0.0035
Intact caterpillar faeces	-0.0006	0.0123
Intact holorganic earthworm faeces	0.0018	-0.0039
Holorganic earthworm faeces tunnelled by enchytraeids	-0.0383	-0.0025
Intact organo-mineral earthworm faeces	0.0031	-0.0265
Compacted organo-mineral earthworm faeces	-0.0040	-0.0140
Organo-mineral earthworm faeces tunnelled by enchytraeids	0.0009	-0.0076
Holorganic milliped faeces	-0.0017	-0.0222
Holorganic milliped faeces tunnelled by enchytraeids	-0.0383	-0.0025
Holorganic milliped faeces tunnelled by phthiracarid mites	-0.0383	-0.0025
Holorganic cranefly faeces	-0.0020	-0.0009
Compacted holorganic sciarid faeces	0.0059	0.0109
Intact holorganic enchytraeid faeces	-0.0083	0.0060
Compacted holorganic enchytraeid faeces	0.0109	0.0235
Organo-mineral enchytraeid faeces	-0.0068	0.0034
Compacted organo-mineral enchytraeid faeces	0.0103	-0.0041
Compacted organic-dominant organo-mineral material	0.0034	-0.0035
Compacted organo-mineral material	0.0070	-0.0078
Compacted mineral-dominant organo-mineral material	0.0048	-0.0048
Unidentified mineral assemblages	0.0051	0.0050
Charcoal	0.0063	-0.0200
Intact stones	0.0036	-0.0269
Weathering stones	0.0092	-0.0065
Weathering stones impregnated with organic matter	0.0077	-0.0002





4

